

## Breast and Prostate Cancer Risks for Male BRCA1 and BRCA2 Pathogenic Variant Carriers Using Polygenic Risk Scores

Daniel R. Barnes , PhD,<sup>1,\*†</sup> Valentina Silvestri , PhD,<sup>2,†</sup> Goska Leslie , MEng,<sup>1</sup> Lesley McGuffog,<sup>1</sup> Joe Dennis , MSc,<sup>1</sup> Xin Yang , PhD,<sup>1</sup> Julian Adlard , MMedSc, PhD,<sup>3</sup> Bjarni A. Agnarsson , MD,<sup>4,5</sup> Munaza Ahmed, MD(Res), FRCP,<sup>6</sup> Kristiina Aittomäki, MD, PhD,<sup>7</sup> Irene L. Andrulis , PhD,<sup>8,9</sup> Adalgeir Arason , BSc,<sup>4,10</sup> Norbert Arnold , PhD,<sup>11,12</sup> Bernd Auber , MD,<sup>13</sup> Jacopo Azzollini , MD,<sup>14</sup> Judith Balmaña, MD, PhD,<sup>15,16</sup> Rosa B. Barkardottir , CandSci,<sup>4,10</sup> Daniel Barrowdale , BSc,<sup>1</sup> Julian Barwell, MRCP, PhD,<sup>17</sup> Muriel Belotti, PhD,<sup>18</sup> Javier Benitez , PhD,<sup>19,20</sup> Pascaline Berthet, MD,<sup>21</sup> Susanne E. Boonen , MD, PhD,<sup>22</sup> Åke Borg, PhD,<sup>23</sup> Aniko Bozsik , PhD,<sup>24</sup> Angela F. Brady, FRCP, PhD,<sup>25</sup> Paul Brennan , MBBS, FRCP,<sup>26</sup> Carole Brewer, MD,<sup>27</sup> Joan Brunet , MD, PhD,<sup>28</sup> Agostino Bucalo , MSc,<sup>2</sup> Saundra S. Buys, MD,<sup>29</sup> Trinidad Caldés, MD,<sup>30</sup> Maria A. Caligo , PhD,<sup>31</sup> Ian Campbell , PhD,<sup>32,33</sup> Hayley Cassingham , MS, CGC,<sup>34</sup> Lise Lotte Christensen, MSc, PhD,<sup>35</sup> Giulia Cini , MSc,<sup>36</sup> Kathleen B.M. Claes , PhD,<sup>37</sup> GEMO Study Collaborators,<sup>38,39,40</sup> EMBRACE Collaborators,<sup>1</sup> Jackie Cook, MD,<sup>41</sup> Anna Coppa , MD,<sup>42,43</sup> Laura Cortesi , MD,<sup>43</sup> Giuseppe Damante, MD,<sup>44</sup> Esther Darder , BS,<sup>28</sup> Rosemarie Davidson, MD,<sup>45</sup> Miguel de la Hoya , PhD,<sup>30</sup> Kim De Leeneer, MD, PhD,<sup>37</sup> Robin de Putter , MD,<sup>37</sup> Jesús Del Valle , PhD,<sup>28</sup> Orland Diez, PhD,<sup>15,46</sup> Yuan Chun Ding, PhD,<sup>47</sup> Susan M. Domchek , MD,<sup>48</sup> Alan Donaldson , MD,<sup>49</sup> Jacqueline Eason , MD,<sup>50</sup> Ros Eeles, MD, PhD,<sup>51</sup> Christoph Engel , MD,<sup>52,53</sup> D. Gareth Evans, MD,<sup>54,55</sup> Lidia Feliubadaló, PhD,<sup>28</sup> Florentia Fostira , PhD,<sup>56</sup> Megan Frone , MS,<sup>57</sup> Debra Frost, ONC,<sup>1</sup> David Gallagher, MD,<sup>58</sup> Andrea Gehrig, MD,<sup>59</sup> Sophie Giraud, MD, PhD,<sup>60</sup> Gord Glendon , MSc,<sup>8</sup> Andrew K. Godwin , PhD,<sup>61</sup> David E. Goldgar, PhD,<sup>62</sup> Mark H. Greene , MD,<sup>57</sup> Helen Gregory, MBBS,<sup>63</sup> Eva Gross, PhD,<sup>64</sup> Eric Hahnen , PhD,<sup>65,66</sup> Ute Hamann, PhD,<sup>67</sup> Thomas V.O. Hansen, PhD,<sup>68</sup> Helen Hanson , MD, FRCP,<sup>69</sup> Julia Hentschel, PhD,<sup>70</sup> Judit Horvath, MD, PhD,<sup>71</sup> KConFab Investigators,<sup>32</sup> HEBON Investigators,<sup>72</sup> Louise Izatt , PhD,<sup>73</sup> Angel Izquierdo , MD, MPH,<sup>28</sup> Paul A. James , MBBS, PhD, FRACP, FRANZCR,<sup>33,74</sup> Ramunas Janavicius, MD, PhD,<sup>75,76</sup> Uffe Birk Jensen, MD, PhD,<sup>77</sup> Oskar Th. Johannsson, MD, PhD,<sup>78</sup> Esther M. John , PhD,<sup>79,80</sup> Gero Kramer, MD,<sup>81</sup> Lone Kroeldrup , MD,<sup>22</sup> Torben A. Kruse , PhD,<sup>22</sup> Charlotte Lautrup, MD, PhD,<sup>82,83</sup> Conxi Lazaro , PhD,<sup>28</sup> Fabienne Lesueur , PhD,<sup>39,40,84</sup> Adria Lopez-Fernández, MSc,<sup>15</sup> Phuong L. Mai, MD, MS,<sup>85</sup> Siranoush Manoukian, MD,<sup>14</sup> Zoltan Matrai , MD, PhD,<sup>86</sup> Laura Matricardi, PhD,<sup>87</sup> Kara N. Maxwell, MD,<sup>88</sup> Noura Mebirouk, PhD,<sup>39,40,84</sup> Alfons Meindl, PhD,<sup>64</sup> Marco Montagna , PhD,<sup>87</sup> Alvaro N. Monteiro , PhD,<sup>89</sup> Patrick J. Morrison , MD,<sup>90</sup> Taru A. Muranen , PhD,<sup>91</sup> Alex Murray, FRCP,<sup>92</sup> Katherine L. Nathanson , PhD,<sup>48</sup> Susan L. Neuhausen , PhD,<sup>47</sup> Heli Nevanlinna , PhD,<sup>91</sup> Tu Nguyen-Dumont , PhD,<sup>93,94</sup> Dieter Niederacher , PhD,<sup>95</sup> Edith Olah, PhD, DSc, HAS-fellow,<sup>24</sup> Olufunmilayo I. Olopade, MD<sup>96</sup> Domenico Palli , MD,<sup>97</sup> Michael T. Parsons , BSc,<sup>98</sup> Inge Sokilde Pedersen, PhD,<sup>83,99,100</sup> Bernard Peissel, MD,<sup>14</sup> Pedro Perez-Segura , MD,<sup>30</sup> Paolo Peterlongo , PhD,<sup>101</sup> Annabeth H. Petersen, MSc, PhD,<sup>102</sup> Pedro Pinto , MSc,<sup>103</sup> Mary E. Porteous, MD,<sup>104</sup> Caroline Pottinger , MD,<sup>92</sup> Miquel Angel Pujana, PhD,<sup>105</sup> Paolo Radice , PhD,<sup>106</sup> Juliane Ramser, PhD,<sup>107</sup> Johanna Rantala, PhD,<sup>108</sup> Mark Robson , MD,<sup>109</sup> Mark T. Rogers, MD,<sup>92</sup>

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Karina Rønlund, MD, PhD,<sup>102</sup> Andreas Rump , PhD,<sup>110</sup> Ana María Sánchez de Abajo, PhD,<sup>111</sup> Payal D. Shah, MD,<sup>88</sup> Saba Sharif , MD,<sup>112</sup> Lucy E. Side, MD,<sup>113</sup> Christian F. Singer, MD, MPH,<sup>114</sup> Zsofia Stadler, MD,<sup>109</sup> Linda Steele , MS,<sup>47</sup> Dominique Stoppa-Lyonnet , MD, PhD,<sup>18,38,115</sup> Christian Sutter, PhD,<sup>116</sup> Yen Yen Tan, PhD,<sup>117</sup> Manuel R. Teixeira , MD, PhD,<sup>103,118</sup> Alex Teulé, MD,<sup>28</sup> Darcy L. Thull , MS,<sup>119</sup> Marc Tischkowitz , MD, PhD,<sup>120,121</sup> Amanda E. Toland , PhD,<sup>122</sup> Stefania Tommasi , PhD,<sup>123</sup> Angela Toss , MD, PhD,<sup>43</sup> Alison H. Trainer, MBBS, PhD, FRACP,<sup>74,124</sup> Vishakha Tripathi , MBBS,<sup>73</sup> Virginia Valentini , PhD,<sup>2</sup> Christi J. van Asperen , MD, PhD,<sup>125</sup> Marta Venturelli , MD,<sup>43</sup> Alessandra Viel , PhD,<sup>36</sup> Joseph Vijai , PhD,<sup>109,126</sup> Lisa Walker, PhD,<sup>127</sup> Shan Wang-Gohrke, MD, PhD,<sup>128</sup> Barbara Wappenschmidt, PhD,<sup>65,66</sup> Anna Whaite , MSc,<sup>129</sup> Ines Zanna, PhD,<sup>97</sup> Kenneth Offit, MD, MPH,<sup>109,126</sup> Mads Thomassen, PhD,<sup>22</sup> Fergus J. Couch, PhD,<sup>130</sup> Rita K. Schmutzler, MD,<sup>65,66,131</sup> Jacques Simard , PhD,<sup>132</sup> Douglas F. Easton , PhD,<sup>1,133</sup> Georgia Chenevix-Trench , PhD,<sup>98</sup> Antonis C. Antoniou, PhD,<sup>1,†</sup> Laura Ottini , MD,<sup>2,\*†</sup> on behalf of the Consortium of Investigators of Modifiers of BRCA1 and BRCA2

<sup>1</sup>Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK; <sup>2</sup>Department of Molecular Medicine, Sapienza University of Rome, Rome, Italy; <sup>3</sup>Yorkshire Regional Genetics Service, Chapel Allerton Hospital, Leeds, UK; <sup>4</sup>Department of Pathology, Landspítali University Hospital, Reykjavik, Iceland; <sup>5</sup>School of Medicine, University of Iceland, Reykjavik, Iceland; <sup>6</sup>North East Thames Regional Genetics Service, Great Ormond Street Hospital for Children NHS Trust, London, UK; <sup>7</sup>Department of Clinical Genetics, Helsinki University Hospital, University of Helsinki, Helsinki, Finland; <sup>8</sup>Fred A. Litwin Center for Cancer Genetics, Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Toronto, ON, Canada; <sup>9</sup>Department of Molecular Genetics, University of Toronto, Toronto, ON, Canada; <sup>10</sup>BMC (Biomedical Centre), Faculty of Medicine, University of Iceland, Reykjavik, Iceland; <sup>11</sup>Department of Gynaecology and Obstetrics, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Kiel, Germany; <sup>12</sup>Institute of Clinical Molecular Biology, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Kiel, Germany; <sup>13</sup>Department of Human Genetics, Hannover Medical School, Hannover, Germany; <sup>14</sup>Unit of Medical Genetics, Department of Medical Oncology and Hematology, Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Milan, Italy; <sup>15</sup>Hereditary Cancer Genetics Group, Vall d'Hebron Institute of Oncology, Vall d'Hebron Hospital Campus, Barcelona, Spain; <sup>16</sup>Department of Medical Oncology, Vall d'Hebron Hospital Universitari, Vall d'Hebron Barcelona Hospital Campus, Barcelona, Spain; <sup>17</sup>Leicestershire Clinical Genetics Service, University Hospitals of Leicester NHS Trust, Leicester, UK; <sup>18</sup>Service de Génétique, Institut Curie, Paris, France; <sup>19</sup>Biomedical Network on Rare Diseases (CIBERER), Madrid, Spain; <sup>20</sup>Human Cancer Genetics Programme, Spanish National Cancer Research Centre (CNIO), Madrid, Spain; <sup>21</sup>Département de Biopathologie, Centre François Baclesse, Caen, France; <sup>22</sup>Department of Clinical Genetics, Odense University Hospital, Odense, Denmark; <sup>23</sup>Division of Oncology and Pathology, Department of Clinical Sciences Lund, Lund University, Lund, Sweden; <sup>24</sup>Department of Molecular Genetics, National Institute of Oncology, Budapest, Hungary; <sup>25</sup>North West Thames Regional Genetics Service, London North West University Healthcare NHS Trust, Northwick Park Hospital, Harrow, UK; <sup>26</sup>Northern Genetics Service, Newcastle Hospitals NHS Foundation Trust, Newcastle, UK; <sup>27</sup>Department of Clinical Genetics, Royal Devon & Exeter Hospital, Exeter, UK; <sup>28</sup>Hereditary Cancer Program, Oncobell-IDIBELL-IGTP, Catalan Institute of Oncology, CIBERONC, Barcelona, Spain; <sup>29</sup>Department of Internal Medicine, Huntsman Cancer Institute at the University of Utah, Salt Lake City, UT, USA; <sup>30</sup>Molecular Oncology Laboratory, CIBERONC, Hospital Clínico San Carlos, IdISSC (Instituto de Investigación Sanitaria del Hospital Clínico San Carlos), Madrid, Spain; <sup>31</sup>SOD Genetica Molecolare, University Hospital, Pisa, Italy; <sup>32</sup>Peter MacCallum Cancer Center, Melbourne, Victoria, Australia; <sup>33</sup>Sir Peter MacCallum Department of Oncology, The University of Melbourne, Melbourne, Victoria, Australia; <sup>34</sup>Department of Internal Medicine, Division of Human Genetics, The Ohio State University Wexner Medical Center, Columbus, OH, USA; <sup>35</sup>Division of Surgical Oncology, National Cancer Centre, Singapore; <sup>36</sup>Division of Functional Onco-Genomics and Genetics, Centro di Riferimento Oncologico di Aviano (CRO), IRCCS, Aviano, Italy; <sup>37</sup>Centre for Medical Genetics, Ghent University, Ghent, Belgium; <sup>38</sup>Department of Tumour Biology, INSERM U830, Paris, France; <sup>39</sup>Institut Curie, Paris, France; <sup>40</sup>Mines ParisTech, Fontainebleau, France; <sup>41</sup>Sheffield Clinical Genetics Service, Sheffield Children's Hospital, Sheffield, UK; <sup>42</sup>Department of Experimental Medicine, Sapienza University of Rome, Rome, Italy; <sup>43</sup>Department of Oncology and Haematology, University of Modena and Reggio Emilia, Modena, Italy; <sup>44</sup>Department of Medicine, University of Udine, Udine, Italy; <sup>45</sup>Department of Clinical Genetics, South Glasgow University Hospitals, Glasgow, UK; <sup>46</sup>Area of Clinical and Molecular Genetics, Vall d'Hebron Hospital Universitari, Vall d'Hebron Barcelona Hospital Campus, Barcelona, Spain; <sup>47</sup>Department of Population Sciences, Beckman Research Institute of City of Hope, Duarte, CA, USA; <sup>48</sup>Basser Center for BRCA, Abramson Cancer Center, University of Pennsylvania, Philadelphia, PA, USA; <sup>49</sup>Clinical Genetics Department, St Michael's Hospital, Bristol, UK; <sup>50</sup>Nottingham Clinical Genetics Service, Nottingham University Hospitals NHS Trust, Nottingham, UK; <sup>51</sup>Oncogenetics Team, The Institute of Cancer Research and Royal Marsden NHS Foundation Trust, London, UK; <sup>52</sup>Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, Leipzig, Germany; <sup>53</sup>LIFE—Leipzig Research Center for Civilization Diseases, University of Leipzig, Leipzig, Germany; <sup>54</sup>Division of Evolution and Genomic Sciences, School of Biological Sciences, Faculty of Biology, Medicine and Health, University of Manchester, Manchester Academic Health Science Centre, Manchester, UK; <sup>55</sup>North West Genomics Laboratory Hub, Manchester Centre for Genomic Medicine, St Mary's Hospital, Manchester University NHS Foundation Trust, Manchester Academic Health Science Centre, Manchester, UK; <sup>56</sup>Molecular Diagnostics Laboratory, INRASTES, National Centre for Scientific Research 'Demokritos', Athens, Greece; <sup>57</sup>Clinical Genetics Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, MD, USA; <sup>58</sup>Academic Unit of Clinical and Molecular Oncology, Trinity College Dublin and St James's Hospital, Dublin, Ireland; <sup>59</sup>Department of Human Genetics, University Würzburg, Würzburg, Germany; <sup>60</sup>Service de Génétique, Groupement Hospitalier Est, Hospices Civils de Lyon, Bron, France; <sup>61</sup>Department of Pathology and Laboratory Medicine, University of Kansas, Medical Center, Kansas City, KS, USA; <sup>62</sup>Department of Dermatology, Huntsman Cancer Institute, University of Utah School of Medicine, Salt Lake City, UT, USA; <sup>63</sup>North of Scotland Regional Genetics Service, NHS Grampian & University of Aberdeen, Foresterhill, Aberdeen, UK; <sup>64</sup>Department of Gynecology and Obstetrics, University of Munich, Munich, Germany; <sup>65</sup>Center for Familial Breast and Ovarian Cancer, Faculty of Medicine and University Hospital Cologne, University of Cologne, Cologne, Germany; <sup>66</sup>Center for Integrated Oncology (CIO), Faculty of Medicine and University Hospital Cologne, University of Cologne, Cologne, Germany; <sup>67</sup>Molecular Genetics of Breast Cancer, German Cancer Research Center (DKFZ), Heidelberg, Germany; <sup>68</sup>Department of Clinical Genetics, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark; <sup>69</sup>Southwest Thames Regional Genetics Service, St George's Hospital, London, UK; <sup>70</sup>Institute of Human Genetics, University Hospital Leipzig, Leipzig, Germany; <sup>71</sup>Institute of Human Genetics, University of Münster, Münster, Germany; <sup>72</sup>The Hereditary Breast and Ovarian Cancer Research Group Netherlands (HEBON), Coordinating Center: The Netherlands Cancer Institute, Amsterdam, The Netherlands; <sup>73</sup>Clinical Genetics, Guy's and St Thomas' NHS Foundation Trust, London, UK; <sup>74</sup>Parkville Familial Cancer Centre, Peter MacCallum Cancer Center, Melbourne, Victoria, Australia; <sup>75</sup>Faculty of Medicine, Institute of Biomedical Sciences, Department of Human and Medical Genetics, Vilnius University, Vilnius, Lithuania; <sup>76</sup>State Research Institute Centre for Innovative Medicine, Vilnius, Lithuania; <sup>77</sup>Department of Clinical Genetics, Aarhus University Hospital, Aarhus N, Denmark; <sup>78</sup>Department of Oncology, Landspítali University Hospital, Reykjavik, Iceland; <sup>79</sup>Department of Epidemiology & Population Health, Stanford University School of Medicine, Stanford, CA, USA; <sup>80</sup>Department of Medicine, Division of Oncology, Stanford Cancer Institute, Stanford University School of Medicine, Stanford, CA, USA; <sup>81</sup>Department of Urology, Medical University of Vienna, Vienna, Austria; <sup>82</sup>Department of Clinical Genetics, Aalborg University Hospital, Aalborg, Denmark; <sup>83</sup>Clinical Cancer Research Center, Aalborg University Hospital, Aalborg, Denmark; <sup>84</sup>Genetic Epidemiology of Cancer Team, Inserm U900, Paris, France; <sup>85</sup> Magee-Womens Hospital, University of Pittsburgh School of Medicine, Pittsburgh, PA, USA; <sup>86</sup>Department of Surgery, National Institute of Oncology, Budapest, Hungary; <sup>87</sup>Immunology and Molecular Oncology Unit, Veneto Institute of Oncology IOV—IRCCS, Padua, Italy; <sup>88</sup>Department of Medicine, Abramson Cancer Center, Perelman School of Medicine at the University of Pennsylvania, Philadelphia, PA, USA; <sup>89</sup>Department of Cancer Epidemiology, Moffitt Cancer Center, Tampa, FL, USA; <sup>90</sup>Northern Ireland Regional Genetics Centre, Belfast City Hospital, Belfast, UK; <sup>91</sup>Department of Obstetrics and Gynecology, Helsinki University Hospital, University of Helsinki, Helsinki, Finland; <sup>92</sup>All Wales Medical Genetics Services, University Hospital of Wales, Cardiff, UK; <sup>93</sup>Precision Medicine, School of Clinical Sciences at Monash Health, Monash University, Clayton, Victoria, Australia; <sup>94</sup>Department of Clinical Pathology, The University of Melbourne, Melbourne, Victoria, Australia; <sup>95</sup>Department of

Gynecology and Obstetrics, University Hospital Düsseldorf, Heinrich-Heine University Düsseldorf, Düsseldorf, Germany; <sup>96</sup>Center for Clinical Cancer Genetics, The University of Chicago, Chicago, IL, USA; <sup>97</sup>Cancer Risk Factors and Life-Style Epidemiology Unit, Institute for Cancer Research, Prevention and Clinical Network (ISPRO), Florence, Italy; <sup>98</sup>Department of Genetics and Computational Biology, QIMR Berghofer Medical Research Institute, Brisbane, Queensland, Australia; <sup>99</sup>Molecular Diagnostics, Aalborg University Hospital, Aalborg, Denmark; <sup>100</sup>Department of Clinical Medicine, Aalborg University, Aalborg, Denmark; <sup>101</sup>Genome Diagnostics Program, IFOM—the FIRCC Institute of Molecular Oncology, Milan, Italy; <sup>102</sup>Department of Clinical Genetics, Vejle Hospital, Vejle, Denmark; <sup>103</sup>Department of Genetics, Portuguese Oncology Institute, Porto, Portugal; <sup>104</sup>South East of Scotland Regional Genetics Service, Western General Hospital, Edinburgh, UK; <sup>105</sup>Translational Research Laboratory, IDIBELL (Bellvitge Biomedical Research Institute), Catalan Institute of Oncology, CIBERONC, Barcelona, Spain; <sup>106</sup>Unit of Molecular Bases of Genetic Risk and Genetic Testing, Department of Research, Fondazione IRCCS Istituto Nazionale dei Tumori (INT), Milan, Italy; <sup>107</sup>Division of Gynaecology and Obstetrics, Klinikum rechts der Isar der Technischen Universität München, Munich, Germany; <sup>108</sup>Clinical Genetics, Karolinska Institutet, Stockholm, Sweden; <sup>109</sup>Clinical Genetics Service, Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, NY, USA; <sup>110</sup>Institute for Clinical Genetics, Faculty of Medicine Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany; <sup>111</sup>Servicio de Análisis Clínicos y Bioquímica Clínica, Complejo Hospitalario Universitario Insular Materno-Infantil de Gran Canaria, Las Palmas de Gran Canaria, Spain; <sup>112</sup>West Midlands Regional Genetics Service, Birmingham Women's Hospital Healthcare NHS Trust, Birmingham, UK; <sup>113</sup>Princess Anne Hospital, Southampton, UK; <sup>114</sup>Department of OB/GYN and Comprehensive Cancer Center, Medical University of Vienna, Vienna, Austria; <sup>115</sup>Université Paris Descartes, Paris, France; <sup>116</sup>Institute of Human Genetics, University Hospital Heidelberg, Heidelberg, Germany; <sup>117</sup>Dept of OB/GYN, Medical University of Vienna, Vienna, Austria; <sup>118</sup>Biomedical Sciences Institute (ICBAS), University of Porto, Porto, Portugal; <sup>119</sup>Department of Medicine, Magee-Womens Hospital, University of Pittsburgh School of Medicine, Pittsburgh, PA, USA; <sup>120</sup>Program in Cancer Genetics, Departments of Human Genetics and Oncology, McGill University, Montréal, QC, Canada; <sup>121</sup>Department of Medical Genetics, University of Cambridge, Cambridge, UK; <sup>122</sup>Department of Cancer Biology and Genetics, The Ohio State University, Columbus, OH, USA; <sup>123</sup>IRCCS Istituto Tumori Giovanni Paolo II, Bari, Italy; <sup>124</sup>Department of Medicine, University of Melbourne, Melbourne, Victoria, Australia; <sup>125</sup>Department of Clinical Genetics, Leiden University Medical Center, Leiden, The Netherlands; <sup>126</sup>Clinical Genetics Research Lab, Department of Cancer Biology and Genetics, Memorial Sloan Kettering Cancer Center, New York, NY, USA; <sup>127</sup>Oxford Regional Genetics Service, Churchill Hospital, Oxford, UK; <sup>128</sup>Department of Gynaecology and Obstetrics, University Hospital Ulm, Ulm, Germany; <sup>129</sup>Liverpool Centre for Genomic Medicine, Liverpool Women's NHS Foundation Trust, Liverpool, UK; <sup>130</sup>Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA; <sup>131</sup>Center for Molecular Medicine Cologne (CMMC), Faculty of Medicine and University Hospital Cologne, University of Cologne, Cologne, Germany; <sup>132</sup>Genomics Center, Centre Hospitalier Universitaire de Québec—Université Laval Research Center, Québec City, QC, Canada and <sup>133</sup>Centre for Cancer Genetic Epidemiology, Department of Oncology, University of Cambridge, Cambridge, UK

†Authors contributed equally as joint senior authors.

‡Authors contributed equally as joint first authors.

\*Correspondence to: Daniel R. Barnes, PhD, Centre for Cancer Genetic Epidemiology, Strangeways Research Laboratory, 2 Worts Causeway, Cambridge, CB1 8RN, UK (e-mail: drb54@medschl.cam.ac.uk) and Laura Ottini, MD, Department of Molecular Medicine, Sapienza University of Rome, Viale Regina Elena, 324, 00161, Rome, Italy (e-mail: laura.ottini@uniroma1.it).

## Abstract

**Background:** Recent population-based female breast cancer and prostate cancer polygenic risk scores (PRS) have been developed. We assessed the associations of these PRS with breast and prostate cancer risks for male *BRCA1* and *BRCA2* pathogenic variant carriers. **Methods:** 483 *BRCA1* and 1318 *BRCA2* European ancestry male carriers were available from the Consortium of Investigators of Modifiers of *BRCA1/2* (CIMBA). A 147-single nucleotide polymorphism (SNP) prostate cancer PRS (PRS<sub>PC</sub>) and a 313-SNP breast cancer PRS were evaluated. There were 3 versions of the breast cancer PRS, optimized to predict overall (PRS<sub>BC</sub>), estrogen receptor (ER)-negative (PRS<sub>ER-</sub>), or ER-positive (PRS<sub>ER+</sub>) breast cancer risk. **Results:** PRS<sub>ER+</sub> yielded the strongest association with breast cancer risk. The odds ratios (ORs) per PRS<sub>ER+</sub> standard deviation estimates were 1.40 (95% confidence interval [CI] = 1.07 to 1.83) for *BRCA1* and 1.33 (95% CI = 1.16 to 1.52) for *BRCA2* carriers. PRS<sub>PC</sub> was associated with prostate cancer risk for *BRCA1* (OR = 1.73, 95% CI = 1.28 to 2.33) and *BRCA2* (OR = 1.60, 95% CI = 1.34 to 1.91) carriers. The estimated breast cancer odds ratios were larger after adjusting for female relative breast cancer family history. By age 85 years, for *BRCA2* carriers, the breast cancer risk varied from 7.7% to 18.4% and prostate cancer risk from 34.1% to 87.6% between the 5th and 95th percentiles of the PRS distributions. **Conclusions:** Population-based prostate and female breast cancer PRS are associated with a wide range of absolute breast and prostate cancer risks for male *BRCA1* and *BRCA2* carriers. These findings warrant further investigation aimed at providing personalized cancer risks for male carriers and informing clinical management.

*BRCA1* and *BRCA2* pathogenic variants are associated with increased male breast cancer and prostate cancer risks (1–4). A recent prospective study estimated the lifetime risk of developing prostate cancer to be 29% for *BRCA1* and 60% for *BRCA2* carriers (5). The risks of developing male breast cancer compared with the general population have been estimated to be 15- to 18-fold higher for *BRCA1* and 80-fold higher for *BRCA2* carriers (6,7). Up to 1 in 10 male *BRCA2* carriers develops breast cancer (8–12) and displays potentially more aggressive disease relative to sporadic cases (8,12,13).

Polygenic risk scores (PRS) that combine the effects of multiple disease-associated single nucleotide polymorphisms (SNPs) provide marked cancer risk stratification in the general population (14,15) and *BRCA1* and *BRCA2* carriers (16–18). Our previous findings suggested the joint effects of PRS and *BRCA1* and *BRCA2* pathogenic variants may identify men at clinically meaningful breast and prostate cancer risk levels (17). Recent studies have identified

additional breast and prostate cancer susceptibility variants (15,19,20) and have refined PRS for these cancers (15,21).

The Breast Cancer Association Consortium recently developed and validated a 313-SNP PRS in European ancestry women, which was further optimized to predict estrogen receptor (ER)-specific disease (21). The estimated per standard deviation odds ratio (OR) for the most predictive (ER-positive) PRS was 1.68 (95% confidence interval [CI] = 1.63 to 1.73) (21). A recent evaluation of this PRS in unselected male breast cancer cases showed similar associations with breast cancer risk in men (22). The most recent prostate cancer PRS was developed using 147-SNPs associated with prostate cancer risk in European-ancestry men from the general population (15). The estimated per standard deviation odds ratio for the prostate cancer PRS was 1.86 (95% CI = 1.83 to 1.89) (15).

Male *BRCA1* and *BRCA2* carriers are likely to benefit from more personalized breast and prostate cancer risk estimates

(23). Investigating the extent to which these PRS modify cancer risks may lead to more precise and gender-specific cancer risk assessment and could assist in optimizing cancer screening.

Here, we assessed the associations of the newly developed 313-SNP breast cancer PRS and 147-SNP prostate cancer PRS derived using population-based data, with breast and prostate cancer risks, respectively, for male *BRCA1* and *BRCA2* carriers. We investigated whether cancer family history influences the associations and if breast cancer associations differed by ER status or tumor grade. Furthermore, we assessed whether associations vary by age or *BRCA1* and *BRCA2* pathogenic variant characteristics (location; functional effect). We used the results to estimate age-specific absolute risks of developing breast and prostate cancers for male carriers by PRS distribution percentiles.

## Methods

Statistical analyses were performed using R-3.6.3 (R Foundation for Statistical Computing, Vienna, Austria) (commands can be found in the [Supplementary Methods](#), available online).

### Study Participants and Genotyping

Male *BRCA1* and *BRCA2* pathogenic variant carriers were recruited through 40 studies from 19 countries participating in the Consortium of Investigators of Modifiers of *BRCA1* and *BRCA2* (CIMBA) (24). The majority of male carriers were ascertained through families attending cancer genetic clinics (96.9%; [Supplementary Tables 1 and 2](#), available online). In this setting, individuals are referred to clinical genetics because of strong family or personal cancer history. The first individual in a family, screened for mutations, tends to be an affected individual diagnosed at a young age, most often a female relative with a young age at breast cancer diagnosis (24). When a pathogenic variant is identified, then other family members are tested for the same variant. All participants were aged 18 years or older and provided written informed consent. All studies were approved by local ethical review committees. A total of 1989 male *BRCA1* and *BRCA2* carriers of European ancestry were included in the present study, by selecting all available men with a breast or prostate cancer diagnosis and matched controls. Details of matching, genotyping, and quality control processes have been described previously (17) and in [Supplementary Table 2](#) (available online).

Data collected included breast or prostate cancer diagnoses; age at diagnosis or interview; prostate cancer Gleason score; breast cancer ER status and grade; and family history of prostate, male breast, and female breast cancers among first- and second-degree relatives. *BRCA1* and *BRCA2* pathogenic variants (detailed pathogenicity description: [http://cimba.ccg.med.schl.cam.ac.uk/files/CIMBA\\_Mutation\\_Classification\\_guidelines\\_May16.pdf](http://cimba.ccg.med.schl.cam.ac.uk/files/CIMBA_Mutation_Classification_guidelines_May16.pdf)) were categorized according to their known or predicted effect on protein function: class I included loss-of-function variants expected to yield unstable or no protein; class II included variants likely to produce stable mutant proteins (25). Pathology data were obtained from pathology reviews; medical, pathology or tumor registry records; or immunohistochemical staining of tissue microarrays (26).

### Polygenic Risk Scores

PRS were constructed as the weighted sums of alleles ([Supplementary Methods](#), available online) for 313-SNPs for

breast cancer (21) and 147-SNPs for prostate cancer (15) ([Supplementary Tables 3 and 4](#), available online). Three breast cancer PRS were evaluated, optimized to predict overall (PRS<sub>BC</sub>), ER-negative (PRS<sub>ER-</sub>), and ER-positive (PRS<sub>ER+</sub>) breast cancer (21). These PRS were scaled to the female population-based control PRS standard deviation (21). The prostate cancer PRS (PRS<sub>PC</sub>) was scaled to the standard deviation calculated from population-based controls (15).

### Associations Between PRS and Cancer Risks

PRS associations with breast and prostate cancer risks were assessed simultaneously using multinomial logistic regression to estimate per standard deviation odds ratios. Men without breast or prostate cancer diagnoses were considered controls. Breast and prostate cancer cases were defined by considering the first occurring cancer. Instances in which breast and prostate cancers were diagnosed simultaneously were considered as breast cancer cases. Statistical models were adjusted for 3 ancestry informative principal components (proxy adjustment for study and/or country, as a direct adjustment would result in too few controls and cases within each study and/or country; [Supplementary Table 1](#), available online) and age. Models using the combined sample of carriers were adjusted for *BRCA1* and *BRCA2* status. To account for relatedness, we estimated robust variances by clustering on family membership (27,28). The primary analyses assumed a continuous PRS. Categorical PRS associations were evaluated using the quartiles of the PRS distributions in the combined *BRCA1* and *BRCA2* carrier controls.

Because the distribution of tumor ER status in male carriers may differ from the distributions in the general population (26), we assessed the associations between all 3 versions of the breast cancer PRS with overall breast cancer risk and ER-specific disease. Associations with ER-positive and ER-negative breast cancer were assessed simultaneously by considering ER negative, ER positive, or unknown as distinct multinomial outcomes. We also assessed the associations with breast cancer grade-specific risk by considering grade 1, grade 2, grade 3, or unknown grade as separate multinomial outcomes. A case-only logistic regression also was undertaken that considered grades 1 and 2 as controls and grade 3 as cases.

To assess the PRS<sub>PC</sub> association with disease aggressiveness, we partitioned prostate cancers into those with Gleason scores less than 7, 7 or greater, or unknown, and these were used as distinct multinomial outcomes. A case-only logistic regression assessed differences in the associations with Gleason scores less than 7 (controls) and Gleason scores of 7 or greater (cases).

Discriminatory ability of each PRS was assessed by calculating the area under the receiver operator characteristic curve (AUC). Under the sampling design, the majority of male carriers were identified through clinical genetics. Therefore, the majority of both affected and unaffected carriers are expected to have family history of cancer. To determine whether this introduces any biases in the PRS associations, we fitted models that were adjusted for family history in first- and second-degree relatives.

To determine whether PRS associations varied by age (continuous), pathogenic variant location, or pathogenic variant effects on protein function (class I or class II variants), we estimated interaction terms between these factors with the PRS, and statistical significance was assessed using likelihood ratio tests (LRT). Pathogenic variants were categorized based on previously reported nucleotide position differences in breast and ovarian, or prostate cancer risks (29–31).

We undertook a sensitivity analysis to test for PRS heterogeneity across study countries ([Supplementary Methods](#), available online).

All statistical tests were 2-sided, and a *P* value of less than .05 was considered statistically significant.

### Predicted Age-Specific Absolute and 10-Year Cancer Risks by PRS

We predicted absolute risks up to age 85 years and 10-year risks of developing breast and prostate cancers by PRS distribution percentiles, assuming the estimated PRS odds ratio follows a log-linear model across the entire PRS range ([Supplementary Methods](#), available online) (32).

## Results

### Study Participants and Genotyping

After quality control, the analyses included 483 BRCA1 (33 breast and 70 prostate cancer cases) and 1318 BRCA2 (244 breast and 141 prostate cancer cases) carriers of European ancestry ([Supplementary Tables 1 and 2](#), available online).

All SNPs from both PRS were well imputed ( $r^2 \geq 0.76$ ; [Supplementary Tables 3 and 4](#), [Supplementary Figures 1 and 2](#), available online). Average PRS were larger for cases compared with controls ([Supplementary Table 2](#), available online).

### Associations With Breast Cancer Risk

The associations between the breast cancer PRS and male breast cancer risk for carriers are shown in [Table 1](#) and [Supplementary Tables 5 and 6](#) (available online). The PRS<sub>ER+</sub> yielded the strongest associations with overall breast cancer risk for BRCA1 (OR = 1.40, 95% CI = 1.07 to 1.83) and BRCA2 (OR = 1.33, 95% CI = 1.16 to 1.52) carriers. The PRS<sub>BC</sub> resulted in nearly identical associations as the PRS<sub>ER+</sub>. There was no statistically significant evidence that the PRS<sub>ER+</sub> associations differed by country ( $P_{\text{heterogeneity}} \geq .48$ ; [Supplementary Figure 3](#), available online). In the joint analysis of BRCA1 and BRCA2 carriers, men in the uppermost PRS<sub>ER+</sub> quartile had approximately twofold increased breast cancer risk (OR = 2.10, 95% CI = 1.43 to 3.08) compared with men in the lowest quartile ([Supplementary Table 6](#), available online).

Most breast cancers among the male carriers were ER positive (95.7%). The odds ratio for the association between the PRS<sub>ER+</sub> and ER-positive breast cancer risk for BRCA1 carriers (OR = 1.79, 95% CI = 1.30 to 2.48; [Table 1](#)) was somewhat higher compared with the odds ratio for overall breast cancer. The number of ER-negative cancers was too small to assess associations with ER-negative disease. There was no statistically significant evidence for differences in the associations of any of the PRS by grade ([Table 1](#); [Supplementary Table 6](#), available online).

The ability of PRS<sub>ER+</sub> to discriminate between controls and breast cancer cases was estimated as an AUC of 0.60 (95% CI = 0.51 to 0.69) for BRCA1 and 0.59 (95% CI = 0.55 to 0.63) for BRCA2 carriers.

### Associations With Prostate Cancer Risk

The estimated associations between the PRS<sub>PC</sub> and prostate cancer risk for male carriers are reported in [Table 2](#) and

[Supplementary Tables 5 and 7](#) (available online). The odds ratios per PRS<sub>PC</sub> standard deviation were estimated to be 1.73 (95% CI = 1.28 to 2.33) for BRCA1 and 1.60 (95% CI = 1.34 to 1.91) for BRCA2 carriers. There was no statistically significant evidence that the PRS<sub>PC</sub> associations differed by country ( $P_{\text{heterogeneity}} \geq .14$ ; [Supplementary Figure 4](#), available online). In the joint analysis of BRCA1 and BRCA2 carriers, men in the top PRS<sub>PC</sub> quartile had a prostate cancer odds ratio of 3.35 (95% CI = 2.06 to 5.42) compared with men in the lowest quartile ([Supplementary Table 7](#), available online).

There was a suggestion of higher risk for aggressive disease for BRCA1 carriers (Gleason score  $\geq 7$ : OR = 2.09, 95% CI = 1.27 to 3.46; Gleason score  $< 7$ : OR = 1.11, 95% CI = 0.70 to 1.77), also supported by the case-only analysis (OR = 1.87, 95% CI = 1.01 to 3.44;  $P = .05$ ; [Table 2](#)). There were no differences in the PRS<sub>PC</sub> associations with high- or low-Gleason score among BRCA2 carriers ([Table 2](#)).

The PRS<sub>PC</sub> discriminatory ability was estimated as an AUC of 0.62 (95% CI = 0.54 to 0.69) for BRCA1 and 0.62 (95% CI = 0.57 to 0.67) for BRCA2 carriers.

### Adjusting for Cancer Family History

Adjusting for family history of male breast cancer did not influence the PRS<sub>ER+</sub> associations with breast cancer risk ([Table 1](#); [Supplementary Table 8](#), available online). However, the odds ratio estimates were somewhat larger when adjusting for female breast cancer family history ([Table 1](#); [Supplementary Table 9](#), available online).

The associations of PRS<sub>PC</sub> with prostate cancer risk remained similar after adjusting for prostate cancer family history ([Table 2](#); [Supplementary Table 10](#), available online).

### PRS Interactions With Age and Gene Pathogenic Variants Characteristics

There was little evidence for odds ratio estimate variability with age, for both the breast and prostate cancer PRS ( $P_{\text{LRT}} \geq .43$ ; [Table 3](#)).

The PRS<sub>ER+</sub> and PRS<sub>PC</sub> odds ratios with breast or prostate cancer risks appeared to be larger for class II variant (pathogenic variants likely to yield stable mutant proteins) carriers compared with class I BRCA1 and BRCA2 variant carriers ([Table 3](#)). However, these differences were not statistically significant ( $P_{\text{LRT}} \geq .26$ ).

There was no statistically significant evidence that the PRS<sub>ER+</sub> ( $P_{\text{LRT}} \geq .61$ ) or PRS<sub>PC</sub> ( $P_{\text{LRT}} = .52$ ) associations differed by the pathogenic variant location in the gene ([Table 3](#)).

### Absolute Risks of Developing Breast and Prostate Cancer

The absolute risks of developing breast cancer by age 85 years for BRCA2 carriers was predicted to be 7.7% at the 5th and 18.4% at the 95th PRS<sub>ER+</sub> distribution percentiles ([Figure 1](#)). The 10-year risks of developing breast cancer at 50 years were 0.8% at the 5th and 2.0% at the 95th PRS<sub>ER+</sub> distribution percentiles for BRCA2 carriers ([Figure 2](#)). The corresponding risks at age 75 years were 3.7% and 9.3%, respectively.

The predicted absolute risks of developing prostate cancer by age 85 years were 13.1% at the 5th and 50.4% at the 95th PRS<sub>PC</sub> distribution percentiles for BRCA1 carriers ([Figure 1](#)). The corresponding risks for BRCA2 carriers were 34.1% and 87.6%. BRCA2 carriers had 10-year risks of 2.1% and 10.1% at

**Table 1.** Breast cancer PRS associations with breast cancer risk for BRCA1 and BRCA2 carriers

PRS investigated and outcome	BRCA1 carriers				BRCA2 carriers			
	No. of controls	No. of cases	OR (95% CI)	P <sup>a</sup>	No. of controls	No. of cases	OR (95% CI)	P <sup>a</sup>
<b>PRS<sub>BC</sub></b>								
PRS <sub>BC</sub> association with breast cancer risk								
Continuous <sup>b</sup>	380	33	1.40 (1.06 to 1.85)	.02	933	244	1.32 (1.15 to 1.52)	<.001
Continuous: adjusted for male relative breast cancer FH <sup>c</sup>	380	33	1.39 (1.05 to 1.84)	.02	933	244	1.33 (1.15 to 1.52)	<.001
Continuous: adjusted for female relative breast cancer FH <sup>c</sup>	380	33	1.44 (1.07 to 1.95)	.02	933	244	1.36 (1.18 to 1.57)	<.001
PRS <sub>BC</sub> association with grade-specific breast cancer risk <sup>d</sup>								
Controls	380	—	1.00 (referent)		933	—	1.00 (referent)	
Grade 1	—	1	1.03 (0.63 to 1.67) <sup>g</sup>	.92	—	11	1.33 (0.74 to 2.36)	.34
Grade 2	—	6			—	68	1.29 (1.04 to 1.60)	.02
Grade 3	—	12	1.56 (1.03 to 2.37)	.04	—	98	1.23 (1.00 to 1.50)	.05
Grade unknown	—	14	1.47 (0.93 to 2.32)	.10	—	67	1.51 (1.18 to 1.93)	.001
Case-only: grade 1 + 2 vs grade 3 <sup>e</sup>	7	12	6.30 (0.88 to 44.87)	.07	79	98	0.95 (0.71 to 1.27)	.73
<b>PRS<sub>ER-</sub></b>								
PRS <sub>ER-</sub> association with breast cancer risk								
Continuous <sup>b</sup>	380	33	1.12 (0.79 to 1.59)	.52	933	244	1.23 (1.07 to 1.41)	.004
Continuous: adjusted for male relative breast cancer FH <sup>c</sup>	380	33	1.12 (0.79 to 1.59)	.53	933	244	1.23 (1.07 to 1.42)	.004
Continuous: adjusted for female relative breast cancer FH <sup>c</sup>	380	33	1.14 (0.80 to 1.63)	.48	933	244	1.25 (1.09 to 1.45)	.002
PRS <sub>ER-</sub> association with ER-specific breast cancer risk <sup>f</sup>								
Controls	380	—	1.00 (referent)		933	—	1.00 (referent)	
ER negative	—	2	0.38 (0.06 to 2.29)	.29	—	7	0.51 (0.27 to 0.98)	.04
ER positive	—	21	1.47 (0.97 to 2.24)	.07	—	178	1.26 (1.08 to 1.47)	.004
ER status unknown	—	10	0.78 (0.46 to 1.30)	.34	—	59	1.24 (0.94 to 1.64)	.13
<b>PRS<sub>ER+</sub></b>								
PRS <sub>ER+</sub> association with breast cancer risk								
Continuous <sup>b</sup>	380	33	1.40 (1.07 to 1.83)	.01	933	244	1.33 (1.16 to 1.52)	<.001
Continuous: adjusted for male relative breast cancer FH <sup>c</sup>	380	33	1.39 (1.06 to 1.82)	.02	933	244	1.33 (1.16 to 1.53)	<.001
Continuous: adjusted for female relative breast cancer FH <sup>c</sup>	380	33	1.46 (1.09 to 1.94)	.01	933	244	1.36 (1.18 to 1.57)	<.001
PRS <sub>ER+</sub> association with ER-specific breast cancer risk <sup>f</sup>								
Controls	380	—	1.00 (referent)		933	—	1.00 (referent)	
ER negative	—	2	0.35 (0.03 to 3.59)	.37	—	7	0.68 (0.38 to 1.22)	.20
ER positive	—	21	1.79 (1.30 to 2.48)	<.001	—	178	1.30 (1.11 to 1.52)	<.001
ER status unknown	—	10	1.00 (0.68 to 1.47)	1.00	—	59	1.52 (1.18 to 1.94)	.001
PRS <sub>ER+</sub> association with grade-specific breast cancer risk <sup>d</sup>								
Controls	380	—	1.00 (referent)		933	—	1.00 (referent)	
Grade 1	—	1	1.03 (0.65 to 1.65) <sup>g</sup>	.89	—	11	1.31 (0.76 to 2.27)	.34
Grade 2	—	6			—	68	1.29 (1.05 to 1.59)	.02
Grade 3	—	12	1.51 (1.04 to 2.19)	.03	—	98	1.23 (1.01 to 1.51)	.04
Grade unknown	—	14	1.51 (0.96 to 2.38)	.07	—	67	1.51 (1.19 to 1.92)	<.001
Case-only: grade 1 + 2 vs grade 3 <sup>e</sup>	7	12	5.41 (0.79 to 37.20)	.09	79	98	0.95 (0.71 to 1.28)	.75

<sup>a</sup>P value was calculated using a 2-sided Wald test. CI = confidence interval; ER = estrogen receptor; FH = family history; OR = odds ratio per PRS standard deviation, estimated from a multinomial logistic regression (unless otherwise stated); PRS = polygenic risk scores PRS<sub>BC</sub> = overall breast cancer PRS; PRS<sub>ER-</sub> = ER-negative breast cancer PRS; PRS<sub>ER+</sub> = ER-positive breast cancer PRS.

<sup>b</sup>The continuous test shows the per PRS standard deviation associations, estimated from a multinomial logistic regression model assuming a continuous PRS.

<sup>c</sup>Association estimates adjusted for family history of (male and female) breast cancer in first- and second-degree relatives. FH was coded as no family history, 1 or more relatives diagnosed with breast cancer, unknown FH or missing FH. [Supplementary Table 8](#) (available online; male breast cancer FH adjusted) and [Supplementary Table 9](#) (available online; female breast cancer FH adjusted) describe the breast cancer FH adjusted analyses in greater detail.

<sup>d</sup>The breast cancer grade specific odds ratios were estimated by partitioning breast cancer status into multinomial outcomes for grade 1, grade 2, grade 3, or grade unknown.

<sup>e</sup>The case-only breast cancer grade analysis was a logistic regression considering grade 1 and grade 2 breast cancers combined as controls and grade 3 breast cancers as cases.

<sup>f</sup>The ER-specific breast cancer odds ratios were estimated by partitioning breast cancer status into distinct multinomial outcomes for ER negative, ER positive, or ER status unknown.

<sup>g</sup>Grade 1 and grade 2 combined for BRCA1 carriers (to ensure adequate sample size to estimate associations).

the 5th and 95th PRS<sub>PC</sub> percentiles at age 50 years, respectively. The corresponding risks at age 75 years were 25.5% and 77.0% ([Figure 2](#)).

## Discussion

We evaluated the associations of the most recently developed breast and prostate cancer PRS with site-specific cancer risks in

the largest case-control study of male BRCA1 and BRCA2 carriers available to date. Our findings showed that these PRS, developed using population-based data, are associated with breast and prostate cancer risks for male BRCA1 and BRCA2 carriers. Despite the modest estimated AUCs, our results demonstrate that because male carriers are already at elevated risks of developing breast and prostate cancers, these PRS can lead to large differences in the absolute cancer risks for carriers across PRS percentiles.

**Table 2.** Prostate cancer PRS associations with prostate cancer risk for BRCA1 and BRCA2 carriers

PRS investigated and outcome	BRCA1 carriers				BRCA2 carriers			
	No. of controls	No. of cases	OR (95% CI)	P <sup>a</sup>	No. of controls	No. of cases	OR (95% CI)	P <sup>a</sup>
Continuous <sup>b</sup>	380	70	1.73 (1.28 to 2.33)	<.001	933	141	1.60 (1.34 to 1.91)	<.001
Continuous: adjusted for FH <sup>c</sup>	380	70	1.74 (1.29 to 2.35)	<.001	933	141	1.59 (1.32 to 1.90)	<.001
PRS <sub>PC</sub> association with Gleason score (GS)-specific prostate cancer risk <sup>d</sup>								
Controls	380	—	1.00 (referent)		933	—	1.00 (referent)	
GS < 7	—	26	1.11 (0.70 to 1.77)	.66	—	27	1.83 (1.29 to 2.58)	<.001
GS ≥ 7	—	21	2.09 (1.27 to 3.46)	.004	—	82	1.68 (1.32 to 2.13)	<.001
GS unknown	—	23	2.38 (1.49 to 3.80)	<.001	—	32	1.26 (0.95 to 1.68)	.11
Case-only analysis: GS ≥ 7 vs GS < 7 <sup>e</sup>	26	21	1.87 (1.01 to 3.44)	.05	27	82	0.93 (0.63 to 1.37)	.72

<sup>a</sup>P value was calculated using a 2-sided Wald test. CI = confidence interval; GS = Gleason score; FH = family history; OR = odds ratio per PRS standard deviation, estimated from a multinomial logistic regression (unless otherwise stated); PRS = polygenic risk scores; PRS<sub>PC</sub> = prostate cancer PRS.

<sup>b</sup>The continuous test shows the per PRS standard deviation associations, estimated from a multinomial logistic regression model assuming a continuous PRS.

<sup>c</sup>Association estimates adjusted for family history of prostate cancer in first- and second-degree relatives. FH was coded as no family history, 1 or more diagnosed relatives, unknown FH, or missing FH. [Supplementary Table 10](#) (available online) describes the prostate cancer FH adjusted analyses in greater detail.

<sup>d</sup>The Gleason score prostate cancer odds ratios were estimated by partitioning prostate cancer status into distinct multinomial outcomes for GS < 7, GS ≥ 7, or GS unknown.

<sup>e</sup>The case-only prostate cancer analysis was a logistic regression considering GS < 7 prostate cancers as “controls” and GS ≥ 7 prostate cancers as “cases”.

PRS<sub>BC</sub> and PRS<sub>ER+</sub> were associated with larger odds ratio estimates than PRS<sub>ER-</sub> in predicting breast cancer risk, consistent with the fact that most breast cancers in men are ER positive, including those harboring BRCA1 and BRCA2 pathogenic variants (26). Similarly, when assessing associations with ER-positive breast cancer risk, PRS<sub>BC</sub> and PRS<sub>ER+</sub> showed the strongest associations for BRCA1 and BRCA2 carriers. There were no differences in PRS associations by breast cancer grade.

The 147-SNP PRS<sub>PC</sub> (15) yielded larger per standard deviation odds ratio estimates than a previously evaluated 103-SNP prostate cancer PRS (17). There was some evidence that PRS<sub>PC</sub> may be associated with a higher odds ratio for more aggressive disease (Gleason score ≥ 7) for BRCA1 carriers. This pattern was not observed for BRCA2 carriers, who tend to develop more aggressive disease (5). If this finding is replicated by larger studies, the PRS may prove to be useful in cancer prevention and surveillance by identifying BRCA1 carriers at greater risk of developing aggressive prostate cancers.

PRS associations with breast or prostate cancer risk, adjusted for family history of male breast or prostate cancer, were similar to unadjusted estimates, suggesting that cancer family history in male relatives does not alter PRS associations. Adjusting for family history of female breast cancer resulted in somewhat larger odds ratio estimates for the breast cancer PRS compared with unadjusted estimates. This observation is consistent with male carriers being identified and recruited into our studies mostly based on their female relatives' breast cancers.

There was little evidence supporting variability in PRS associations by age or pathogenic variant characteristics. However, larger sample sizes are required to reliably assess such differences, and the current analyses were likely underpowered.

Previous studies (18,33) suggest the magnitude of the breast cancer PRS associations is attenuated in female BRCA1 and BRCA2 carriers compared with associations seen in the general population (21). As seen for female carriers, the estimated breast cancer odds ratios for male carriers were attenuated compared with estimates for women in the general population (21). Similarly, the estimated prostate cancer odds ratio estimate for male carriers was attenuated compared with

population-based data (15). Taken together, these observations suggest there is a deviation from the multiplicative model for the joint effects of BRCA1 and BRCA2 pathogenic variants and the PRS for male and female carriers. These observed attenuations for BRCA1 and BRCA2 carriers are unlikely to be an overestimation of the effects in the general population [“winner's curse” (34)], as they have been validated in independent prospective cohorts (21). The lower odds ratios for the breast and prostate cancer PRS in male BRCA1 and BRCA2 carriers, compared with the general population, may reflect a general attenuation of the effect sizes of common variants on genetic risk in the presence of a pathogenic variant in a high-risk gene (35,36). This supposition may also explain the larger PRS odds ratios for BRCA1 carriers, who are at lower risk compared with BRCA2 carriers (37). However, given the current study design, we cannot rule out that the observed attenuations in effect size are related to ascertainment biases. Although adjusting for family history did not change the odds ratio estimates substantially, residual confounding may still remain. Large-scale population studies will be required to address this. If the attenuations in the PRS effect size are real, they would result in a smaller range of cancer risks for BRCA1 and BRCA2 carriers compared with using the PRS effect sizes estimated from general population data.

Although breast cancer risk stratification might not currently be feasible for men in the general population, male BRCA1 and BRCA2 carriers may represent a group likely to benefit from a more refined stratification of their individual breast and prostate cancer risks, to better inform their clinical management. At present, limited recommendations based on low-level evidence or expert opinion are available for male carriers. Current guidelines recommend clinical breast examinations beginning at ages 30–35 years and suggest mammographic screening on an individual basis, whereas clinical prostate cancer screening, particularly for BRCA2 carriers, is recommended from ages 40 to 45 years (38–40).

The PRS percentile-specific absolute risks varied substantially over the PRS distribution, consistent with previous studies in male (17) and female (16,18) BRCA1 and BRCA2 carriers. At least twofold increased risk is often considered a clinically

**Table 3.** PRS interactions with age and BRCA1 and BRCA2 pathogenic variant characteristics for BRCA1 and BRCA2 carriers with breast cancer risk and prostate cancer risk.

Model and category	Breast cancer (PRS <sub>ER+</sub> ) <sup>a</sup>				Prostate cancer (PRS <sub>PC</sub> )			
	BRCA1 carriers		BRCA2 carriers		BRCA1 carriers		BRCA2 carriers	
	OR (95% CI)	p <sup>b</sup>	OR (95% CI)	p <sup>b</sup>	OR (95% CI)	p <sup>b</sup>	OR (95% CI)	p <sup>b</sup>
<b>PRS x age interaction<sup>c</sup></b>								
PRS	1.88 (0.68 to 5.18)	.22	1.34 (0.71 to 2.53)	.37	0.64 (0.20 to 2.04)	.45	2.03 (0.91 to 4.52)	.08
PRS x age	1.00 (0.98 to 1.01)	.56	1.00 (0.99 to 1.01)	.98	1.02 (1.00 to 1.03)	.09	1.00 (0.98 to 1.01)	.55
P <sub>LRT</sub> <sup>d</sup>		.90		.86		.43		.79
<b>Gene pathogenic variant class<sup>e</sup></b>								
Class I	1.38 (1.03 to 1.84)	.03	1.31 (1.13 to 1.52)	<.001	1.57 (1.13 to 2.19)	.008	1.57 (1.31 to 1.89)	<.001
Class II	1.71 (0.72 to 4.07)	.23	1.39 (0.67 to 2.86)	.38	3.00 (1.36 to 6.60)	.006	2.04 (0.63 to 6.55)	.23
P <sub>LRT</sub> <sup>d</sup>		.76		.69		.26		.97
<b>BRCA1 pathogenic variant location (OCCR)</b>								
5' to c.2281	1.50 (1.00 to 2.26)	.05	NA		NA		NA	
c.2282 to c.4071	1.17 (0.79 to 1.72)	.44	NA		NA		NA	
c.4072 to 3'	1.61 (0.87 to 2.98)	.13	NA		NA		NA	
P <sub>LRT</sub> <sup>d</sup>		.85						
<b>BRCA2 pathogenic variant location (OCCR)</b>								
5' to c.2830	NA		1.43 (1.09 to 1.88)	.009	NA		NA	
c.2831 to c.6401	NA		1.24 (0.99 to 1.55)	.06	NA		NA	
c.6402 to 3'	NA		1.33 (1.04 to 1.70)	.02	NA		NA	
P <sub>LRT</sub> <sup>d</sup>				.61				
<b>BRCA2 pathogenic variant location (PCCR)</b>								
5' to c.755	NA		NA		NA		1.67 (1.06 to 2.62)	.03
c.756 to c.1000	NA		NA		NA		1.77 (1.07 to 2.95)	.03
c.1001 to c.7913	NA		NA		NA		1.49 (1.18 to 1.89)	<.001
c.7914 to 3'	NA		NA		NA		1.76 (1.24 to 2.50)	.002
P <sub>LRT</sub> <sup>d</sup>								.52

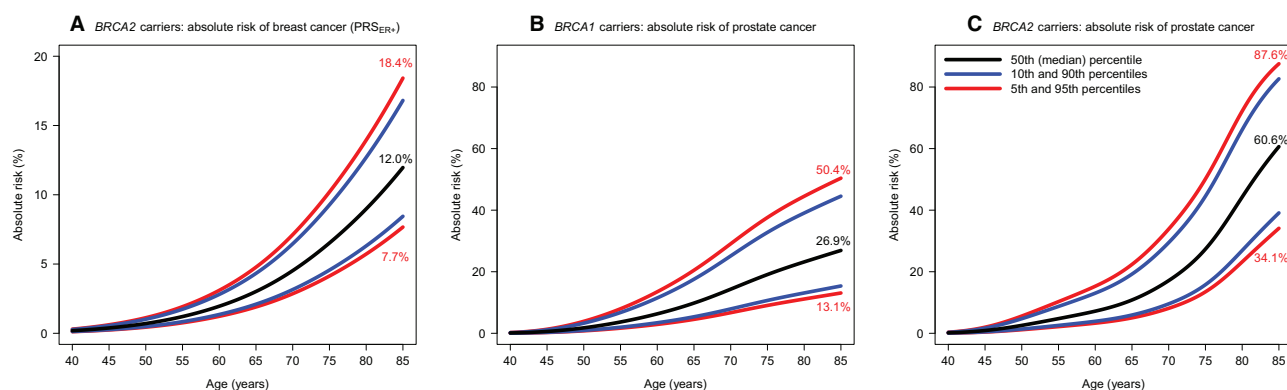
<sup>a</sup>The associations with breast cancer risk are reported for the ER-positive breast cancer PRS (PRS<sub>ER+</sub>). CI = confidence interval; OCCR = ovarian cancer cluster region; OR = odds ratio per PRS standard deviation, estimated from a multinomial logistic regression; PCCR = prostate cancer cluster region; PRS = polygenic risk score; NA = not applicable.

<sup>b</sup>P value was calculated using a 2-sided Wald test, unless otherwise indicated.

<sup>c</sup>The PRS term is applicable at age 0 years and the PRS x age interaction term is a per-year effect. Age in years.

<sup>d</sup>P values were calculated using a 2-sided likelihood ratio test. The likelihood ratio test compared the model that estimated the interaction term with a nested model that omitted the interaction term.

<sup>e</sup>Class I pathogenic variant = loss-of-function pathogenic variants expected to result in unstable or no protein; class II pathogenic variant = pathogenic variants likely to yield stable mutant proteins.

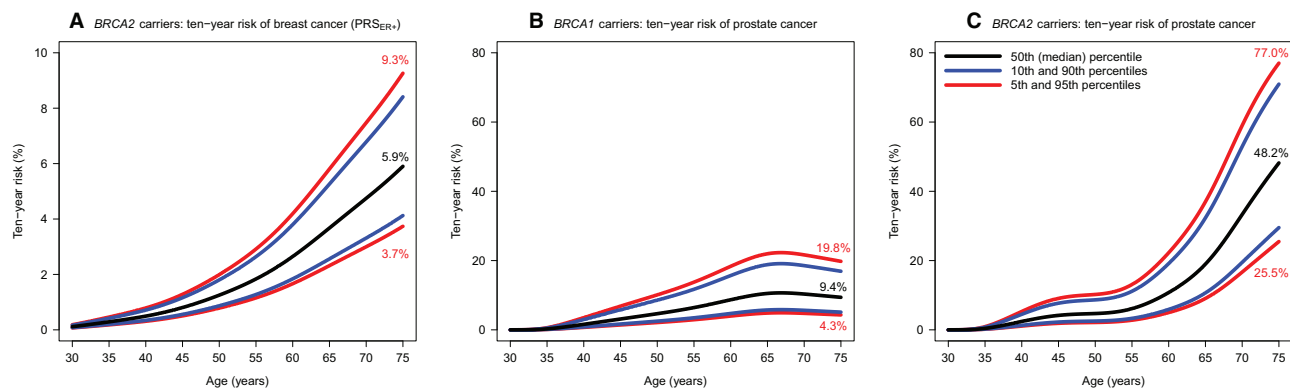


**Figure 1.** The predicted absolute risks of developing breast cancer and prostate cancer by PRS percentile. Risks were calculated assuming the per standard deviation ratio estimates in the combined sample of BRCA1 and BRCA2 carriers (Supplementary Tables 6 and 7). (A) The absolute risks of developing breast cancer for BRCA2 carriers by PRS<sub>ER+</sub> percentiles. (B) The absolute risks of developing prostate cancer for BRCA1 carriers by PRS<sub>PC</sub> percentiles. (C) The absolute risks of developing prostate cancer for BRCA2 carriers by PRS<sub>PC</sub> percentiles. PRS = polygenic risk scores; PRS<sub>ER+</sub> = ER-positive breast cancer PRS.

actionable level for breast and prostate cancers (41). Our findings may inform the development of age-specific clinical recommendations and provide guidance on when to start risk-adapted screening, based on their PRS percentile-specific 10-

year risks. Overall, refined risk estimates may be useful to distinguish male carriers at higher risk, who may benefit from enhanced and/or earlier screening, and identify carriers at lower risk, who may opt for more limited or postponed surveillance.





**Figure 2.** The predicted 10-year risks of developing breast cancer and prostate cancer by PRS percentile. Ten-year risks were calculated from the absolute risks of developing breast cancer or prostate cancer (Figure 1). (A) The 10-year risks of developing breast cancer for BRCA2 carriers by PRS<sub>ER+</sub> percentiles. (B) The 10-year risks of developing prostate cancer for BRCA1 pathogenic variant carriers by PRS<sub>PC</sub> percentiles. (C) The 10-year risks of developing prostate cancer for BRCA2 pathogenic variant carriers by PRS<sub>PC</sub> percentiles. PRS = polygenic risk scores PRS<sub>ER+</sub> = ER-positive breast cancer PRS.

Identification of men at lower risk of prostate cancer by PRS stratification has been shown to be useful in reducing overdiagnosis in the general population, resulting in a reduction in the harms associated with prostate-specific antigen testing (42). Similar arguments may apply to male carriers in whom PRS prediction may further improve screening efficacy.

Strengths of this study include the fact that this is an independent validation of the most recently derived breast (21) and prostate (15) cancer PRS derived from population-based data. We benefited from the availability of Gleason scores and breast cancer ER status and grade; hence, we could assess subtype-specific associations. Finally, we assumed recent prospectively estimated prostate cancer incidence rates (5) to predict absolute prostate cancer risks, which may be more representative of risks for carriers currently seen in clinical genetics centers.

Study limitations include the limited sample size to assess PRS associations with cancer risks for subgroups of male carriers. However, these data remain the largest male BRCA1 and BRCA2 carrier case-control study with available genotype data. The breast (21) and prostate (15) cancer PRS do not include male breast cancer-specific risk-associated SNPs or SNPs that may specifically be associated with prostate cancer risk for carriers. If such SNPs exist, further improvement may be gained in risk prediction by including them in PRS. The absolute risk calculations assumed that the PRS odds ratio behaves log linearly over the PRS range. It was difficult to evaluate this assumption in the present analyses because of the limited sample size of male carriers. However, empirical evidence based on larger sample sizes of female carriers (18) or in the general population (15,21) suggests that this assumption is plausible. Additionally, the absolute breast and prostate cancer risk predictions by PRS will require validation in large prospective studies of male carriers with long-term follow-up, although such studies remain a challenge. Finally, the PRS that we investigated were derived using European ancestry data; hence, our estimated associations and predicted risks may not be applicable to non-European ancestry carriers.

PRS are now used in cancer risk-stratified screening trials and implementation studies in the general population (43–47). They are commercially available and are used in multifactorial cancer-risk prediction models for women (48,49). We found that PRS derived from population-based data are associated with breast and prostate cancer risks and lead to meaningful risk stratification for male carriers. These findings may potentially

be used to provide more personalized cancer risk predictions and therefore assist clinical management decisions. Future implementation studies should determine if optimal strategies exist for incorporating these PRS into genetic counseling and risk assessment to clarify whether they can influence the clinical management decisions of male BRCA1 or BRCA2 carriers.

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## Data Availability

The complete dataset is not publicly available because of restraints imposed by the ethical committees of individual studies. Requests to access the complete dataset, which is subject to General Data Protection Regulation (GDPR) rules, can be made to the Data Access Coordinating Committee (DACC) of CIMBA, following the process described on the CIMBA website (<http://cimba.ccge.medschl.cam.ac.uk/projects/data-access-requests/>). Submitted applications are reviewed by the CIMBA DACC every 3 months.

## References

- Castro E, Eeles R. The role of BRCA1 and BRCA2 in prostate cancer. *Asian J Androl*. 2012;14(3):409–414.
- Rizzolo P, Silvestri V, Tommasi S, et al. Male breast cancer: genetics, epigenetics, and ethical aspects. *Ann Oncol*. 2013;24(suppl 8):viii75–viii82.
- Leongamornlert D, Mahmud N, Tymrakiewicz M, et al.; for the UKGPCS Collaborators. Germline BRCA1 mutations increase prostate cancer risk. *Br J Cancer*. 2012;106(10):1697–1701.
- Kote-Jarai Z, Leongamornlert D, Saunders E, et al.; for the UKGPCS Collaborators. BRCA2 is a moderate penetrance gene contributing to young-onset prostate cancer: implications for genetic testing in prostate cancer patients. *Br J Cancer*. 2011;105(8):1230–1234.
- Nyberg T, Frost D, Barrowdale D, et al. Prostate cancer risks for male BRCA1 and BRCA2 mutation carriers: a prospective cohort study. *Eur Urol*. 2020;77(1):24–35.
- Liede A, Karlan BY, Narod SA. Cancer risks for male carriers of germline mutations in BRCA1 or BRCA2: a review of the literature. *J Clin Oncol*. 2004;22(4):735–742.
- Weiss JR, Moysich KB, Swede H. Epidemiology of male breast cancer. *Cancer Epidemiol Biomarkers Prev*. 2005;14(1):20–26.
- Basham VM, Lipscombe JM, Ward JM, et al. BRCA1 and BRCA2 mutations in a population-based study of male breast cancer. *Breast Cancer Res*. 2002;4(1):R2.
- Ottini L, Masala G, D'Amico C, et al. BRCA1 and BRCA2 mutation status and tumor characteristics in male breast cancer: a population-based study in Italy. *Cancer Res*. 2003;63(2):342–347.
- Easton DF, Steele L, Fields P, et al. Cancer risks in two large breast cancer families linked to BRCA2 on chromosome 13q12–13. *Am J Hum Genet*. 1997;61(1):120–128.
- Thompson D, Easton D. Breast cancer linkage C. variation in cancer risks, by mutation position, in BRCA2 mutation carriers. *Am J Hum Genet*. 2001;68(2):410–419.
- Ferzoco RM, Ruddy KJ. The epidemiology of male breast cancer. *Curr Oncol Rep*. 2016;18(1):1.
- Kwiatkowska E, Teresiak M, Filas V, Karczewska A, Breborowicz D, Mackiewicz A. BRCA2 mutations and androgen receptor expression as independent predictors of outcome of male breast cancer patients. *Clin Cancer Res*. 2003;9(12):4452–4459.
- Mavaddat N, Pharoah PD, Michailidou K, et al. Prediction of breast cancer risk based on profiling with common genetic variants. *J Natl Cancer Inst*. 2015;107(5):dju036.
- Schumacher FR, Al Olama AA, Berndt SI, et al.; for the Genetic Associations and Mechanisms in Oncology (GAME-ON)/Elucidating Loci Involved in Prostate Cancer Susceptibility (ELLIPSE) Consortium. Association analyses of more than 140,000 men identify 63 new prostate cancer susceptibility loci. *Nat Genet*. 2018;50(7):928–936.
- Kuchenbaecker KB, McGuffog L, Barrowdale D, et al. Evaluation of polygenic risk scores for breast and ovarian cancer risk prediction in BRCA1 and BRCA2 mutation carriers. *J Natl Cancer Inst*. 2017;109(7):djw302.
- Lecarpentier J, Silvestri V, Kuchenbaecker KB, et al.; for the KConFab Investigators. Prediction of breast and prostate cancer risks in male BRCA1 and BRCA2 mutation carriers using polygenic risk scores. *J Clin Oncol*. 2017;35(20):2240–2250.
- Barnes DR, Rookus MA, McGuffog L, et al.; for the Consortium of Investigators of Modifiers of BRCA1 and BRCA2. Polygenic risk scores and breast and epithelial ovarian cancer risks for carriers of BRCA1 and BRCA2 pathogenic variants. *Genet Med*. 2020;22(10):1653–1666.
- Michailidou K, Lindstrom S, Dennis J, et al.; for the ConFab/AOCS Investigators. Association analysis identifies 65 new breast cancer risk loci. *Nature*. 2017;551(7678):92–94.
- Milne RL, Kuchenbaecker KB, Michailidou K, et al.; for the ABCTB Investigators. Identification of ten variants associated with risk of estrogen-receptor-negative breast cancer. *Nat Genet*. 2017;49(12):1767–1778.
- Mavaddat N, Michailidou K, Dennis J, et al.; for the NBCS Collaborators. Polygenic risk scores for prediction of breast cancer and breast cancer subtypes. *Am J Hum Genet*. 2019;104(1):21–34.
- Maguire S, Perraki E, Tomczyk K, et al. Common susceptibility loci for male breast cancer. *J Natl Cancer Inst*. 2020;113(4):453–61.
- Gaddam S, Heller SL, Babb JS, Gao Y. Male breast cancer risk assessment and screening recommendations in high-risk men who undergo genetic counseling and multigene panel testing. *Clin Breast Cancer*. 2021;21(1):74–79.
- Chenevix-Trench G, Milne RL, Antoniou AC, et al.; for CIMBA. An international initiative to identify genetic modifiers of cancer risk in BRCA1 and BRCA2 mutation carriers: the Consortium of Investigators of Modifiers of BRCA1 and BRCA2 (CIMBA). *Breast Cancer Res*. 2007;9(2):104.
- Antoniou AC, Sinilnikova OM, Simard J, et al.; for the Consortium of Investigators of Modifiers of BRCA1/2 (CIMBA). RAD51 135G→C modifies breast cancer risk among BRCA2 mutation carriers: results from a combined analysis of 19 studies. *Am J Hum Genet*. 2007;81(6):1186–1200.
- Silvestri V, Barrowdale D, Mulligan AM, et al.; for EMBRACE. Male breast cancer in BRCA1 and BRCA2 mutation carriers: pathology data from the Consortium of Investigators of Modifiers of BRCA1/2. *Breast Cancer Res*. 2016;18(1):15.
- Huber PJ. The behavior of maximum likelihood estimates under nonstandard conditions. Paper presented at Proceedings of the Fifth Berkeley Symposium on Mathematical Statistics and Probability, Statistical Laboratory of the University of California, Berkeley, California, USA (June 21–July 18, 1965 and December 27, 1965–January 7, 1966); 1967.
- White H. A heteroskedasticity-consistent covariance matrix estimator and a direct test for heteroskedasticity. *Econometrica*. 1980;48(4):817–838.
- Kuchenbaecker KB, Hopper JL, Barnes DR, et al.; and the BRCA1 and BRCA2 Cohort Consortium. Risks of breast, ovarian, and contralateral breast cancer for BRCA1 and BRCA2 mutation carriers. *JAMA*. 2017;317(23):2402–2416.
- Rebbeck TR, Mitra N, Wan F, et al.; for the CIMBA Consortium. Association of type and location of BRCA1 and BRCA2 mutations with risk of breast and ovarian cancer. *JAMA*. 2015;313(13):1347–1361.
- Patel VL, Busch EL, Friebe TM, et al. Association of genomic domains in BRCA1 and BRCA2 with prostate cancer risk and aggressiveness. *Cancer Res*. 2020;80(3):624–638.
- Antoniou AC, Beesley J, McGuffog L, et al.; for CIMBA. Common breast cancer susceptibility alleles and the risk of breast cancer for BRCA1 and BRCA2 mutation carriers: implications for risk prediction. *Cancer Res*. 2010;70(23):9742–9754.
- Gallagher S, Hughes E, Wagner S, et al. Association of a polygenic risk score with breast cancer among women carriers of high- and moderate-risk breast cancer genes. *JAMA Netw Open*. 2020;3(7):e208501.
- Xiao R, Boehnke M. Quantifying and correcting for the winner's curse in genetic association studies. *Genet Epidemiol*. 2009;33(5):453–462.
- Sawyer S, Mitchell G, McKinley J, et al. A role for common genomic variants in the assessment of familial breast cancer. *J Clin Oncol*. 2012;30(35):4330–4336.
- Coignard J, Lush M, Beesley J, et al.; for the ABCTB Investigators. A case-only study to identify genetic modifiers of breast cancer risk for BRCA1/BRCA2 mutation carriers. *Nat Commun*. 2021;12(1):1078.
- Silvestri V, Leslie G, Barnes DR, et al.; and the CIMBA Group. Characterization of the cancer spectrum in men with germline BRCA1 and BRCA2 pathogenic variants: results from the Consortium of Investigators of Modifiers of BRCA1/2 (CIMBA). *JAMA Oncol*. 2020;6(8):1218.
- Daly MB, Pal T, Berry MP, et al. Genetic/Familial High-Risk Assessment: Breast, Ovarian, and Pancreatic, Version 2.2021, NCCN Clinical Practice Guidelines in Oncology. *J Natl Compr Canc Netw*. 2021;19(1):77–102.
- Paluch-Shimon S, Cardoso F, Sessa C, et al.; for the ESMO Guidelines Committee. Prevention and screening in BRCA mutation carriers and other breast/ovarian hereditary cancer syndromes: ESMO Clinical Practice Guidelines for cancer prevention and screening. *Ann Oncol*. 2016;27(suppl 5):v103–v110.
- American society of Clinical Oncology. Hereditary breast and ovarian cancer guidelines: screening for men with a BRCA1 or BRCA2 gene mutation; 2017. <https://www.cancer.net/cancer-types/hereditary-breast-and-ovarian-cancer>. Accessed September 30, 2019.
- Jia G, Lu Y, Wen W, et al. Evaluating the utility of polygenic risk scores in identifying high-risk individuals for eight common cancers. *JNCI Cancer Spectr*. 2020;4(3):pkaa021.

42. Toland AE. Polygenic risk scores for prostate cancer: testing considerations. *Can J Urol*. 2019;26(5, suppl 2):17–18.
43. Eeles RA, Raghallaigh H; for the Group TBS. BARCODE 1: a pilot study investigating the use of genetic profiling to identify men in the general population with the highest risk of prostate cancer to invite for targeted screening. *J Clin Oncol*. 2020;38(suppl 15):1505–1505.
44. Institute of Cancer Research. BARCODE 1: The use of genetic profiling to guide prostate cancer targeted screening; 2020. <http://www.icr.ac.uk/our-research/research-divisions/division-of-genetics-and-epidemiology/oncogenetics/research-projects/barcode-1>. Accessed October 21, 2020.
45. Castro E, Mikropoulos C, Bancroft EK, et al.; for the PROFILE Study Steering Committee. The PROFILE feasibility study: targeted screening of men with a family history of prostate cancer. *Oncologist*. 2016;21(6):716–722.
46. Institute of Cancer Research. The PROFILE Study: germline genetic profiling: correlation with targeted prostate cancer screening and treatment; 2020. <http://www.icr.ac.uk/our-research/research-divisions/division-of-genetics-and-epidemiology/oncogenetics/research-projects/profile>. Accessed October 21, 2020.
47. Pashayan N, Antoniou AC, Ivanus U, et al. Personalized early detection and prevention of breast cancer: ENVISION consensus statement. *Nat Rev Clin Oncol*. 2020;17(11):687–705.
48. Lee A, Mavaddat N, Wilcox AN, et al. BOADICEA: a comprehensive breast cancer risk prediction model incorporating genetic and nongenetic risk factors. *Genet Med*. 2019;21(8):1708–1718.
49. IBIS. IBIS breast cancer risk evaluation tool; 2017. <http://www.ems-trials.org/riskevaluator/>. Accessed May 21, 2020.