

Exercise-mediated reinnervation of skeletal muscle in elderly people: An update

Claudia Coletti (1), Gilberto F Acosta (1), Stefan Keslacy (1), Dario Coletti (2,3,4)

(1) School of Kinesiology, Nutrition and Food Science, California State University Los Angeles, Los Angeles, CA, USA; (2) DAHFMO - Unit of Histology and Medical Embryology, Sapienza University of Rome, Rome, Italy; (3) Biological Adaptation and Ageing, CNRS UMR 8256, Inserm U1164, Institut de Biologie Paris-Seine, Sorbonne Université, Paris, France; (4) Interuniversity institute of Myology, Rome, Italy.

This article is distributed under the terms of the Creative Commons Attribution Commercial License (CC BY NC), which permits any commercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

Abstract

Sarcopenia is defined by the loss of muscle mass and function. In aging sarcopenia is due to mild chronic inflammation but also to fiber-intrinsic defects, such as mitochondrial dysfunction. Age-related sarcopenia is associated with physical disability and lowered quality of life. In addition to skeletal muscle, the nervous tissue is also affected in elderly people. With aging, type 2 fast fibers preferentially undergo denervation and are reinnervated by slow-twitch motor neurons. They spread forming new neuro-muscular junctions with the denervated fibers: the result is an increased proportion of slow fibers that group together since they are associated in the same motor unit. Grouping and fiber type shifting are indeed major histological features of aging skeletal muscle. Exercise has been proposed as an intervention for age-related sarcopenia due to its numerous beneficial effects on muscle mechanical and biochemical features. In 2013, a precursor study in humans was published in the European Journal of Translation Myology (formerly known as Basic and Applied Myology), highlighting the occurrence of reinnervation in the musculature of aged, exercise-trained individuals as compared to the matching control. This paper, entitled «Reinnervation of Vastus lateralis is increased significantly in seniors (70-years old) with a lifelong history of high-level exercise», is now being reprinted for the second issue of the «Ejtm Seminal Paper Series». In this short review we discuss those results in the light of the most recent advances confirming the occurrence of exercise-mediated reinnervation, ultimately preserving muscle structure and function in elderly people who exercise.

Key Words: Sarcopenia; denervation; muscle atrophy; skeletal muscle; physical activity.

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

The definition of sarcopenia, initially meant to indicate the progressive loss of skeletal muscle mass associated with aging, has recently been updated by stressing the importance of the loss of muscle strength and physical performance.¹ Indeed, functional assessments are now part of the definition of sarcopenia, similarly to other forms of muscle atrophy such as cachexia,² which reckons the importance of muscle functionality in defining a muscle disease state. The association of sarcopenia with a significant impairment of function such as slowing of movement and loss of neuromuscular control is linked to decreased quality of life and increased risk of falls.³ With the global aging population, sarcopenia becomes increasingly prevalent, rising major concerns for the health-related authorities,⁴ and an increased awareness of the importance of research aimed to improve the preservation of the muscle mass and its related function in elderly individuals. A major avenue of

research in the field of age-related sarcopenia deals with the process of muscle denervation in aging and the countermeasures that prevent this phenomenon or favor recovery in the elderly. A pioneer report on this subject is the article by Mosole et al.,⁵ which is re-published in the current issue of European Journal of Translational Myology. Their findings on the beneficial effects of exercise on muscle reinnervation with age have been confirmed and extended by many others⁶⁻⁹ and are discussed in this mini-review in the light of the more recent literature.

Muscle fiber type and fiber shifting

Fiber diversity reflects the plethora of muscle functional and metabolic roles. Myosin has a prominent position both as a marker for fiber typing and as a molecular motor characterizing the function of the muscle tissue. The conventional class II of muscle myosin encompasses 18 genes driving the expression of several myosin heavy

Exercise-mediated reinnervation of skeletal muscle in elderly

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

Table 1. Major myosin isoforms expressed in mammal muscle

Gene	Protein	Functional properties	Expression pattern
<i>MYH1</i>	MyHC-2X	Fast contracting	Multiple skeletal muscles, Type 2X fibers
<i>MYH2</i>	MyHC-2A	Fast contracting	Multiple skeletal muscles, Type 2A fibers
<i>MYH3</i>	MyHC-emb	Slow contracting ⁸⁷	Developing and regenerating muscle
<i>MYH4</i>	MyHC-2B	Fast contracting	Not in humans, multiple skeletal muscles, Type 2B fibers
<i>MYH6</i>	MyHC-alpha	Intermediate speed ⁸⁸	Cardiac and skeletal muscle of the jaws
<i>MYH7</i>	MyHC-beta/slow	Slow contracting	Cardiac and slow skeletal muscles, Type 1 fibers
<i>MYH8</i>	MyHC-neo	NA	Developing and regenerating muscle
<i>MYH13</i>	MyHC-EO	Superfast cont. ⁸⁹	Extraocular (EO) skeletal muscles
<i>MYH14*</i>	MyHC-slow/tonic	Slow contracting ⁹⁰	Muscle spindles, Extraocular skeletal muscles
<i>MYH15</i>	MyHC-15	Slow contracting ⁹⁰	Muscle spindles, Extraocular skeletal muscles
<i>MYH16</i>	MyHC-M		Not in humans, ^o Translational Myolog it is evident Skeletal muscle of the jaws

The table summarizes the nomenclature and expression pattern of the myosin heavy chain (MyHC) genes (MYH) and the corresponding sarcomeric protein product in mammals. The expression pattern in the striated muscles, as well as the functional properties in terms of contraction speed, are also reported. Hybrid fibers containing two MyHC

chain (MyHC) isoforms.¹⁰ These genes are evolutionary conserved in mammals and expressed to a variable extent and in different combinations in adult striated muscles, while special isoforms are expressed during embryogenesis, perinatal life, and muscle regeneration (Table 1). Distinct myosin gene expression determines four major muscle fiber types, as exhaustively highlighted by Schiaffino and Reggiani.¹⁰ Briefly: there are two types of fast-contracting fibers, called type 2A and 2B, which differ for being fatigue-resistant and fatigable, respectively, the type 1 fibers predominant in slow-twitch muscles, the most resistant to fatigue, which form mostly postural muscles; a fourth type of fiber, the 2X, has twitch properties (contraction and half-relaxation time) similar to those of 2A and 2B units, and has an intermediate resistance to fatigue.^{10,11} These fiber types are determined during development by hormonal and neurogenic factors but are susceptible of extensive remodeling in postnatal life. The default phenotype of a muscle fiber is the fast one,¹⁰ as confirmed by the fact that denervated, regenerating muscle fibers activate the default expression of fast-like myosin expression.¹² Worth noting, regenerating muscle fibers following trauma or pathological cues re-enter a developmental program in which they re-express embryonic or perinatal

MyHC isoforms, such as the embryonic, fetal or neonatal myosins.^{13–16} Regenerating muscle fibers also share morphological features with pre-natal muscle fibers, such as the central nuclei,¹⁷ and ultimately mature acquiring features that are not necessarily those prior to damage, which accounts for muscle plasticity. The final MyHC isoform expressed by an adult, regenerated muscle fiber is ultimately conditioned by the microenvironment, including the type of motoneuron that innervates the fiber.¹⁸ Muscle fiber types also differ with respect to resistance to fatigue and numerous additional features, as highlighted by the differential content of succinate dehydrogenase (SDH) and other metabolic enzymes. Interestingly, fiber shifts in MyHC with aging are associated to a remodulation of these enzymatic activities.¹⁹ Additional factors, such as the increase in resting muscle stiffness, affect age-related muscle impairment and will not be discussed here.²⁰

Effects of aging on the skeletal muscle

Aging is characterized by a progressive decline in skeletal muscle mass, ultimately leading to decreased strength and functionality.¹ The two major mechanisms underpinning the decline in muscle mass are muscle fiber atrophy²¹ and muscle fiber loss, even though there is not

consensus on the latter.²² Type 2 fibers are preferentially sensitive to atrophy in aging.²³ Two additional phenomena characteristic of aging are fiber type grouping and shifting. Grouping is the presence of clusters of fibers of the same type, defined as a group of fibers with at least one enclosed fiber.²⁴ Grouping typically occurs for type 1 fibers in aging²⁴ and is associated to an enrichment in the percentage of type 1 fibers in the musculature.²⁵ The latter is a paradoxical phenomenon, since, as state above, the default muscle fiber phenotype is the fast, type 2 fiber¹² therefore, aging actively and selectively promote the appearance of type 1 fibers in the musculature. Indeed, in murine models it has been shown that regenerating fibers express by default the fast MyHC-2X and -2B transcripts in absence of innervation, even in slow muscle such as the Soleus. If slow nerve activity is restored or mimicked by electrodes, this induces a switch toward a slow, MyHC-beta/slow isoform in the targeted muscle fibers,²⁶ through a pathway involving RAS and ERK activation.²⁷ Thus, the fiber type essentially depends on the innervation of the muscle fibers themselves. Taken together these reports helped to state the hypothesis that, with aging, type 2, fast fibers preferentially undergo denervation to then form new NMJ by reinnervation from slow-twitch motor neurons. The result is an increased proportion of slow fibers, which also group with each other, since they are associated in the same motor unit. These phenomena, in our opinion, would also explain the slower movements typical of elderly individuals. If reinnervation is insufficient, then fibers undergo atrophy or apoptosis.²⁸ The sarcopenia-associated changes in motor unit numbers has been demonstrated using electromyographic motor unit recordings through surface-recorded compound muscle action potentials (CMAP) and indirectly through MRI and histochemistry (reviewed by Piasecki et al.).²⁹ Motor unit loss and the alterations characterizing the aging neurons are superbly summarized in a recent review by Larsson et al. and will not be further discussed here, for lack of space.³ In addition to denervation and atrophy, other alterations characterize the aging muscle, such as a reduced regenerative capacity and fiber-intrinsic defects: protein oxidation, organelle dysfunction (including lysosomes and mitochondria), changes in sarcomeres and endoplasmic reticulum, ultimately leading to defects in calcium handling.³⁰ Collectively, these events synergize, leading to loss of muscle function during aging.³⁰ For instance, calcium homeostasis, which is very important for the myogenic development and muscle regeneration,^{31,32} is also essential in the adult muscle fiber and calcium leakage is a major responsible for the diminished contraction capacity observed in aging.³³ In the 2010 European consensus article “Report of the European working group on Sarcopenia in older people”¹ the authors described the major muscle issues associated with aging and proposed strategies that include treatments and changes in lifestyle in order to prevent

age-associated sarcopenia. These interventions are as diverse as hormone therapy and exercise/physical activity, caloric intake control and lowering inflammation.³ The findings of Mosole et al. summarized below, paved the way to exercise-based interventions to spare muscle wasting and pinpointed the peculiar role of denervation-reinnervation processes.

Major findings by Mosole et al. and more recent developments

In the seminal work by Mosole et al. the group demonstrated that long-term high-level exercise promotes muscle reinnervation with age⁵ (republished in the current issue of Eur J Transl Myol). Indeed, this has been the focus of their research activities for years and led to three publications - first in 2013 and then, extending the original findings, in 2014 and 2016 - that are discussed in details below. Researchers at the University of Padua are among those who propose to apply *strictu sensu* the concept of “use it or lose it” to sarcopenic muscles.⁸ Since the 90s there was an interest on skeletal muscle in master athletes and Westerterp in 2000 was probably one of the very first to extensively demonstrate and discuss the benefits deriving from physical activity for elderly individuals.^{34,35} Nonetheless, Mosole and coworkers were the first ones to demonstrate a clear effect of exercise on muscle fiber reinnervation, thus paving the road for a new avenue of investigation. Indeed, in 2013 Mosole et al. compared muscle biopsies from sedentary seniors, physically active (i.e. sportsmen) seniors and young people in order to prove that exercise protects against age-related denervation.⁵ In this study they clearly demonstrated the occurrence of selective denervation in the aged muscle, as highlighted by the occurrence of angular, atrophic muscle fibers, a phenomenon prevented in life-long exercised muscles. In addition, the authors demonstrated that senior sportsmen show a greater level of type 1-fiber grouping, as if their muscle were undergoing a higher rate of reinnervation.⁵ One year later, in 2014, Mosole et al. further developed this idea, by exploiting a technically improved approach to analyze skeletal muscle biopsies, consisting in immunofluorescence for MyHC types.³⁶ They clearly showed the effect of exercise on bona fide denervation markers (such as N-CAM expression by the muscle fibers), as well as fiber type transition (by double immunofluorescence showing the co-expression of fast and slow MyHC). Worth noting, while histology was comparable between the two studies, in the 2014 article the force of the quadriceps was measured, instead of that of the vastus lateralis that was analyzed in the 2013 article. The authors also compared slow fibers with the type of training undertaken by the physically active seniors but they did not establish a correlation. The authors concluded that a long-term, intense exercise promotes reinnervation of muscle fibers, with positive consequences on the muscle structure and function, ultimately leading to a delay in mobility decline.³⁶

Exercise-mediated reinnervation of skeletal muscle in elderly

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

Mosole's 2013 article has been inspirational and the reference for several other papers over the years, confirming that heavy training improves skeletal muscles in very old individuals. In particular, the pivotal role of motor units in the maintenance of muscle mass with aging has been confirmed, on the basis of observations in both male and female master athletes.³⁷ To this regard, the group of Holm demonstrated a very innovative idea, i.e. that just 12 weeks of heavy resistance training are enough to improve muscle strength in old individuals and, suggesting that lifelong exercise may not be strictly required to generate beneficial effects.³⁸ Messi et al. extended these observations to the obese population – indeed, sarcopenic obesity is a common condition observed in a major percentage of elderly individuals, characterized by the combination of being significantly overweight with the age-related loss of muscle mass and strength.³⁹ In particular, Messi et al. showed that resistance training diminished the expression of N-CAM (a denervation marker) in muscle fibers and affected fiber grouping even in obese patients. This group expanded their analysis to muscle stem cells, which account for the remarkable skeletal muscle plasticity, showing that exercise does not modify either the total number of satellite cells or their relative abundance in every fiber type.⁴⁰ The histological examination alone may not reveal the full extent of ageing-related motor unit remodeling; for instance, it was reported that the fiber grouping did not correlate with aging. Taken alone, these data would indicate that the examined population did not show a major feature of sarcopenia,²⁴ further suggesting the importance of the functional characterization of the

aged musculature. Notably, Messa et al. examined the vastus lateralis in athletes, albeit aged, which is a major difference in respect to the previous studies, that focused on very active people, but not athletes. A study by Kletzien et al. reported changes in thyroarytenoid muscle associated to both aging and to exercise training in rats. These changes, affecting MyHC isoforms, were consistent with a glycolytic to oxidative shift in muscle fiber type.⁴¹ The conclusion by the authors is that because thyroarytenoid muscle is active during vocalization – it is actually part of the laryngeal muscle structure – the approach of doing exercise is crucial in order to prevent “vocal function deficits, dysphagia, and aspiration observed in elderly people”.⁴¹ In addition, we believe that this study represents a striking evidence that the upper airways muscles are affected by treadmill running, even though they are not directly involved, offering a proof of principle of the occurrence of systemic effects of physical activity on muscle fiber type shifting. Another breakthrough article was the comparison of muscle status and performance on monozygotic twins with decades of discordant exercise habits. Here, the authors showed that the trained twins exhibited better endurance features, more slow-twitch MyHC, and increased level of pro-myogenic markers than their counterpart. They concluded that skeletal muscle as a high plasticity depending on lifestyle which overrides the genetic background.⁴² Intriguingly, an indirect but solid confirmation of Mosole's data comes from the group of Carraro and co-workers that in several studies have actually exploited Functional Electrical Stimulation (FES) to modify “muscle fibers by increasing

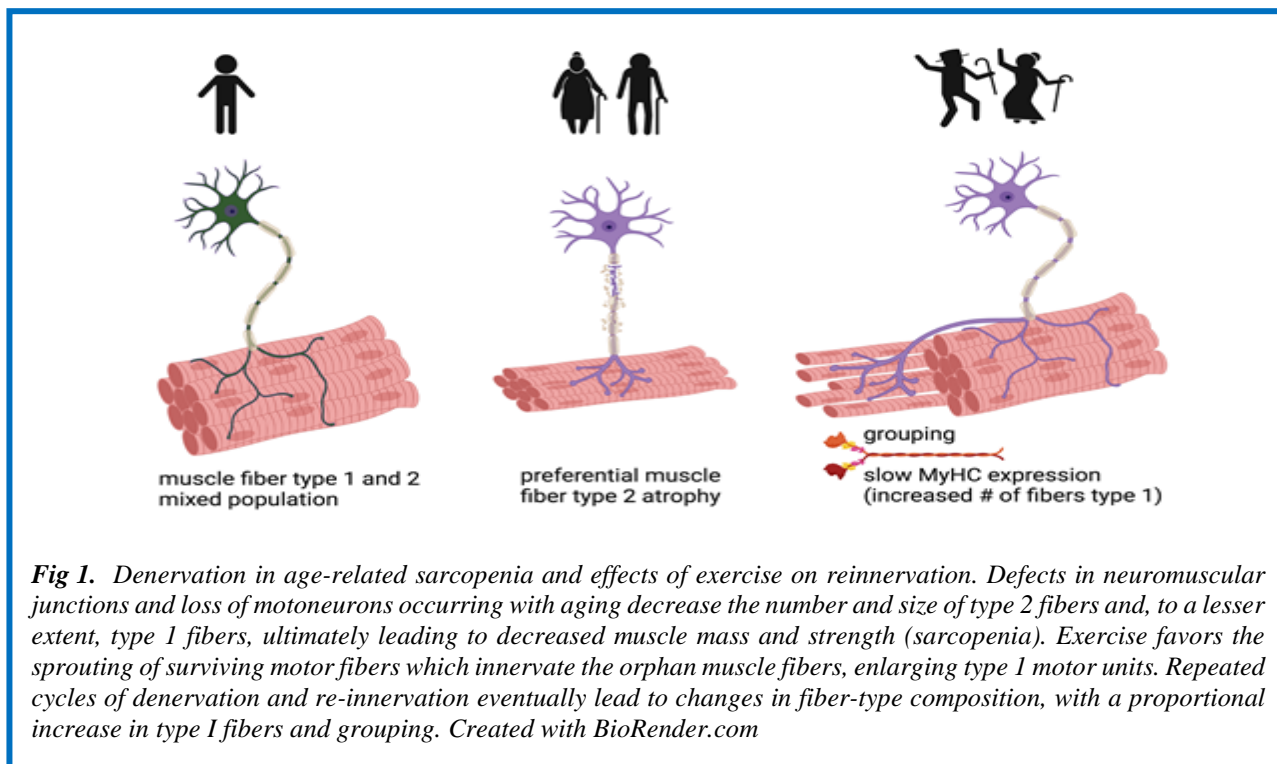


Fig 1. Denervation in age-related sarcopenia and effects of exercise on reinnervation. Defects in neuromuscular junctions and loss of motoneurons occurring with aging decrease the number and size of type 2 fibers and, to a lesser extent, type 1 fibers, ultimately leading to decreased muscle mass and strength (sarcopenia). Exercise favors the sprouting of surviving motor fibers which innervate the orphan muscle fibers, enlarging type 1 motor units. Repeated cycles of denervation and re-innervation eventually lead to changes in fiber-type composition, with a proportional increase in type I fibers and grouping. Created with BioRender.com

contractions per day^{43,44} in the absence of physical activity. While primarily used to counteract neurogenic atrophy,⁴⁵ FES has been successfully applied to age-related sarcopenia, demonstrating that muscle contraction *per se* is beneficial against atrophy.⁴⁶ Current trials are optimizing protocols for neuromuscular electrical stimulation in humans.⁴⁷ Whether FES is sufficient to promote reinnervation has not been established yet.⁴⁴ However, very promising results have been recently reported on the use of FES to promote regeneration of a transplanted nerve and muscle functional recovery in a rat model.⁴⁸ Based on all of the above, we conclude that muscle contraction, induced by physical activity or by electrical stimulation, protects motoneurons from age-related sarcopenia and that peripheral reinnervation might occur due to sprouting of slow motoneurons, ultimately preserving muscle structure and function, in elderly, exercised individuals (Figure 1).

The dark side of the moon: sensory nerves and intrafusal fibers in muscle spindles

The skeletal muscle possesses nociceptors⁴⁹ that are believed to mediate muscle pain via group III and IV afferent fibers. In addition, muscle has proprioceptive neurons, that innervate intrafusal fibers, in the muscle spindles, conveying length information to the central nervous system via primary type Ia and secondary type II sensory fibers.^{50,51} Very little is known on the effects of aging on the sensory fibers and nociceptors in the aging muscle. Nonetheless, research in this field is of the utmost importance. While being the effector system, the musculoskeletal system participate in the integration of information responsible for balance which also declines with aging involving major social and clinical consequences.⁵² Such decline in balance is likely due to both reduced proprioception and ability of the cortex to process sensory information,⁵³ resulting in an increased likelihood of falling when fatigued. The relevance of leg proprioception in postural control and its age-related alterations have been notably summarized by Henry and Baudry.⁵⁴ Aging alters the intrafusal fibers by increasing their number and modifying the content of intrafusal slow and developmental isoforms of MyHC.⁵⁵ Interestingly, intrafusal muscle fibers undergo degeneration and regeneration cycles during aging in rodents,⁵⁶ so one would expect the nervous fibers innervating the muscle spindle to also be affected by aging. While it has been reported that bone sensory fibers are spared by aging,⁵⁷ how aging affects proprioception in muscle remains poorly explored. Recently, it has been shown that, similarly to what happens in the bulk muscle fiber population, proprioceptive sensory neurons degenerate prior to atrophy of the intrafusal muscle fibers with aging.⁵⁸ However, not all the sensory neurons are affected by aging to the same extent: in aged rats the discharge frequency in response to muscle stretch (dynamic sensitivity) diminishes,^{59,60} whereas the static

sensitivity does not exhibit such an effect. While the effects of exercise on position sensing, including differential effects induced by different muscle contraction types, have been investigated in detail - for a review, see Fortier 2012,⁶¹ - to the best of our knowledge, nothing is known on whether exercise has any effect on aged nociceptive or proprioceptive fibers. This issue definitely deserves further investigation.

Countermeasures: which type of exercise is more efficacious against sarcopenia?

Physical activity has been shown to decrease with age with only an estimated 28-34% of adults age 65 and older participating in any leisure time physical activity.⁶² In regard to current trends in age-related changes in physical activity, the COVID-19 pandemic has demonstrated that the young and elderly population have experienced the greatest decline in physical activity.⁶³ The link between loss of muscle strength and mass is clearly dependent on physical inactivity,⁶⁴ which is a modifiable risk factor that can partially reverse skeletal muscle dysfunction related to age.⁶⁵ Physical activity, and more specifically resistance training, has been demonstrated to be a modality to enhance muscle function in older sarcopenic adults.⁶⁶ Overall the generalized recommendation in resistance training for sarcopenia includes dynamic movements involving the facilitation of both concentric and eccentric contractions of major muscle groups.⁶⁷ These movements can be accomplished by a variety of exercise modalities which include strength training, muscular endurance training, power training, and high intensity interval training. Some of the key factors that distinguish the difference between these modalities are the intensity and volume prescribed. In order for resistance training to induce a physiological response that will improve muscle mass and strength, a training load of greater than 70% of the one-repetition max (1-RM) is needed.⁶⁸ The adaptations from strength training has been demonstrated to increase muscle strength and muscle mass. However, increases in muscular strength or mass may not always relate to a direct improvement in functional muscle performance.⁶⁵ Worth noting, power training seems to not only elicit a positive effect on muscle mass³⁷ but may more directly improve muscle function.⁶⁹ There is also evidence of aerobic exercise, balance training and flexibility to have beneficial effects in sarcopenia^{70,71} and recent findings demonstrate that endurance exercise induces the appearance of hybrid fiber alterations in seniors consistent with what previously observed Mosole et al.⁷² Sarcopenia is a condition with multiple factors involved^{67,73} and aerobic exercise seems to be very efficient in eliciting a plethora of beneficial effects on the organism.⁷⁴ Indeed, a major advantage of endurance exercise are the metabolic and biochemical adaptations specifically linked to this type of exercise, for instance the stimulation of the muscle endocrine activity.⁷⁵⁻⁷⁸ The combination of various modes of exercise may benefit elderly individuals, as this

provides variety and allows the exercise practitioner to periodize the training program in order to improve the physiological factors associated with sarcopenia.

Aging is typically associated with burden of late-life diseases.⁷⁹ In many cases, such as cachexia,^{2,80} sarcopenia is associated with a primary disease and is possibly exacerbated by aging in a synergistic manner. By using exercise, in principle, it is possible to target the morbidity deriving from the primary disease and the muscle wasting occurring at the same time.^{77,81,82} However, it is not known whether exercise promotes reinnervation in pathological conditions but still is characterized by NMJ loss in age-related sarcopenia, as it is seen in cancer cachexia.^{15,83,84}

Conclusions

Sarcopenia is often associated to aging, likely due to undergoing mild inflammation,⁸⁵ but also to more subtle phenomena, such as selective denervation of the fast fibers, ultimately accounting for pronounced muscle fiber atrophy, functional deficit and the acquisition of a slow phenotype of the musculature in elderly people. Since the publication of the seminal work by Mosole et al. republished in this issue of European Journal of Translational Myology it is evident that physical activity promotes muscle reinnervation, thus preserving not only muscle mass but also the capability of the muscle to contract. Therefore, both endurance and resistance training are recommended throughout life or at least for elderly individuals, including patients suffering from various pathologies. The recommendations for exercise should be a multimodal approach as to maximize the benefits from the physical activity.⁸⁶

List of acronyms

CMAP - Compound Muscle Action Potentials

MyHC - Myosin Heavy Chain

N-CAM - Neuronal-Cell Adhesion Molecule

SDH - Succinate Dehydrogenase

Contributions of Author

DC conceived the original idea of the manuscript; CC, GFA, SK, and DC participated to the writing of the text..

Acknowledgments

None.

Funding

DC is funded by Emergence SiRIC - CURAMUS 2020 Sorbonne Université and Fondi di Ateneo 2019 Sapienza University of Rome.

Conflict of Interest

The authors declare no conflict of interest s.

Ethical Publication Statement

I confirm that I have read the Journal's position on ethical publication issues and affirms that this report is consistent with those guidelines.

Corresponding Author

Prof. Dario Coletti, SAIMLAL Dept.- Section of Histology, 16 Via Scarpa, 00161 Roma, Italy
Tel. +39 0649766756, fax +39 06 44628 54,
ORCID ID: 0000-0001-7373-1953
Email: dario.coletti@uniroma1.it

Emails and ORCIDiD of Coauthors

Claudia Coletti: ccolett@calstatela.edu

ORCID iD: 000-0002-2960-3046

Gilberto F Acosta: gacost14@calstatela.edu

ORCID iD: 0000-0003-2783-1122

Stefan Keslacy: skeslac@calstatela.edu

ORCID iD: 0000-0002-5225-3590

References

1. Cruz-Jentoft AJ, Baeyens JP, Bauer JM, Boirie Y, Cederholm T, Landi F, Martin FC, Michel JP, Rolland Y, Schneider SM, Topinková E, Vandewoude M, Zamboni M; European Working Group on Sarcopenia in Older People. Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. *Age Ageing*. 2010 Jul;39(4):412-23. doi: 10.1093/ageing/afq034. Epub 2010 Apr 13.
2. Berardi E, Madaro L, Lozanoska-Ochser B, Adamo S, Thorrez L, Bouche M, Coletti D. A Pound of Flesh: What Cachexia Is and What It Is Not. *Diagnostics (Basel)*. 2021 Jan 12;11(1):116. doi: 10.3390/diagnostics11010116.
3. Larsson L, Degens H, Li M, Salviati L, Lee YI, Thompson W, Kirkland JL, Sandri M. Sarcopenia: Aging-Related Loss of Muscle Mass and Function. *Physiol Rev*. 2019 Jan 1;99(1):427-511. doi: 10.1152/physrev.00061.2017.
4. Dodds RM, Roberts HC, Cooper C, Sayer AA. The Epidemiology of Sarcopenia. *J Clin Densitom*. 2015 Oct-Dec;18(4):461-6. doi: 10.1016/j.jocd.2015.04.012. Epub 2015 Jun 12.
5. Mosole S, Rossini K, Kern H, Löffler S, Simone Fruhmann H, Vogelauer M, Burggraf S, Grim-Stieger M, Cvečka J, Hamar D, Sedliak M, Šarabon N, Pond A, Biral D, Carraro U, Zampieri S. Reinnervation of Vastus lateralis is increased significantly in seniors (70-years old) with a lifelong history of high-level exercise. *Eur J Transl Myol Basic Appl Myol*. 2013;23 (4):205–10.
6. Power GA, Dalton BH, Gilmore KJ, Allen MD, Doherty TJ, Rice CL. Maintaining Motor Units into Old Age: Running the Final Common Pathway. *Eur J Transl Myol*. 2017 Mar 24;27(1):6597. doi: 10.4081/ejtm.2017.6597.
7. Zampieri S, Mosole S, Löffler S, Fruhmann H, Burggraf S, Cvečka J, Hamar D, Sedliak M, Tirtakova V, Šarabon N, Mayr W, Kern H. Physical Exercise in Aging: Nine Weeks of Leg Press or Electrical Stimulation Training in 70 Years

Exercise-mediated reinnervation of skeletal muscle in elderly

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

- Old Sedentary Elderly People. *Eur J Transl Myol.* 2015 Aug 25;25(4):237-42. doi: 10.4081/ejtm.2015.5374.
8. Mosole S, Carraro U, Kern H, Loeffler S, Zampieri S. Use it or Lose It: Tonic Activity of Slow Motoneurons Promotes Their Survival and Preferentially Increases Slow Fiber-Type Groupings in Muscles of Old Lifelong Recreational Sportsmen. *Eur J Transl Myol.* 2016 Nov 25;26(4):5972. doi: 10.4081/ejtm.2016.5972. eCollection 2016 Sep 15.
 9. Galloza J, Castillo B, Micheo W. Benefits of Exercise in the Older Population. *Phys Med Rehabil Clin N Am.* 2017 Nov;28(4):659-669. doi: 10.1016/j.pmr.2017.06.001.
 10. Schiaffino S, Reggiani C. Fiber types in mammalian skeletal muscles. *Physiol Rev.* 2011 Oct;91(4):1447-531. doi: 10.1152/physrev.00031.2010.
 11. Larsson L, Edström L, Lindegren B, Gorza L, Schiaffino S. MHC composition and enzyme-histochemical and physiological properties of a novel fast-twitch motor unit type. *Am J Physiol.* 1991 Jul;261(1 Pt 1):C93-101. doi: 10.1152/ajpcell.1991.261.1.C93.
 12. Esser K, Gunning P, Hardeman E. Nerve-dependent and -independent patterns of mRNA expression in regenerating skeletal muscle. *Dev Biol.* 1993 Sep;159(1):173-83. doi: 10.1006/dbio.1993.1231.
 13. Coletti D, Daou N, Hassani M, Li Z, Parlakian A. Serum Response Factor in Muscle Tissues: From Development to Ageing. *Eur J Transl Myol.* 2016 Jun 22;26(2):6008. doi: 10.4081/ejtm.2016.6008.
 14. Coletti D, Teodori L, Lin Z, Beranudin JF, Adamo S. Restoration versus reconstruction: cellular mechanisms of skin, nerve and muscle regeneration compared. *Regen Med Res.* 2013 Oct 1;1(1):4. doi: 10.1186/2050-490X-1-4.
 15. Daou N, Hassani M, Matos E, De Castro GS, Costa RGF, Seelaender M, Moresi V, Rocchi M, Adamo S, Li Z, Agbulut O, Coletti D. Displaced Myonuclei in Cancer Cachexia Suggest Altered Innervation. *Int J Mol Sci.* 2020 Feb 6;21(3):1092. doi: 10.3390/ijms21031092.
 16. Grifone R, Saquet A, Desgres M, Sangiorgi C, Gargano C, Li Z, Coletti D, Shi DL. Rbm24 displays dynamic functions required for myogenic differentiation during muscle regeneration. *Sci Rep.* 2021 May 3;11(1):9423. doi: 10.1038/s41598-021-88563-3.
 17. Mazzotti AL, Coletti D. The Need for a Consensus on the Locution "Central Nuclei" in Striated Muscle Myopathies. *Front Physiol.* 2016 Nov 23;7:577. doi: 10.3389/fphys.2016.00577.
 18. Blaauw B, Schiaffino S, Reggiani C. Mechanisms modulating skeletal muscle phenotype. *Compr Physiol.* 2013 Oct;3(4):1645-87. doi: 10.1002/cphy.c130009.
 19. Punkt K, Krug H, Huse J, Punkt J. Age-dependent changes of enzyme activities in the different fibre types of rat extensor digitorum longus and gastrocnemius muscles. *Acta Histochem.* 1993 Sep;95(1):97-110. doi: 10.1016/S0065-1281(11)80395-8.
 20. Marcucci L, Reggiani C. Increase of resting muscle stiffness, a less considered component of age-related skeletal muscle impairment. *Eur J Transl Myol.* 2020 Jun 17;30(2):8982. doi: 10.4081/ejtm.2019.8982.
 21. Wilkinson DJ, Piasecki M, Atherton PJ. The age-related loss of skeletal muscle mass and function: Measurement and physiology of muscle fibre atrophy and muscle fibre loss in humans. *Ageing Res Rev.* 2018 Nov;47:123-132. doi: 10.1016/j.arr.2018.07.005. Epub 2018 Jul 23.
 22. Klein CS, Marsh GD, Petrella RJ, Rice CL. Muscle fiber number in the biceps brachii muscle of young and old men. *Muscle Nerve.* 2003 Jul;28(1):62-8. doi: 10.1002/mus.10386.
 23. Nilwik R, Snijders T, Leenders M, Groen BB, van Kranenburg J, Verdijk LB, van Loon LJ. The decline in skeletal muscle mass with aging is mainly attributed to a reduction in type II muscle fiber size. *Exp Gerontol.* 2013 May;48(5):492-8. doi: 10.1016/j.exger.2013.02.012. Epub 2013 Feb 17.
 24. Messa GAM, Piasecki M, Rittweger J, McPhee JS, Koltai E, Radak Z, Simunic B, Heinonen A, Suominen H, Korhonen MT, Degens H. Absence of an aging-related increase in fiber type grouping in athletes and non-athletes. *Scand J Med Sci Sports.* 2020 Nov;30(11):2057-2069. doi: 10.1111/sms.13778. Epub 2020 Aug 28.
 25. Kelly NA, Hammond KG, Stec MJ, Bickel CS, Windham ST, Tuggle SC, Bamman MM. Quantification and characterization of grouped type I myofibers in human aging. *Muscle Nerve.* 2018 Jan;57(1):E52-E59. doi: 10.1002/mus.25711. Epub 2017 Sep 7.
 26. Jerkovic R, Argentini C, Serrano-Sanchez A, Cordonnier C, Schiaffino S. Early myosin switching induced by nerve activity in regenerating slow skeletal muscle. *Cell Struct Funct.* 1997 Feb;22(1):147-53. doi: 10.1247/csf.22.147.
 27. Murgia M, Serrano AL, Calabria E, Pallafacchina G, Lomo T, Schiaffino S. Ras is involved in nerve-activity-dependent regulation of muscle genes. *Nat Cell Biol.* 2000 Mar;2(3):142-7. doi: 10.1038/35004013.
 28. Larsson L, Ansved T. Effects of ageing on the motor unit. *Prog Neurobiol.* 1995 Apr;45(5):397-458. doi: 10.1016/0301-0082(95)98601-z.
 29. Piasecki M, Ireland A, Jones DA, McPhee JS. Age-dependent motor unit remodelling in human limb muscles. *Biogerontology.* 2016 Jun;17(3):485-96. doi: 10.1007/s10522-015-9627-3. Epub 2015 Dec 14.

Exercise-mediated reinnervation of skeletal muscle in elderly

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

30. Demontis F, Piccirillo R, Goldberg AL, Perrimon N. Mechanisms of skeletal muscle aging: insights from *Drosophila* and mammalian models. *Dis Model Mech*. 2013 Nov;6(6):1339-52. doi: 10.1242/dmm.012559. Epub 2013 Oct 2.
31. Naro F, De Arcangelis V, Coletti D, Molinaro M, Zani B, Vassanelli S, Reggiani C, Teti A, Adamo S. Increase in cytosolic Ca²⁺ induced by elevation of extracellular Ca²⁺ in skeletal myogenic cells. *Am J Physiol Cell Physiol*. 2003 Apr;284(4):C969-76. doi: 10.1152/ajpcell.00237.2002. Epub 2002 Dec 18.
32. De Arcangelis V, Coletti D, Canato M, Molinaro M, Adamo S, Reggiani C, Naro F. Hypertrophy and transcriptional regulation induced in myogenic cell line L6-C5 by an increase of extracellular calcium. *J Cell Physiol*. 2005 Mar;202(3):787-95. doi: 10.1002/jcp.20174.
33. Lamboley CR, Wyckelsma VL, McKenna MJ, Murphy RM, Lamb GD. Ca(2+) leakage out of the sarcoplasmic reticulum is increased in type I skeletal muscle fibres in aged humans. *J Physiol*. 2016 Jan 15;594(2):469-81. doi: 10.1113/JP271382. Epub 2015 Dec 14.
34. Westerterp KR. Daily physical activity and ageing. *Curr Opin Clin Nutr Metab Care*. 2000 Nov;3(6):485-8. doi: 10.1097/00075197-200011000-00011.
35. Meijer EP, Westerterp KR, Verstappen FT. Effect of exercise training on physical activity and substrate utilization in the elderly. *Int J Sports Med*. 2000 Oct;21(7):499-504. doi: 10.1055/s-2000-7419.
36. Mosole S, Carraro U, Kern H, Loeffler S, Fruhmann H, Vogelauer M, Burggraf S, Mayr W, Krenn M, Paternostro-Sluga T, Hamar D, Cvecka J, Sedliak M, Tirpakova V, Sarabon N, Musarò A, Sandri M, Protasi F, Nori A, Pond A, Zampieri S. Long-term high-level exercise promotes muscle reinnervation with age. *J Neuropathol Exp Neurol*. 2014 Apr;73(4):284-94. doi: 10.1097/NEN.0000000000000032.
37. Drey M, Sieber CC, Degens H, McPhee J, Korhonen MT, Müller K, Ganse B, Rittweger J. Relation between muscle mass, motor units and type of training in master athletes. *Clin Physiol Funct Imaging*. 2016 Jan;36(1):70-6. doi: 10.1111/cpf.12195. Epub 2014 Oct 24.
38. Bechshøft RL, Malmgaard-Clausen NM, Gliese B, Beyer N, Mackey AL, Andersen JL, Kjær M, Holm L. Improved skeletal muscle mass and strength after heavy strength training in very old individuals. *Exp Gerontol*. 2017 Jun;92:96-105. doi: 10.1016/j.exger.2017.03.014. Epub 2017 Mar 28.
39. Batsis JA, Villareal DT. Sarcopenic obesity in older adults: aetiology, epidemiology and treatment strategies. *Nat Rev Endocrinol*. 2018 Sep;14(9):513-537. doi: 10.1038/s41574-018-0062-9.
40. Messi ML, Li T, Wang ZM, Marsh AP, Nicklas B, Delbono O. Resistance Training Enhances Skeletal Muscle Innervation Without Modifying the Number of Satellite Cells or their Myofiber Association in Obese Older Adults. *J Gerontol A Biol Sci Med Sci*. 2016 Oct;71(10):1273-80. doi: 10.1093/gerona/glv176. Epub 2015 Oct 7.
41. Kletzien H, Russell JA, Connor NP. The effects of treadmill running on aging laryngeal muscle structure. *Laryngoscope*. 2016 Mar;126(3):672-7. doi: 10.1002/lary.25520. Epub 2015 Aug 8.
42. Bathgate KE, Bagley JR, Jo E, Talmadge RJ, Tobias IS, Brown LE, Coburn JW, Arevalo JA, Segal NL, Galpin AJ. Muscle health and performance in monozygotic twins with 30 years of discordant exercise habits. *Eur J Appl Physiol*. 2018 Oct;118(10):2097-2110. doi: 10.1007/s00421-018-3943-7. Epub 2018 Jul 14.
43. Carraro U, Kern H, Gava P, Hofer C, Loeffler S, Gargiulo P, Mosole S, Zampieri S, Gobbo V, Ravara B, Piccione F, Marcante A, Baba A, Schils S, Pond A, Gava F. Biology of Muscle Atrophy and of its Recovery by FES in Aging and Mobility Impairments: Roots and By-Products. *Eur J Transl Myol*. 2015 Aug 25;25(4):221-30. doi: 10.4081/ejtm.2015.5272.
44. Carraro U, Kern H, Gava P, Hofer C, Loeffler S, Gargiulo P, Edmunds K, Árnadóttir ÍD, Zampieri S, Ravara B, Gava F, Nori A, Gobbo V, Masiero S, Marcante A, Baba A, Piccione F, Schils S, Pond A, Mosole S. Recovery from muscle weakness by exercise and FES: lessons from Masters, active or sedentary seniors and SCI patients. *Aging Clin Exp Res*. 2017 Aug;29(4):579-590. doi: 10.1007/s40520-016-0619-1. Epub 2016 Sep 3.
45. Carraro U, Coletti D, Kern H. The Ejtm Specials "The Long-Term Denervated Muscle". *Eur J Transl Myol*. 2014 Mar 27;24(1):3292. doi: 10.4081/ejtm.2014.3292.
46. Kern H, Hofer C, Loeffler S, Zampieri S, Gargiulo P, Baba A, Marcante A, Piccione F, Pond A, Carraro U. Atrophy, ultra-structural disorders, severe atrophy and degeneration of denervated human muscle in SCI and Aging. Implications for their recovery by Functional Electrical Stimulation, updated 2017. *Neurol Res*. 2017 Jul;39(7):660-666. doi: 10.1080/01616412.2017.1314906. Epub 2017 Apr 13.
47. Kern H, Hofer C, Loeffler S, Zampieri S, Gargiulo P, Baba A, Marcante A, Piccione F, Pond A, Carraro U. Atrophy, ultra-structural disorders, severe atrophy and degeneration of denervated human muscle in SCI and Aging. Implications for their recovery by Functional Electrical Stimulation, updated 2017. *Neurol Res*. 2017 Jul;39(7):660-666. doi: 10.1080/01616412.2017.1314906. Epub 2017 Apr 13.
48. Wang P, Li Y, Zhang Z, Lin Y, Jiang Z, Ding X,

Exercise-mediated reinnervation of skeletal muscle in elderly

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

- Yang L. Effects of functional electrical stimulation on neuromuscular function after targeted muscle reinnervation surgery in rats. *Annu Int Conf IEEE Eng Med Biol Soc.* 2020 Jul;2020:3823-3826. doi: 10.1109/EMBC44109.2020.9175836.
49. Mense S. Muscle Nociceptors, Neurochemistry. In: Gebhart GF, Schmidt RF, editors. *Encyclopedia of Pain.* Berlin, Heidelberg: Springer Berlin Heidelberg; 2013. p. 1944–50. doi:10.1007/978-3-642-28753-4_2529.
50. Giuriati W, Ravara B, Porzionato A, Albertin G, Stecco C, Macchi V, De Caro R, Martinello T, Gomiero C, Patruno M, Coletti D, Zampieri S, Nori A. Muscle spindles of the rat sternomastoid muscle. *Eur J Transl Myol.* 2018 Dec 13;28(4):7904. doi: 10.4081/ejtm.2018.7904.
51. Ravara B, Gobbo V, Incendi D, Porzionato A, Macchi V, Caro R, Coletti D, Martinello T, Patruno M. Revisiting the peculiar regional distribution of muscle fiber types in rat Sternomastoid Muscle. *Eur J Transl Myol.* 2018 Mar 1;28(1):7302. doi: 10.4081/ejtm.2018.7302.
52. Konrad HR, Girardi M, Helfert R. Balance and aging. *Laryngoscope.* 1999 Sep;109(9):1454-60. doi: 10.1097/00005537-199909000-00019.
53. Piitulainen H, Seipäjärvi S, Avela J, Parviainen T, Walker S. Cortical Proprioceptive Processing Is Altered by Aging. *Front Aging Neurosci.* 2018 Jun 14;10:147. doi: 10.3389/fnagi.2018.00147.
54. Henry M, Baudry S. Age-related changes in leg proprioception: implications for postural control. *J Neurophysiol.* 2019 Aug 1;122(2):525-538. doi: 10.1152/jn.00067.2019. Epub 2019 Jun 5.
55. Rosant C, Nagel MD, Pérot C. Aging affects passive stiffness and spindle function of the rat soleus muscle. *Exp Gerontol.* 2007 Apr;42(4):301-8. doi: 10.1016/j.exger.2006.10.007. Epub 2006 Nov 21.
56. Desaki J, Nishida N. A further observation of muscle spindles in the extensor digitorum longus muscle of the aged rat. *J Electron Microsc (Tokyo).* 2010;59(1):79-86. doi: 10.1093/jmicro/dfp038. Epub 2009 Jul 31. PMID: 19648233.
57. Chartier SR, Mitchell SAT, Majuta LA, Mantyh PW. The Changing Sensory and Sympathetic Innervation of the Young, Adult and Aging Mouse Femur. *Neuroscience.* 2018 Sep 1;387:178-190. doi: 10.1016/j.neuroscience.2018.01.047. Epub 2018 Feb 10.
58. Vaughan SK, Stanley OL, Valdez G. Impact of Aging on Proprioceptive Sensory Neurons and Intrafusal Muscle Fibers in Mice. *J Gerontol A Biol Sci Med Sci.* 2017 Jun 1;72(6):771-779. doi: 10.1093/gerona/glw175.
59. Miwa T, Miwa Y, Kanda K. Dynamic and static sensitivities of muscle spindle primary endings in aged rats to ramp stretch. *Neurosci Lett.* 1995 Dec 8;201(2):179-82. doi: 10.1016/0304-3940(95)12165-x.
60. Kim GH, Suzuki S, Kanda K. Age-related physiological and morphological changes of muscle spindles in rats. *J Physiol.* 2007 Jul 15;582(Pt 2):525-38. doi: 10.1113/jphysiol.2007.130120. Epub 2007 May 10.
61. Fortier S, Basset FA. The effects of exercise on limb proprioceptive signals. *J Electromyogr Kinesiol.* 2012 Dec;22(6):795-802. doi: 10.1016/j.jelekin.2012.04.001. Epub 2012 May 11.
62. Nascimento CM, Ingles M, Salvador-Pascual A, Cominetti MR, Gomez-Cabrera MC, Viña J. Sarcopenia, frailty and their prevention by exercise. *Free Radic Biol Med.* 2019 Feb 20;132:42-49. doi: 10.1016/j.freeradbiomed.2018.08.035. Epub 2018 Aug 31.
63. Wilke J, Mohr L, Tenforde AS, Edouard P, Fossati C, González-Gross M, Sánchez Ramírez C, Laiño F, Tan B, Pillay JD, Pigozzi F, Jimenez-Pavon D, Novak B, Jaunig J, Zhang M, van Poppel M, Heidt C, Willwacher S, Yuki G, Lieberman DE, Vogt L, Verhagen E, Hespanhol L, Hollander K. A Pandemic within the Pandemic? Physical Activity Levels Substantially Decreased in Countries Affected by COVID-19. *Int J Environ Res Public Health.* 2021 Feb 24;18(5):2235. doi: 10.3390/ijerph18052235.
64. Denison HJ, Cooper C, Sayer AA, Robinson SM. Prevention and optimal management of sarcopenia: a review of combined exercise and nutrition interventions to improve muscle outcomes in older people. *Clin Interv Aging.* 2015 May 11;10:859-69. doi: 10.2147/CIA.S55842.
65. Phu S, Boersma D, Duque G. Exercise and Sarcopenia. *J Clin Densitom.* 2015 Oct-Dec;18(4):488-92. doi: 10.1016/j.jocd.2015.04.011. Epub 2015 Jun 10.
66. Papadopoulou SK. Sarcopenia: A Contemporary Health Problem among Older Adult Populations. *Nutrients.* 2020 May 1;12(5):1293. doi: 10.3390/nu12051293.
67. Giallauria F, Cittadini A, Smart NA, Vigorito C. Resistance training and sarcopenia. *Monaldi Arch Chest Dis.* 2016 Jun 22;84(1-2):738. doi: 10.4081/monaldi.2015.738.
68. Lichtenberg T, von Stengel S, Sieber C, Kemmler W. The Favorable Effects of a High-Intensity Resistance Training on Sarcopenia in Older Community-Dwelling Men with Osteosarcopenia: The Randomized Controlled FrOST Study. *Clin Interv Aging.* 2019 Dec 16;14:2173-2186. doi: 10.2147/CIA.S225618.
69. Freiberger E, Sieber C, Pfeifer K. Physical activity, exercise, and sarcopenia - future challenges. *Wien Med Wochenschr.* 2011 Sep;161(17-18):416-25. doi: 10.1007/s10354-011-0001-z. Epub 2011 Jul 29
70. Carraro U. Thirty years of translational research in Mobility Medicine: Collection of abstracts of the

Exercise-mediated reinnervation of skeletal muscle in elderly

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

- 2020 Padua Muscle Days. Eur J Transl Myol. 2020 Apr 1;30(1):8826. doi: 10.4081/ejtm.2019.8826.
71. Furst T, Massaro A, Miller C, Williams BT, LaMacchia ZM, Horvath PJ. β -Alanine supplementation increased physical performance and improved executive function following endurance exercise in middle aged individuals. J Int Soc Sports Nutr. 2018 Jul 11;15(1):32. doi: 10.1186/s12970-018-0238-7.
 72. Moreillon M, Conde Alonso S, Broskey NT, Greggio C, Besson C, Rousson V, Amati F. Hybrid fiber alterations in exercising seniors suggest contribution to fast-to-slow muscle fiber shift. J Cachexia Sarcopenia Muscle. 2019 Jun;10(3):687-695. doi: 10.1002/jcsm.12410. Epub 2019 Mar 25.
 73. Garcia M, Seelaender M, Sotiropoulos A, Coletti D, Lancha AH Jr. Vitamin D, muscle recovery, sarcopenia, cachexia, and muscle atrophy. Nutrition. 2019 Apr;60:66-69. doi: 10.1016/j.nut.2018.09.031. Epub 2018 Oct 7.
 74. Barbieri E, Agostini D, Polidori E, Potenza L, Guescini M, Lucertini F, Annibali G, Stocchi L, De Santi M, Stocchi V. The pleiotropic effect of physical exercise on mitochondrial dynamics in aging skeletal muscle. Oxid Med Cell Longev. 2015;2015:917085. doi: 10.1155/2015/917085. Epub 2015 Apr 5
 75. Di Felice V, Coletti D, Seelaender M. Editorial: Myokines, Adipokines, Cytokines in Muscle Pathophysiology. Front Physiol. 2020 Oct 23;11:592856. doi: 10.3389/fphys.2020.592856.
 76. Coletti D, Aulino P, Pigna E, Barteri F, Moresi V, Annibali D, Adamo S, Berardi E. Spontaneous Physical Activity Downregulates Pax7 in Cancer Cachexia. Stem Cells Int. 2016; 2016: 6729268. Published online 2015 Dec 20. doi: 10.1155/2016/6729268.
 77. Alves de Lima E Jr, Teixeira AAS, Biondo LA, Diniz TA, Silveira LS, Coletti D, Busquets Rius S, Rosa Neto JC. Exercise Reduces the Resumption of Tumor Growth and Proteolytic Pathways in the Skeletal Muscle of Mice Following Chemotherapy. Cancers (Basel). 2020 Nov 20;12(11):3466. doi: 10.3390/cancers12113466.
 78. Consitt LA, Dudley C, Saxena G. Impact of Endurance and Resistance Training on Skeletal Muscle Glucose Metabolism in Older Adults. Nutrients. 2019 Nov 3;11(11):2636. doi: 10.3390/nu11112636.
 79. Partridge L, Deelen J, Slagboom PE. Facing up to the global challenges of ageing. Nature. 2018 Sep;561(7721):45-56. doi: 10.1038/s41586-018-0457-8. Epub 2018 Sep 5.
 80. de Castro GS, Simoes E, Lima JDCC, Ortiz-Silva M, Festuccia WT, Tokeshi F, Alcântara PS, Otoch JP, Coletti D, Seelaender M. Human Cachexia Induces Changes in Mitochondria, Autophagy and Apoptosis in the Skeletal Muscle. Cancers (Basel). 2019 Aug 28;11(9):1264. doi: 10.3390/cancers11091264.
 81. Pigna E, Berardi E, Aulino P, Rizzuto E, Zampieri S, Carraro U, Kern H, Merigliano S, Gruppo M, Mericskay M, Li Z, Rocchi M, Barone R, Macaluso F, Di Felice V, Adamo S, Coletti D, Moresi V. Aerobic Exercise and Pharmacological Treatments Counteract Cachexia by Modulating Autophagy in Colon Cancer. Sci Rep. 2016 May 31;6:26991. doi: 10.1038/srep26991.
 82. Brunjes DL, Kennel PJ, Christian Schulze P. Exercise capacity, physical activity, and morbidity. Heart Fail Rev. 2017 Mar;22(2):133-139. doi: 10.1007/s10741-016-9592-1.
 83. Sartori R, Hagg A, Zampieri S, Armani A, Winbanks CE, Viana LR, Haidar M, Watt KI, Qian H, Pezzini C, Zanganeh P, Turner BJ, Larsson A, Zanchettin G, Pierobon ES, Moletta L, Valmasoni M, Ponzoni A, Attar S, Da Dalt G, Sperti C, Kustermann M, Thomson RE, Larsson L, Loveland KL, Costelli P, Megighian A, Merigliano S, Penna F, Gregorevic P, Sandri M. Perturbed BMP signaling and denervation promote muscle wasting in cancer cachexia. Sci Transl Med. 2021 Aug 4;13(605):eaay9592. doi: 10.1126/scitranslmed.aay9592.
 84. Huot JR, Pin F, Bonetto A. Muscle weakness caused by cancer and chemotherapy is associated with loss of motor unit connectivity. Am J Cancer Res. 2021 Jun 15;11(6):2990-3001
 85. Bouché M, Muñoz-Cánoves P, Rossi F, Coletti D. Inflammation in muscle repair, aging, and myopathies. Biomed Res Int. 2014;2014:821950. doi: 10.1155/2014/821950. Epub 2014 Aug 4.
 86. Beckwée D, Delaere A, Aelbrecht S, Baert V, Beaudart C, Bruyere O, de Saint-Hubert M, Bautmans I. Exercise Interventions for the Prevention and Treatment of Sarcopenia. A Systematic Umbrella Review. J Nutr Health Aging. 2019;23(6):494-502. doi: 10.1007/s12603-019-1196-8.
 87. Racca AW, Beck AE, Rao VS, Flint GV, Lundy SD, Born DE, Bamshad MJ, Regnier M. Contractility and kinetics of human fetal and human adult skeletal muscle. J Physiol. 2013 Jun 15;591(12):3049-61. doi: 10.1113/jphysiol.2013.252650. Epub 2013 Apr 29.
 88. Wall CE, Holmes M, Soderblom EJ, Taylor AB. Proteomics and immunohistochemistry identify the expression of α -cardiac myosin heavy chain in the jaw-closing muscles of sooty mangabeys (order Primates). Arch Oral Biol. 2018 Jul;91:103-108. doi: 10.1016/j.archoralbio.2018.01.019. Epub 2018 Feb 3.
 89. Briggs MM, Schachat F. The superfast extraocular myosin (MYH13) is localized to the innervation zone in both the global and orbital layers of rabbit extraocular muscle. J Exp Biol. 2002 Oct;205(Pt

Exercise-mediated reinnervation of skeletal muscle in elderly

Eur J Transl Myol 32 (1): 10416, 2022 doi: 10.4081/ejtm.2022.10416

- 20):3133-42. doi: 10.1242/jeb.205.20.3133.
90. Rossi AC, Mammucari C, Argentini C, Reggiani C, Schiaffino S. Two novel/ancient myosins in mammalian skeletal muscles: MYH14/7b and MYH15 are expressed in extraocular muscles and muscle spindles. *J Physiol.* 2010 Jan 15;588(Pt 2):353-64. doi: 10.1113/jphysiol.2009.181008. Epub 2009 Nov 30.
91. Mosole S, Rossini K, Kern H, Löfler S, Simone Fruhmann H, Vogelauer M, Burggraf S, Grim-

Stieger M, Cvečka J, Hamar D, Sedliak M, Šarabon N, Pond A, Biral D, Carraro U, Zampieri S. Reinnervation of Vastus lateralis is increased significantly in seniors (70-years old) with a lifelong history of high-level exercise (2013, revisited here in 2022). *Eur J Transl Myol.* 32 (1): 10420, 2022. doi: 10.4081/ejtm.2022.10420.

Submitted: February 17, 2022

Accepted for publication: February 17, 2022