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Article

Molecular Modeling of the Deamidation Reaction in Solution: A Theoretical-Computational Study

Maria Laura De Sciscio, Alessandro Nicola Nardi, Fabio Centola, Mara Rossi, Enrico Guarnera,* and Marco D'Abramo*

Cite This: J. F	Phys. Chem. B 2023, 127, 9550–955	59	Read Online			
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ABSTRACT: In this work, a theoretical-computational method is applied to study the deamidation reaction, a critical post-translational modification in proteins, using a simple						

deamidation reaction, a critical post-translational modification in proteins, using a simple model molecule in solution. The method allows one to comprehensively address the environmental effect, thereby enabling one to accurately derive the kinetic rate constants for the three main steps of the deamidation process. The results presented, in rather good agreement with the available experimental data, underline the necessity for a rigorous treatment of environmental factors and a precise kinetic model to correctly assess the overall kinetics of the deamidation reaction.



1. INTRODUCTION

Amino acids bearing an amide group on the side chain, i.e., asparagine (Asn) and glutamine (Gln), can undergo spontaneous nonenzymatic deamidation, yielding aspartic acid (Asp) and glutamic acid (Glu), respectively. This post-translational modification (PTM) entails the conversion of a neutral side-chain to a negatively charged one, i.e., Asp and Glu acidic functions are deprotonated at physiological pH, and it represents a common pathway of chemical degradation of peptides and proteins.^{1,2} Among these two residues, Gln deamidation is significantly slower than Asn one and has been detected mainly in long-lived proteins.^{3,4} On the other hand, deamidation of Asn residues, due to its high rate and frequency in proteins, is involved in several pathological events,⁵ and the increasing content of deamidated products in proteins is related to aging conditions and possible loss of biological activities.^{6,7}

Furthermore, deamidation is a common PTM in biopharmaceutical proteins that can affect the stability, biological activity, and efficacy of the product, making it an important critical quality attribute (CQA) in drug development.^{8,9} The rate of deamidation reactions is influenced by various factors, such as protein structure, temperature, and pH.⁹ For instance, deamidation rates are greatly enriched in asparagine if followed by glycine and, to a lesser extent, if followed by serine.¹⁰

Although the reaction mainly alters the amino acid side chain, an essential role is played by the C-carboxyl residue (residue n + 1) at physiological pH. In fact, according to the literature,¹¹ deamidation proceeds through succinimide formation via a three-step reaction. As reported in Figure 1, the

process starts with the deprotonation of the nitrogen atom belonging to the (n + 1) residue, and it is followed by a spontaneous attack of the latter atom, now negatively charged, on the Asn side-chain amide carbon atom, leading to the formation of a cyclic tetrahedral intermediate (Tet⁻). The metastable product naturally evolves into the succinimide intermediate (Suc), which is an experimentally characterized product. Thereafter, the hydrolysis of both sides of the imide can occur, leading to Asp or *iso*-Asp.

Succinimide formation (ring closure and deamination stages) and, in particular, the conversion of Tet⁻ to imide, entailing deamination, were identified as the rate-determining step (RDS) at pH = 6-8.¹¹ Therefore, the molecular features of these stages directly correlate with the overall deamidation rate.

In neutral-to-basic conditions, deamidation rates are influenced by several factors linked to the different reaction stages: (i) deprotonation is affected by acid—base equilibrium and the type of residue in position n + 1, upon which pK_a depends¹² (ii) ring closure and deamination stages are strongly affected by the torsional angles of Asn and of the n + 1 residue,^{10,13,14} as well as by any factors able to modify the side-chain amide carbon atom and the n + 1 nitrogen electro-

Received:July 11, 2023Revised:September 15, 2023Accepted:October 6, 2023Published:October 30, 2023





Figure 1. Deamidation mechanism. The reaction begins with the deprotonation of the Asn, resulting in Asn⁻ formation, which leads to the intramolecular ring closure by attacking the Asn carbon atom of the amide side-chain. The metastable tetrahedral intermediate (Tet⁻) loses the negatively charged NH₂ group, leading to a succinimide intermediate (Suc). Then, two hydrolytic paths are possible: one leads to Asp (blue) and the other to iso-Asp (green), via the gendiol (Gem) or iso-gendiol (iso-Gem) intermediate formation. Atoms in red represent n + 1 amino acid, while X and Y are the C-terminal residues and the N-terminal one from the Ca atom, respectively; in this work, X = H and Y = CH₃.

philicity (iii) hydrolysis stage mainly depends on water molecules/hydroxide ion proximity to imide hydrolytic sites.

Several computational studies focused on deamidation in proteins used to evaluate the reaction through classical descriptors-such as Asn averaged Solvent Accessible Surface Area (SASA), C_{γ} -N_{n+1} distance, root means square fluctuations (RMSF), backbone and side-chain dihedrals (e.g., χ and ψ), and H-bond analysis—by means of classical molecular dynamics simulation and (static) structure-based approaches.^{1,10,13-19} The limited quantum mechanics/molecular mechanics (QM/MM) works often approximate the deamidation reaction to a single-stage Asn \rightarrow Suc reaction,^{13,14} which may affect the accuracy of the prediction.¹⁴ Furthermore, for computational reasons, more approximate QM semiempirical methods are usually employed to describe the reaction. On the other hand, QM works-focused on mechanistic investigations-do not include an explicit treatment of the solvent, which leads to an overestimation of the energy barriers of the reaction (higher than 30-40 kcal/mol),²⁰⁻²² not in a good agreement with the experimental estimates of reaction half-life times and Arrhenius activation energies (~20-25 kcal/ mol).^{2,23}

Although several efforts were made to understand the deamidation reaction features, the structural determinants allowing the correct prediction of Asn residues prone to deamidate are not yet completely understood. This work aims to give new insights into the deamidation mechanism by means of an accurate theoretical-computational QM/MM approach that can provide a quantitative estimate of the kinetics of the

deamidation reaction in a simple model system (NH₂-Asn-NMe, Figure 2).



Figure 2. 2D representation of the model system, i.e., NH2-Asn-NMe, used in this study.

Hence, we reconstruct the free energy barriers of the three steps leading to the formation of the Asp-i.e., ring-closure, deamination, and hydrolysis stages-in a fully solvated condition. With such an approach, we were able to model the kinetics and, therefore, predict the overall reaction behavior, which is in rather good agreement with the available experimental data on similar systems.

2. THEORY

2.1. Perturbed Matrix Method. In this work, we applied the Perturbed Matrix Method (PMM)-a theoreticalcomputational quantum mechanics/molecular mechanics (QM/MM) approach-to study the deamidation reaction. As the PMM theory has been fully detailed in previous papers,²⁴⁻²⁹ here we provide only the main features and the relevant equations used in this work. According to the PMM scheme and similar to other QM/MM approaches, the system

is subdivided into two regions, one composed of the atoms directly involved in the process of interest and known as the Quantum Center (QC), and the complementary part, which constitutes the environment. The former is treated by means of quantum-chemical calculations, while the latter, acting as the perturbing environment, is described by classical molecular dynamics (MD) simulations. Unlike "on the fly" QM/MM methods, within the PMM approach, the effect of the environment is added a posteriori, allowing for the inclusion of the statistical fluctuation of the QC environment at a reasonable computational cost. In the PMM scheme, for each QC nuclear configuration, the electronic Hamiltonian (\widehat{H}) of the QC can be expressed as the sum of the isolated QC electronic Hamiltonian, i.e., the unperturbed gas-phase electronic Hamiltonian of the QC, \widehat{H}^0 , and the perturbation operator, \widehat{H}^0 , as reported in eq 1.

$$\widehat{H} = \widehat{H}^0 + \widehat{V} \tag{1}$$

The corresponding electronic Hamiltonian matrix elements $[\tilde{H}]_{l,l,\nu}$ expressed in the basis set of the unperturbed electronic QC eigenstates Φ_l^0 and eigenvalues v_l^0 , can be obtained as

$$[\widetilde{H}]_{l,l'} = \langle \Phi_l^0 | \widehat{H} | \Phi_{l'}^0 \rangle = v_l^0 \delta_{l,l'} + \langle \Phi_l^0 | \widehat{V} | \Phi_{l'}^0 \rangle \tag{2}$$

The perturbation operator, describing QC-environment interaction, can be obtained via a multipolar expansion centered in the QC center of mass (\mathbf{r}_0)

$$\widehat{V} \cong \sum_{j} \left[\mathcal{V}(\mathbf{r}_{0}) - \mathbf{E}(\mathbf{r}_{0}) \cdot (\mathbf{r}_{j} - \mathbf{r}_{0}) + \dots \right] q_{j}$$
(3)

where $\mathcal{V}(\mathbf{r}_0)$ and $\mathbf{E}(\mathbf{r}_0)$ are the perturbing electric potential and the electric field produced by the environment, respectively; the *j* index runs over all QC electrons and nuclei, q_j is the charge of *j*-th particle, and \mathbf{r}_j is the associated coordinates. The multipolar expansion, reported in eq 3, is explicitly treated up to the dipolar term for the off-diagonal elements of the electric Hamiltonian matrix. The diagonal elements of the \widetilde{H} matrix are calculated within the atom-based expansion PMM level of theory, according to which \widehat{V} is obtained as follows

$$\widehat{V} \cong \sum_{N} \sum_{j} \Omega_{N}(\mathbf{r}_{j}) [\mathcal{V}(\mathbf{R}_{N}) - \mathbf{E}(\mathbf{R}_{N}) \cdot (\mathbf{r}_{j} - \mathbf{R}_{N}) + \dots]$$

$$q_{j} \qquad (4)$$

where N refers to all QC atoms, Ω_N is a step function that is null outside and unitary inside the N-th atomic region, and \mathbf{R}_N is the N-th nucleus position. Hence, through the diagonalization of \widetilde{H} , at each MD frame, a trajectory of QC perturbed electronic eigenstates and eigenvalues is obtained. When the calculation is performed along the reaction coordinate (ξ), as in the present work, the Helmholtz free energy of the reaction can be calculated as a function of such a coordinate, as previously reported.^{25,30–32} Thus, the perturbed free energy change (ΔA), due to the movement from an initial position (i) to a subsequent point (i + 1), along the reaction coordinate is

$$\Delta A_{i \to i+1}(\xi) \simeq -k_{\rm B}T \langle {\rm e}^{-\beta \Delta U} \rangle_{i \to i+1} \tag{5}$$

where ΔU is the perturbed energy change, $k_{\rm B}$ is the Boltzmann's constant, and $\beta = 1/k_{\rm B}T$. The sum of each free energy variation leads to the $\Delta A_{\rm tot}$.

2.2. Reaction Kinetics. A general first-order reaction, starting from a reactant species (R) and spontaneously evolving to a different compound, the product (P), can be generally expressed as

$$R \underset{k'}{\stackrel{k}{\rightleftharpoons}} P \tag{6}$$

When k and k' are of the same order of magnitude, the reaction rate $\left(\frac{dx}{dt}\right)$, defined as the time (t) rate of the increase in the reaction extent (x), or the speed at which reaction occurs for the law of mass action, is

$$\frac{\mathrm{d}x}{\mathrm{d}t} = -\frac{\mathrm{d}[\mathrm{R}]}{\mathrm{d}t} = \frac{\mathrm{d}[\mathrm{P}]}{\mathrm{d}t} = k[\mathrm{R}] - k'[\mathrm{P}] \tag{7}$$

Each of the possible reaction paths, i.e., the forward and reverse ones, reasonably goes through a unique transition state (TS), such that the reaction can be schematized for the forward path as

$$R \xrightarrow{k} TS \xrightarrow{k_0} P \tag{8}$$

In the case of $k \ll k_0$, the steady state condition for TS and an equilibrium within the tiny TS region, which implies that the rate constants of the inward and outward fluxes are equal, can be assumed. Thus, the TS population has the same probability of falling into R or P, and the rate constant k_0 times $\frac{1}{2}$ is the TS rate constant, i.e., $k_0' \simeq k_0/2$. It can therefore be written

$$\frac{\mathrm{d}[\mathrm{R}]}{\mathrm{d}t} = -k[\mathrm{R}] \tag{9}$$

and

$$\frac{d[P]}{dt} \simeq Kk_0'[R] = \frac{Q_{\rm TS}}{Q_{\rm R}}k_0'[R] = k_{\rm L}[R]$$
(10)

where $K = k/k'_{0}$, while Q_{TS} and Q_{R} are the canonical distribution functions of the TS and reactant, respectively. As a reaction can be followed along a reaction coordinate, defined as ξ , it is possible to express and obtain the reactant and transition state partition functions within ξ ensembles. Finally, assuming a homogeneous density in the TS region, the Landau kinetic constant can be written as

$$k_{\rm L} = \frac{{\rm e}^{-\beta\Delta A^*}}{\int_{\xi_{\rm min}}^{\xi_{\rm max} - \delta/2} {\rm e}^{-\beta\Delta A(\xi)} {\rm d}\xi} \frac{\nu}{2}$$
(11)

where the integral limits $(\xi_{\min}, \xi_{\max} - \delta/2)$ define the reactant ensemble ξ area—from the energy minimum, i.e., the reactant, to the TS immediately preceding point—and $\Delta A(\xi)$ is the Landau free energy variation along ξ , while ΔA^{\ddagger} is the free energy variation between the transition state and the reactant. $\beta = 1/k_{\rm B}T$, and ν represents the mean velocity at which TS transits toward the product.

A more detailed explanation of the mathematical derivations of eqs 10 and 11, as well as the meaning and calculation of Landau free energy from the Helmholtz one, can be found in our previous work.³⁰

When the reactant area shape is fully quadratic and the TS traversing velocity (v) is approximated to a normal mode fluctuation, eq 11 becomes the well-known Eyring equation

$$k_{\rm E} = \kappa \frac{k_{\rm B}T}{h} {\rm e}^{-\Delta A^{\dagger}/RT}$$
(12)

where $k_{\rm B}$ is the Boltzmann's constant, *T* the temperature, *R* the ideal gas constant, and κ the transmission coefficient.

2.3. Deamidation Kinetic Model. A full description of the deamidation reaction and associated kinetic rate constants, considering only one hydrolysis path (i.e., the blue one in Figure 1), is as follows

$$\operatorname{Asn} \underset{k_{H-1}}{\overset{k_{H}}{\longleftrightarrow}} \operatorname{Asn}^{-} \underset{k_{-1}}{\overset{k_{1}}{\rightleftharpoons}} \operatorname{Tet}^{-} \overset{k_{2}}{\to} \operatorname{Suc} + \operatorname{NH}_{2}^{-} \overset{k_{3}}{\to} \operatorname{Gem} \overset{k_{4}}{\to} \operatorname{Asp}$$
(13)

To simplify the complex resulting kinetic scheme, we applied two approximations: (i) the first stage, that is, deprotonation, can be considered a fast pre-equilibrium with a high $K_{\rm Eq} = k_{\rm H}/k_{\rm H-1}$, i.e., product formation is favored; (ii) k_3 and k_4 are far higher, i.e., hydrolysis is faster, than k_1 and k_2 .

Accordingly, the deamidation reaction scheme becomes

$$\operatorname{Asn}^{-} \underset{k_{-1}}{\overset{k_{1}}{\rightleftharpoons}} \operatorname{Tet}^{-} \underset{k_{-1}}{\overset{k_{2}}{\to}} \operatorname{Suc} + \operatorname{NH}_{2}^{-}$$
(14)

The associated differential equations are

$$\frac{d[Asn^{-}]}{dt} = -k_{1}[Asn^{-}] + k_{-1}[Tet^{-}]$$
(15)

$$\frac{d[\text{Tet}^-]}{dt} = k_1[\text{Asn}^-] - k_{-1}[\text{Tet}^-] - k_2[\text{Tet}^-]$$
(16)

$$\frac{\mathrm{d[Suc + NH_2^-]}}{\mathrm{d}t} = k_2[\mathrm{Tet}^-]$$
(17)

The exact solutions of these equations,³³ taking both [Tet⁻] and [Suc + NH_2^-] to be equal to 0 and [Asn⁻] equal to 1 at t = 0, are

$$[\operatorname{Asn}^{-}] = \frac{k_1(\lambda_2 - k_2)}{\lambda_2(\lambda_2 - \lambda_3)} e^{-\lambda_2 t} + \frac{k_1(k_2 - \lambda_3)}{\lambda_3(\lambda_2 - \lambda_3)} e^{-\lambda_3 t}$$
(18)

$$[\text{Tet}^-] = -\frac{k_1}{(\lambda_2 - \lambda_3)} e^{-\lambda_2 t} + \frac{k_1}{(\lambda_2 - \lambda_3)} e^{-\lambda_3 t}$$
(19)

$$[\operatorname{Suc} + \operatorname{NH}_{2}^{-}] = \frac{k_{1}k_{2}}{\lambda_{2}\lambda_{3}} + \frac{k_{1}k_{2}}{\lambda_{2}(\lambda_{2} - \lambda_{3})}e^{-\lambda_{2}t} - \frac{k_{1}k_{2}}{\lambda_{3}(\lambda_{2} - \lambda_{3})}e^{-\lambda_{3}t}$$
(20)

where the eigenvalues λ_2 and λ_3 are obtained from the kinetic matrix determinant, defined as follows

$$\lambda_{2} = \frac{1}{2} \cdot (k_{1} + k_{-1} + k_{2} + \sqrt{(k_{1} + k_{-1} + k_{2})^{2} - 4k_{1}k_{2}})$$
(21)
$$\lambda_{2} = \frac{1}{2} \cdot (k_{1} + k_{2} + k_{2} - \sqrt{(k_{1} + k_{-1} + k_{2})^{2} - 4k_{1}k_{2}})$$

$$\lambda_3 = \frac{1}{2} \cdot (k_1 + k_{-1} + k_2 - \sqrt{(k_1 + k_{-1} + k_2)^2 - 4k_1 k_2})$$
(22)

3. METHODS

The estimation of the free energy profiles, kinetic rate constants, and deamidation half-times required several operative steps. For each stage of the deamidation reaction (Figure 1), starting after the deprotonation, the calculated

electronic properties in vacuum and the effect of the perturbing environment, evaluated by means of classical molecular dynamics simulations, were combined (via eq 1) to obtain the free energy variation along the selected reaction coordinate. The free energy variations were always obtained by means of the PMM, as shown in eq 5. The rate constants associated with forward and backward reactions were calculated through eqs 11 and 12. The evaluation of the profile shape and rate constants allowed us to choose the most representative kinetic scheme for the deamidation reaction. Using eqs 18-20, the populations of the involved species versus time and the corresponding half-life $(\tau_{1/2})$ were estimated. The block-averaging procedure was applied for the estimation of ΔA statistical error, while $k_{\rm L}$, $k_{\rm E}$, and $\tau_{1/2}$ were evaluated by recalculation of the values according to ΔA statistical fluctuations. The system used to study the reaction of interest is composed of an N-terminal Asn where the carboxylic function is capped with a methyl amine (NH₂-Asn-NMe), as can be seen in Figure 2.

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3.1. Quantum-Chemical Calculations. The QM calculations were performed with the Gaussian 16 software³⁴ using density functional theory (DFT) and time-dependent density functional theory (TD-DFT) with the B3LYP functional and 6-311++G(2d,2p) as the basis set. The starting geometry was obtained by a structural analysis of the MD simulation of Ace-Gly-Asn-Gly-Gly-NMe (details are given in the following section). For each reaction step, the reactant was first optimized without constraints and subsequently a relaxed scan was run to achieve the energy profile; the last point, i.e., the product, was optimized in the absence of restraints, while TS search was performed using QST2 or QST3 routine and checked for the correct characteristic imaginary frequency. Finally, the IRC was calculated to verify the reactant and product associated with the TS as well as the path to them. Once the profile was obtained, for every optimized geometry (to use in PMM calculation), gas-phase properties, such as electronic energies, dipoles, and ESP charges, were obtained for the ground state and the first three excited states. The QC is made up of NH₂-Asn-NMe for the first two investigated reaction stages (i.e., Ring-Closure and Deamination, Figure 1), while for the hydrolytic steps were also added two water molecules, in the case of the water-mediated mechanism, and a hydroxide ion with an additional water molecule to stabilize the OH⁻ anionic charge in vacuum for the alkaline catalysis. The following distances were chosen as reaction coordinates (ξ) for the corresponding reaction steps:

- (i) $N_{n+1}-C_{\gamma}$ for the ring-closure process (Asn⁻ \rightarrow Tet⁻).
- (ii) $C_{\gamma} N_{\gamma}$ for deamination stage (Tet⁻ \rightarrow Suc + NH $_2^{-}$).
- (iii) C_{γ}-O of OH⁻/H₂O in the first hydrolysis stage (Suc \rightarrow Gem).
- (iv-1) difference of $N_{n+1}-C_{\gamma}$ and $N_{n+1}-H$ of H_2O for the second hydrolysis stage within the water-mediated mechanism (Gem \rightarrow Asp).
- (iv-2) $N_{n+1}-C_{\gamma}$ for the second hydrolysis stage within the alkaline catalysis mechanism (Gem \rightarrow Int).
 - (v) N_{n+1} -H of H₂O for the third hydrolysis stage according to the alkaline catalysis mechanism (Int \rightarrow Asp).

3.2. Molecular Dynamics Simulations. Molecular dynamics simulations were performed using Gromacs-2021³⁵ and Q-FORCE/OPLS as the force field.^{36–38} The Lennard-Jones parameters are derived from the OPLS FF, while all the bonded parameters for the solute molecules are calculated by

ab initio calculations, following the procedure described by Sami and co-workers.³⁶ Atomic charges were estimated by the ESP procedure.^{39,40} The QC, coinciding with the solute in our work, was placed in a cubic simulation box large enough to avoid boundary effects, which was then filled with water molecules (using the TIP3P water model⁴¹) and neutralizing ions (Na⁺). For the solvent molecules, OPLSAA force field parameters were used.³⁸ For all the systems, the production runs were performed in the NVT ensemble after an energy minimization of the system with the steepest descent algorithm and short equilibration steps, in which the box size was adjusted to reproduce the experimental water density.⁴² The temperature was kept constant (at 300 K) by the velocity rescaling algorithm,⁴³ with a $\tau_{\rm T}$ of 0.01 ps. Simulations, lasting at least 150 ns, were carried out with a time step of 2 fs; longrange interactions were evaluated with the particle mesh Ewald method,^{44,45} and the Verlet cutoff scheme was used to generate the neighbor lists.⁴⁶ The cutoff for electrostatic and van der Waals interactions was 1.1 nm. To ensure the calculation of $\Delta A(\xi)$, i.e., the free energy variation at fixed values of the reaction coordinate, the QM optimized geometry was used as the starting configuration for MD simulation, constraining it by freezing its atomic coordinates or adding a harmonic potential to the distance of interest. In particular, for the Asnsimulation, the solute atomic coordinates were fixed, while in all the other cases, the distance between the atoms involved in the bond formation or cleavage was kept fixed. An additional simulation of the capped tetrapeptide Ace-Gly-Asn-Gly-Gly-NMe¹¹ was performed to obtain the initial conformation that is used for the QM calculations in our model system. A cluster analysis,⁴⁷ using a cutoff of 0.15 nm on the 500 ns simulation, allowed us to extract the most sampled Asn-Gly reactive-like conformation, as defined by χ dihedral (C-C_a- $C_{\beta}-C_{\gamma}$) values and $C_{\gamma}-N_{n+1}$ distance of ~70° and 3.4 Å, respectively.^{1,10,13,14,19}

4. RESULTS AND DISCUSSION

The deamidation reaction, shown in Figure 1, was studied in a fully solvated system. Such a reaction consists of four main consecutive steps: deprotonation, ring closure, deamination, and hydrolysis. As Asn deprotonation is usually very fast, we disregarded this step in the calculation of the reaction kinetics.

4.1. Succinimide Formation in Solution. According to the proposed deamidation mechanism,¹¹ once Asn is deprotonated, succinimide intermediate is obtained through the ring closure and the NH₂ expulsion as a leaving group via Tet⁻ intermediate. Hence, for the Asn⁻ \rightarrow Tet⁻ step, the C_{γ}-N_{n+1} distance was selected as the reaction coordinate (ξ) to describe the ring closure process, while for the deamination step, the reaction coordinate is represented by the C_{γ}-N_{γ} bond length (see Figure 3).

The initial reactant conformation was obtained from an additional MD simulation of the Ace–Gly–Asn–Gly–Gly–NHMe tetrapeptide,¹¹ taking the central structure of the most sampled cluster. The free energy profile of succinimide formation was obtained according to the PMM scheme discussed in the Theory and Methods section, with an estimated error on $\Delta A(\xi) < 0.1\%$.

As can be seen in Figure 3, the free energy path associated with the succinimide formation clearly represents a two-step reaction, where the first relative minimum ($\xi \approx 12$ a.u.) is associated with the metastable product Tet⁻. In the second step, the crossing of a similar free energy barrier leads to the



Figure 3. Two-step succinimide formation (free) energy pathway in the gas phase and in solution. The horizontal axis reaction coordinate ξ (a.u.) in the range 1–12 represents the C_{γ} – N_{n+1} distance, while in the deamination stage it is C_{γ} – N_{γ} ($\xi = 12-20$ a.u.). At left, middle, and right are reported the graphical 2D representation of Asn⁻, Tet⁻, and Suc + NH $_2^-$, highlighting the reaction coordinate ξ used in the two reaction steps.

succinimide intermediate ($\xi \approx 20$ a.u.). The effect of the solvent is to stabilize the intermediates, while the transition state for both reactions is only slightly affected by the presence of water molecules.

It is worth noting that in this model system, $\Delta A_{Asn \rightarrow Tet}^{\ddagger}$ is equal to 18.7 kcal/mol, while $\Delta A_{Tet \rightarrow Suc}^{\ddagger}$ is 16.5 kcal/mol, in rather good agreement with a previous computational study.⁴⁸ From the overall free energy profile of succinimide formation, it is possible to evaluate the kinetic rate constants of the process. From Figure 3, it comes out that $\Delta A_{Tet \rightarrow Asn}^{\ddagger} < \Delta A_{Asn \rightarrow Tet}^{\ddagger}$, meaning that when the tetrahedral intermediate is formed, the reaction is able to go back to the reactant, affecting the overall kinetics of the process. Hence, to estimate the time required for succinimide formation, we apply the kinetic scheme described in the Deamidation kinetic model section. The resulting population profiles of the species involved in the reaction versus time are reported in Figure 4.

The calculated $\tau_{1/2}$ of succinimide formation—the time at which half of the product is formed—is, at 300 K, 1.7, or 5.1 days using Landau or Eyring equations, respectively. It is worth noting that, considering the reaction leading to the formation of the Tet⁻ intermediate as irreversible, remarkable differences in the reactions' kinetics arise (see Figure S1). Therefore, for proper treatment of the reaction kinetics, it is mandatory to identify the correct kinetic scheme.

4.2. Succinimide Hydrolysis: Water-Mediated vs Hydroxide Ion Catalysis. Due to the imidic nature of the species of interest, whose reactivity is known to be slightly lower than the corresponding amide and, of particular interest here, more resistant to hydrolysis, two hydrolysis mechanisms were taken into account: the alkaline catalysis and the water-mediated mechanism. A direct water attack on the poor electrophilic carbon was considered inappropriate, while acidic hydrolysis was not included because at pH < 5 deamidation

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Figure 4. Time evolution of Asn⁻, Tet⁻, and Suc + NH₂⁻ normalized populations within the kinetic model used in the present work (eqs 18–20) with (a) Landau kinetic constants: $k_{L1} = 7.0 \times 10^{-1}$, $k_{L-1} = 4.3 \times 10^{6}$, and $k_{L2} = 28.9 \text{ s}^{-1}$; (b) Eyring kinetic constants: $k_{E1} = 1.40 \times 10^{-1}$, $k_{E-1} = 5.12 \times 10^{5}$, and $k_{E2} = 5.79 \text{ s}^{-1}$.



Figure 5. Succinimide hydrolysis mechanisms: water-mediated (upper panel) and alkaline catalysis (lower panel). Optimized structures of reactants, transition states (TSs), intermediates, and products are reported for each mechanism.

proceeds via a direct reaction, which does not involve the formation of the succinimide intermediate. 2,11

Therefore, we evaluated the two possible mechanisms for Asp generation from succinimide in the gas phase, while only the RDS for this stage has been studied in solution. In the water-mediated mechanism, two water molecules were included in the QM calculations,^{20,49} where one acts as a nucleophile while the other activates both the electrophilic and nucleophilic species, catalyzing the reaction. The second water molecule is, in fact, essential to form the active nucleophilic species, the hydroxide ion, obtained by an exchange of protons between the three oxygen atoms (i.e., the attacking oxygen transfers a proton to the second water molecule, which, acting as a proton shuttle, makes the attacked carbonyl an alcohol; Figure 5, upper panel).

When the gemdiol (Gem) intermediate is formed, the reaction proceeds through the concerted five-term ring opening and the N protonation by the second water molecules (the bidimensional potential energy surface is shown in Figure S2), which is restored via a proton transfer from one of the side chain hydroxyls, leading to the final product (Figure 5, upper panel).

On the other hand, alkaline catalysis shows lower energy barriers than the former process, reported in Table 1, as the stronger nucleophile easily attacks the inactive imidic carbon. In this way, the anionic form of Gem is produced, which subsequently leads to the ring opening. The final, fast

Table 1. Hydrolysis Energy Barriers and Kinetic Rate Constants in the Gas phase (Calculated Using eq 12)

	reaction step	ΔE^{\ddagger} (kcal/mol)	$k (s^{-1})$
water-mediated	$Suc \to Gem$	37.2	5.1×10^{-15}
	$\mathrm{Gem} \to \mathrm{Asp}$	19.7	2.6×10^{-2}
alkaline catalysis	$Suc \to Gem$	2.1	1.7×10^{4a}
	$\text{Gem} \to \text{Int}$	7.5	1.4×10^{7}
	$\mathrm{Int} \to \mathrm{Asp}$	0.38	3.2×10^{12}

^{*a*}Second order rate constant, pH = 7.



Figure 6. Free energy of the RDS of succinimide hydrolysis considering the water-mediated mechanism (left) and alkaline catalysis (right).

protonation of the nitrogen atom in position n + 1 by a water molecule leads to the deamidated product Asp in its carboxylated form. The reactant, transition states, and products are reported in Figure 5. From that Figure, it can be seen that alkaline catalysis requires an additional step with respect to the water-mediated case but appears mainly barrier-less. The gasphase energy barriers for both hydrolysis mechanisms and the corresponding rate constants are reported in Table 1.

In the water-mediated mechanism, water attack requires a larger activation energy, in agreement with literature data,²⁰ while in alkaline catalysis, the ring-opening step limits the reaction kinetics (Gem \rightarrow Int). However, from a kinetic point of view, these processes are bimolecular, and thus, the concentration of the other reactants needs to be taken into account. In water, the dependency of the rate constant on its concentration can be neglected, while the alkaline catalysis is affected by the concentration of the nucleophilic species (i.e., the hydroxide ion). In light of such considerations, as can be seen from the kinetic constant reported in Table 1, the nucleophilic attack is the RDS for both the hydrolysis mechanisms. Therefore, the Corresponding free energy profiles were evaluated by means of the PMM approach (Figure 6).

In alkaline catalysis, the reactant, which featured a net charge separation, is strongly stabilized by the solvent. The effect is less pronounced in the transition state, resulting in an increase in the activation barrier with respect to the in-vacuum condition.

As reported in Table 2, the RDS rate constant is significantly higher in the alkaline catalysis (Suc \rightarrow Gem step) with respect

Table 2. Free Energy and Kinetic Rate Constants for Suc \rightarrow Gem Transformation (300 K and pH = 7)

	ΔA^{\ddagger} (kcal/mol)	$k_{\rm L} ({\rm s}^{-1})$	$k_{\rm E}~({\rm s}^{-1})$
water-mediated	31.2	2.7×10^{-14}	1.2×10^{-10}
alkaline catalysis	5.3	2.0×10^{2}	9.2×10

to the water-mediated mechanism, suggesting that the succinimide hydrolysis proceeds mainly via alkaline hydrolysis, even at neutral pH. Such a hypothesis is fully consistent with the strong correlation observed between deamidation rates and pH values, where a slope of $\simeq 1$ in a log(k) vs pH plot above pH 6 was observed.^{50,51}

4.3. Overall Deamidation Process. In light of the findings discussed above, the formation of succinimide and its hydrolysis need to be taken into account to evaluate the deamidation reaction kinetics. The kinetic rate constants and the activation free energy values for the overall reaction Asn⁻ \rightarrow Gem (see data in Table 3) suggest that the deamidation kinetic is, at least in the studied system, mainly determined by the succinimide formation.

Table 3. Calculated Activation Free Energies and Kinetic Rate Constants for the Sub-Reactions Needed to Transform Asn^- in Gem at 300 K and Neutral pH^a

reaction step	ΔA^{\ddagger} (kcal/mol)	$k_{\rm E}~({ m s}^{-1})$	$k_{\rm L}~({\rm s}^{-1})$
Asn \rightarrow Tet $^-$	18.7	1.40×10^{-1}	7.0×10^{-1}
$Tet^- \rightarrow Asn^-$	9.73	5.12×10^{5}	4.3×10^{6}
$\mathrm{Tet}^{-}\!$	16.5	5.79	28.9
$Suc \to Gem$	5.26	9.20×10	2.0×10^{2}
^a The estimated statisti	cal error is within	0.5% for ΔA^{\ddagger}	, while it i

under 1% for $k_{\rm L}$ and 4% for $k_{\rm E}$.

To validate these results, the calculated kinetic data were compared with the experimental results available in the literature. First of all, as no data are available for our model system, we considered experimental data on oligopeptides containing the Asn-Gly sequence. Among the several experimental works focused on deamidation kinetics, some are centered on the kinetics of succinimide formation and/or its hydrolysis,^{11,23,50,52} while others refer to the overall deamidation process, following the disappearance (onset) of Asn (Asp).^{2,53} Notably, regarding the succinimide hydrolysis kinetics, we cannot compare our results to the experimental ones due to the two parallel competitive reactions that occur (see Figure 1) under experimental conditions. However, Xie and co-workers results on succinimide hydrolysis mechanism are in good agreement with our alkaline catalysis hypothesis at neutral pH.5

To the best of our knowledge, there are no experimentally determined k_1 , k_{-1} , and k_2 ; thus, we could only compare deamidation half-life times, as reported in Table 4.

Table 4. Comparison of Experimental and Calculated Deamidation $\tau_{1/2}$, Obtained by the Propagation of eqs $18-20^a$

$ au_{1/2}$ (days)	$\operatorname{Calc}_{\operatorname{L}}$	$\operatorname{Calc}_{\operatorname{E}}$	Exp	Exp	Exp	Exp	Exp
	1.70	5.07	1.41 ^b	1.03 ^c	4.96 ^d	1.89 ^e	1.14 ^f

^{*a*}Calc_L (Calc_E) is the reaction half-life times considering Landau (Eyring)-derived rate constants; its statistical error never exceeds 3% (5%). All the experimental results were obtained at 310 K and neutral pH (7.4–7.5) with TrisHCl as buffer, unless 4.96¹¹ and 1.89² extrapolated at zero buffer concentration and in phosphate buffer, respectively.^{2,11} ^{*b*} ref 23. ^{*c*} ref 53. ^{*d*} ref 11. ^{*e*} ref 24.

5. CONCLUSIONS

As a common post-translational modification (PTM), deamidation holds significant importance in the development of biologics. This chemical process, involving the conversion of asparagine or glutamine residues to aspartic acid or glutamic acid, respectively, can occur spontaneously under various physiological conditions.⁵⁵ Deamidation can profoundly impact the stability, efficacy, and immunogenicity of therapeutic proteins, making it a critical consideration in biopharmaceutical development. To understand and mitigate the effects of deamidation, several computational approaches were proposed for estimating the rates and exploring the underlying mechanisms of this PTM.^{10,13–15,48}

In this work, we presented a theoretical-computational approach to reconstruct the overall deamidation kinetics in solution, evaluating the kinetic rate constants of the three reaction steps, e.g., ring closure, deamination, and hydrolysis. From the explicit treatment of these stages in solution, it comes out that the Tet⁻ \rightarrow Asn⁻ backward reaction plays a central role in deamidation kinetics. In fact, the rate of succinimide formation, which accounts for the overall deamidation process, is strictly affected by k_1 and k_{-1} . Notably, the forward and backward ring-closure reactions are often neglected by approximating the succinimide formation to a single irreversible step. Our data, in line with available experimental data evaluated in similar systems (i.e., short peptides), represent the first step toward the possibility to predict the PTM rate in different contexts, i.e., proteins, by theoretical-computational procedures based on an accurate treatment of the environment.

ASSOCIATED CONTENT

1 Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.jpcb.3c04662.

Absolute energies, imaginary frequencies, and geometries of stationary points in gas phase; population profiles of the species involved in succinimide formation applying the two consecutive irreversible reaction kinetic scheme; bidimensional in-vacuum potential energy surface of Gem to Asp subreaction within the watermediated mechanism. (PDF)

AUTHOR INFORMATION

Corresponding Authors

Enrico Guarnera – Global Analytical Development, 00012 Guidonia Montecelio, Italy; Antibody Discovery and Protein Engineering, Merck Healthcare KGaA, 64293 Darmstadt, *Germany;* orcid.org/0000-0002-6780-3313; Email: enrico.guarnera@merckgroup.com

Marco D'Abramo – Department of Chemistry, University of Rome, 00185 Rome, Italy; • orcid.org/0000-0001-6020-8581; Email: marco.dabramo@uniroma1.it

Authors

- Maria Laura De Sciscio Department of Chemistry, University of Rome, 00185 Rome, Italy Alessandro Nicola Nardi – Department of Chemistry,
- University of Rome, 00185 Rome, Italy
- Fabio Centola Global Analytical Development, 00012 Guidonia Montecelio, Italy
- **Mara Rossi** Global Analytical Development, 00012 Guidonia Montecelio, Italy

Complete contact information is available at: https://pubs.acs.org/10.1021/acs.jpcb.3c04662

Notes

The authors declare no competing financial interest.

REFERENCES

(1) Yan, Q.; Huang, M.; Lewis, M. J.; Hu, P. Structure Based Prediction of Asparagine Deamidation Propensity in Monoclonal Antibodies. *mAbs* **2018**, *10*, 901–912.

(2) Patel, K.; Borchardt, R. T. Chemical pathways of peptide degradation. III. Effect of primary sequence on the pathways of deamidation of asparaginyl residues in hexapeptides. *Pharm. Res.* **1990**, 07, 787–793.

(3) Robinson, N.; Robinson, A. Prediction of primary structure deamidation rates of asparaginyl and glutaminyl peptides through steric and catalytic effects. *J. Pept. Res.* **2004**, *63*, 437–448.

(4) Robinson, N.; Robinson, Z.; Robinson, B.; Robinson, A.; Robinson, J.; Robinson, M.; Robinson, A. Structure-dependent nonenzymatic deamidation of glutaminyl and asparaginyl pentapeptides. J. Pept. Res. 2004, 63, 426–436.

(5) Weintraub, S. J.; Deverman, B. E. Chronoregulation by Asparagine Deamidation. *Sci. STKE* 2007, 2007, re7.

(6) Forsythe, H. M.; Vetter, C. J.; Jara, K. A.; Reardon, P. N.; David, L. L.; Barbar, E. J.; Lampi, K. J. Altered Protein Dynamics and Increased Aggregation of Human γ S-Crystallin Due to Cataract-Associated Deamidations. *Biochem* **2019**, *58*, 4112–4124.

(7) Wilmarth, P. A.; Tanner, S.; Dasari, S.; Nagalla, S. R.; Riviere, M. A.; Bafna, V.; Pevzner, P. A.; David, L. L. Age-Related Changes in Human Crystallins Determined from Comparative Analysis of Post-translational Modifications in Young and Aged Lens: Does Deamidation Contribute to Crystallin Insolubility? *J. Proteome Res.* **2006**, *5*, 2554–2566.

(8) Gupta, S.; Jiskoot, W.; Schöneich, C.; Rathore, A. S. Oxidation and Deamidation of Monoclonal Antibody Products: Potential Impact on Stability, Biological Activity, and Efficacy. *J. Pharm. Sci.* **2022**, *111*, 903–918.

(9) Gervais, D. Protein deamidation in biopharmaceutical manufacture: understanding, control and impact. J. Chem. Technol. Biotechnol. 2016, 91, 569–575.

(10) Irudayanathan, F. J.; Zarzar, J.; Lin, J.; Izadi, S. Deciphering deamidation and isomerization in therapeutic proteins: Effect of neighboring residue. *mAbs* **2022**, *14*, 2143006.

(11) Capasso, S.; Mazzarella, L.; Sica, F.; Zagari, A.; Salvadori, S. Kinetics and mechanism of succinimide ring formation in the deamidation process of asparagine residues. *J. Chem. Soc., Perkin Trans.* 2 **1993**, 679.

(12) Radkiewicz, J. L.; Zipse, H.; Clarke, S.; Houk, K. N. Neighboring Side Chain Effects on Asparaginyl and Aspartyl Degradation: An Ab Initio Study of the Relationship between Peptide Conformation and Backbone NH Acidity. *J. Am. Chem. Soc.* 2001, 123, 3499–3506.

(13) Ugur, I.; Marion, A.; Aviyente, V.; Monard, G. Why Does Asn71 Deamidate Faster Than Asn15 in the Enzyme Triosephosphate Isomerase? Answers from Microsecond Molecular Dynamics Simulation and QM/MM Free Energy Calculations. *Biochemistry* **2015**, *54*, 1429–1439.

(14) Plotnikov, N. V.; Singh, S. K.; Rouse, J. C.; Kumar, S. Quantifying the Risks of Asparagine Deamidation and Aspartate Isomerization in Biopharmaceuticals by Computing Reaction Free-Energy Surfaces. J. Phys. Chem. B 2017, 121, 719–730.

(15) Vatsa, S. In silico prediction of post-translational modifications in therapeutic antibodies. *mAbs* **2022**, *14*, 2023938.

(16) Ugur, I.; Aviyente, V.; Monard, G. Initiation of the Reaction of Deamidation in Triosephosphate Isomerase: Investigations by Means of Molecular Dynamics Simulations. *J. Phys. Chem. B* **2012**, *116*, 6288–6301.

(17) Kumar, S.; Plotnikov, N. V.; Rouse, J. C.; Singh, S. K. Biopharmaceutical Informatics: supporting biologic drug development *via* molecular modelling and informatics. *J. Pharm. Pharmacol.* **2018**, 70, 595–608.

(18) Sydow, J. F.; Lipsmeier, F.; Larraillet, V.; Hilger, M.; Mautz, B.; Mølhøj, M.; Kuentzer, J.; Klostermann, S.; Schoch, J.; Voelger, H. R.; et al. Structure-Based Prediction of Asparagine and Aspartate Degradation Sites in Antibody Variable Regions. *PLoS One* **2014**, *9*, No. e100736.

(19) Delmar, J. A.; Wang, J.; Choi, S. W.; Martins, J. A.; Mikhail, J. P. Machine Learning Enables Accurate Prediction of Asparagine Deamidation Probability and Rate. *Mol. Ther.*—*Methods Clin. Dev.* **2019**, *15*, 264–274.

(20) Catak, S.; Monard, G.; Aviyente, V.; Ruiz-López, M. F. Deamidation of Asparagine Residues: Direct Hydrolysis versus Succinimide-Mediated Deamidation Mechanisms. *J. Phys. Chem. A* **2009**, *113*, 1111–1120.

(21) Peters, B.; Trout, B. L. Asparagine Deamidation: pH-Dependent Mechanism from Density Functional Theory. *Biochem* 2006, 45, 5384–5392.

(22) Lawson, K. E.; Dekle, J. K.; Evans, M. N.; Adamczyk, A. J. Deamidation reaction network mapping of pharmacologic and related proteins: impact of solvation dielectric on the degradation energetics of asparagine dipeptides. *React. Chem. Eng.* **2022**, *7*, 1525–1543.

(23) Geiger, T.; Clarke, S. Deamidation, isomerization, and racemization at asparaginyl and aspartyl residues in peptides. Succinimide-linked reactions that contribute to protein degradation. *J. Biol. Chem.* **1987**, *262*, 785–794.

(24) Zanetti-Polzi, L.; Del Galdo, S.; Daidone, I.; D'Abramo, M.; Barone, V.; Aschi, M.; Amadei, A. Extending the perturbed matrix method beyond the dipolar approximation: comparison of different levels of theory. *Phys. Chem. Chem. Phys.* **2018**, *20*, 24369–24378.

(25) Amadei, A.; D'Alessandro, M.; Aschi, M. Statistical Mechanical Modeling of Chemical Reactions in Complex Systems: The Reaction Free Energy Surface. *J. Phys. Chem. B* **2004**, *108*, 16250–16254.

(26) Amadei, A.; D'Abramo, M.; Daidone, I.; D'Alessandro, M.; Nola, A. D.; Aschi, M. Statistical mechanical modelling of chemical reactions in complex systems: the kinetics of the Haem carbon monoxide binding–unbinding reaction in Myoglobin. *Theor. Chem. Acc.* **2007**, *117*, 637–647.

(27) Amadei, A.; D'Abramo, M.; Zazza, C.; Aschi, M. Electronic properties of formaldehyde in water: a theoretical study. *Chem. Phys. Lett.* **2003**, 381, 187–193.

(28) Aschi, M.; D'Abramo, M.; Ramondo, F.; Daidone, I.; D'Alessandro, M.; Di Nola, A.; Amadei, A. Theoretical modeling of chemical reactions in complex environments: the intramolecular proton transfer in aqueous malonaldehyde. *J. Phys. Org. Chem.* **2006**, *19*, 518–530.

(29) Chen, C. G.; Nardi, A. N.; Amadei, A.; D'Abramo, M. PyMM: An Open-Source Python Program for QM/MM Simulations Based on the Perturbed Matrix Method. *J. Chem. Theory Comput.* **2023**, *19*, 33– 41. (30) Nardi, A. N.; Olivieri, A.; Amadei, A.; Salvio, R.; D'Abramo, M. Modelling Complex Bimolecular Reactions in a Condensed Phase: The Case of Phosphodiester Hydrolysis. *Molecules* **2023**, *28*, 2152.

(31) Segreto, G. E.; Alba, J.; Salvio, R.; D'Abramo, M. DNA cleavage by endonuclease I-DmoI: a QM/MM study and comparison with experimental data provide indications on the environmental effects. *Theor. Chem. Acc.* **2020**, *139*, 68.

(32) De Sciscio, M. L.; D'Annibale, V.; D'Abramo, M. Theoretical Evaluation of Sulfur-Based Reactions as a Model for Biological Antioxidant Defense. *Int. J. Mol. Sci.* **2022**, 23, 14515.

(33) Moore, J. W.; Pearson, R. G. Kinetics and Mechanism; John Wiley & Sons, 1981.

(34) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Petersson, G. A.; Nakatsuji, H.; et al. *Gaussian 16*. Revision C.01; Gaussian Inc: Wallingford CT, 2016;.

(35) Abraham, M. J.; Murtola, T.; Schulz, R.; Pall, S.; Smith, J. C.; Hess, B.; Lindahl, E. GROMACS: High performance molecular simulations through multi-level parallelism from laptops to supercomputers. *SoftwareX* **2015**, *1–2*, 19–25.

(36) Sami, S.; Menger, M. F.; Faraji, S.; Broer, R.; Havenith, R. W. A. Q-Force: Quantum Mechanically Augmented Molecular Force Fields. *J. Chem. Theory Comput.* **2021**, *17*, 4946–4960.

(37) Jorgensen, W. L.; Tirado-Rives, J. The OPLS [optimized potentials for liquid simulations] potential functions for proteins, energy minimizations for crystals of cyclic peptides and crambin. *J. Am. Chem. Soc.* **1988**, *110*, 1657–1666.

(38) Robertson, M. J.; Tirado-Rives, J.; Jorgensen, W. L. Improved Peptide and Protein Torsional Energetics with the OPLS-AA Force Field. J. Chem. Theory Comput. **2015**, 11, 3499–3509.

(39) Singh, U. C.; Kollman, P. A. An approach to computing electrostatic charges for molecules. *J. Comput. Chem.* **1984**, *5*, 129–145.

(40) Besler, B. H.; Merz, K. M., Jr.; Kollman, P. A. Atomic charges derived from semiempirical methods. *J. Comput. Chem.* **1990**, *11*, 431–439.

(41) Mark, P.; Nilsson, L. Structure and Dynamics of the TIP3P, SPC, and SPC/E Water Models at 298 K. J. Phys. Chem. A 2001, 105, 9954–9960.

(42) Del Galdo, S.; Marracino, P.; D'Abramo, M.; Amadei, A. In silico characterization of protein partial molecular volumes and hydration shells. *Phys. Chem. Chem. Phys.* **2015**, *17*, 31270–31277.

(43) Bussi, G.; Donadio, D.; Parrinello, M. Canonical sampling through velocity rescaling. J. Chem. Phys. 2007, 126, 014101.

(44) Essmann, U.; Perera, L.; Berkowitz, M. L.; Darden, T.; Lee, H.; Pedersen, L. G. A smooth particle mesh Ewald method. *J. Chem. Phys.* **1995**, *103*, 8577–8593.

(45) Darden, T.; York, D.; Pedersen, L. G. Particle mesh Ewald: An N-log(N) method for Ewald sums in large systems. *J. Chem. Phys.* **1993**, *98*, 10089–10092.

(46) Páll, S.; Hess, B. A flexible algorithm for calculating pair interactions on SIMD architectures. *Comput. Phys. Commun.* **2013**, *184*, 2641–2650.

(47) Daura, X.; Gademann, K.; Jaun, B.; Seebach, D.; van Gunsteren, W. F.; Mark, A. E. Peptide Folding: When Simulation Meets Experiment. *Angew. Chem., Int. Ed.* **1999**, *38*, 236–240.

(48) Kaliman, I.; Nemukhin, A.; Varfolomeev, S. Free Energy Barriers for the N-Terminal Asparagine to Succinimide Conversion: Quantum Molecular Dynamics Simulations for the Fully Solvated Model. J. Chem. Theory Comput. **2010**, *6*, 184–189.

(49) Antonczak, S.; Ruiz-Lopez, M. F.; Rivail, J. L. Ab Initio Analysis of Water-Assisted Reaction Mechanisms in Amide Hydrolysis. *J. Am. Chem. Soc.* **1994**, *116*, 3912–3921.

(50) Capasso, S.; Kirby, A. J.; Salvadori, S.; Sica, F.; Zagari, A. Kinetics and mechanism of the reversible isomerization of aspartic acid residues in tetrapeptides. *J. Chem. Soc., Perkin Trans.* 2 **1995**, 437–442.

(51) Xie, M.; Vander Velde, D.; Morton, M.; Borchardt, R. T.; Schowen, R. L. pH-Induced Change in the Rate-Determining Step for the Hydrolysis of the Asp/Asn-Derived Cyclic-Imide Intermediate in Protein Degradation. J. Am. Chem. Soc. **1996**, 118, 8955–8956.

(52) Capasso, S.; Di Cerbo, P. Kinetic and thermodynamic control of the relative yield of the deamidation of asparagine and isomerization of aspartic acid residues: Control of deamidation and isomerization yields. *Pept. Res.* **2000**, *56*, 382–387.

(53) Robinson, N. E.; Robinson, A. B. Molecular clocks. Proc. Natl. Acad. Sci. U.S.A. 2001, 98, 944–949.

(54) Stephenson, R. C.; Clarke, S. Succinimide Formation from Aspartyl and Asparaginyl Peptides as a Model for the Spontaneous Degradation of Proteins. J. Biol. Chem. **1989**, 264, 6164–6170.

(55) Pace, A. L.; Wong, R. L.; Zhang, Y. T.; Kao, Y.-H.; Wang, Y. J. Asparagine Deamidation Dependence on Buffer Type, pH, and Temperature. *J. Pharm. Sci.* **2013**, *102*, 1712–1723.