

Research Article

Histological *Ex Vivo* Evaluation of the Suitability of a 976 nm Diode Laser in Oral Soft Tissue Biopsies

Gaspare Palaia,¹ Federico Renzi,¹ Daniele Pergolini ,¹ Alessandro Del Vecchio,¹ Paolo Visca,² Gianluca Tenore,¹ and Umberto Romeo¹

¹“Sapienza” University of Rome, Department of Oral and Maxillofacial Sciences, Via Caserta 6, Rome 00161, Italy

²Department of Cytology and Cellular Diagnostics, Regina Elena Institute, Via Elio Chianesi 53, Rome 00144, Italy

Correspondence should be addressed to Daniele Pergolini; daniele.pergolini@uniroma1.it

Received 29 November 2020; Revised 24 March 2021; Accepted 19 April 2021; Published 28 April 2021

Academic Editor: Roberta Gasparro

Copyright © 2021 Gaspare Palaia et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction. Laser-induced thermal effects can preclude a safe histological evaluation of biopsy resection margins. The aim of this study was to evaluate the suitability of a 976 nm diode laser in oral soft tissue biopsies in an *ex vivo* study. **Materials and Methods.** A 976 nm diode laser (Solase®, Lazon Medical Laser, China) has been used in the contact mode, using a 400 μm fiber tip, at different parameters from 4 to 6 W in the continuous wave (CW), with a fluence between 3184 and 4777 J/cm², and pulsed wave (PW) mode, with a fluence between 318,4 and 477,7 J/cm², to obtain 30 samples from fresh pig cadaver tongues. All specimens were subdivided into 6 groups (from A to F), and each group consisted of 5 samples. Two sections were obtained from each sample. A histological analysis was performed using an optical microscope at magnifications of 5x and 10x. Statistical analysis was carried out using Kruskal–Wallis and Dunn’s tests. **Results.** The results showed that histological readability was optimal in all the samples. The thermal damage was negligible in all groups. The average thermal damage was 208.40 ± 133.81 μm in the epithelial tissue and 330.14 ± 147.45 μm in the connective tissue. The statistical analysis showed no differences between the groups ($p > 0.05$). **Conclusion.** A 976 nm diode laser demonstrated good surgical effectiveness that provoked little peripheral damage in the cut edges and allowed a safe histological diagnosis. **Clinical Relevance.** In oral pathology, many times, there is fear in using the laser to remove some lesions due to its thermal effect on the tissues close to the lesion. This effect is always present in the use of the laser, but the intent is to minimize this effect to have as little alteration as possible on the surrounding tissues.

1. Introduction

Due to their various advantages, lasers have been recently introduced into dental practice, and they have applications in many fields of dentistry [1, 2]. The most used lasers are the diode (600–980 nm), potassium titanyl phosphate (KTP, 532 nm), carbon dioxide (CO₂, 10,600 nm), neodymium-doped yttrium aluminium garnet (Nd:YAG, 1064 nm), and erbium-doped yttrium aluminium garnet (Er:YAG, 2940 nm) lasers [3].

As regards the diode laser, it is a versatile tool that finds a wide range of applications such as in soft tissue surgery, periodontology [4], endodontics [5], teeth whitening, and photobiostimulation (PBM) [6].

This kind of laser emits a beam of light that selectively interacts with tissue chromophores such as haemoglobin and melanin to transform light energy into thermal energy. At the point of incidence of the laser beam, the temperature exceeds 100°C, which vaporizes the tissue and generates a haemostatic cut [7, 8]. In this way, it is possible to safely create access incisions (operculectomy, exposure of impacted teeth, and access for cystic lesions), gingivectomies, gingivoplasties, frenectomies, and fornix elongations.

As regards nonsurgical periodontal treatment, the diode laser showed good results if combined with conventional scaling and root planing (SRP), reducing the probing pocket depth, increasing the clinical attachment level, and decreasing the red complex bacteria [9].

Even other oral diseases, such as osteonecrosis of the jaws [10], burning mouth syndrome [11, 12], temporomandibular joint disorder [6], and vascular lesions [13] can be managed using lasers.

However, there is still a debate regarding the possibility to use lasers to make biopsies, and this is because of the surrounding area of the cut in which the laser-induced overheating may create histological artifacts on the resection margins that, especially for clinically suspected lesions, could interfere with the histological reading of the sample, making it difficult to make a clear and safe anatomopathological diagnosis and casting doubts about the true effectiveness of laser biopsies [14, 15].

According to the actual literature, it is always possible to make a diagnosis when performing laser biopsies, but to determine the real extension of the sick tissue, a good operator's skill and knowledge in laser surgery are required [16–18].

Our group has previously tested many other wavelengths that showed encouraging results; the aim of this study was to evaluate the histological effects of a 976 nm diode laser (a wavelength recently introduced on the market) and also to determine the exact extension of the laser-produced histological alterations in order to establish whether it is possible or not to make safe biopsies with such a kind of laser and the clinical protocol to follow to minimize the risk of unreadable samples.

2. Materials and Methods

This study was performed *ex vivo* in six swine tongues obtained from animals that had died less than 24 hours before. Pig tongue was chosen because of its similar histological and physiological structures as the human one [7]. A diode laser with a wavelength of 976 nm (Solase®, Lazon Medical Laser, China) and a 400 μm fiber tip in the contact mode was used for this purpose. This laser operates with a frequency of up to 50 kHz, a pulse length between 10 μs and 0.9 s, and a maximum power of 16 W.

Thirty specimens were taken from the tongue dorsum by the same expert operator. They were divided into six groups (A, B, C, D, E, and F) that corresponded to different settings that have been used (Table 1). Groups A, B, and C were treated using a continuous wave (CW) modality at 4 W with a fluence of 3184 J/cm², 5 W with a fluence of 3980 J/cm², and 6 W with a fluence of 4777 J/cm², respectively. In contrast, groups D, E, and F were treated with a pulsed wave (PW) mode at 4 W with a fluence of 318,4 J/cm², 5 W with a fluence of 398 J/cm², and 6 W with a fluence of 477,7 J/cm² ($t_{\text{on}} - t_{\text{off}}$: 100 ms–100 ms), respectively. All the samples were fixed in a 10% formalin solution.

Afterwards, 2 sections for each specimen, for a total of 60 sections, were stained with haematoxylin and eosin for histological evaluation performed by a pathologist who was blinded to the purpose of the study (single-blinded study) and measured in μm the extension of the laser-induced thermal effects. The histological analysis was performed using an optical microscope (Leica DM2000) at a magnification of 5x and 10x; the width of thermal damage in the

TABLE 1: The laser parameters used in the study.

Samples' number	Power (W)	Group	Mode
5	4	A	CW
5	5	B	
5	6	C	
5	4	D	PW ($t_{\text{on}} - t_{\text{off}}$ 100 ms)
5	5	E	
5	6	F	

peri-incisional epithelial and connective tissue was measured using Leica Suite 3.4 software.

Finally, a statistical analysis was carried out with the Kruskal–Wallis and multiple comparison Dunn's test using GraphPad Prism 8.4.2 software to understand whether there were statistically significant differences between the groups.

3. Results

Different thermal effects emerged in the epithelial and connective tissue layers; the epithelium showed vacuolization, cell coarctation, and carbonization, while collagen homogenization and carbonization were noted in the connective tissue (Figure 1). In the epithelium, the damage extension was always smaller in size, ranging from 46.679 μm to 689.575 μm , compared to the connective tissue, which showed histological alterations from 99.425 μm to 964.24 μm ($p < 0.01$). All samples demonstrated clear and well-readable cut margins with only a small damaged area (Table 2 and Figures 2–7).

The highest damage amounts recorded in this study were 689.575 μm in the epithelium in a sample taken at 6 W in the PW mode and 964.24 μm in the connective tissue in a sample taken at 5 W in the CW mode. No statistically significant differences were found among the groups according to the Kruskal–Wallis and Dunn's tests ($p > 0.05$).

4. Discussion

Laser-assisted soft tissue surgery has many advantages over traditional soft tissue surgery for both the operator and the patient. The cut generated by the laser fiber generates hemostasis, because of the affinity of the wavelength for haemoglobin, and contemporary disinfection of the surgical wound due to the laser's photothermal effects. The benefits for the patient include a faster postoperative course, with a faster healing process and less intra- and postoperative pain. In addition, suture is not necessary in many occasions, and this is particularly useful when approaching areas such as the soft palate, where, by healing the tissue for the second intention, the risk of anatomical distortion is minimized [19–22].

However, in the particular case of biopsies, the possibility of using the laser is debated because of the laser's photothermal effects that produce histological artifacts in the marginal portion of the sample that may interfere with the reading of the transition portion between the sick and healthy tissue, and this aspect is critically important in excisional biopsies of clinically malignant lesions [14, 23].

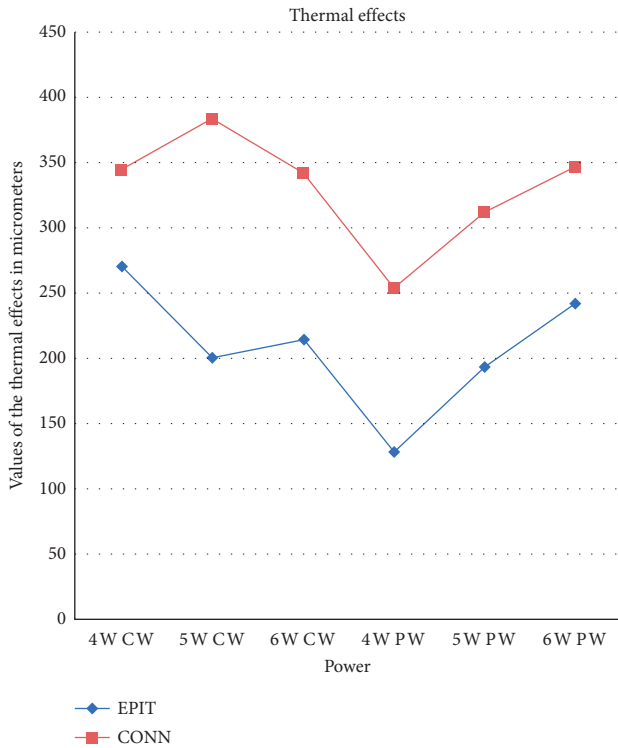


FIGURE 1: A line chart showing the average values in μm obtained in the epithelium and connective tissue.

TABLE 2: The mean peri-incisional effects.

Average thermal effects	
Average epithelial peri-incisional effects of all samples	$0.2 \pm 0.13 \text{ mm}$
Average connectival peri-incisional effects of all samples	$0.3 \pm 0.14 \text{ mm}$
Average epithelial peri-incisional effects of PW samples	$0.1 \pm 0.14 \text{ mm}$
Average connectival peri-incisional effects of PW samples	$0.3 \pm 0.13 \text{ mm}$
Average epithelial peri-incisional effects of CW samples	$0.2 \pm 0.13 \text{ mm}$
Average connectival peri-incisional effects of CW samples	$0.3 \pm 0.16 \text{ mm}$

Over time, studies have been carried out to quantify the portion of tissue damaged by the heat produced by the laser both in *in vivo* and *ex vivo* conditions, using different kinds of lasers. Especially, the most recent ones have reported promising results regarding the possibility of carrying out biopsies, with very small peripheral damage that does not affect the diagnosis and that could potentially ensure a correct interpretation of the peripheral margins if used correctly and by a skilled operator.

Studies conducted by Romeo et al. [16–18, 24] using different wavelengths, both in *ex vivo* and *in vivo* conditions, came to the conclusion that all types of lasers tested (diode 980 nm, diode 808 nm, diode 445 nm, Nd:YAG, Er,Cr:YSGG, KTP, and CO₂) can be used to perform biopsies safely if used properly. Monteiro came to the similar

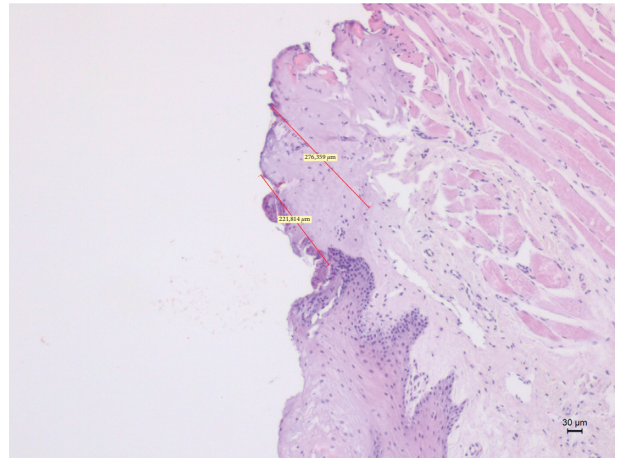


FIGURE 2: The histological specimen obtained using 4W in CW with colour EE at 10x magnification (group A).

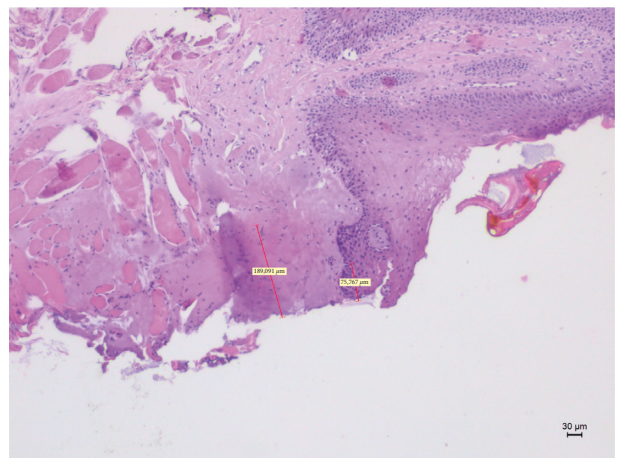


FIGURE 3: The histological specimen obtained using 4W in PW with colour EE at 10x magnification (group D).

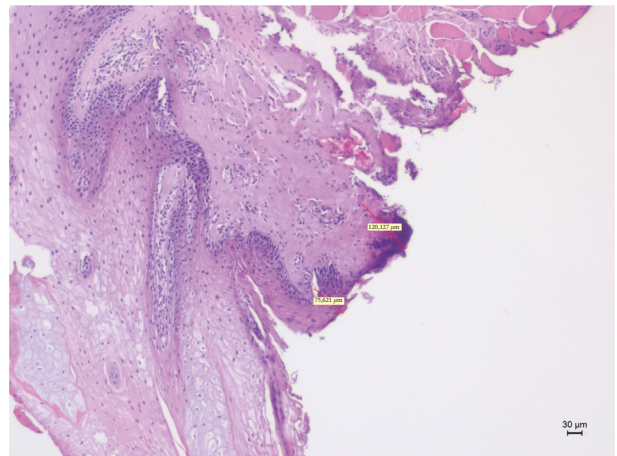


FIGURE 4: The histological specimen obtained using 5W in CW with colour EE at 10x magnification (group B).

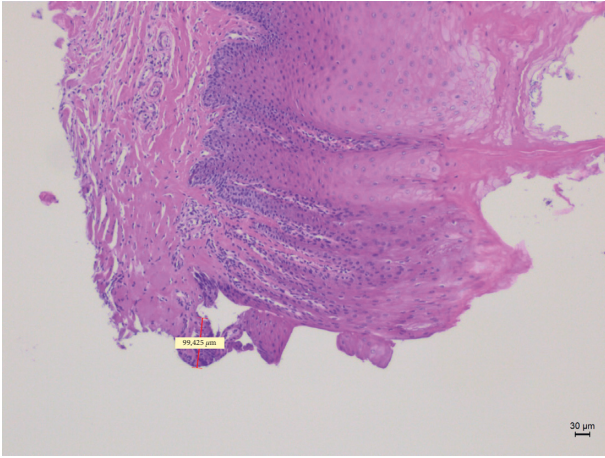


FIGURE 5: The histological specimen obtained using 5 W in PW with colour EE at 10x magnification (group E).

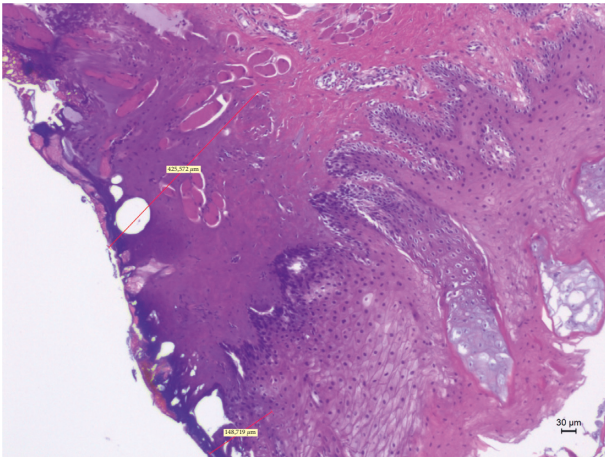


FIGURE 6: The histological specimen obtained using 6 W in CW with colour EE at 10x magnification (group C).

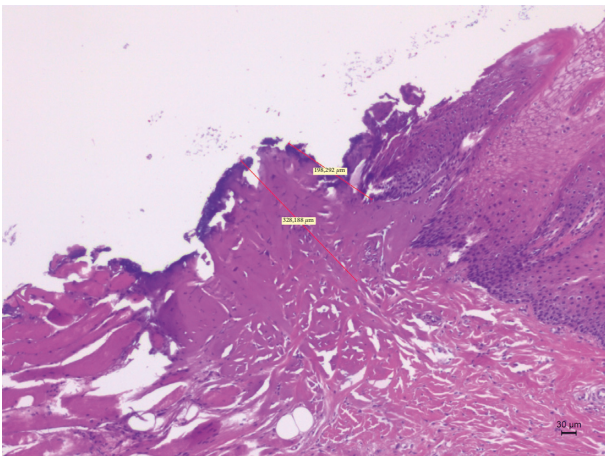


FIGURE 7: The histological specimen obtained using 6 W in PW with colour EE at 10x magnification (group F).

conclusions in a study in which they analyzed the resection margins of fibroepithelial lesions obtained using CO₂ lasers, diode lasers, Er:YAG, and Nd:YAG *in vivo* compared with the electrosurgical scalpel and cold scalpel, and they assessed that none of the tested lasers showed limitations regarding histological diagnosis [25]. Merigo et al. instead analyzed the temperature increase by using a thermal camera and the histological quality of samples taken through different wavelengths, obtaining positive results in terms of readability and diagnostic reliability [26].

In a study conducted *in vivo* using both KTP and diode laser, it was noted, as was to be expected, that the thermal damage was greater in some lesions such as oral lichen planus characterized by increased cellularity and inflammation compared to lesions such as mucocele that instead suffered minor thermal damage [3].

Angiero et al., in a retrospective study, noted a relationship between the size of the bioptic fragment prelevated with a diode laser and the presence of artifacts. Samples with a size of less than 3 mm led to limitations in diagnosis [27]. Also, Vescovi et al. found that, using a Nd:YAG laser, epithelial and stromal changes were significantly more frequent in specimens with a mean size less than 7 mm ($p < 0.0001$) [28].

The results of our study are in line with those of other reported studies; the size of the thermally damaged area has always been very small (less than a millimetre) both in the epithelium and in the connective tissue, and the peripheral margins were well readable, allowing a good histological evaluation of the samples.

Independent of the setting, the mean thermal damage was $208.40 \pm 133.81 \mu\text{m}$ in the epithelium and $330.14 \pm 147.45 \mu\text{m}$ in the connective tissue. In the PW samples, the average thermal damage was $188.32 \pm 137.28 \mu\text{m}$ in the epithelium and $303.88 \pm 129.15 \mu\text{m}$ in the connective tissue. The thermal damage size was larger in the CW samples, with a mean of $228.47 \pm 129.40 \mu\text{m}$ in the epithelium and $356.39 \pm 161.63 \mu\text{m}$ in the connective tissue. Compared to the CO₂, KTP, and 445 nm diode lasers in *ex vivo* conditions [7, 17, 18], the 976 nm diode laser generated greater thermal damage but did not compromise the quality of the histological evaluation.

It is interesting to note that, in our study, the damage to the epithelium has always been smaller than that of the connective tissue ($p < 0.01$). This finding is explainable by the selective absorption of the light radiation emitted by the 976 nm diode laser by haemoglobin, which implies not only a greater cutting effectiveness in more vascularized tissues but also greater thermal effects. Therefore, it seems that lasers could work well for obtaining biopsies. However, when using lasers, it is important to be well versed in the physical principles on which they are based to avoid many iatrogenic mistakes.

Based on the data from this study, the PW mode showed better results than the CW one ($p < 0.05$) in the epithelium and therefore is to be preferred allowing the tissue to recover from the thermal shock; moreover, it is important to enlarge the resection margins by about 1–2 mm to avoid that the heat-induced alterations could interfere with the histological evaluation of the transitional mucosa. According to our experience, it is also very

important to carefully calibrate the parameters, especially the power (W) and the frequency (Hz). If this is done, it may be possible to safely perform soft tissue biopsies with an optimal histological readability.

5. Conclusions

This study showed that a 976 nm diode laser may be safely used in the excision of oral lesions with a suitable width of resection margins. The highest amount of damage recorded was 689.575 μm in the epithelium and 964.24 μm in the connective tissue. No statistically significant differences were found among the groups. We suggest the use of a lower power that allows a good clinical cut in the PW mode to minimize the risk of generating heavy thermal damage in the treated tissue.

Although lasers appear to be an excellent alternative in the treatment of oral benign lesions, they still cannot be considered the first choice in the treatment of suspicious dysplastic or neoplastic lesions. However, their use may not be considered an absolute contraindication for such types of lesions. In these cases, it is particularly important to enlarge the resection margins by about 1 mm compared to the cold blade in order to avoid thermal effects that might disguise the real extent of the pathology and also to use controlled laser settings.

The limitations of this study include that it was conducted *ex vivo*, with all the related limits, and in healthy mucosa. In contrast, pathologic mucosa is characterized by different degrees of inflammation, hyperaemia, and increased cellularity. All these characteristics may affect the laser-tissue interaction and therefore could alter the results obtained.

Further studies, especially *in vivo* investigations, are needed to confirm these results in order to clarify how the anatomopathological alterations of pathological mucosa modify the laser-tissue interaction.

Data Availability

The data used to support the findings of this study were supplied by Dr. Daniele Pergolini under license and so cannot be made freely available. Requests for access to these data should be made to Dr. Daniele Pergolini (daniele.pergolini@uniroma1.it).

Ethical Approval

This article does not contain any studies with human participants or animals performed by any of the authors. This study was performed *ex vivo* using pig cadaver tongues, deceased within 24 h, purchased at the butcher's.

Consent

For this type of study, formal consent is not required.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] L. Walsh, "The current status of laser applications in dentistry," *Australian Dental Journal*, vol. 48, no. 3, pp. 146–155, 2003.
- [2] S. Verma, P. Chaudhari, S. Maheshwari, and R. Singh, "Laser in dentistry: an innovative tool in modern dental practice," *National Journal of Maxillofacial Surgery*, vol. 3, no. 2, p. 124, 2012.
- [3] U. Romeo, C. Russo, G. Palaia et al., "Biopsy of different oral soft tissues lesions by KTP and diode laser: histological evaluation," *The Scientific World Journal*, vol. 2014, Article ID 761704, 6 pages, 2014.
- [4] S. Andreana, "The use of diode lasers in periodontal therapy: literature review and suggested technique," *Dentistry Today*, vol. 24, no. 11, pp. 132–135, 2005.
- [5] S. Ulrich, W. Kluger, S. Dervisbegovi et al., "Innovative wavelengths in endodontic treatment," *Lasers in Surgery and Medicine*, vol. 38, p. 6, 2006.
- [6] A. Del Vecchio, M. Floravanti, A. Boccassini et al., "Evaluation of the efficacy of a new low-level laser therapy home protocol in the treatment of temporomandibular joint disorder-related pain: a randomized, double-blind, placebo-controlled clinical trial," *Cranio*, vol. 39, no. 2, pp. 141–150, 2019.
- [7] G. Palaia, A. Del Vecchio, A. Impellizzeri et al., "Histological *ex vivo* evaluation of peri-incisional thermal effect created by a new-generation CO₂ superpulsed laser," *The Scientific World Journal*, vol. 2014, Article ID 345685, 6 pages, 2014.
- [8] I. Tuncer, C. Özçakır-Tomruk, K. Şencift, and S. Çoğlu, "Comparison of conventional surgery and CO₂ laser on intraoral soft tissue pathologies and evaluation of the collateral thermal damage," *Photomedicine and Laser Surgery*, vol. 28, no. 1, pp. 75–79, 2010.
- [9] S. Nammour, M. El Mobadder, E. Maalouf et al., "Clinical evaluation of diode (980 nm) laser-assisted nonsurgical periodontal pocket therapy: a randomized comparative clinical trial and bacteriological study," *Photobiomodulation, Photomedicine, and Laser Surgery*, vol. 39, no. 1, pp. 10–22, 2021.
- [10] G. Tenore, A. Zimbalatti, F. Rocchetti et al., "Management of medication-related osteonecrosis of the jaw (MRONJ) using leukocyte- and platelet-rich fibrin (L-PRF) and photobiomodulation: a retrospective study," *Journal of Clinical Medicine*, vol. 9, no. 11, p. 3505, 2020.
- [11] U. Romeo, A. Del Vecchio, M. Capocci, C. Maggiore, and M. Ripari, "The low level laser therapy in the management of neurological burning mouth syndrome. A pilot study," *Ann Stomatol (Roma)*, vol. 1, no. 1, pp. 14–18, 2010.
- [12] H.-W. Yang and Y.-F. Huang, "Treatment of burning mouth syndrome with a low-level energy diode laser," *Photomedicine and Laser Surgery*, vol. 29, no. 2, pp. 123–125, 2011.
- [13] U. Romeo, A. Del Vecchio, C. Russo et al., "Laser treatment of 13 benign oral vascular lesions by three different surgical techniques," *Medicina Oral Patología Oral Y Cirugía Bucal*, vol. 67, pp. e279–e284, 2013.
- [14] L. R. Eversole, "Laser artifacts and diagnostic biopsy," *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontics*, vol. 83, no. 6, pp. 639–640, 1997.
- [15] P. Sahni, "Lasers in oral pathology—a review," *Medico-Legal Update*, vol. 12, no. 1, pp. 65–67, 2012.
- [16] U. Romeo, F. Libotte, G. Palaia, G. Tenore, A. Galanakis, and S. Annibaldi, "Is erbium: yttrium-aluminum-garnet laser versus conventional rotary osteotomy better in the

- postoperative period for lower third molar surgery? Randomized split-mouth clinical study,” *Journal of Oral and Maxillofacial Surgery*, vol. 73, no. 2, pp. 211–218, 2015.
- [17] G. Palaia, A. Impellizzeri, G. Tenore et al., “Ex vivo histological analysis of the thermal effects created by a 445 nm diode laser in oral soft tissue biopsy,” *Clinical Oral Investigations*, vol. 24, no. 8, pp. 2645–2652, 2019.
- [18] U. Romeo, G. Palaia, A. Del Vecchio et al., “Effects of KTP laser on oral soft tissues. An in vitro study,” *Lasers in Medical Science*, vol. 25, no. 4, pp. 539–543, 2010.
- [19] P. Vescovi, M. Manfredi, E. Merigo et al., “Surgical approach with Er: YAG laser on osteonecrosis of the jaws (ONJ) in patients under bisphosphonate therapy (BPT),” *Lasers in Medical Science*, vol. 25, pp. 101–113, 2010.
- [20] M. Tamarit Borràs, E. Delgado Molina, L. Berini Aytés, and C. Gay Escoda, “Removal of hyperplastic lesions of the oral cavity. a retrospective study of 128 cases [Exéresis de las lesiones hiperplásicas de la cavidad bucal. Estudio retrospectivo de 128 casos],” *Med Oral Patol Oral Cir Bucal*, vol. 1, pp. 1–45, 2005.
- [21] J. Yagüe-García, A. J. España-Tost, L. Berini-Aytés, and C. Gay-Escoda, “Treatment of oral mucocele - scalpel versus CO₂ laser,” *Medicina Oral, Patología Oral Y Cirugía Bucal*, vol. 144 pages, 2009.
- [22] S. Nammour, T. Zeinoun, A. Nammour et al., “Evaluation of different laser-supported surgical protocols for the treatment of oral leukoplakia: a long-term follow-up,” *Photomedicine and Laser Surgery*, vol. 35, no. 11, pp. 629–638, 2017.
- [23] R. A. Convissar, “Laser biopsy artifact,” *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontics*, vol. 84, no. 5, p. 458, 1997.
- [24] U. Romeo, F. Libotte, G. Palaia et al., “Histological in vitro evaluation of the effects of Er:YAG laser on oral soft tissues,” *Lasers in Medical Science*, vol. 27, no. 4, pp. 749–753, 2012.
- [25] L. Monteiro, M. -L. Delgado, F. Garcès et al., “A histological evaluation of the surgical margins from human oral fibrous-epithelial lesions excised with CO₂ laser, diode laser, er:Yag laser, nd:Yag laser, electrosurgical scalpel and cold scalpel,” *Medicina Oral, Patología Oral y Cirugía Bucal*, vol. 24, no. 2, pp. e271–e280, 2019.
- [26] E. Merigo, F. Clini, C. Fornaini et al., “Laser-assisted surgery with different wavelengths: a preliminary ex vivo study on thermal increase and histological evaluation,” *Lasers in Medical Science*, vol. 30, pp. 1–9, 2013.
- [27] F. Angiero, L. Parma, R. Crippa, and S. Benedicenti, “Diode laser (808 nm) applied to oral soft tissue lesions: a retrospective study to assess histopathological diagnosis and evaluate physical damage,” *Lasers in Medical Science*, vol. 27, pp. 383–388, 2012.
- [28] P. Vescovi, L. Corcione, M. Meleti et al., “Nd:YAG laser versus traditional scalpel. a preliminary histological analysis of specimens from the human oral mucosa,” *Lasers in Medical Science*, vol. 25, p. 685, 2010.