DOI: 10.1111/ina.12949

REVIEW

WILEY

Occupational scenarios and exposure assessment to formaldehyde: A systematic review

Vittoria Cammalleri¹ | Roberta Noemi Pocino¹ | Daniela Marotta¹ | Carmela Protano¹ | Federica Sinibaldi² | Stefano Simonazzi² | Marta Petyx³ | Sergio Iavicoli³ | Matteo Vitali¹

¹Department of Public Health and Infectious Diseases, University of Rome "La Sapienza", Rome, Italy

²Department of Anatomical Histological Medical Legal Sciences and Locomotor Apparatus, University of Rome "La Sapienza", Rome, Italy

³Department of Occupational and Environmental Medicine, Epidemiology and Hygiene, INAIL Research, Rome, Italy

Correspondence

Matteo Vitali, Department of Public Health and Infectious Diseases, University of Rome "La Sapienza", Piazzale Aldo Moro 5, 00185, Rome, Italy. Email: matteo.vitali@uniroma1.it

Funding information

Istituto Nazionale per l'Assicurazione Contro Gli Infortuni sul Lavoro, Grant/ Award Number: B86C19000070001

Abstract

Revised: 7 October 2021

The objectives of the systematic review were to: identify the work sectors at risk for exposure to formaldehyde; investigate the procedures applied to assess occupational exposure; evaluate the reported exposure levels among the different settings. An electronic search of Pubmed, Scopus, Web of Science and ToxNet was carried out for collecting all the articles on the investigated issue published from January 1, 2004 to September 30, 2019. Forty-three papers were included in the review, and evidenced a great number of occupational scenarios at risk for formaldehyde exposure. All the included studies collected data on formaldehyde exposure levels by a similar approach: environmental and personal sampling followed by chromatographic analyses. Results ranged from not detectable values until to some mg m^{-3} of airborne formaldehyde. The riskiest occupational settings for formaldehyde exposure were the gross anatomy and pathology laboratories, the hairdressing salons and some specific productive settings, such as wooden furniture factories, dairy facilities and fish hatcheries. Notice that formaldehyde, a well-known carcinogen, was recovered in air at levels higher than outdoor in almost all the studied scenarios/activities; thus, when formaldehyde cannot be removed or substituted, targeted strategies for exposure elimination or mitigation must be adopted.

KEYWORDS

analytical approach, exposure assessment, exposure levels, formaldehyde, occupational settings

1 | INTRODUCTION

Formaldehyde is an organic compound that, at room temperature and standard atmospheric pressure, occurs in the form of a colorless, pungent and irritating gas, extremely volatile and highly soluble in water.¹ It is present as a natural product in many living systems, in the environment, in some foods and in the organism of mammals, including humans, as a product of oxidative metabolism.²

Although formaldehyde is naturally present in the troposphere, due to its formation during the oxidation of hydrocarbons,³ the main sources determining human exposure are anthropogenic. Among these, some are present in indoor environments such as

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited. © 2021 The Authors. *Indoor Air* published by John Wiley & Sons Ltd wiley

products containing and releasing formaldehyde (insulating materials, resins, glues, chipboard, plywood, fabrics, etc),⁴ while others are related to activities involving combustion processes, tobacco and e-cigarettes active and passive smoking and cooking (especially frying).^{3,5} Formaldehyde is a well-known occupational carcinogen and a recognized sensory irritant compound, especially for sensitive individuals,^{6,7} present in many different working scenarios.⁸ Indeed, formaldehyde is widely used in numerous production processes and sanitary applications due to its chemical-physical characteristics and broad spectrum microbicide activity.^{9,10} The International Agency for the Research on Cancer (IARC) has identified three main occupational scenarios where workers may be exposed to formaldehyde at air concentrations significantly higher than the indoor and outdoor background levels: (i) the production of formaldehyde and/or its solutions; (ii) the production of products containing formaldehyde or during their use and (iii) the combustion of products generating formaldehyde.⁸ Thus, workers in industrial production processes (resins, plastics, semi-finished wood products, furnishing accessories and textiles),^{11,12} professionals of gross anatomy and pathology laboratories, veterinarians, embalmers,^{9,10,13-15} breeders,¹⁶ carpenters, industrial launderers,¹⁷ firefighters, beauticians and printingrooms workers¹⁸⁻²⁰ are the categories at higher risk of exposure to formaldehyde.

In this regard, a robust scientific evidence has highlighted over the years several acute and chronic adverse health effects deriving from such exposure.²¹⁻²⁵ Moreover, after a revision of the scientific literature, IARC in 2004 has classified formaldehyde as group I carcinogen with sufficient evidence for nasopharyngeal carcinoma²⁶ and, afterward, also for leukaemia.⁸ Then, given the evidences, in 2011 the listing status of formaldehyde was changed also from "reasonably anticipated to be a human carcinogen" to "known to be a human carcinogen based on sufficient evidence of carcinogenicity" in the Twelfth Annual Report on Carcinogens of the National Toxicology Program (NTP).²⁷ More recently, the European Commission (EC) has reclassified formaldehyde to carcinogenic category 1B (may cause cancer by inhalation) and mutagen category 2 (suspected of causing genetic defects).²⁸ Following this reclassification, the EC Regulation No. 1272/2008²⁹ on classification, labelling and packaging of substances was amended, and the hazard classification of formaldehyde labelling was modified, as shown in Table 1.

The classification of formaldehyde as a carcinogen has led to the need to re-evaluate the risk management systems for potentially exposed workers, as implemented in the various occupational settings. In particular, the typical chemical risk assessment had to move toward a carcinogenic one. This implied the obligation to first evaluate the replacement of formaldehyde with other non-carcinogenic substances or, if not possible due to technical reasons (often due to cost-benefit constraints), to mitigate any exposure.

The need to carry out accurate occupational risk assessments for formaldehyde has therefore led to the improvement of sampling and analysis methods. In particular, occupational exposure is usually evaluated by active or passive sampling carried out in fixed positions (environmental sampling) and/or through personal samplers. As regards analysis methods, the airborne formaldehyde can currently be

Practical Implications

- The findings of this systematic review provide an overall picture of the worldwide occupational scenarios at potential exposure to formaldehyde and trace evidences for targeted prevention and mitigation actions.
- Personal samplings show higher levels of airborne formaldehyde than environmental ones. Thus, both monitoring modalities should be performed for an accurate risk assessment.

measured at ng m³ levels by sampling air with specific sorbent tubes containing 2,4-dinitrophenyhydrazine or 2-(hydroxymethyl)piperidine as derivatizer with a built-in ozone scrubber, and quantifying it by High Performance Liquid Chromatography Mass Spectrometry (HPLC-MS/MS) or Gas Chromatography Mass Spectrometry (GC-MS); alternatively, there are portable samplers/analyzers, equipped with photoacoustic spectroscopy detectors or electrochemical detectors, which have a sensitivity of the order of few μ g m³.

The aim of the present systematic review was to examine the scientific literature reporting experimental data on occupational exposure to formaldehyde from 2004 to the time of the review's conduction (until to September 30, 2019); 2004 was chosen as the first year of classification of formaldehyde as a carcinogen²⁴ and, consequently, the year from which the analytical methods used must surely be reliable and sensitive. In particular, specific objectives were to (i) identify the work sectors at risk of exposure to formaldehyde; (ii) investigate the managing procedures used to assess occupational exposure; (iii) evaluate the reported exposure levels among the different settings.

2 | MATERIALS AND METHODS

2.1 | Search strategy

This systematic review was performed according to the PRISMA statement. $^{\rm 30}$

Three investigators (V.C., D.M. and R.N.P.) searched published studies from January 1, 2004 to September 30, 2019 through the electronic databases MEDLINE via PubMed, SCOPUS and TOXNET. The search terms "(occupational OR workplace OR professional) AND exposure AND formaldehyde" were used. The results obtained by the three different researchers were merged by EndNote X9 software and then all duplicates were removed.

2.2 | Inclusion/Exclusion criteria

We included all the original studies, published in Italian or English in the fixed period, reporting experimental data obtained directly by the authors on occupational exposure to formaldehyde. In vitro and animal studies as well as all kind of reviews, reports, monographs,

TABLE 1 Classification of formaldehyde hazard statements

Hazard categories	Hazard statements
Carc. 1B	H350: may cause cancer by inhalation
Muta. 2	H341: suspected of causing genetic defects
Acute Tox. 3	H301: toxic if swallowed
Acute Tox. 3	H311: toxic in contact with skin
Acute Tox. 3	H331: toxic if inhaled
Skin Corr. 1B	H314: causes severe skin burn and eye damage
Skin Sens. 1	H317: may cause an allergic skin reaction

book chapters and conference acts were excluded. Then, the title and the abstract of the included articles were independently reviewed by three investigators (V.C., D.M. and R.N.P.). Articles which did not fall within the inclusion criteria were excluded during this phase.

In the following phase, the full text of the remaining potentially eligible papers was independently examined by the same three investigators for final decision on their inclusion or not in the review.

During this multi-step exclusion process, any disagreement in the decision on the examined studies was discussed until consensus was reached among the three investigators. All the process was supervised by other two different investigators (C.P. and M.V.).

2.3 | Study quality and evaluation

Study quality was independently assessed by three investigators (V.C., D.M. and R.N.P.) using the Joanna Briggs Institute (JBI) Critical Appraisal Checklist for Analytical Cross-Sectional Studies.³¹ This tool was used to evaluate the risk of bias through a checklist of eight questions, which include sample selection (two questions), exposure evaluation (two questions), confounding factors (two questions), outcomes and appropriate statistical analysis (two questions). The possible answers for each question were "yes," "no," "unclear" and "not applicable." According to a previously bias assessment that has been already described, ³² if the answer "yes" was \geq 50% of all questions, the evaluated paper was considered with low risk of bias; on the contrary, if the answer "no" was \geq 50%, the risk of bias was high. Finally, if the answer "unclear" was \geq 50%, the risk of bias was considered uncertain. Studies that presented high or uncertain risk of bias was were excluded from the qualitative synthesis of the present review.

All the studies included in the review were synthetized according to year of publication, country, occupational setting, exposure assessment methodology, and main results.

3 | RESULTS

3.1 | Study selection process

Figure 1 shows the flow chart summarizing the selection steps for the systematic review.

In total, we recovered 1581 studies from all searched databases (584 from PubMed, 889 from Scopus, 108 from ToxNet) and, after removing the 461 duplicates, 1120 articles remained. Out of the remaining 1120 papers, 1051 were excluded after review of their titles and abstracts. Thus, the full-text of 69 papers were searched and evaluated considering the inclusion/exclusion criteria and the quality assessment. After the evaluation of the full-text, 18 articles were excluded for the following reasons: 2 articles evaluated the occupational exposure to other chemicals, 2 did not assess occupational exposure, estimated the exposure to formaldehyde based on data reported in national/institutional databases, reported measured performed before the year 2004, 1 just reported data on biological monitoring, 1 was a proceeding.^{6,10–12,18,33–45} In Figure 2 are reported the results of risk of bias assessment for all the 51 articles, considering the percentage of the responses to each question of the checklist.

Finally, 8 articles were excluded because at high risk of bias.⁴⁶⁻⁵³

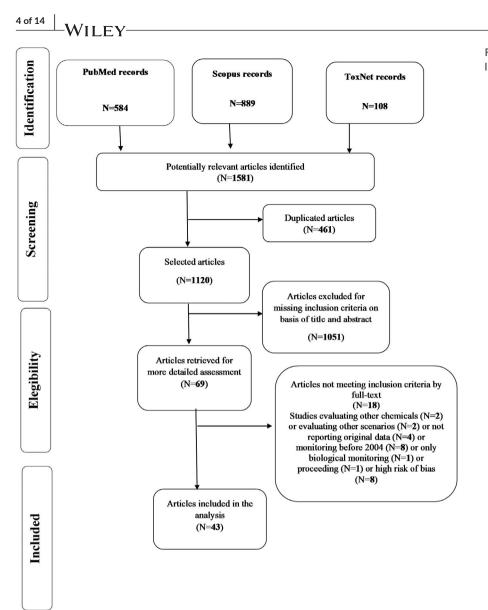
At the end of these steps, 43 articles, all with a low risk of bias, were included in the systematic review. In particular, all the papers described the results of cross-sectional studies performed in several countries worldwide: 12 in USA, 4 in Brazil, 3 in Australia, 3 in Japan, 3 in Taiwan, 3 in Thailand (1 conducted at the same time in Malaysia), 2 in Malaysia (1 conducted at the same time in Thailand), 2 in Portugal, 2 in Italy, 2 in China, 1 in Saudi Arabia, 1 in Iran, 1 in Greece, 1 in Spain, 1 in Egypt, 1 in Sweden and 1 in Korea, all summarized in Tables 2-5.

All the included articles were grouped according to the studied occupational scenarios as follows: healthcare and research (Table 2), esthetic and wellness (Table 3), industrial (Table 4), fire fighters' and other settings (Table 5).

3.2 | Main characteristics of the studies involving healthcare and research settings

In Table 2 are reported the main characteristics of the included studies performed in healthcare and research settings.

In total, 12 papers reported the results of studies performed in healthcare and research settings.^{13,15,54-63} Most of these studies $^{13,15,55-59,62}$ evaluated the occupational exposure to formal dehyde in gross anatomy laboratories (dissection room) and pathology or histology laboratories during a typical working day, with a variable sampling period according to the specific work activities. Other occupational healthcare and research scenarios included different indoor environments of university research institutes⁵⁴ or of learning institute⁶³ and hospital operating theaters.⁶⁰ Sampling and analyses were carried out in the greatest part of the studies by the use of active and/or passive environmental sampling and/or active and/or passive personal sampling with 2,4-DNPH cartridges and HPLC with UV/DAD or UV/VIS. The exceptions to these cases were the studies of Lakchayapakor et al.⁵⁶ and of Kwong et al.⁶² The first study evaluated formaldehyde exposure performing active environmental and personal sampling by the use of 2-hydroxymethyl piperidine



tive environmental monitoring with a formaldehyde meter monitor. The results of the included studied recovered exposure values

cartridges and GC-FID technique, while the second performed an ac-

ranging from not detectable levels to concentrations in the order of about 1–3 mg m⁻³, with the highest level recovered in the hospital pathology and histology laboratories (maximum value for passive personal monitoring equal to 2.70 mg m⁻³).⁶² Notice that personal sampling involved higher concentrations compared to workplace sampling in all cases.

3.3 | Main characteristics of the studies involving esthetic and wellness settings

Table 3 shows the main characteristics of the studies carried out in esthetic and wellness settings.

Eleven papers included in the present review evaluated formaldehyde occupational exposure in esthetic and wellness scenarios, including beauty salons⁶⁴⁻⁶⁶ hair salons,⁶⁷⁻⁶⁹ nail salons⁷⁰⁻⁷³ and aromatherapy spa.⁷⁴ Similarly, to the monitoring performed in research and healthcare scenarios, even in these settings exposure assessment was performed during a typical working day with variable sampling periods. The main procedures used for assessing formaldehyde occupational exposure were active and/or passive environmental and active and/or passive personal sampling with 2,4-DNPH cartridges or tapes and HPLC with UV/VIS or UV/DAD detector. In the other cases, the exposure was evaluated by the use of active environmental monitoring with chromatometric detector tubes,⁶⁴ active environmental monitoring with formaldehyde meter monitor,67,71 active environmental and personal sampling with 2-hydroxymethyl piperidine cartridge and GC-MS.⁶⁹ active environmental monitoring with a formaldehyde colorimetric/photoelectric sensor.73 In one study the exposure was also evaluated by the use of biological monitoring.⁶⁵ The levels of environmental formaldehydes ranged from not detectable to more than 4 mg m^{-3} , with the highest levels recovered in hair saloons.66,67

CAMMALLERI ET AL.

FIGURE 1 PRISMA flow diagram of the literature search

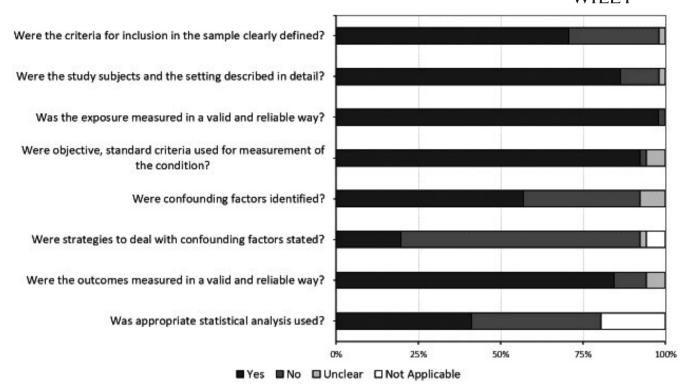


FIGURE 2 Results of the risk of bias assessment for each question of the checklist

3.4 | Main characteristics of the studies involving industrial settings

In Table 4 are reported the studies performed in industrial settings.

Ten studies included in the systematic review were performed in industrial settings, involving different sectors: machine shops,⁷⁵ wooden furniture manufacturing factories,⁷⁶ commercial-scale processing of polyethylene,⁷⁷ tire manufacturing plants,⁷⁸ a wastewater treatment plant,⁷⁹ a materials recovery facility,⁸⁰ dairy facilities,⁸¹ decorating workplace during the decorating engineering,⁸² fish hatchery incubation room,⁸³ wood pellets industry.⁸⁴ Exposure assessment to formaldehyde was performed with active environmental and active and passive personal sampling with 2,4-DNPH cartridge and HPLC-UV/VIS or GC-MS with NPD, or with active environmental monitoring with formaldehyde meter monitor^{79,81-83} or, in one case, by the use of active personal sampling with 2-(hydroxymethyl) piperidine cartridge and GC-MS technique.⁸⁴ Exposure levels were in the order of $\mu g m^{-3}$, but in the cases of wooden furniture manufacturing factories⁷⁶ and fish hatchery incubation room,⁸³ exposure concentrations achieves until to $2-3 \text{ mg m}^{-3}$.

3.5 | Main characteristics of the studies involving fire fighters' and other settings

In Table 5 are reported the studies carried out in fire fighters' and other settings and a miscellanea of other occupational scenarios.

As shown in Table 5, three studies⁸⁵⁻⁸⁷ assessed firefighters' exposure to formaldehyde during their occupational activities. All

the studies were performed by the same procedures: active environmental and passive personal sampling with 2,4-DNPH filters and HPLC with UV/VIS detector. Exposure levels ranged from <LOD to 5 mg m⁻³.

Other monitored occupational activities and/or scenarios included gardening activities,⁸⁸ vehicles such as taxi, bus and subway,⁸⁹ highway tollbooth,⁹⁰ gas station,⁹¹ school campus as a micro-scale society,⁹² dry cleaning shops,⁹³ copy centers.⁹⁴ In most cases, exposure to formaldehyde was evaluated with active and/or passive environmental and active and/or passive personal sampling with 2,4-DNPH cartridges and HPLC with UV/VIS or UV/DAD detector. In one study the active environmental monitoring was performed with a formaldehyde meter monitor⁹⁴ while in another study the evaluation was performed by the use of active environmental and personal sampling with 2-(hydroxymethyl)piperidine cartridges and GC-FID technique.⁹³ The levels of exposure resulted in the order of μ g m⁻³, with the exception of the concentrations recovered during garden activities, until to about 4 mg m⁻³.⁸⁸

4 | DISCUSSION

The present systematic review was conducted to recover scientific evidences on occupational exposure to formaldehyde, in order to define the occupational settings at risk of exposure and the procedures applied to assess exposure levels. These two aims are even more relevant since formaldehyde was recognized as a carcinogen, making mandatory to carry out workers' health surveillance profiled on exposure data.

of 14	-WH	LEY									CAMMALLE
	Exposure levels	Environmental (min-max) = 0.023-0.162 mg·m ⁻³ Personal = ND	Environmental (min-max) = 0.27-1.36 mg·m ⁻³ Personal (min-max) = 0.40-1.84 mg·m ⁻³	Environmental (min-max) = 0.017-2.048 mg·m ⁻³ Personal = ND	Environmental (min-max) = 0.501-0.726 mg·m ⁻³ Personal (min-max) = 0.590-1.059 mg·m ⁻³	Environmental (min-max) = 0.66-1.61 mg·m ⁻³ Personal (min-max) = 0.77-2.15 mg·m ⁻³	Environmental (min-max) = 0.257-0.714 mg·m ⁻³ Personal (min-max) = 0.184-1.115 mg·m ⁻³	Environmental (min-max) = 0.03- 2.52 mg m ⁻³ Personal (min-max) = 1.89-4.82 mg m ⁻³	Environmental (min-max) =0.146- 0.518 mg m ⁻³ Personal (min-max) =0.157-1.469 mg m ⁻³	Environmental = ND Personal (mean \pm SD) = 0.034 \pm 0.008 mg m^{-3}	Environmental (min- max) = <lod-2.65 m<sup="" mg="">-3 Personal = ND</lod-2.65>
	Sampling period	Typical working day	Typical working day in the dissection room (from 1.1 to 6 h)	Typical working day	Typical working day	Typical working day in the dissection room (from 2 to 3 h)	Typical working day in the dissection room (usually 2 h)	Typical working day (from 0.5 h in embalming room to 4 h in other sites)	Typical working day in the dissection room (typical dissection sessions)	Typical working day (8 h)	24 h for pathology laboratory and 10 min for other sites
Exposure assessment	Analytical methodology (sampling and analysis)	Active environmental sampling with 2,4-DNPH cartridge; HPLC-UV/ DAD	Passive environmental and personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Passive environmental sampling with 2,4-DNPH cartridge; HPLC-UV/ DAD	Active environmental and personal sampling with 2-hydroxymethyl piperidine cartridge; GC-FID	Passive environmental and personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Active environmental and personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Active environmental and personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Active environmental and personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Passive personal sampling with 2,4- DNPH cartridge; HPLC-UV/VIS	Active environmental sampling with 2,4-DNPH cartridge; HPLC-UV/ DAD
	Sampling setting and sites	University research institute (offices, classrooms, laboratories, library, print rooms)	Gross anatomy laboratory of a medical school (dissection room)	Pathology departments (offices, laboratories, processing rooms, reduction rooms, storage rooms)	University gross anatomy laboratory (dissection room)	University gross anatomy laboratory (dissection room)	University gross anatomy laboratory (indoor laboratory, corridor, moulage, classroom)	University morphology department (corridor, entrance hall, embalming room, anatomy laboratory)	University gross anatomy laboratory (dissection room)	Hospital operating theater	Medical facilities of a prefecture (pathology laboratories, anatomy laboratories, organ preservation facilities, disinfection facilities, dissecting room)
	First author (Year) Country	Cavalcante, (2005) ⁵⁴ Brazil	Ohmichi (2006) ¹³ Japan	Perdelli (2006) ⁵⁵ Italy	Lakchayapakor (2010) ⁵⁶ Thailand	Vohra (2011) ⁵⁷ Saudi Arabia	Azari (2012) ¹⁵ Iran	De Ochs (2012) Brazil	Saowakon (2015) ⁵⁹ Thailand	Bellisario (2016) ⁶⁰ Italy	Higashikubo (2017) ⁶¹ Japan

max) = 0.01

		Exposure assessment		
First author (Year) Country	Sampling setting and sites	Analytical methodology (sampling and analysis)	Sampling period	Exposure levels
Lee (2017) ⁶² West Virginia and Texas (USA)	Hospital pathology laboratory and hospital histology laboratory	Active and passive environmental and Typical working day (117–515 min) personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Typical working day (117–515 min)	Active environmental (min-m 0.31 mg m ⁻³ Passive environmental (min- max) = 0.02-0.36 mg m ⁻³ Active personal (min-max) = 0.04 mm ⁻³

(Continued)

TABLE 2

Abbreviations: 2,4-DNPH, 2,4-dinitrophenylhydrazine; HPLC, high performance liquid chromatography; GC-FID, gas chromatography - flame ionization detector; LOD, limit of detection; ND, not Personal = ND with formaldehyde meter monitor (electrochemical sensor) determined; UV/DAD, ultraviolet/diode array detector; UV/VIS, ultraviolet/visible detector workshops) Malesia

Environmental (min-max) = 0-1.973 mg

Fypical working day

Active environmental monitoring

Learning institute (laboratories,

Kwong (2018)⁶³

Passive personal (min-max) = 0.01-

 2.70 mg m^{-3}

= 0.01-

ιņ

The first relevant result is related to the great number of occupational scenarios linked to the potential exposure to formaldehyde; indeed, although formaldehyde is a known carcinogen, we found several workplaces and work activities involving a potential exposure to formaldehyde both for its use or its emission during different thermal processes. In total, we grouped four main scenarios: healthcare and research, esthetic and wellness, industrial, firefighters' and a miscellanea of other occupational places/ activities.

Healthcare and research settings included gross anatomy, pathology or histology laboratories^{13,15,55-59,61,62} or also operating theaters⁶⁰ and other indoor environments of universities and research or training institutes.^{54,63} In particular, some work activities performed in gross anatomy, pathology or histology laboratories and in operating rooms involve the use of solutions containing formaldehyde for fixing and preserving biological tissues and for preparing cadavers. Thus, formaldehyde vapors can pollute the indoor air of these environments, resulting in a risk of occupational exposure in hospital settings, research laboratories and medical schools. Besides, it is also demonstrated that formaldehyde exposure can occur not only during the handling of formaldehyde and formaldehyde- treated materials, but also through inappropriate storage of this substance or treated materials and through an ineffective local exhaust ventilation system.⁶¹

The second main recognized sector include esthetic and wellness settings, such as beauty, hair or nail salons and spa.⁶⁴⁻⁷⁴ In particular, hair dressing activities exposed to higher levels of formaldehyde respect to nail and beauty salons and spas. Probably hair dressers use routinely specific products containing formaldehyde, consciously or because not clearly reported in the label.^{68,95} Likewise, detectable levels of formaldehyde were found also in nail and beauty salons due to its presence in care products used in these settings.

As regard to the industrial scenarios,^{75–84} airborne formaldehyde is generally present at low levels (micrograms), except where it is directly released from resins⁷⁶ or used for its biocidal properties in dairy facilities^{76,81} and fish hatcheries.⁸³

In addition to the well-known exposure profile of firefighters,⁸⁵⁻⁸⁷ other investigated scenarios are very diversified, both in terms of settings (school campuses, laundries, copy centers etc)⁸⁸⁻⁹⁴ and in terms of formaldehyde levels (from not detectable to some mg m⁻³).

The methodological approach and the analytical methods are similar in the greatest part of the studies and, thus, their results are comparable. Notice that, in general, the results of personal monitoring are higher than environmental ones. This finding should be taken into account when a survey strategy is planned: both modalities should be performed to define risk levels and elaborate a risk assessment document. For this purpose, a comparison of data obtained with regulatory limits must be carried out. At today, however, United States, EU and others countries have adopted different approaches for the toxicological evaluation of experimental and epidemiological data on formaldehyde exposure and effects resulting in different limit values both for long and short term exposure.

IABLE 3 Selected	I Characteristics of stu	I ABLE 3 Selected characteristics of studies (n = 11) involving eschedic and weilness settings included in the systematic review	ded in the systematic re	view
First author (Year)	Sampling setting	Exposure assessment		
Country	and sites	Analytical methodology	Sampling period	Exposure levels
Tsignonia (2010) Greece	Beauty salons	Active environmental monitoring with chromatometric detector tubes	Typical working day	Environmental = <lod Personal = ND</lod
Pierce (2011) ⁶⁶ Illinois, California (USA)	Hair salon	Active environmental and active and passive personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS Active environmental monitoring with formaldehyde meter monitor (UV detector)	Typical working day	Environmental (min-max) = 0.06-4.48 mg m ⁻³ Personal = ND
Huang (2012) Taiwan	Aromatherapy spa	Active environmental sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	90–120 min	Environmental (min-max) = 0.026-0.030 mg m ⁻³ Personal = ND
Alaves (2013) ⁷⁰ Utah (USA)	Nail salons	Active environmental sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Typical working day (8 h)	Environmental (min-max) = 0.011-0.040 mg m ⁻³ Personal = ND
Peteffi (2016) ⁶⁵ Brazil	Beauty salons	Passive environmental sampling with 2,4-DNPH tape; HPLC-UV/VIS Biological sampling (urine); headspace GC-FID	Typical working day	Environmental (min-max) = 0.09-0.20 mg m ⁻³ Personal = ND Biological (min-max) = 2.72-53.91 mg L ⁻¹
Aglan (2020) Egypt	Hair salons	Passive environmental sampling with 2,4-DNPH tape; HPLC-UV/VIS	15 min	Environmental (mean \pm SD) = 2.10 \pm 0.34 mg m ⁻³ hairstylists involved in hair straightening procedures for <5 years and 2.29 \pm 0.20 mg m ⁻³ for hairstylists involved in hair straightening procedures for >5 years. Personal = ND
Chang (2018) ⁶⁹ Taiwan	Hair salons	Active environmental and personal sampling with 2-hydroxymethyl piperidine cartridge; GC-MS	5 h	Environmental (min-max) = 0.012-1.040 mg m ⁻³ Personal = 0.295-0.468 mg·m ⁻³
Heaton (2019) ⁷¹ Alabama (USA)	Nail salon (experimental chamber)	Active environmental monitoring with formaldehyde meter monitor (electrochemical sensor)	15 min	Environmental (min-max) = 0.15-0.27 mg m ⁻³ Personal = ND
Lamplugh (2019) ⁷² Colorado (USA)	Nail salons	Active environmental sampling with 2,4-DNPH cartridge; HPLC-UV/DAD	8 h	Environmental (min-max) = 0.005-0.021 mg·m ⁻³ Personal = ND
Pexe (2019) ⁶⁶ Brazil	Beauty and haidresser salons	Active personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS Passive environmental sampling with 2,4-DNPH tape; HPLC-UV/VIS	15 min for actives 8 h for passive	Environmental (min-max) (passive) = 0.10-2.40 mg m ⁻³ Environmental (min-max) (active) = <lod-5.15 m<sup="" mg="">-3 Personal = ND</lod-5.15>
Zhong (2019) ⁷³ Michigan (USA)	17 nail salons	Active environmental monitoring with a formaldehyde colorimetric/photoelectric sensor	At least 30 min	Environmental (min-max) = <lod-0.040 mg·m<sup="">-3 Personal = ND</lod-0.040>
Abbreviations: 2,4-DN	IPH, 2,4-dinitrophenylh	vdrazine: HPLC. high performance liguid chromatography: GC	C-FID, gas chromatograph	Abbreviations: 2.4-DNPH, 2.4-dinitrophenylhydrazine; HPLC, high performance liquid chromatography; GC-FID, gas chromatography - flame ionization detector; GC-MS, gas chromatography - mass

TABLE 3 Selected characteristics of studies (n = 11) involving esthetic and wellness settings included in the systematic review

Abbreviations: 2,4-DNPH, 2,4-dinitrophenylhydrazine; HPLC, high performance liquid chromatography; GC-FID, gas chromatography - flame ionization detector; GC-MS, gas chromatography - mass spectrometry; LOD, limit of detection; ND, not determined; UV/VIS, ultraviolet/visible detector; UV/DAD, ultraviolet/diode array detector.

MILEY

Eirst author (Vear)		Exposure assessment		
Country	Sampling setting and sites	Analytical methodology	Sampling period	Exposure levels
Lillienberg (2008) ⁷⁵ Sweden	Machine shops	Active and passive personal sampling with 2,4- DNPH cartridge; HPLC-UV/VIS	6-8 h	Environmental = ND Personal = 0.001-0.154 mg m ⁻³
Ratnasingam (2010) ⁷⁶ Malaysia, Thailand	Wooden furniture manufacturing factories	Active personal sampling with 2,4-DNPH cartridge; HPLC (detector not specified)	30 min	Environmental = ND Personal = 2.07-2.72 mg m ⁻³
Traviss (2010) ⁸⁰ New Hampshire (USA)	Materials recovery facility	Active environmental sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	One work shift per fuel type	Environmental (min-max) = about 0.0003-about 0.0025 mg m ⁻³ Personal = ND
Tikuisis (2010) ⁷⁷ Canada	Commercial-scale processing of polyethylene	Active environmental and personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Full-shift workplace	Environmental = <lod Personal = <lod< td=""></lod<></lod
Lee (2012) ⁷⁸ Korea	2 tire manufacturing plants	Active environmental sampling with 2,4-DNPH cartridge; GC-NPD	Full-shift workplace	Environmental (min-max) = 0.011-0.036 mg m ⁻³ Personal = ND
Teixeira (2013) ⁷⁹ Portugal	Wastewater treatment plant	Active environmental monitoring with a formaldehyde colorimetric/photoelectric sensor	30 min	Environmental = always <lod Personal = ND</lod
Doane (2014) ⁸¹ New York (USA)	Two dairy facilities	Active environmental monitoring with formaldehyde meter monitor (electrochemical sensor)	Three consecutive days	Environmental (min-max) = 0-about 3.0 mg m ⁻³ Personal = ND
Wang (2014) ⁸² Taiwan	Decorating workplace during the decorating engineering.	Active environmental monitoring with formaldehyde meter monitor (electrochemical sensor)	5 min	Environmental (min-max) = 0.10 \pm 0.03- 0.86 \pm 0.54 mg m^{-3} Personal = ND
Voorhees (2016) ⁸³ South Dakota (USA)	Fish hatchery incubation room	Active environmental monitoring with formaldehyde meter monitor (electrochemical sensor)	60 min + additional 30 min if the values were elevated above the basal level	Environmental = <lod-about 2.5="" m<sup="" mg="">-3 Personal = ND</lod-about>
Rahman (2017) ⁸⁴ New York State (USA)	Manufacturing and storage of wood pellets in a warehouse and enclosed test chambers of wood pellets industry	Active personal sampling with 2-(hydroxymethyl)piperidine cartridge; GC-MS	30 min	Environmental = ND Personal (min-max) = $0.16-0.19$ mg m ⁻³ (drums) and 0.01-0.65 mg m ⁻³ (warehouse)
Abbreviations: 2,4-DN	PH, 2,4-dinitrophenylhydrazine; H LOD limit of detection: ND not d	Abbreviations: 2,4-DNPH, 2,4-dinitrophenylhydrazine; HPLC, high performance liquid chromatography; GC-MS, gas chromatography/mass spectrometry; GC-NPD, gas chromatography/nitrogen phosphorus detectors: I OD. limit of detection: ND. not determined: UV/VIS. ultraviolet/visible detector.	MS, gas chromatography/mass spectrometry	; GC-NPD, gas chromatography/nitrogen

WILEY-

OPSmpling setting and siteAutotationeloySempling period1Carcinu (USA)Carcinu (SA)Carcinu (SA)Comin-2 hCarcinu (USA)Carcinu (SA)Carcinu (SA)Comin-2 hComin-2 hCarcinu (USA)2 vericles including skiPasive vericommental sampling with 2.4-DNH fitterComin-2 h0Live (SA)Pasive personal sampling with 2.4-DNH fi	First author		Exposure assessment			- vv
ofine districts devices and sampling with 24-DNPH cartridge dom-0.1 onine USA) 2 verkietes including taxi Pasive environmental sampling with 24-DNPH filter Demire rush hours 2 verkietes including taxi Pasive environmental sampling with 24-DNPH filter Bening rush hours 2 verkietes including taxi Pasive personal sampling with 24-DNPH filter Bening rush hours 2 verkietes including taxi Pasive personal sampling with 24-DNPH filter Bening rush hours 2 verkietes including Pasive personal sampling with 24-DNPH filter Bening rush hours 2 verkietes including Pasive personal sampling with 24-DNPH filter Bening rush hours 2 verkietes including rush Pasive personal sampling with 24-DNPH filter Pasive personal sampling with 24-DNPH filter 2 verkietes including rush Parter personal sampling with 24-DNPH filter Pasive rush rush rush rush rush Pasive rush rush rush rush rush 2 verkieter including rush 24-DNPH filter Pasive rush rush rush rush rush rush rush rush	(rear) Country	Sampling setting and sites	Analytical methodology	Sampling period	Exposure levels	ILE
29 whicles including taxi Desire environmental sampling with 24-DNPH filter Density territy is carridge: HPLC-UV/VIS Centring territy is carried and suppling with 24-DNPH filter Service match and personal sampling with 24-DNPH filter Ab University campus office. University campus office. Aller environmental sampling with 24-DNPH filter Campus office. University campus office. Aller environmental sampling wit	Baldauf (2006) ⁸⁸ North Carolina (USA)	Gardening activities	Active personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	30 min -2 h	Environmental = ND Personal (min-max) = 0.01-about 4 mg m ⁻³	. Y ——
Freferets Erecipaters Ensisters Ensister Personal sampling with 2.4-DNPH filter 5-360 min ubstiftie snoke Basers HPLC-UV/NS Basers HPLC-UV/NS Basers HPLC-UV/NS Basers	Pang (2007) ⁸⁹ China	29 vehicles including taxi, bus and subway	Passive environmental sampling with 2,4-DNPH cartridge; HPLC-UV/VIS	Evening rush hours	Environmental = 0.015-0.094 mg m ⁻³ Personal = ND	
Firefighters exposure to bubblifies smoke Cub denoting stand passive personal sampling with 2.4-DNPH filter	Reisen (2009) ⁸⁵ Australia	Firefighters' exposure to bushfire smoke	Passive personal sampling with 2,4-DNPH filter paper; HPLC-UV/VIS Active environmental sampling with 2,4-DNPH filter cassette; HPLC-UV/VIS	35-360 min	Environmental (min-max) = 0.07-0.65 mg m ⁻³ Personal (min-max) = 0.07-0.71 mg m ⁻³	_
Intellecta Highway tollbooth Ceive personal sampling with 2.4-DNPH filter paper. HPLC-UV/NS 7-Bh A) Four dry cleaning shops Ceitive environmental and personal sampling with 2.4-DNPH filter 7-Bh A) Four dry cleaning shops Ceive environmental and personal sampling with 2.4-DNPH filter 7-Bh A) University campus (offices, offices, offices, pleuc-UV/DAD Ceive environmental and personal sampling with 2.4-DNPH filter Pilter A) University campus (offices, offices, pleuc-UV/DAD Ceive environmental monitoring with formal deby of spending time domination, jubrary, prime dominated by the domination of the domination, jubrary, prime domination, ju	Reisen (2011) ⁸⁶ Australia	Firefighters' exposure to bushfire smoke	Active and passive personal sampling with 2,4-DNPH filter paper; HPLC-UV/VIS		Environmental = ND Personal (min-max) = <lod-0.817 m<sup="" mg="">-3</lod-0.817>	
A Eur dry cleaning shops Cative environmental and personal sampling with 2-(hydroxymethy)biperidine cartridge; GC-FID Eul-shift A) University campus (offices, dining room, student ofming room, student ofming room, student A Piperidine cartridge; GC-FID Piperidine cartridge; FID-CUV/DAD Pip	Belloc-Santaliestra (2015) ⁹⁰ Spain	Highway tollbooth	Active personal sampling with 2,4-DNPH filter paper; HPLC-UV/VIS	7-8 h	Environmental = ND Personal (min-max) = <lod-0.0162 m<sup="" mg="">-3</lod-0.0162>	
University campus (offices, dining room, student dormitory, library, print nooms)Cative environmental sampling with 2,4-DNPH spending day or typical spending time spending time spending time(2016)Two copy centers Two copy centersActive environmental monitoring with 1,4-DNPH tages entroled encodencial sensor)Papeading day or typical spending time(2016)Two copy centersActive environmental monitoring with 1,4-DNPH tape: HPLC-UV/VIS20-35 min for passive personal sampling with 2,4-DNPH tape: HPLC-UV/VISaGas stationActive personal sampling with 2,4-DNPH cartridge; HPLC-UV/VIS20-35 min for passive personal sampling and 12-18 min for active personal sampling with 2,4-DNPH cartridge; HPLC-UV/VISaGas stationActive environmental and personal sampling with 2,4-DNPH cartridge; HPLC (detector not specified)	Ceballos (2016) ⁹³ Ohio (USA)	Four dry cleaning shops	Active environmental and personal sampling with 2-(hydroxymethyl)piperidine cartridge; GC-FID	Full-shift	Environmental (min-max) = <lod-0.054 mg<br="">m⁻³ Personal (min-max) = <lod-0.109 m<sup="" mg="">-3</lod-0.109></lod-0.054>	
(2016) Two copy centers Active environmental monitoring with formaldehyde 24 h En Per meter monitor (electrochemical sensor) Per Per Compartment fire behavior Passive personal sampling with 2,4- DNPH tape; 20-35 min for passive personal En Training HPLC-UV/VIS 20-35 min for passive personal Per Active personal sampling with 2,4- DNPH tape; 20-35 min for passive personal Per Active personal sampling with 2,4- DNPH cartridge; active personal sampling Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4- DNPH cartridge; Per Per Meter on of the personal sampling with 2,4-	Ho (2016) ⁹² China	University campus (offices, dining room, student dormitory, library, print rooms)	Active environmental sampling with 2,4-DNPH cartridge; HPLC-UV/DAD	Typical working day or typical spending time	Environmental (mean \pm SD) = 0.009 \pm 0.009 mg m^{-3} Personal = ND	
Compartment fire behavior training Passive personal sampling with 2,4- DNPH tape; 20-35 min for passive personal sampling and 12-18 min for sampling and 12-18 min for appling and 12-18 min for active personal sampling hPLC-UV/VIS a Gas station Active personal sampling with 2,4- DNPH cartridge; HPLC-UV/VIS 20-35 min for passive personal sampling and 12-18 min for active personal sampling with 2,4- DNPH cartridge; HPLC (detector not specified)	Vincente (2016) Portugal	Two copy centers	Active environmental monitoring with formaldehyde meter monitor (electrochemical sensor)	24 h	Environmental (mean) = 0.04 \pm 0.01 and 0.03 \pm 0.01 mg m^{-3} for copy centers A and B Personal (min-max) = ND	
a Gas station Active environmental and personal sampling 2 h in a spring day and 2 h in a with 2,4-DNPH cartridge; HPLC (detector not winter day specified)	Kirk (2019) ⁸⁷ Australia	Compartment fire behavior training	Passive personal sampling with 2,4- DNPH tape; HPLC-UV/VIS Active personal sampling with 2,4- DNPH cartridge; HPLC-UV/VIS	20–35 min for passive personal sampling and 12–18 min for active personal sampling	Environmental (min-max) = <lod-0.043 mg<br="">m⁻³ Personal (min-max) = <lod-0.0087 m<sup="" mg="">-3</lod-0.0087></lod-0.043>	
	Shinohara (2019) ⁹¹ Japan	Gas station	Active environmental and personal sampling with 2,4-DNPH cartridge; HPLC (detector not specified)	2 h in a spring day and 2 h in a winter day	$ \begin{array}{l} \mbox{Environmental (mean) = 0.010 mg m^{-3} (spring) \\ \mbox{or } 0.024 mg m^{-3} (winter) \\ \mbox{Personal (mean) = 0.005 mg } m^{-3} (spring) or \\ \mbox{0.012 } mg m^{-3} (winter) \\ \end{array} $	

10 of 14

This review presents some limitations. Firstly, several studies only report average or min-max air levels; secondly, only few studies measured short time or peak air concentrations; besides, the time intervals of the monitoring and the number of samples were different from a study to another. Thus, we could not compare exposure data reported by included articles with actual regulatory limits.

Finally, given the differences of the studies included in this review in terms of exposed populations and procedures used for assessing airborne formaldehyde levels, we decided to review and summarize the results of the selected studies rather than to carry out a formal meta-analysis. Thus, statistical heterogeneity and publication bias were not assessed.

5 | CONCLUSIONS

The results of the present review demonstrate that there is a great number of diversified occupational scenarios at risk for formaldehyde exposure. Nevertheless, the monitoring approaches are very similar each other and based on environmental and personal sampling followed by chromatographic analyses, thus allowing data comparison. The settings at higher levels of airborne formaldehyde resulted the gross anatomy and pathology laboratories, the hairdressing salons and some specific productive settings such as wooden furniture factories, dairy facilities and fish hatcheries. However, it is important to highlight that in almost all the studied scenarios/activities, formaldehyde was recovered in air at levels higher than outdoor. Considering that formaldehyde is a well-known carcinogen, targeted strategies for exposure elimination or mitigation (when formaldehyde cannot be removed or substituted) must be adopted.

ACKNOWLEDGEMENTS

This work was carried out within the BRiC-2018 research projects financially supported by INAIL (Project number B86C19000070001).

CONFLICT OF INTERESTS

None of the authors declares any conflict of interest.

AUTHOR CONTRIBUTIONS

Vittoria Cammalleri was involved in data curation, formal analysis, methodology, investigation, validation, and writing the original draft. Roberta Noemi Pocino was involved in data curation, formal analysis, methodology, investigation, and writing the original draft. Daniela Marotta was involved in data curation, formal analysis, investigation, and writing the original draft. Carmela Protano was involved in conceptualization, project administration, validation, writing the original draft, and writing, review and editing. Federica Sinibaldi was involved in data curation, investigation, and writing the original draft. Stefano Simonazzi was involved in formal analysis, and writing the original draft. Marta Petyx was involved in validation, and writing, review and editing. Sergio lavicoli was involved in validation, and writing, review and editing. Matteo Vitali was involved in conceptualization, data curation, formal analysis, funding acquisition, methodology, project administration, resources, supervision, visualization, writing the original draft, and writing, review and editing.

PEER REVIEW

The peer review history for this article is available at https://publo ns.com/publon/10.1111/ina.12949.

ORCID

Matteo Vitali D https://orcid.org/0000-0002-1001-6343

REFERENCES

- Franz AW, Kronemayer H, Pfeiffer D, et al. Formaldehyde. Ullmann's Encyclopedia of Industrial Chemistry. Wiley-VCH Verlag GmbH & Co. KGaA; 2016:1-34.
- Yang M, Ospina M, Tse C, Toth S, Caudill SP, Vesper HW. Ultraperformance liquid chromatography tandem mass spectrometry method to determine formaldehyde hemoglobin adducts in humans as biomarker for formaldehyde exposure. *Chem Res Toxicol.* 2017;30(8):1592-1598.
- WHO Regional Office for Europe Copenhagen. In Air Quality guidelines for Europe, 2nd ed, 2000. https://www.euro.who.int/__ data/assets/pdf_file/0005/74732/E71922.pdf Accessed February 9, 2021.
- Nielsen GD, Larsen ST, Wolkoff P. Re-evaluation of the WHO (2010) formaldehyde indoor air quality guideline for cancer risk assessment. Arch Toxicol. 2017;91(1):35-61.
- Kosmider L, Cox S, Zaciera M, et al. Daily exposure to formaldehyde and acetaldehyde and potential health risk associated with use of high and low nicotine e-liquid concentrations. *Sci Rep.* 2020;10(1):6546.
- Salonen H, Pasanen A-L, Lappalainen S, et al. Airborne concentrations of volatile organic compounds, formaldehyde and ammonia in Finnish office buildings with suspected indoor air problems. J Occup Environ Hyg. 2009;6(3):200-209.
- Salonen H, Pasanen A-L, Lappalainen S, et al. Volatile organic compounds and formaldehyde as explaining factor on reported complaints and symptoms in the office environment. J Occup Environ Hyg. 2009;6(4):239-247.
- IARC, Working Group on the Evaluation of Carcinogenic Risks to Humans. Chemical Agents and Related Occupations. IARC Monogr. Eval. Carcinog. Risks Hum; 2012;401-430.
- Dumas O, Wiley AS, Quinot C, et al. Occupational exposure to disinfectants and asthma control in US nurses. *Eur Respir J.* 2017;50(4):1700237.
- Xu W, Stewart EJ. A comparison of engineering controls for formaldehyde exposure during grossing activities in health care anatomic pathology laboratories. J Occup Environ Hyg. 2016;13(7):529-537.
- Vaizoğlu SA, Aycan S, Akin L, et al. Determination of formaldehyde levels in 100 furniture workshops in Ankara. *Tohoku J Exp Med*. 2005;207(2):157-163.
- Thorud S, Gjolstad M, Ellingsen DG, Molander P. Air formaldehyde and solvent concentrations during surface coating with acid-curing lacquers and paints in the woodworking and furniture industry. J Environ Monit. 2005;7(6):586-591.
- Ohmichi K, Komiyama M, Matsuno Y, et al. Formaldehyde exposure in a gross anatomy laboratory-personal exposure level is higher than indoor concentration. *Environ Sci Pollut Res Int.* 2006;13(2):120-124.
- 14. Yahyaei E, Majlesi B, Naimi Joubani M, et al. Occupational exposure and risk assessment of formaldehyde in the pathology departments of hospitals. *Asian Pac J Cancer Prev.* 2020;21(5):1303-1309.

12 of 14 | WILEY

- Azari MR, Asadi P, Jafari MJ, Soori H, Hosseini V. Occupational exposure of a medical school staff to formaldehyde in Tehran. *Tanaffos*. 2012;11(3):36-41.
- Ricke SC, Richardson K, Dittoe DK. Formaldehydes in feed and their potential interaction with the poultry gastrointestinal tract microbial community-a review. *Front Vet Sci.* 2019;6:188.
- Siew SS, Kauppinen T, Kyyrönen P, Heikkilä P, Pukkala E. Occupational exposure to wood dust and formaldehyde and risk of nasal, nasopharyngeal, and lung cancer among Finnish men. *Cancer Manag Res.* 2012;4:223-232.
- Reinhardt TE, Ottmar RD. Baseline measurements of smoke exposure among wildland firefighters. J Occup Environ Hyg. 2004;1(9):593-606.
- Barbosa E, Dos Santos ALA, Peteffi GP, et al. Increase of global DNA methylation patterns in beauty salon workers exposed to low levels of formaldehyde. *Environ Sci Pollut Res Int*. 2019;26(2):1304-1314.
- Betha R, Selvam V, Blake DR, Balasubramanian R. Emission characteristics of ultrafine particles and volatile organic compounds in a commercial printing center. J Air Waste Manag Assoc. 2011;61(11):1093-1101.
- Solomons K, Cochrane JW. Formaldehyde toxicity. Part II. Review of acute and chronic effects on health. S Afr Med J. 1984;66(3):103-106.
- Liteplo RG, Meek ME. Inhaled formaldehyde: exposure estimation, hazard characterization, and exposure-response analysis. J Toxicol Environ Health B Crit Rev. 2003;6(1):85-114.
- Mathur N, Rastogi SK. Respiratory effects due to occupational exposure to formaldehyde: systematic review with meta-analysis. *Indian J Occup Environ Med.* 2007;11(1):26-31.
- Swenberg JA, Moeller BC, Lu K, Rager JE, Fry RC, Starr TB. Formaldehyde carcinogenicity research: 30 years and counting for mode of action, epidemiology, and cancer risk assessment. *Toxicol Pathol.* 2013;41(2):181-189.
- Tang X, Bai Y, Duong A, Smith MT, Li L, Zhang L. Formaldehyde in China: production, consumption, exposure levels, and health effects. [published correction appears in Environ Int. 2010;36(3):308]. *Environ Int.* 2009;35(8):1210-1224.
- IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Formaldehyde, 2-Butoxyethanol and 1-tert- Butoxy-2-propanol, Volume 88. IARC Monogr Eval Carcinog Risks Hum; 2006.
- 27. National Toxicology Program, Department of Health and Human Services. Report on Carcinogens, Fourteenth Edition; 2016. https://ntp.niehs.nih.gov/ntp/roc/content/profiles/formaldehy de.pdf Accessed May 13, 2021.
- 28. Commission Regulation (EU) No 605/2014 of 5 June 2014 amending, for the purposes of introducing hazard and precautionary statements in the Croatian language and its adaptation to technical and scientific progress, Regulation (EC) No 1272/2008 of the European Parliament and of the Council on classification, labelling and packaging of substances and mixtures. https://eur-lex.europa. eu/eli/reg/2014/605/oj Accessed February 9, 2021.
- Regulation (EC) No 1272/2008 of the European Parliament and of the Council of 16 December 2008 on classification, labelling and packaging of substances and mixtures, amending and repealing Directives 67/548/EEC and 1999/45/EC, and amending Regulation (EC) No 1907/2006. https://eur-lex.europa.eu/legal-content/EN/ ALL/?uri=CELEX%3A32008R1272 Accessed February 9, 2021.
- Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Medicine*. 2009;6(7):e1000097.
- Moola SMZ, Tufanaru C, Aromataris E, Sears K. Chapter 7: Systematic reviews of etiology and risk. In: Aromataris E, Munn Z, eds. *Joanna Briggs Institute Reviewer's Manual*. The Joanna Briggs Institute; 2017:2-7.

- Yu L, Wang B, Cheng M, et al. Association between indoor formaldehyde exposure and asthma: a systematic review and metaanalysis of observational studies. *Indoor Air.* 2020;30(4):682-690.
- Kurose T, Kodera H, Aoyama H, Kawamata S. Formaldehyde concentration in the air and in cadavers at the gross anatomy laboratory in Hiroshima University. *Hiroshima J Med Sci.* 2004;53(3–4):33-37.
- Priha E, Pennanen S, Rantio T, Uitti J, Liesivuori J. Exposure to and acute effects of medium-density fiber board dust. J Occup Environ Hyg. 2004;1(11):738-744.
- Ghasemkhani M, Jahanpeyma F, Azam K. Formaldehyde exposure in some educational hospitals of Tehran. *Ind Health*. 2005;43(4):703-707.
- Lavoué J, Beaudry C, Goyer N, Perrault G, Gérin M. Investigation of determinants of past and current exposures to formaldehyde in the reconstituted wood panel industry in Quebec. *Ann Occup Hyg.* 2005;49(7):587-602.
- Posniak M, Makhniashvili I, Koziel E. Volatile organic compounds in the indoor air of Warsaw Office Buildings. *Indoor Built Environ*. 2005;3(4):269-275.
- McNary JE, Jackson EM. Inhalation exposure to formaldehyde and toluene in the same occupational and consumer setting. *Inhal Toxicol*. 2007;19(6–7):573-576.
- Steinsvåg K, Bråtveit M, Moen BE. Exposure to carcinogens for defined job categories in Norway's offshore petroleum industry, 1970 to 2005. Occup Environ Med. 2007;64(4):250-258.
- Vimercati L, Carrus A, Dell'Erba A, Assennato G. Occupational exposure to formaldehyde in autopsy room and in pathologic anatomy laboratories. *G Ital Med Lav Ergon*. 2007;29(3 Suppl):266-268.
- Hui PS, Mui KW, Wong LT. Influence of indoor air quality (IAQ) objectives on air-conditioned offices in Hong Kong. *Environ Monit Assess*. 2008;144(1-3):315-322.
- Rezai-Basiri M, Samini M, Ghazi-Khansari M, Rezayat M, Sahebgharani M, Partoazar A. Monitoring Ames assay on urine of clinical pathology laboratories tecnhnicians. *J of Pharma and Tox*. 2008;3(3):230-235.
- 43. Jung JH, Youn SU, Kwon E, et al. Emission rates of air pollutants from portable gas ranges and nitrogen dioxide exposure assessment in restaurants. *J UOEH*. 2009;31(1):13-22.
- Tsakas MP, Siskos PA. Indoor air quality in the control tower of Athens International Airport, Greece. *Indoor Built Environ*. 2011;20(2):284-289.
- 45. Que DE, Chao HR, Hsu YC, et al. Emission of carbonyl compounds from cooking oli fumes in the night market areas. *Aerosol Air Qual Res.* 2019;19:1566-1578.
- Westberg H, Löfstedt H, Seldén A, Lilja BG, Nayström P. Exposure to low molecular weight isocyanates and formaldehyde in foundries using hot box core binders. *Ann Occup Hyg.* 2005;49(8):719-725.
- Godderis L, Deschuyffeleer T, Roelandt H, Veulemans H, Moens G. Exposure to metalworking fluids and respiratory and dermatological complaints in a secondary aluminium plant. *Int Arch Occup Environ Health.* 2008;81(7):845-853.
- Baur X, Poschadel B, Budnik LT. High frequency of fumigants and other toxic gases in imported freight containers-an underestimated occupational and community health risk. Occup Environ Med. 2010;67(3):207-212.
- Laitinen J, Mäkelä M, Mikkola J, Huttu I. Fire fighting trainers' exposure to carcinogenic agents in smoke diving simulators. *Toxicol Lett*. 2010;192(1):61-65.
- Ahmed HO. Preliminary study: formaldehyde exposure in laboratories of Sharjah University in UAE. *Indian J Occup Environ Med*. 2011;15(1):33-37.
- Yang TT, Chuang KJ, Chang NY, et al. Exposure assessment of particulate and gaseous pollutants emitted during surgery in operating rooms of different specialties. *Air Qual Atmos Health*. 2018;11:937-947.

- 52. Su M, Sun R, Zhang X, et al. Assessment of the inhalation risks associated with working in printing rooms: a study on the staff of eight printing rooms in Beijing, China. *Environ Sci Pollut Res Int.* 2018;25(17):17137-17143.
- Väisänen AJK, Hyttinen M, Ylönen S, Alonen L. Occupational exposure to gaseous and particulate contaminants originating from additive manufacturing of liquid, powdered, and filament plastic materials and related post-processes. J Occup Environ Hyg. 2019;16(3):258-271.
- 54. Cavalcante RM, Seyffert BH, Montes D'Oca MG, et al. Exposure assessment for formaldehyde and acetaldehyde in the workplace. *Indoor Built Environ*. 2005;14(2):165-172.
- 55. Perdelli F, Spagnolo AM, Cristina ML, et al. Occupational exposure to formaldehyde in three pathology departments. *Ann Ig.* 2006;18(6):481-490.
- Lakchayapakorn K, Watchalayarn P. Formaldehyde exposure of medical students and instructors and clinical symptoms during gross anatomy laboratory in Thammasat University. J Med Assoc Thai. 2010;93(Suppl 7):S92-S98.
- Vohra MS. Personal formaldehyde exposure level in the gross anatomy dissecting room at College of Medicine King Saud University Riyadh. Int J Occup Med Environ Health. 2011;24(1):108-113.
- De Ochs SM, Grotz Lde O, Factorine LS, Rodrigues MR, Pereira Netto AD. Occupational exposure to formaldehyde in an institute of morphology in Brazil: a comparison of area and personal sampling. *Environ Sci Pollut Res Int.* 2012;19(7):2813-2819.
- Saowakon N, Ngernsoungnern P, Watcharavitoon P, Ngernsoungnern A, Kosanlavit R. Formaldehyde exposure in gross anatomy laboratory of Suranaree University of Technology: a comparison of area and personal sampling. *Environ Sci Pollut Res Int.* 2015;22(23):19002-19012.
- 60. Bellisario V, Mengozzi G, Grignani E, et al. Towards a formalin-free hospital. Levels of 15-F2t-isoprostane and malondialdehyde to monitor exposure to formaldehyde in nurses from operating theatres. *Toxicol Res (Camb)*. 2016;5(4):1122-1129.
- Higashikubo I, Miyauchi H, Yoshida S, et al. Assessment of workplace air concentrations of formaldehyde during and before working hours in medical facilities. *Ind Health.* 2017;55(2):192-198.
- Lee EG, Magrm R, Kusti M, et al. Comparison between active (pumped) and passive (diffusive) sampling methods for formaldehyde in pathology and histology laboratories. *J Occup Environ Hyg.* 2017;14(1):31-39.
- Kwong QJ, Abdullah J, Tan SC, Thio THG, Yeaw WS. A field study of indoor air quality and occupant perception in experimental laboratories and workshops. *Manag Environ Qual*. 2018;30(2):467-482.
- 64. Tsigonia A, Lagoudi A, Chandrinou S, Linos A, Evlogias N, Alexopoulos EC. Indoor air in beauty salons and occupational health exposure of cosmetologists to chemical substances. Int J Environ Res Public Health. 2010;7(1):314-324.
- Peteffi GP, Antunes MV, Carrer C, et al. Environmental and biological monitoring of occupational formaldehyde exposure resulting from the use of products for hair straightening. *Environ Sci Pollut Res Int*. 2016;23(1):908-917.
- 66. Pexe ME, Marcante A, Luz MS, et al. Hairdressers are exposed to high concentrations of formaldehyde during the hair straightening procedure. *Environ Sci Pollut Res Int.* 2019;26(26):27319-27329.
- 67. Pierce JS, Abelmann A, Spicer LJ, et al. Characterization of formaldehyde exposure resulting from the use of four professional hair straightening products. J Occup Environ Hyg. 2011;8(11):686-699.
- Aglan MA, Mansour GN. Hair straightening products and the risk of occupational formaldehyde exposure in hairstylists. *Drug Chem Toxicol*. 2020;43(5):488-495.
- 69. Chang CJ, Cheng SF, Chang PT, Tsai SW. Indoor air quality in hairdressing salons in Taipei. *Indoor Air.* 2018;28(1):173-180.
- Alaves VM, Sleeth DK, Thiese MS, Larson RR. Characterization of indoor air contaminants in a randomly selected set of commercial

nail salons in Salt Lake County, Utah, USA. Int J Environ Health Res. 2013;23(5):419-433.

- Heaton T, Hurst LK, Amiri A, Lungu CT, Oh J. Laboratory estimation of occupational exposures to volatile organic compounds during nail polish application. Workplace Health Saf. 2019;67(6):288-293.
- Lamplugh A, Harries M, Xiang F, Trinh J, Hecobian A, Montoya LD. Occupational exposure to volatile organic compounds and health risks in Colorado nail salons. *Environ Pollut*. 2019;249:518-526.
- Zhong L, Batterman S, Milando CW. VOC sources and exposures in nail salons: a pilot study in Michigan, USA. Int Arch Occup Environ Health. 2019;92(1):141-153.
- 74. Huang HL, Tsai TJ, Hsu NY, Lee CC, Wu PC, Su HJ. Effects of essential oils on the formation of formaldehyde and secondary organic aerosols in an aromatherapy environment. *Build Environ*. 2010;57:120-125.
- Lillienberg L, Burdorf A, Mathiasson L, Thörneby L. Exposure to metalworking fluid aerosols and determinants of exposure. Ann Occup Hyg. 2008;52(7):597-605.
- 76. Ratnasingam J, Natthondan V, Ioras F, McNulty T. Dust, noise and chemical solvents exposure of workers in the wooden furniture industry in South East Asia. *J Appl Sci.* 2010;10(14):1413-1420.
- Tikuisis T, Taylor G, Sonnenberg K, Cavanagh J. Quantification of emission compounds generated during commercial-scale processing of advanced SCLAIRTECHTM polyethylene. J Plast Film Sheeting. 2010;26(2):167-191.
- Lee N, Lee BK, Jeong S, Yi GY, Shin J. Work environments and exposure to hazardous substances in korean tire manufacturing. Saf Health Work. 2012;3(2):130-139.
- Teixeira JV, Miranda S, Monteiro RA, et al. Assessment of indoor airborne contamination in a wastewater treatment plant. *Environ Monit Assess*. 2013;185(1):59-72.
- Traviss N, Thelen BA, Ingalls JK, Treadwell MD. Biodiesel versus diesel: a pilot study comparing exhaust exposures for employees at a rural municipal facility. J Air Waste Manag Assoc. 2010;60(9):1026-1033.
- Doane M, Sarenbo S. Exposure of farm laborers and dairy cattle to formaldehyde from footbath use at a dairy farm in New York State. *Sci Total Environ*. 2014;487:65-71.
- Wang YF, Tsai CH, Lin CH, Chen SH. Measurement of air quality during a decorating engineering. *Aerosol Air Qual Res.* 2014;14:2029-2039.
- 83. Voorhees JM, Barnes ME. Airborne formaldehyde levels during simulated formalin egg treatments in vertical-flow tray incubators at a production fish hatchery. J Agric Saf Health. 2016;22(3):199-207.
- Rahman MA, Rossner A, Hopke PK. Occupational exposure of aldehydes resulting from the storage of wood pellets. *J Occup Environ Hyg.* 2017;14(6):417-426.
- Reisen F, Brown SK. Australian firefighters' exposure to air toxics during bushfire burns of autumn 2005 and 2006. Environ Int. 2009;35(2):342-352.
- Reisen F, Hansen D, Meyer CP. Exposure to bushfire smoke during prescribed burns and wildfires: firefighters' exposure risks and options. *Environ Int*. 2011;37(2):314-321.
- Kirk KM, Logan MB. Exposures to air contaminants in compartment fire behavior training (CFBT) using particleboard fuel. J Occup Environ Hyg. 2019;16(7):432-439.
- Baldauf R, Fortune C, Weinstein J, Wheeler M, Blanchard F. Air contaminant exposures during the operation of lawn and garden equipment. J Expo Sci Environ Epidemiol. 2006;16(4):362-370.
- Pang X, Mu Y. Characteristics of carbonyl compounds in public vehicles of Beijing city: concentrations, sources, and personal exposures. Atmos Environ. 2007;41:1819-1824.
- Belloc-Santaliestra M, van der Haar R, Molinero-Ruiz E. Occupational exposure assessment of highway toll station workers to vehicle engine exhaust. J Occup Environ Hyg. 2015;12(1):51-61.

HILEY

- Shinohara N, Okazaki Y, Mizukoshi A, Wakamatsu S. Exposure to benzene, toluene, ethylbenzene, xylene, formaldehyde, and acetaldehyde in and around gas stations in Japan. *Chemosphere*. 2019;222:923-931.
- 92. Ho SSH, Cheng Y, Bai Y, et al. Risk assessment of indoor formaldehyde and other carbonyls in campus environments in Northwestern China. *Aerosol Air Qual Res.* 2016;16:1967-1980.
- Ceballos DM, Whittaker SG, Lee EG, et al. Occupational exposures to new dry cleaning solvents: high-flashpoint hydrocarbons and butylal. J Occup Environ Hyg. 2016;13(10):759-769.
- 94. Vincente ED, Ribeiro JP, Custòdio D, Alves CA. Assessment of the indoor air quality in copy centres at Aveiro, Portugal. *Air Qual Atmos Health*. 2017;10:117-127.
- 95. U.S. Food and Drug Administration. Hair-smoothing products that release formaldehyde when heated. https://www.fda.gov/cosme tics/cosmetic-products/hair-smoothing-products-release-forma ldehyde-when-heated. Accessed February 9, 2021.

How to cite this article: Cammalleri V, Pocino RN, Marotta D, et al. Occupational scenarios and exposure assessment to formaldehyde: A systematic review. *Indoor Air.* 2022;32:e12949. doi:10.1111/ina.12949