

# Small GTPases in megakaryocyte and platelet biology

Lucia Stefanini, & Wolfgang Bergmeier

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*Lucia Stefanini and Wolfgang Bergmeier*



## Small GTPases in megakaryocyte and platelet biology

Lucia Stefanini<sup>1</sup> & Wolfgang Bergmeier<sup>2,3</sup>

<sup>1</sup>Department of Internal Medicine and Medical Specialties, Sapienza University of Rome, Rome, Italy, <sup>2</sup>Department of Biochemistry and Biophysics, University of North Carolina, Chapel Hill, NC, USA, and <sup>3</sup>McAllister Heart Institute, University of North Carolina, Chapel Hill, NC, USA

**Q2**  
5 Blood platelets are critical for diverse physiological processes  
**Q3** such as hemostasis, angiogenesis, and innate immunity [1].  
Every day ~100 billion platelets are produced by megakaryocytes  
(MKs) in the bone marrow and destroyed by phagocytic cells of  
the reticulo-endothelial system [2]. Defects in either process can  
lead to thrombocytopenia (low platelet count) or thrombocytosis  
(high platelet count). While thrombocytosis is a risk factor for  
various clinical complications, including venous thrombosis and  
cancer [3,4], thrombocytopenia is a common cause of bleeding  
complications [1]. Hemostatic plug formation under shear stress  
conditions at sites of vascular injury depends on the platelets'  
ability to sense and to rapidly respond to disruptions in the  
vascular wall. Small GTPases are critical in the cellular signaling  
required for proper MK and platelet function. They are binary  
signaling proteins that rapidly switch between an active, GTP-  
bound state, and an inactive, GDP-bound state. They are activated  
by guanine-nucleotide exchange factors (GEFs), inactivated by  
GTPase-activating proteins (GAPs), or held in an inactive state  
by guanine dissociation inhibitors, as for RHO-family GTPases  
[5]. In humans small GTPases belong to a large superfamily that  
includes 153 members and is divided into 5 branches based on  
sequence and functional similarities: RAS (cell proliferation,  
growth, adhesion), RHO (cytoskeletal dynamics), RAB (mem-  
brane trafficking), ARF (vesicular transport and actin remodel-  
ing), and RAN (nuclear transport and microtubule regulation).  
Despite the highly conserved structure, variations in sequence,  
post-translational modifications, subcellular localization and  
upstream regulatory proteins allow this large family to control  
a wide range of cellular responses. This review series discusses  
the current state-of-the-art in our understanding of how small  
GTPases control MK and platelet biology.

The best studied small GTPases in platelets are members of  
the RHO family, key regulators of the actin cytoskeleton [6].  
RHO GTPases regulate actin-binding proteins directly or indir-  
ectly to control actin dynamics and filament geometry [7]. They  
can have both unique and overlapping functions, and they can  
regulate one another through feedback and feedforward  
mechanisms [8]. The review by Pleines et al. summarizes the  
recent literature documenting a critical role for RHO GTPases  
in platelet release from MKs, with a focus on mouse lines in  
which knockout strategies have been applied to study the func-  
tion of the best characterized members RAC1, CDC42, RHOA,  
and their downstream effector proteins. The review by Aslan  
et al. summarizes our current understanding of how platelet

RHO GTPase activity is controlled by a very complex signaling  
network, and how proper regulation or dysfunction of these  
signaling pathways affects platelet function in health and dis-  
ease. The review by Thomas et al. will discuss the key role of  
formins, downstream targets of RHO GTPases, in the organiza-  
tion and interaction of the tubulin and actin cytoskeleton in  
MKs and platelets.

Small GTPases are also critical in the regulation of intracel-  
lular membrane trafficking and integrin-mediated adhesion.  
Membrane trafficking is a complex process regulated by the  
coordinated interplay of several members of the Rab, Arf, and  
Ras subfamilies [9]. In platelets, these processes are critical for  
the loading and unloading of granules, intracellular storage ves-  
icles that contain a variety of hemostatic and vasoactive factors,  
proteases, and small molecule compounds. The review by Poole  
et al. will summarize the most pertinent findings published in this  
nascent field of study. Members of the RAP subfamily of RAS  
GTPases are important regulators of integrin signaling [10], the  
key event during platelet adhesion and hemostatic plug formation  
[1]. Independent yet synergistic pathways control the activity of  
the main RAP-GEF and –GAP proteins in platelets, thereby  
facilitating rapid and sustained RAP activation. The review by  
Stefanini & Bergmeier will summarize this important topic,  
including recent studies in transgenic mice that helped identify  
the key regulators of RAP1 activity in platelets and ongoing work  
on the downstream targets and spatio-temporal activity control of  
individual RAP isoforms.

Taken together, the goal of this series of reviews is to provide  
the readership of *Platelets* a state-of-the-art overview of the role  
of small GTPases in MK and platelet biology. While the focus is  
on recent advances in the area of basic science, the clinical  
relevance of alterations in small GTPase signaling for platelet  
count and function will also be discussed.

### Disclosure statement

The authors report no declaration of interest.

### Funding

This work was supported by NIH grants R01 HL121650 and P01  
HL120846 (W.B.) and the Ministero della Istruzione e della Ricerca  
Young Researchers Program Rita Levi Montalcini (L.S.).

### References

1. Broos K, Feys HB, De Meyer SF, Vanhoorelbeke K, Deckmyn H. Platelets at work in primary hemostasis. *Blood Rev* 2011;25:155–167. doi:10.1016/j.blre.2011.03.002

2. Grozovsky R, Giannini S, Falet H, Hoffmeister KM. Novel mechanisms of platelet clearance and thrombopoietin regulation. *Curr Opin Hematol* 2015;22:445–451. doi:10.1097/MOH.0000000000000170
- 95 3. Zakai NA, Wright J, Cushman M. Risk factors for venous thrombosis in medical inpatients: validation of a thrombosis risk score. *J Thromb Haemost* 2004;2:2156–2161. doi:10.1111/j.1538-7836.2004.00991.x
- 100 4. Bailey SE, Ukoumunne OC, Shephard EA, Hamilton W. Clinical relevance of thrombocytosis in primary care: a prospective cohort study of cancer incidence using English electronic medical records and cancer registry data. *Br J Gen Pract* 2017;67:e405–e13. doi:10.3399/bjgp17X691109
5. Bar-Sagi D, Ras HA. Rho GTPases: a family reunion. *Cell* 2000;103:227–238.
6. Jaffe AB, Hall A. Rho GTPases: biochemistry and biology. *Annu Rev Cell Dev Biol* 2005;21:247–269. doi:10.1146/annurev.cellbio.21.020604.150721 105
7. Sit ST, Manser E. Rho GTPases and their role in organizing the actin cytoskeleton. *J Cell Sci* 2011;124:679–683. doi:10.1242/jcs.064964 110
8. Ridley AJ. Life at the leading edge. *Cell* 2011;145:1012–1022. doi:10.1016/j.cell.2011.06.010
9. Mizuno-Yamasaki E, Rivera-Molina F, Novick P. GTPase networks in membrane traffic. *Annu Rev Biochem* 2012;81:637–659. doi:10.1146/annurev-biochem-052810-093700 115
10. Bos JL. Linking rap to cell adhesion. *Curr Opin Cell Biol* 2005;17:123–128. doi:10.1016/j.ceb.2005.02.009

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