



Review

Interleukin 1 α : a comprehensive review on the role of IL-1 α in the pathogenesis and treatment of autoimmune and inflammatory diseases

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ABSTRACT

The interleukin (IL)-1 family member IL-1 α is a ubiquitous and pivotal pro-inflammatory cytokine. The IL-1 α precursor is constitutively present in nearly all cell types in health, but is released upon necrotic cell death as a bioactive mediator. IL-1 α is also expressed by infiltrating myeloid cells within injured tissues. The cytokine binds the IL-1 receptor 1 (IL-1R1), as does IL-1 β , and induces the same pro-inflammatory effects. Being a bioactive precursor released upon tissue damage and necrotic cell death, IL-1 α is central to the pathogenesis of numerous conditions characterized by organ or tissue inflammation. These include conditions affecting the lung and respiratory tract, dermatoses and inflammatory skin disorders, systemic sclerosis, myocarditis, pericarditis, myocardial infarction, coronary artery disease, inflammatory thrombosis, as well as complex multifactorial conditions such as COVID-19, vasculitis and Kawasaki disease, Behçet's syndrome, Sjogren Syndrome, and cancer.

This review illustrates the clinical relevance of IL-1 α to the pathogenesis of inflammatory diseases, as well as the rationale for the targeted inhibition of this cytokine for treatment of these conditions. Three biologics are available to reduce the activities of IL-1 α : the monoclonal antibody bermekimab, the IL-1 soluble receptor rilonacept, and the IL-1 receptor antagonist anakinra. These advances in mechanistic understanding and therapeutic management make it incumbent on physicians to be aware of IL-1 α and of the opportunity for therapeutic inhibition of this cytokine in a broad spectrum of diseases.

1. Introduction

Members of the interleukin 1 (IL-1) family of cytokines are cardinal mediators of inflammation [1]. Within this family, IL-1 α and IL-1 β are the archetypical pro-inflammatory cytokines. Following identification of these two cytokines in 1974, many studies explored the molecular biology and clinical effects of these molecules. However, IL-1 α and IL-1 β are not equally established players in the body of clinical medicine. At present, the inflammatory properties of IL-1 β are common knowledge in

the biomedical field, and many are familiar with molecular mechanisms of IL-1 β secretion (i.e. processing by the inflammasome). Conversely, and perhaps surprisingly, the role of IL-1 α in the pathogenesis of autoimmune and inflammatory diseases has remained relatively overlooked, perhaps because IL-1 α is rarely observed in the circulation of patients with inflammatory diseases.

Excellent reviews on the biology of IL-1 α are available, and we refer the reader to these previous publications for the molecular features and pre-clinical studies of this cytokine [2–5]. While using these excellent

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works as a foundation, this review is primarily written for clinical audiences. Specifically, this review aims at illustrating the clinical relevance of IL-1 α to the pathogenesis of inflammatory and autoimmune diseases, as well as the rationale for the targeted inhibition of this cytokine for the clinical treatment of these conditions.

Several factors make this a timely and clinically relevant topic. On the one hand, the clinical arena has welcomed biologics inhibiting IL-1 α either selectively (monoclonal antibodies) or non-selectively (IL-1 receptor antagonist), which reveal the extent and relevance of IL-1 α in the pathogenesis of human disease. On the other, the recent outbreak of COVID-19, as well as advances in the understanding and therapeutic management of different diseases, make it incumbent on physicians to be aware of IL-1 α and of the opportunity for therapeutic inhibition of this cytokine.

2. Biology Of IL-1 α

2.1. The IL-1 family of cytokines

Immune responses are orchestrated by cytokines, mediators produced by cells in order to induce a change in the function of different, target cells expressing specific receptors. The IL-1 family comprises 10 receptors and 11 cytokines, which can be classified into three subgroups based on the prevalent functional effect: I) secreted cytokines with agonistic activity (IL-1 α , IL-1 β , IL-18, IL-33, IL-36 α , IL-36 β , and IL-36 γ); II) receptor antagonists (IL-1Ra, IL-36Ra); III) anti-inflammatory cytokines (IL-37, IL-38) [1,6]. Interleukin 1 (IL-1) is the prototypical pro-inflammatory cytokine. Although the term IL-1, coined in 1979 [7], suggested a single molecule, previous studies on 'endogenous pyrogens' had already revealed the existence of two cytokines with different isoelectric points and molecular weight [8]. Afterwards, in the gene-cloning era, these two cytokines were unambiguously identified as two biological forms of IL-1 (α and β).

2.2. IL-1 α and IL-1 β bind the same receptor and induce identical pro-inflammatory effects

Distinct genes encode IL-1 α and IL-1 β , and the sequence homology is less than 26%, but the region of homology accounts for binding of both cytokines to the same IL-1 family receptor, namely IL-1R1 [4]. Since they bind the same receptor, IL-1 α and IL-1 β induce the same pro-inflammatory effects. These include activation of key transcription factors associated with inflammatory and immune responses, such as nuclear factor-kB (NF-kB), activator protein-1 (AP-1), c-Jun N-terminal kinase (JNK), p38 and other mitogen-associated protein kinases (MAPKs), extracellular signal-regulated kinases (ERKs), and interferon-regulating genes [9]. Once activated, these signaling cascades lead to the expression of myriad mediators, which collectively orchestrate the development of an immune response. Chief mechanisms include the synthesis of other cytokines, such as IL-6, IL-2, interferons or chemokines; production of prostaglandins; vasodilation and expression of adhesion molecules by endothelial cells, which enable migration of immune cells into tissues; activation of T-helper cells and maturation and clonal expansion of B cells, which activate adaptive immunity [9]. In inducing these pro-inflammatory effects, IL-1 α and IL-1 β act synergistically with other mediators such as TNF α , bradykinin, or growth factors. Activation of this interconnected network of soluble and cellular mediators has both local and systemic effects. For example, local effects are characterized by inflammation-mediated organ dysfunction and, eventually, damage. On the other hand, systemic effects include fever, vasodilation (rash), and musculoskeletal pain, as well as hypotension and shock for increasing concentrations.

2.3. IL-1 α and IL-1 β : differences

Although the shared IL-1R1 receptor transduces identical pro-

inflammatory effects, there are key differences in the biology of IL-1 α and IL-1 β , which account for the unique contribution of each cytokine to the orchestration of inflammatory responses (Table 1).

2.3.1. Expression by different tissues and cell types

IL-1 α is constitutively present as a precursor in all healthy tissues of mesenchymal origin, in particular, cells rich in IL-1 α constitute tissues with a barrier function, such as keratinocytes in the skin, type 2 epithelial cells in the lung, the epithelium of the entire gastrointestinal tract, endothelial cells in blood vessels, and astrocytes in the brain [3]. In addition to constitutive presence in barrier epithelia, production of IL-1 α precursor can be induced in myeloid cells during inflammation. Conversely, IL-1 β is not constitutively expressed in healthy states and is exclusively produced by myeloid cells during inflammation.

2.3.2. Activity of the precursor molecule and need for processing by the NLRP3 inflammasome

Both IL-1 α and IL-1 β are synthesized as intracellular precursors. However, the IL-1 α precursor is biologically active, whereas the IL-1 β precursor is not [10]. Inside the cell, the activity of the IL-1 α precursor is kept in check by the type 2 IL-1R (IL-1R2), a decoy receptor which binds the cytokine but does not transduce inflammatory effects, thus functioning as a 'sink' [11,12]. However, IL-1 α exiting from the cell can directly bind the IL-1R1 and exert its biologic effects. Conversely, the IL-1 β precursor must undergo intracellular processing and activating cleavage by the NLRP3 inflammasome in order to generate biologically active IL-1 β [13].

2.3.3. Role as 'alarmin'

Alarmins are proteins or molecular components normally found inside the cell, whose presence in the extracellular compartment signals loss of membrane integrity to nearby cell [3]. Thus, alarmins function as *de facto* danger-associated molecular patterns (DAMPs), and trigger sterile inflammation. Notable examples include HMGB1 [14], as well as IL-1 α [10]. Since the IL-1 α precursor is constitutively present in a biologically active form in epithelial cells, necrosis (i.e. following trauma, ischemia or viral infection) results in the immediate release of IL-1 α in the extracellular space, and consequent engagement of IL-1R1 on adjacent live cells. This results in induction of tissue inflammation (Figure 1). Conversely, IL-1 β is neither present in most cells nor biologically active as a precursor, and thereby does not function as an alarmin.

2.3.4. Juxtacrine versus paracrine signaling

Both IL-1 α and IL-1 β can be released into the extracellular space and bind IL-1R1 on nearby cells (paracrine signaling). However, IL-1 α also

Table 1
Differences in biological properties and function of IL-1 α and IL-1 β

IL-1 α	IL-1 β
IL-1 α is constitutively present in a wide variety of cells; expression can be induced in hematopoietic and nonhematopoietic cells in response to inflammatory stimuli.	IL-1 β is absent in cells at homeostasis and is expressed upon stimulation only in myelomonocytic cells.
Both the precursor (pro-IL-1 α) and the cleaved form of IL-1 α are biologically active IL-1R1 ligands. Caspase-1 and the inflammasome have no direct role in cleaving pro-IL-1 α .	Only the cleaved mature form of IL-1 β is biologically active and a ligand for IL-1R1. Pro-IL-1 β is cleaved by caspase-1 following activation of the inflammasome.
Preformed, biologically active IL-1 α is released by necrotic cells and acts as an alarmin.	Not being functionally active as a precursor, IL-1 β does not function as an alarmin.
IL-1 α functions both as a soluble mediator and as a membrane-bound cytokine.	Mature IL-1 β only functions as a soluble mediator.
Dual function: extracellular binding to the IL-1R1 and nuclear function as a transcription factor.	Extracellular binding to the IL-1R1 only.

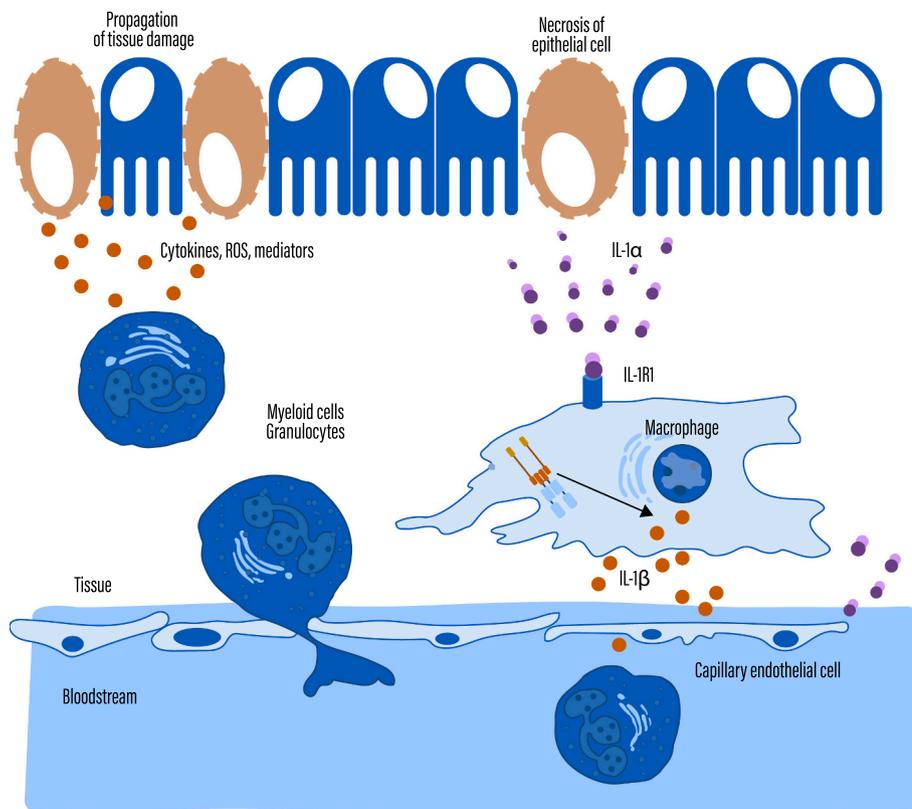


Fig. 1. The IL-1-driven inflammatory loop. Tissue damage triggers release of intracellular IL-1 α from dying cells. This generates an IL-1 α -containing milieu, which is sensed by infiltrating myeloid cells (i.e. monocytes, macrophages) expressing the IL-1R1. These cells in turn activate inflammasome-dependent production of IL-1 β , which initiates a cascade leading to the recruitment of inflammatory myeloid cells (i.e. granulocytes) to the site of tissue damage. Influx of inflammatory cells producing cytokines, reactive oxygen species, and effector mediators propagates tissue damage.

functions as an integral membrane protein, particularly on macrophages [15,16]. Membrane-bound IL-1 α exposed on the cell membrane by activated macrophages binds the IL-1R1 on adjacent cells (juxtacrine signaling) [17], a mechanism instrumental to the induction of local –or even focal– inflammation.

2.3.5. Dual function of IL-1 α

Dual functions of proteins allow the relatively few human genes expressed in adult life to accomplish a multitude of functions. Unlike IL-1 β , IL-1 α is a ‘dual function’ cytokine. In the extracellular space, released or membrane-bound IL-1 α binds to the IL-1R1 on the surface of the cells and triggers a pro-inflammatory cascade. Inside the cell, the IL-1 α precursor has its own function. In health, the IL-1 α precursor shuttles between the cytosol and the nucleus: due to a very strong nuclear localization sequence (KKRR) in the N terminus pro-piece, IL-1 α binds to DNA [18]. In cells undergoing non-inflammatory apoptosis, IL-1 α remains tightly bound to DNA and is not available to bind to IL-1R1 and trigger inflammation. In cells undergoing a necrotic cell death, the IL-1 α precursor remains in the cytosol and is released into the extracellular space and triggers inflammation [18]. In stimulated cells, for example skin keratinocytes exposed to ultraviolet light, the IL-1 α precursor functions as a nuclear transcription factor [9,18–20]. The ‘dual function’

of IL-1 α in the extracellular and cytosolic compartments is also found in IL-1 family member IL-33. In IL-33, a similar phenomenon occurs; failure of the IL-33 precursor to bind to DNA results in release of the IL-33 precursor into the extracellular space and marked systemic inflammation [21]. IL-1 family member IL-37 also has dual (intracellular and extracellular) functions. However, since IL-37 is an anti-inflammatory cytokine, failure to localize to the nucleus results in a loss of its anti-inflammatory properties [22,23].

2.4. Functional properties in inflammation: α is local, β is systemic

The binding to the same receptor by IL-1 α and IL-1 β raises the question as to why there are two IL-1s: either redundancy is needed for backup and robustness, or perhaps each cytokine has a specialized function [24]. The conundrum of an apparent functional excess and redundancy of IL-1 α and IL-1 β production at sites of inflammation is explained by the ‘IL-1 α -driven inflammatory loop’ (Figure 1). In this model, IL-1 α is either released from dying cells, or exposed on the surface of cells undergoing oxidative or metabolic stress. This generates an IL-1 α -containing milieu, which is sensed by myeloid cells expressing the IL-1R1. These cells in turn activate IL-1 β production downstream of IL-1R1, which initiates a cascade leading to recruitment of inflammatory

hematopoietic cells to the site of damage or stress.

Thus, *IL-1 α* is a *primum movens* of local inflammation, affecting the local microenvironment either through exposure on the membrane of stressed cells followed by juxtacrine binding of IL-1R1 on nearby cells, or through release upon cell death followed by paracrine binding of IL-1R1 on surrounding cells. Conversely, *IL-1 β* acts as a master regulator of systemic inflammation, as production of IL-1 β by infiltrating macrophages and circulating monocytes takes inflammation beyond the local level and causes widespread inflammation.

The non-redundant functions of IL-1 α as an alarmin and mediator of local inflammation also emerge during highly conserved immune mechanisms against invading pathogens, such as granuloma formation and responses to immune-evasive viruses. Granulomas function as focal inflammatory lesions comprising both hematopoietic and non-hematopoietic cells, which typically surround poorly degradable particulate matter or microbial, viral, fungal, protozoan or helminthic pathogens [25]. Because granulomas are fundamentally a local inflammatory response, it is not surprising that IL-1 α , with its specific role in inducing an inflammatory milieu upon exposure on the cell membrane, is involved in granuloma formation. Exposure of macrophages as well as lung epithelial cells to *M. tuberculosis* results in rapid expression of biologically active membrane-bound IL-1 α ; in experimental models of *M. tuberculosis* infection, IL-1 α -deficient mice did not develop protective granuloma structures and succumbed to the infection earlier than wild-type mice [25]. An analogous role has emerged for IL-1 α in granulomas induced by *Cryptococcus neoformans* and *Leishmania major* infections [26,27]. Granuloma formation is thereby an example of an evolutionarily non-redundant role of IL-1 α in inflammatory responses. Indeed, the expression of membrane-bound IL-1 α by myeloid cells surrounding a pathogen flags a restricted, infected area within a tissue, thus fostering targeted accumulation of recruited immune cells on the circumferential borders of the forming granuloma. This process results in efficient confinement and trapping of the invading pathogen (or foreign body). This same result could not be achieved by production of soluble mediators, which would result in widespread tissue inflammation or even inappropriate and detrimental systemic inflammatory responses.

In addition, non-redundant functions of IL-1 α are important for protection against immune-evasive viruses, which are poorly immunogenic or actively suppress inflammatory gene expression. For example, vesicular stomatitis virus infects keratinocytes and prevents induction of interferon responses, thereby being immune-evasive. However, infection and virus-induced lysis of keratinocytes prompts the release of preformed IL-1 α , whose immunogenic function is similar to and vicariates that of interferons [28]. IL-1 α is most important to control immune-evasive virus replication in fibroblasts and other barrier cell types, and represents a functional backup antiviral system to ensure barrier defense. In addition, neutralization of IL-1 α inhibits the antiviral activity of IFN- γ by 90%, whereas no inhibition of type I IFN activity was observed. Indeed, the antiviral activity of IFN- γ depends largely on the basal level of NF- κ B, which is maintained by constitutively expressed IL-1 α [29].

2.5. Evolutionary origin of IL-1 α accounts for intracellular localization and dual function

The IL-1 family of cytokines is evolutionarily ancient, and likely originated from duplications of a common ancestral gene related to fibroblast growth factors (FGFs) [30,31]. Indeed, a related IL-1-like molecule is found in the starfish and acts as a growth factor for the repair of severed limbs [32]. IL-1 α likely arose as a result of an ancestral gene duplication of IL-1 β [33]. Within the IL-1 family, IL-1 α has the closest homology to FGF [34]. FGF proteins localize in the nucleus, and this nuclear localization of FGF progenitors provides an evolutionary explanation for the role of IL-1 α , as well as other IL-1 family members IL-33 and IL-37, as nuclear factors [35,36]. Later in evolution, appearance of surface receptors of the immunoglobulin family enabled extracellular signaling [37]. In parallel, molecular machinery evolved in order to

expose IL-1 α on the cell surface and initiate paracrine inflammation upon binding of IL-1R1 on nearby cells (juxtacrine signaling). On the cell surface, extracellular proteases cleave pro-IL-1 α , leading to its release as a soluble mediator. At the same time, cell death due to injury or infection results in a passive leakage of cytosolic pro-IL-1 α into the surrounding milieu and activation of inflammation in an IL-1R1-dependent manner.

2.6. Mechanisms for control of inflammation mediated by IL-1 α

To accomplish the task of preventing unwanted activation and minimizing collateral damage of inflammation, the innate immune system relies on feedback regulatory circuits provided by suppressive mediators. Within the IL-1 family of cytokines, several inhibitory mechanisms are in place to prevent runaway inflammation induced by IL-1 α or IL-1 β . The main mechanism is the IL-1 receptor antagonist (IL-1Ra), which blocks the IL-1R1 and prevents binding of IL-1 α and IL-1 β [38].

In addition, the decoy receptor IL-1R2 binds to both IL-1 α and IL-1 β , but does not induce cytoplasmic signaling, thus inhibiting inflammation by functioning as a 'sink' for these mediators [39]. Specifically, the IL-1R2 genomic structure is similar to that coding for the extracellular portion of IL-1R1; however, IL-1R2 lacks a TIR domain and only has a short 29 amino acid-long cytoplasmic tail, and indeed, it does not transduce an inflammatory signal upon binding of IL-1 α and IL-1 β , thus acting as a molecular trap [40,41]. IL-1R2 is found both in the cytosol where it binds pro-IL-1 α and limits excessive IL-1 α -dependent sterile-inflammation during necrosis [11,12], as well as in the extracellular space and circulation, where it binds IL-1 β and limits systemic inflammation [42,43].

An additional way of taming the inflammatory response is provided by regulatory cytokines [44], which orchestrate a complex network of mechanisms leading to resolution of inflammation, including reduced cell infiltration and suppression of pro-inflammatory mediators. Within the IL-1 family, regulatory cytokines involved in the regulation of IL-1-driven inflammation include IL-37 and IL-38 [1,45–47].

Finally, non-immunological mechanisms are also in place to prevent IL-1 α release and initiation of inflammatory responses during programmed, non-inflammatory cell death. Release of cytosolic pro-IL-1 α during apoptosis is prevented by sequestration into the nucleus, followed by inclusion into apoptotic bodies and clearance by scavenging phagocytes [18,48]. In addition, the decoy receptor IL-1R2 binds and sequesters cytosolic IL-1 α , thus preventing inflammatory signaling during programmed cell death [11].

3. Role of IL-1 α in the pathogenesis of autoimmune and inflammatory diseases

3.1. The role of IL-1 α in the pathogenesis of human diseases

Given its role as a bioactive precursor released upon tissue damage and cell death, IL-1 α is centrally involved in diseases primarily characterized by tissue or organ inflammation (i.e. pericarditis, myositis), by disruption of barrier epithelia (i.e. dermatoses, airway diseases such as COVID-19, colitis), and by endothelial damage or thromboembolic activation (i.e. Behçet's syndrome, vasculitides). There is an overlap in the effects of IL-1 α and IL-1 β in many diseases, as local inflammation initiated by IL-1 α eventually triggers systemic inflammatory responses (see the IL-1 α -driven inflammatory loop in Figure 1). In this section, we describe the contribution of IL-1 α to the pathogenesis of different diseases, subdivided by tissue or organ system.

3.2. Skin

The skin is the prototypical IL-1 α 'organ'. Skin keratinocytes contain abundant IL-1 α , which is released upon injury, trauma, or infection. Prompt release of IL-1 α ensures rapid induction of local inflammatory

responses, which are essential both to the maintenance of skin integrity (i.e. by inducing wound healing) and to the deployment of effective barrier functions (i.e. by enabling clearance of invading pathogens). Several human diseases illustrate the relevance of IL-1 α to skin inflammation.

3.2.1. Deficiency of IL-1Ra (DIRA)

DIRA is a rare, neonatal-onset monogenic disorder characterized by constitutive lack of IL-1Ra [49]. Newborns develop severe, multisystem inflammation due to the unopposed activity of both IL-1 α and IL-1 β . A striking clinical manifestation of DIRA is severe, widespread neutrophilic dermatosis, which resembles pustular psoriasis. This manifestation is mediated by IL-1 α , as it is not observed in other auto-inflammatory conditions exclusively characterized by excessive release or activity of IL-1 β , such as cryopyrin-associated periodic syndromes or familial Mediterranean fever [50]. These clinical observations pinpoint the role of IL-1 α in the development of neutrophilic dermatoses. Treatment with anakinra is rapidly effective and leads to a reversal of clinical manifestations.

3.2.2. Hidradenitis suppurativa and other neutrophilic dermatoses

Neutrophilic dermatoses are a heterogeneous group of auto-inflammatory skin disorders, which are associated with cutaneous inflammatory lesions and rich infiltration with granulocytes at histology evaluation [51]. This group includes hidradenitis suppurativa (HS), pyoderma gangrenosum, Sweet's syndrome, and amicrobial pustulosis of the skinfolds. HS is a common, chronic disorder of skin areas rich in apocrine glands, characterized by formation of painful, pus-containing inflammatory nodules, followed by rupture and scarring. The causal role of IL-1 α in the pathogenesis of hidradenitis suppurativa is demonstrated by the clinical efficacy of anakinra [52], as well as by specific blockade of IL-1 α with monoclonal antibodies, which was associated with even more robust responses in clinical trials [53]. A central involvement of IL-1 α is also likely in the pathogenesis of pyoderma gangrenosum, yet evidence in this context is primarily derived from mouse models, in which caspase recruitment domain family member 9 (CARD9) signaling was identified as inducing IL-1 α [54]; however, reports on the efficacy of anakinra in patients with pyoderma gangrenosum are mixed [55,56]. Amicrobial pustulosis of the skinfolds is also characterized by elevated IL-1 α in skin biopsies and by response to treatment with anakinra [55,56].

3.2.3. Systemic sclerosis

Systemic sclerosis (SSc) is a severe autoimmune condition characterized by progressive vascular damage and fibrosis of tissues. Fibroblasts purified from lesional skin of patients with SSc express abundant amounts of IL-1 α , which is localized on the plasma membrane, in the cytosol, and in the nucleus [57]. Compared to fibroblasts from normal skin, SSc fibroblasts also express high amounts of IL-6, the growth factor PDGF- α , IL-1R1, and collagen [58]. Suppression of IL-1 α expression in SSc fibroblasts reduces amounts of secreted IL-6 and pro-collagen, whereas overexpression of pro-IL-1 α in normal fibroblasts increases IL-6 and pro-collagen production [57]. Moreover, IL-1-dependent PDGF- α production has direct mitogenic effects on fibroblasts and smooth muscle cells [59]. Collectively considered, these observations link excessive IL-1 α activity with a pathological tissue response leading to excessive deposition of collagen and fibrosis.

3.3. Lung and respiratory tract

IL-1 α is constitutively expressed in the lung epithelium [4], and is released upon cell death in conditions characterized by alveolar or airway epithelium inflammation. In addition, IL-1 α released by dying epithelial cells has a role in priming pro-inflammatory responses in fibroblasts, which leads to progressive fibrosis, maladaptive tissue remodeling, and damage.

3.3.1. Chronic obstructive pulmonary diseases (COPD)

COPD is a common disease characterized by chronic airway and alveolar inflammation, typically induced by cigarette smoking [60–62] and exacerbated by common viral infections [63,64]. COPD leads to maladaptive tissue remodeling, clinically manifested with emphysema and respiratory insufficiency. Overall, epithelial-derived IL-1 α mediates a pathologic cross talk between damaged lung epithelium and fibroblasts [65–67]. *In vitro* studies showed that IL-1 α released by damaged epithelium is both necessary and sufficient to promote a switch of primary human lung fibroblasts towards an inflammatory phenotype [65,68,69], with consequent up-regulation of IL-6, IL-8, monocyte chemoattractant protein-1, and GM-CSF. Interestingly, this effect is reduced with the administration of anti-IL-1 α or IL-1ra but not with anti-IL-1 β antibodies, thus supporting the causal role of IL-1 α [65]. In addition, lung epithelial cells infected by human rhinovirus release IL-1 α upon death, thus prompting a pro-inflammatory response in fibroblasts [66,67]. Airway epithelial cells obtained from severe COPD patients co-cultured with fibroblasts induce production of the neutrophil recruiting factor IL-8/CXCL8 by fibroblasts. Neutralizing antibodies against IL-1 α but not IL-1 β completely abrogated IL-8/CXCL8, suggesting a key role of IL-1 α in lung inflammation and tissue remodeling in COPD [68].

Altogether, these observations provided a biologic rationale for exploring the therapeutic potential of anti-IL-1 α agents in COPD, which remains a major public health problem due to disability and frequent virus-triggered exacerbations [63,64]. At present, experience with IL-1 α inhibition in COPD is limited. Treatment with MEDI8968, a fully human immunoglobulin G2 monoclonal antibody against the IL-1R1, did not yield statistically significant improvements in acute exacerbation rate, lung function and quality of life in a prospective trial of COPD patients [70].

3.3.2. Pulmonary fibrosis

Pulmonary fibrosis is the common end-stage of several inflammatory or autoimmune conditions characterized by progressive structural damage of the lungs [71,72]. Interstitial lung disease (ILD) is a form of pulmonary fibrosis that can be encountered in the context of systemic immune-mediated diseases such as connective tissue disease and rheumatoid arthritis [73]. In a study of 70 patients with rheumatoid arthritis, increased serum levels of naturally occurring antibodies against IL-1 α were detected in patients with ILD compared to patients without ILD. IL-1 α autoantibody positivity was also observed in a separate study of patients with rapidly progressive idiopathic pulmonary fibrosis [72]. Generation of autoantibodies against IL-1 α is indicative of the presence of extracellular IL-1 α , and may also represent a regulatory mechanism aimed at limiting IL-1 α bioactivity *in vivo*, in order to limit lung injury [74]. An open label extension of a randomized, double blind controlled study investigated the safety profile of anakinra compared to placebo in 1346 patients with rheumatoid arthritis. Only two patients developed interstitial lung disease, which indirectly suggest a protective effect of anakinra against the occurrence of ILD [75].

Lung involvement with fibrosis can occur in systemic sclerosis (SSc) and is associated with a poor prognosis [76]. The T-889C SNP in the promoter of the gene encoding IL-1 α is associated with pulmonary restrictive disease and with poor responses to cyclophosphamide in SSc-related lung fibrosis [77]. Studies investigating cytokine production in SSc patients revealed significantly increased IL-1 α plasma levels [78,79], which correlated with pulmonary impairment [80].

However, therapeutic inhibition of IL-1 yielded mixed results in pre-clinical models. IL-1Ra administration effectively antagonized activation of fibroblasts co-cultured with epidermal cells from SSc patients [81]. However, treatment with anakinra of mouse models of interstitial lung disease was associated with a worsening of pulmonary damage and with increased amounts of T cells, B cells and eosinophils in bronchoalveolar lavage fluid, possibly as a result of a polarization of the immune response towards a Th2 phenotype inducing pro-fibrotic alternatively activated macrophages [82]. Although management of

SSc-associated ILD still relies on non-selective immunosuppressants, targeted inhibition of cytokines remain an attractive therapeutic perspective and investigations in this field are ongoing [83].

3.3.3. Cystic fibrosis

Cystic fibrosis is characterized by thickening of secretions in the lungs, but also the pancreas, liver, kidneys, and intestine. Recurrent pulmonary infections result in progressive lung damage. Bronchoalveolar lavage fluid of patients with cystic fibrosis is rich in IL-1 α , IL-1 β , and IL8, and is characterized by high neutrophil elastase activity [84]. Concentrations of these mediators, as well as neutrophil elastase activity, correlate with the extent of pulmonary structural damage on computed tomography; however, the association is strongest for IL-1 α [84]. These observations point at IL-1 α as a driver of neutrophilic inflammation, particularly in early stages of cystic fibrosis, even in germ-free conditions [84]. An interventional, Phase 2a trial evaluating safety and efficacy of anakinra in patients with cystic fibrosis is ongoing (NCT03925194).

3.3.4. Bronchopulmonary dysplasia

Bronchopulmonary dysplasia is one of the most common disorders associated with preterm birth. Experimental models show that the developing lung of mice subjected to endotoxemia exhibit a robust NF κ B-dependent IL-1 α expression [85], and that the developing lung is susceptible to the downstream inflammatory effects of IL-1 α . In humans, evaluation of repeated tracheal aspirates of intubated, premature infants revealed a dramatic, sustained upregulation of IL-1 α mRNA (65-fold) and, to a lesser extent, of IL-1 β and IL-1Ra. These findings indicate that, at least in intubated preterm infants, there is pulmonary IL-1-mediated inflammation [86]. Blockade of IL-1R1 may attenuate lung injury and subsequent abnormal development following neonatal inflammatory stress. However, this therapeutic opportunity presently remains unexplored.

3.4. Gastrointestinal tract

The epithelial layers of the intestine contain abundant pro-IL-1 α . The role of IL-1 α as an alarmin is particularly relevant in the gut microenvironment, where constant exposure to pathogenic bacteria requires prompt induction of protective, host-defensive inflammatory responses. However, excessive epithelial cell injury and disruption of the mucosal barrier result in invasion of bacteria into the sub-epithelial *lamina propria* and in subsequent colon inflammation. These pathological features characterize chronic inflammatory bowel diseases (IBD), such as Crohn's disease and ulcerative colitis.

3.4.1. Colitis

The expression of both IL-1 α by epithelial cells and IL-1 β by myeloid cells in the intestinal mucosa correlates with the severity of inflammation in experimental models of IBD and in affected patients [87,88]. However, the two cytokines have different, non-redundant functions in intestinal inflammation: specifically, IL-1 α is released from damaged intestinal epithelial cells and functions as an alarmin by initiating and propagating inflammation, whereas IL-1 β is primarily involved in repair and reconstitution of the epithelial barrier in the resolution phase of intestinal inflammation [89,90]. Consistent with this mechanistic model, mice lacking IL-1 α subjected to experimental colitis exhibit mild disease symptoms, whereas mice lacking IL-1 β develop a more severe disease [89,90]. Neutralization of IL-1 α with monoclonal antibodies effectively reduced intestinal inflammation and damage in SAMP mice, which spontaneously develop a Crohn's-like ileitis, and in mice subjected to an established model of chemical (dextran sulfate sodium-induced) colitis [91]. In both models, the protective effects of IL-1 α neutralization were associated with taxonomic divergence of the fecal gut microbiome: most notably, a decreased ratio of *Proteobacteria* to *Bacteroidetes* heralds the anti-inflammatory effects of IL-1 α

neutralization. These findings confirm the clinically relevant interconnectedness of IL-1-mediated inflammation and the microbiota in intestinal homeostasis. Despite strong conceptual and pre-clinical evidence, no clinical trials have tested the efficacy of anti-IL-1 strategies and of IL-1 α blockade in particular in patients with IBD.

3.5. Muscle tissue

3.5.1. Myositis

Inflammatory myopathies are a heterogeneous group of conditions including dermatomyositis, polymyositis and inclusion body myositis, collectively characterized by inflammation-mediated progressive muscle damage and loss of strength. Different pro-inflammatory mediators are involved in the pathogenesis [92], particularly Th1-related cytokines [93]. Muscle cells constitutively contain abundant IL-1 α . Accordingly, in patients with inflammatory myopathies (i.e. dermatomyositis and sarcoid myopathy granulomas) there is a marked release of IL-1 α [94]. This mediator is focally expressed by muscle fibres characterized by myosinolysis, atrophy, ischemia, or regeneration [95]. IL-1 α is also abundantly expressed by endothelial cells of capillaries and venules of muscle biopsies from patients with inflammatory myopathies [94]; of note, marked expression of IL-1 α on endothelial cells is found even in patients with symptomatic DM and PM without clear evidence of lymphocytic infiltrates. This finding indicates that IL-1 α is likely implicated in the development of clinically relevant inflammation independently of muscle infiltration [96]. Also consistent with these observations, some patients display high expression of IL-1 α on muscle fibers and persistent muscle weakness even following treatment with corticosteroids, which results in disappearance of lymphocytic infiltrates [97]. It is plausible that expression of IL-1 α results in low-grade, persistent, inflammation-mediated muscle weakness [98]. In turn, IL-1 α induces production of IL-1 β by infiltrating myeloid cells: in line with the IL-1 α -driven inflammatory loop concept, IL-1 α is expressed on endothelial, muscle, and inflammatory cells, whereas expression of IL-1 β is restricted to infiltrating immune cells [99].

At present, limited data is available supporting treatment of inflammatory myopathies with anakinra. In mouse models, muscle inflammation is prevented in mice lacking IL-1 α and IL-1 β [100]; in a separate study, IL-1 α levels in muscle tissue correlated with disease severity and treatment with IL-1ra resulted in a significant suppression of muscle inflammation [101]. In humans, the efficacy of anakinra was evaluated in 15 patients with myositis refractory to conventional immunosuppressive therapies: seven out of 15 patients exhibited a clinical improvement [102]. In a separate study, anakinra was administered with benefit in a case of refractory of amyopathic dermatomyositis associated with anti-MDA5 antibodies [103]. In summary, inhibition of IL-1 α seems promising in difficult-to-treat inflammatory myopathies, or in patients with chronic muscle weakness and low-grade inflammation with mild or absent inflammatory infiltrates.

3.6. Cardiovascular system: heart and pericardium

3.6.1. Myocarditis

Myocarditis is an inflammatory disease affecting the myocardium, clinically manifested with heart failure and arrhythmia and diagnosed using imaging and/or histopathologic criteria. This condition can develop in isolated forms, or as part of infectious and autoimmune disorders [104]. Regardless of the initiating agent, the pathogenesis revolves around deregulated IL-1-mediated inflammation and innate immune system activation. During myocardial injury, dying myocytes release IL-1 α , together with several intracellular debris and other inflammatory proteins such as high-mobility group box 1 protein, IL-33 and the Ca²⁺-binding S100 proteins [105,106]. Following release of these alarmins, infiltrating immune cells are alerted of tissue damage and initiate local and systemic inflammatory responses. Binding of IL-1 α by IL-1R1 expressed on the cell membrane of immune cells results in

activation of nuclear factor- κ B with subsequent transcription of several proinflammatory genes. These notably include the precursors of IL-1 β and IL-18, as well as components of the NLRP3 inflammasome, which generates bioactive IL-1 β and further enhances the inflammatory process [107]. The findings of exaggerated inflammasome activation were confirmed both in humans with acute myocarditis following a viral infection [107] and in murine models of coxsackievirus-induced myocarditis [108]. Further evidence was also found in the heart of patients with acute transplant rejection suggesting that inflammasome formation correlates with the severity of rejection and with heart failure [109]. This IL-1-rich milieu causes further apoptosis of cardiomyocytes, loss of contractile tissue, fibrosis, cardiomyopathy, heart failure, and arrhythmic outburst. A close relationship exists between IL-1 α , IL-1 β , and heart function: both cytokines worsen contractile function by inhibiting L-type calcium channels, uncoupling β -adrenergic receptors from the adenylyl-cyclase [110] and causing transcriptional and post-translational changes in phospholamban and sarcoplasmic/endoplasmic reticulum calcium ATPase [111]. Moreover, effects of IL-1-mediated inflammation in the myocardium include disruption of calcium and β -adrenergic receptors signaling, mitochondrial dysfunction, and increased nitric oxide production [112].

Preliminary clinical experience with anakinra provided unequivocal proofs that IL-1 plays a pivotal role in both myocardial inflammation and systolic impairment. The effectiveness of IL-1 blockade was first shown in a randomized double-blind trial enrolling 23 rheumatoid arthritis patients with impaired vascular and left ventricular function: a prompt amelioration of cardiac function including myocardial contractility and relaxation, coronary flow reserve, and brachial artery flow-mediated dilatation was observed within 3 hours of anakinra administration [113]. More recently, a double-blind, placebo-controlled trial of anakinra in patients with diastolic heart failure and systemic inflammation confirmed beneficial effects of IL-1 inhibition; contractile function improved also in cases of life-threatening fulminant myocarditis or dilated cardiomyopathy [114–117]. The clinical trials ARAMIS and MYTH-1 are recruiting participants to assess superiority of add-on anakinra compared to conventional therapy in patients with acute and chronic myocarditis (NCT03018834 and EudraCT 2018-003472-13, respectively).

3.6.2. Myocardial infarction and ischemia-reperfusion injury

Acute myocardial infarction, characterized by myocardial ischemia followed by necrosis [118], is a common cardiac emergency and a leading cause of morbidity and mortality worldwide [119]. Following a myocardial infarction, a robust inflammatory response develops in the heart, which regulates infarct healing but may also promote maladaptive ventricular remodeling resulting in dilation and systolic dysfunction [120].

IL-1 α is strongly expressed in cardiomyocytes. Similar to myositis and myocarditis, necrosis of cardiomyocytes results in the release of IL-1 α and other danger associated molecular patterns, including high mobility group box-1 (HMGB-1), heat shock proteins, myosin, hyaluronic acid, fibrinogen, fibronectin and nucleic acids. These are sensed by neighboring cells, such as infiltrating myeloid cells and cardiac fibroblasts, which differentiate into myofibroblasts. In these cells, IL-1 α induces different pro-inflammatory cytokines including IL-1 β , TNF- α and IL-6 in a concentration-dependent manner by activation of different inflammatory pathways [121]. Upon sensing of IL-1 α and other DAMPs, myofibroblasts also initiate tissue repair and scar formation [122].

Various animal models showed that binding of either IL-1 α or IL-1 β to the IL-1R1 receptor has detrimental consequences for ischemia-reperfusion injury and myocardial remodeling [123,124]. Following ischemia, mice lacking IL-1 α or IL-1 β exhibit reduced tissue inflammation [125], whereas mice lacking IL-1Ra exhibit increased tissue inflammation [126]. Moreover, animals lacking the IL-1R1 or overexpressing IL-1Ra [124] exhibit reduced inflammation, neutrophil infiltration, and ventricular dilatation following myocardial infarction

[127].

There is also evidence for a specific, non-redundant effect of IL-1 α . Specifically, a study showed that treatment with anti-IL-1 α antibodies administered immediately after reperfusion resulted in reduced ischemic penumbra and in preservation of cardiac function [128]; notably, this study also revealed a decreased activity of caspase-1, thus substantiating the role of IL-1 α as an upstream and crucial danger signal triggering secretion of IL-1 β . Conversely, in a different study from the same group, IL-1 β inhibition alone neither prevented caspase-1 activation nor reduced the extent of the infarcted area [129]. Also of note, IL-1 α and IL-1R1, but not IL-1 β , are required for neutrophilic influx [130].

In humans, there is clinical evidence that IL-1 inhibition is beneficial in ischemic heart disease. The efficacy of IL-1 inhibition with anakinra has been shown in 3 sequential studies enrolling patients with ST-segment-elevation myocardial infarction (STEMI) and elevated C-reactive protein. In these studies, patients treated with anakinra for two weeks experienced a significant reduction in C-reactive protein [131,132] as well as a lower incidence of new-onset heart failure and hospitalization for heart failure [131]. However, another randomized, placebo-controlled study enrolling patients with non-STEMI acute myocardial infarction failed to show an improvement in cardiac clinical outcomes despite a decrease in inflammatory markers with anakinra [133]. In the massive CANTOS trial, selective blockade of IL-1 β prevented recurrent myocardial infarction in patients with previous ischemic events and systemic inflammation [134]. Overall, it appears that distinct roles of IL-1 α and IL-1 β during myocardial infarction reflect different timing of activation, with IL-1 α found earlier and more upstream in the inflammatory process [128]. Altogether, these observations provide a conceptual framework for blocking IL-1 in patients with active acute myocardial infarction, also in order to prevent incident heart failure [135].

3.6.3. Pericarditis

Recurrent pericarditis affects 15-30% of patients after a first episode of acute pericarditis [136,137]. Treatment regimen consists of a combination of non-steroidal anti-inflammatory drugs (NSAIDs) and colchicine, with the addition of corticosteroids in resistant or intolerant cases [136–139]. Patients often become corticosteroid-dependent and are unable to taper or discontinue steroids without experiencing a relapse. The necessity to avoid chronic corticosteroid treatment led to first evaluation of IL-1 receptor blockade in children with recurrent pericarditis [140]. Ever since, anakinra emerged as a safe and effective therapy to obtain or maintain remission while discontinuing corticosteroids, both in children and in adults [141]. On this basis, the European guidelines on the management of pericardial diseases gave a class IIb recommendation to administer anakinra to corticosteroid-dependent, colchicine-resistant patients with recurrent pericarditis [142].

The efficacy of anakinra for recurrent pericarditis was confirmed in a randomized clinical trial: the AIRTRIP study [140] evaluated 21 patients with recurrent pericarditis refractory to colchicine resistance and dependent on corticosteroids; anakinra was administered at a dose of 2 mg/kg/day (maximum 100 mg/day) for 2 months, then responders were randomized to continue anakinra ($n = 11$) or switch to placebo ($n = 10$) for 6 months or until a pericarditis recurrence. The primary outcomes were recurrent pericarditis and time to recurrence after randomization. Recurrent pericarditis occurred in 90% patients receiving placebo compared to 18% patients receiving anakinra; median time to recurrence was 72 days in patients receiving placebo, whereas this parameter could not be calculated due to too few events in the anakinra group [140]. A subsequent international multicenter registry of anakinra in recurrent pericarditis, the IRAP registry (International Registry of Anakinra for Pericarditis), confirmed the findings of the AIRTRIP trial in a real-world population [141]. On these bases, anakinra is currently the treatment of choice for recurrent pericarditis after failure of conventional anti-inflammatory therapies including non-steroidal anti-

inflammatory drugs, colchicine, and corticosteroids.

Recently, the efficacy of rilonacept, an interleukin-1 α and interleukin-1 β cytokine trap, was also evaluated in the RHAPSODY study, a phase 3, randomized clinical trial of 61 patients with recurrent pericarditis. Patients presenting with pericarditis recurrence while receiving standard therapy were enrolled in a 12-week run-in period, during which rilonacept was administered subcutaneously once weekly and background medications were discontinued. Clinical responders were further randomly allocated to receive continued rilonacept monotherapy or placebo. The primary efficacy end point was the time to the first pericarditis recurrence. In this study, 2 of 30 patients (7%) in the rilonacept group had a pericarditis recurrence, compared to 23 of 31 patients (74%) in the placebo group. The median time to recurrence in the placebo group was 8.6 weeks, whereas in the rilonacept group recurrences were too few to allow for this outcome to be evaluated ([ClinicalTrials.gov NCT03737110](https://clinicaltrials.gov/ct2/show/study/NCT03737110)).

In recent reports, pediatric patients with refractory recurrent pericarditis, who were receiving treatment with anakinra with optimal disease control, were switched to canakinumab, a monoclonal antibody neutralizing IL-1 β , and experienced disease relapses [143]. In these cases, treatment with anakinra was re-established and again led to disease control. These preliminary observations of differential efficacy of IL-1 β and IL-1 receptor blockade support a central role of IL-1 α in the pathogenesis of pericarditis.

3.7. Vascular system and thrombosis

IL-1 α is expressed abundantly by endothelial cells, activated platelets, and circulating monocytes during inflammation [144,145]. Growing evidence suggests that IL-1 α acts as a link between inflammatory responses and the coagulation cascade. Mechanistically, during wounding or hemorrhage, surface pro-IL-1 α expressed on the surface of endothelial cells undergoes activating cleavage by thrombin [145]. Thrombin-cleaved IL-1 α is detectable in human conditions characterized by thrombo-inflammatory activation, such as sepsis [145]. Locally expressed IL-1 α promotes thrombosis by increasing platelet activity, time of clot lysis [146,147], and endothelial activation [148]; in addition, circulating IL-1 α induces thrombopoiesis and megakaryocyte fragmentation, thus increasing the number of circulating platelets [149]. In mice models of vascular inflammation, IL-1 α but not IL-1 β expressed by platelets induces endothelial activation, with expression of adhesion molecules (ICAM-1 and VCAM-1), release of CXCL1, and production of tissue factor [150]. These effects promote recruitment of inflammatory cells (particularly neutrophils), endothelial cell dysfunction, and thrombosis, and are relevant to several human inflammatory diseases.

3.7.1. Behçet's syndrome

Behçet's syndrome is a systemic vasculitis, clinically characterized by muco-cutaneous, ocular, vascular, neurological, pulmonary and gastrointestinal manifestations [151,152]. Inflammatory thrombosis is a hallmark of Behçet's syndrome, and up to 40% of patients develops vascular complications involving the venous or arterial vascular tree [153]. Superficial and deep venous thrombosis of the limbs is common, but thrombosis in atypical sites such as the vena cava, the hepatic veins with Budd-Chiari syndrome, the portal vein, the cerebral venous sinus, or the right ventricle, may also occur [151]. Arterial inflammation and thrombosis is not uncommon, which makes Behçet's syndrome one of the few disorders causing aneurysms both in the peripheral, visceral and pulmonary districts [154,155]. The term "angio-Behçet" identifies a specific group subtype of the disease characterized by recurrent thrombosis [156,157], often with concomitant arterial aneurysms and venous involvement [158].

Behçet's syndrome is considered a model of inflammation-induced thrombosis [159–161], and indeed few conventional thrombogenic risk factors have been described in Behçet's syndrome patients. IL-1

gene cluster polymorphisms (IL-1A 2889C and IL-1B +5887T haplotype) are associated with increased susceptibility to vascular inflammation and Behçet's syndrome [162,163]. Moreover, IL-1 α is highly expressed in neutrophils, lymphocytes and endothelial cells in the *vasa vasorum* of patients with Behçet's syndrome [163], and serum levels of IL-1 α are significantly higher in patients with Behçet's syndrome compared to healthy individuals [164]. The IL-1 α -rich milieu of inflamed blood vessels in Behçet's syndrome also promotes neutrophil recruitment, whose aberrant activation is followed by programmed cell death and release of structures containing condensed chromatin (Neutrophils Extracellular Traps, NETs) [165,166]. NETs contain proteases such as cathepsin G and elastase, which promote vascular inflammation and damage [167] while also serving as extracellular platforms for the processing and activation of IL-1 family cytokines, thus fostering inflammation [168]. Activated neutrophils also release abundant reactive oxygen species (ROS), which are responsible for post-translational modifications of fibrinogen leading to generation of more resistant clots [169]. Overall, these studies indicate that IL-1 α represents a potential therapeutic target for the treatment of arterial inflammation in vasculitis associated with Behçet's syndrome [170]. Factors suggesting the suitability of IL-1 α inhibition specifically in Behçet's syndrome include concomitant venous and arterial inflammation, as well as other clinical manifestations typically mediated by IL-1 α (i.e. pustular skin or mucosal involvement).

3.7.2. Arterial aneurysms and Kawasaki disease

Besides inflammatory thrombosis, IL-1 α is involved in aneurysm formation. In a study evaluating changes in the concentration of serum cytokines following repair of abdominal aortic aneurysm, only IL-1 α was significantly reduced, which likely implicates this cytokine in the molecular pathogenesis of aneurysms [171].

In addition, IL-1 α is involved in the formation of arterial lesions associated with Kawasaki disease (KD), the main cause of acquired heart disease in childhood [172,173]. KD is a systemic vasculitis of medium-sized arteries, characterized by fever, rash, conjunctival injection, cervical lymphadenopathy, and inflammation of the coronary arteries. In 40-80% of children, KD leads to persistent coronary artery inflammation, aneurysmal dilation of coronary arteries, or myocarditis. The cause of KD remains undefined, but several infectious triggers (i.e. *Retroviruses*, *Coronaviruses*, *Epstein-Barr virus*, *Staphylococcal superantigen*) have been postulated to act on a predisposing genetic background [174,175]. Notable polymorphisms include Inositol 1,4,5- trisphosphate 3- kinase C (ITPKC) [176], which leads to persistent intracellular increase of Ca²⁺, and hence to enhanced activation of NLRP3 inflammasome.

Mouse models of Kawasaki disease showed that both IL-1 α and IL-1 β play a non-dispensable role in the development of coronary arteritis [170], and that inhibition of either mediator by gene deletion or pharmacological blockade can prevent aneurysm formation. IL-1 α , released as an alarmin in the microenvironment or expressed by neighbouring fibroblasts, triggers the inflammatory response in the initial phase of KD and likely regulate IL-1 β secretion, whose role prevails in later phases of KD [170]. Both IL-1 α and IL-1 β trigger vasodilatation and promote the influx of myeloid cells to affected tissues. Genetic and transcriptomic data in humans confirmed the role of IL-1 α and IL-1 β in the pathogenesis of KD vasculitis [177,178].

Treatment of KD is currently based on corticosteroids and intravenous immunoglobulins (IVIG). High-dose IVIG significantly inhibits coronary arterial endothelial cells and macrophage activation [179]. The mechanisms underlying the therapeutic efficacy of IVIG in KD remain undetermined [180]. Of note, previous study indicated that IVIG treatment suppresses IL-1 α and IL-1 β and stimulates IL-1Ra production, both *in vitro* and *in vivo* [181]; decreased levels of IL-1 receptor antagonist (IL-1Ra) are documented in IVIG-resistant KD patients [182].

Treatment with anti-IL-1 α or IL-1 β mAb alone protects mice against KD vasculitis and myocarditis, but the protection is partial unless the

two mAbs are used together or unless both cytokines are inhibited via receptor blockade [170]. In humans, treatment of KD with anakinra is under consideration in clinical trials [183,] and showed efficacy in case series [184]. In these studies, IL-1 receptor blockade led to the resolution of the clinical manifestations in difficult-to-treat patients [175,184,185].

4. COVID-19

In 2020, a new infectious disease denominated coronavirus disease 2019 (COVID-19), caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), resulted in an unprecedented pandemic outbreak [186]. SARS-CoV-2 is an airborne pathogen, which infects lung epithelial cells as well as endothelial cells causing inflammation of the airway tract and thrombotic microangiopathy. Since COVID-19 is a new disease, limited mechanistic data is available on the pathogenesis. However, IL-1 α is constitutively present in epithelial cells, endothelial cells, and infiltrating myeloid cells in the lung. This implies a massive release of IL-1 α upon the lytic death of these cells due to the viral infection, with subsequent sensing by alveolar macrophages [187]. In some patients, a life-threatening hyper-inflammatory reaction ensues, which resembles a macrophage activation syndrome (MAS) or a cytokine storm [188] and culminates in acute respiratory distress syndrome (ARDS) [189]. Inflammatory thrombosis also typically occurs in the lung of COVID-19 patients [190]. Advanced stages of COVID-19 ARDS are characterized by diffuse lung damage (with both alveolar epithelial and endothelial injury), accumulation of inflammatory exudate fluid in alveolar spaces, extensive thrombosis, and maladaptive tissue repair with fibrotic changes [191].

Overall, the pathogenesis of COVID-19 seems to recapitulate several key steps of IL-1 α -mediated inflammation. Specifically, release of IL-1 α as an alarmin by epithelial cells is followed by sensing by inflammatory myeloid cells and activation of the inflammasome, which results in amplification of the inflammatory cascade; at the same time, expression of IL-1 α by endothelial cells results in granulocyte recruitment and inflammatory thrombosis.

Production of IL-1Ra is a physiologic mechanism to limit excess inflammation and tissue damage in ARDS. IL-1Ra is elevated in the broncho-alveolar fluid of ARDS patients, particularly in the resolution phase [192]. Moreover, IL-1Ra serum levels are also associated with disease severity in pediatric ARDS [193]; in patients with community-acquired pneumonia, polymorphisms are associated with adverse outcomes [194].

Given these evidences, as well as similarities between severe COVID-19 and hyper-inflammation in the context of IL-1-mediated autoimmune or autoinflammatory conditions, IL-1-blockade was evaluated for the treatment of COVID-19 [195]. In a landmark study, high-dose intravenous anakinra (5 mg/kg twice a day) reduced mortality and ameliorated systemic inflammation and respiratory function in 29 patients with ARDS and on non-invasive ventilation, compared to standard management [196]. Subsequent studies independently reported comparable reductions in mortality and/or need for invasive mechanical ventilation [197–204]. To date, 14 and 3 different clinical trials are evaluating treatment of COVID-19 patients with anakinra and canakinumab, respectively (<https://clinicaltrials.gov/>). Meanwhile, studies evaluating monoclonal antibodies blocking IL-6 yielded more conflicting results [205–209].

Given the bi-directional relationship between IL-1-mediated inflammation and coagulation, blockade of IL-1 α and IL-1 β might also reduce inflammatory thrombosis and thromboembolic complications in COVID-19. In previous studies of disseminated intravascular coagulation during sepsis or MAS, anakinra treatment was associated with survival benefits [210,211]. However, this opportunity remains to be evaluated in COVID-19.

4.1. Pediatric inflammatory multisystem syndrome temporarily associated to SARS-CoV-2 infection

During the COVID-19 pandemic, a severe pediatric inflammatory multisystem syndrome temporarily associated to SARS-CoV-2 infection (PIMS-TS) was described. Specifically, some children and adolescents with confirmed COVID-19 or close contact with affected relatives developed predominantly gastrointestinal symptoms followed by an acute hyper-inflammatory disease and myocardial insufficiency, mimicking acute KD [212,213].

The clinical overlap between PIMS-TS and KD was first proposed by Jones *et al.*, who described a patient with classical KD, who had a positive nasopharyngeal swab for SARS-CoV-2 [214]. Children with PIMS-TS have more often antibodies against SARS-CoV-2 rather than a positive nasopharyngeal swab. This finding suggests a latency (up to one month) between COVID-19 infection and the development of PIMS-TS, which is consistent with a maladaptive immune response rather than a direct causative role of the virus. Given pathogenic and clinical similarities with both KD and COVID-19 hyper-inflammation, IL-1 α is plausibly implicated in the pathogenesis of PIMS-TS. In analogy with KD, the proposed first-line treatment for PIMS-TS was IVIG or corticosteroids. In non-responders or in patients with hyper-inflammation resembling macrophage activation syndrome, high-dose intravenous anakinra can be effective, consistent with use in KD and COVID-19 hyper-inflammation.

5. Sjögren's syndrome

Sjögren's Syndrome (SS) is an autoimmune condition characterized by chronic inflammation of exocrine glands leading to sicca symptoms [215,216]. Epithelial cells of lacrimal and salivary glands are both the target and the instigator of inflammation in SS [217,218]. Indeed, epithelial cell activation and death result in the release of pro-inflammatory mediators, including IL-1 α as well as other IL-1 family members, which prompt recruitment of immune cells [219–221] and activation of the inflammasome [222]. In this IL-1-rich milieu, innate inflammation leads to progressive tissue damage and exocrine gland dysfunction in the early phases of the disease, whereas lymphocytic infiltrates are the hallmark of established SS [223].

Evidence for a role of IL-1 α in SS comes from pre-clinical and clinical studies. In experimental animals, direct instillation of IL-1 α in salivary [224] or lacrimal glands [225] causes inflammation, dysfunction, and loss of acinar epithelial cells. Consistently, deficiency of IL-1R1 prevents eye keratinization and epitheliopathy [226], without affecting lymphocytic infiltration of glands. The conjunctival epithelium of patients with SS exhibits high concentrations of IL-1 α as well as other cytokines [227]. In dry-eye disease, tear fluids display an increase in IL-1 α and mature IL-1 β and a decrease in the biologically inactive precursor of IL-1 β [228]. High levels of IL-1Ra are also observed in tears of patients with SS, which likely represents an insufficient attempt at curbing excess inflammation [229]. Indeed, an imbalance in the IL-1/IL-1Ra ratio is observed in the saliva of patients with SS compared to controls [230]. Exogenous administration of anakinra is clinically effective in dry-eye syndrome, both in experimental models [231] and humans receiving topical treatment [232]. A randomized, double-blind, placebo-controlled study evaluated systemic administration of anakinra in 26 patients with SS. Unfortunately, this study did not evaluate sicca symptoms, yet a decrease in inflammation-mediated fatigue was reported [233], which substantiates a systemic role of IL-1 in SS. Given the central role of epithelial inflammation and damage, leading to release of IL-1 α followed by active production of IL-1 β in affected glands, IL-1 inhibition deserves consideration as a therapeutic strategy in SS, particularly in light of the lack of effective therapies for sicca symptoms.

6. Erdheim-Chester disease

Erdheim-Chester disease is a rare inflammatory neoplasm characterized by infiltration of multiple tissues by foamy macrophages [234]. These cells bear activating mutations along the MAPK pathway (most commonly BRAFV600E) and produce abundant cytokines [235–238]. Clinical manifestations are protean, and are caused both by the development of mass-like lesions as well as by systemic inflammation and fibrosis [239,240]. Mutated macrophages infiltrating tissues in ECD express abundant membrane-bound IL-1 α , which is likely instrumental to local recruitment and activation of inflammatory cells and progressive lesion growth [241]. Indeed, treatment with anakinra can be effective both in suppressing systemic inflammation and in reducing lesion size in ECD [242–244].

7. Cancer

Since IL-1 α is constitutively expressed in mesenchymal cells, malignant transformed cells of mesenchymal origin contain the IL-1 α precursor. IL-1 α is also observed within the nuclei of cancer cell lines, as a result of the nuclear localization sequence. However, whether IL-1 α in the nucleus has the potential to contribute to malignant transformation is unknown [245]. The current understanding of the role of IL-1 α in cancer development is more complex. In the early phases of carcinogenesis, IL-1 α expressed on the cell membrane of dendritic cells promotes the development of an immune response aimed at halting tumor formation by facilitating neoantigen recognition in the context of MHC [4]: for example, initial carcinogenesis induced by cigarette tars in the lung is often halted by neoantigen recognition and immune elimination. However, once tumors escape immune surveillance and a mass develops, high rates of cellular turnover result in abundant release of IL-1 α and in the development of chronic inflammation in the tumor microenvironment [4]. Additional sources of secreted IL-1 α found in the extracellular compartment include tumor-associated macrophages and plasmacytoid dendritic cells [246,247]. This IL-1 α -rich milieu has detrimental, clinically relevant consequences both at the local and systemic level. Locally, a pro-inflammatory tumor microenvironment promotes inflammation-mediated immunosuppression, cancer cell survival, and tumor progression [24,248]. Systemically, IL-1 α -driven inflammation mediates muscle loss and cachexia, clinically manifested as the loss of lean body mass and wasting syndrome typical of end-stage cancer patients [249].

Based on these observations, inhibition of IL-1 α was evaluated as a therapeutic strategy to reduce cancer growth and improve patients' clinical status. The first trials were proof-of-concept studies that evaluated the efficacy of bermekimab, a monoclonal neutralizing antibody targeting IL-1 α for the treatment of patients with advanced cancers of various origins [250,251]. Bermekimab was evaluated in an open label, Phase 1 dose escalation and expansion cohort study in patients with advanced NSCLC cancer refractory to standard therapies. Despite the small sample size limiting the statistical power, self-reported outcomes revealed pain and fatigue reduction as well as improvement in appetite, reduction in resting energy expenditure, and increase in lean body mass [250]. Following these encouraging observations, a randomized, placebo-controlled study evaluated the efficacy of this treatment in 333 patients with advanced, metastatic colorectal cancer [252]. The study met its primary and secondary endpoints, following an 8-week monotherapy course of the IL-1 α neutralizing human antibody, which resulted in increased lean body mass, decreased constitutional symptoms, decreased pain, improved quality of life, and lower circulating IL-6 and platelet counts in treated patients. Increased survival was also observed in responders compared with non-responders.

The role of IL-1 α is also likely relevant to the cytotoxic effects of chemotherapy, which can result in additional and accelerated release of IL-1 α by dying cells. Anakinra, which blocks IL-1 α binding to IL-1R1, was evaluated in patients with advanced metastatic colorectal cancer

[253] and resulted in improved survival. In addition, anakinra was administered to patients with hormone negative, metastatic breast cancer [254]. Anakinra also emerged as a suitable option for the treatment patients with advanced pancreatic cancer receiving the highly toxic FOLFIRINOX therapy [255]. Specifically, anakinra added to treatment with gemcitabine, nab-paclitaxel and cisplatin in patients with non-metastatic pancreatic ductal adenocarcinoma prior to resection resulted in reduced local cancer spread compared to historical controls receiving gemcitabine, nab-paclitaxel and cisplatin alone (NCT 0255037).

8. Conclusions

The IL-1 family member IL-1 α is a ubiquitous and pivotal pro-inflammatory cytokine. It is constitutively present in nearly all cell types as a bioactive mediator, and is released upon cell death or expressed by infiltrating myeloid cells within injured tissues. It binds the IL-1R1 receptor, shared with IL-1 β , and induces identical pro-inflammatory effects. However, IL-1 α and IL-1 β maintain non-redundant inflammatory functions. IL-1 α triggers local inflammation, which rapidly escalates into an IL-1-mediated pro-inflammatory loop characterized by secretion of IL-1 β by nearby myeloid cells.

Given its role as a bioactive precursor released upon tissue damage and cell death, IL-1 α is central to the pathogenesis of numerous conditions characterized by organ or tissue inflammation. These include disorders of the skin, lung and respiratory tract, heart and pericardium, and blood vessels, but also complex multifactorial conditions, such as COVID-19, Behcet's syndrome, or Sjogren Syndrome. Among available therapies, anakinra and riloncept target IL-1 α but also reduce IL-1 β activities; only bermekimab specifically targets IL-1 α . Unlike emerging oral NLRP3 inhibitors for reducing the activities of IL-1 β and IL-18, presently there are no oral therapies to treat IL-1 α -mediated diseases. Targeting cytosolic IL-1 α with orally active synthetic molecules could also be used to suppress inflammation.

Declaration of Competing Interest

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References

- [1] Dinarello CA. Overview of the IL-1 family in innate inflammation and acquired immunity. *Immunol Rev* 2018 Jan;281(1):8–27.
- [2] Di Paolo NC, Shayakhmetov DM. Interleukin-1 α and the inflammatory process. *Nat Immunol* 2016;17:906–13.
- [3] Rider P, Voronov E, Dinarello CA, Apte RN, Cohen I. Alarmins: feel the stress. *J Immunol* 2017 Feb 15;198(4):1395–402.
- [4] Rider P, Carmi Y, Voronov E, Apte RN. Interleukin-1 α . *Semin Immunol* 2013 Dec 15;25(6):430–8.
- [5] Malik A, Kanneganti T-D. Function and regulation of IL-1 α in inflammatory diseases and cancer. *Immunol Rev* 2018 Jan;281(1):124–37.

- [6] van de Veerdonk FL, de Graaf DM, Ab Joosten L, Dinarello CA. Biology of IL-38 and its role in disease. *Immunol Rev* 2018 Jan;281(1):191–6.
- [7] Aarden L, Brunner T, Cerottini J, Dayer J, de Weck A, Dinarello C, et al. Revised nomenclature for antigen-nonspecific T cell proliferation and helper factors. *J Immunol* 1979 Dec;123(6):2928–9.
- [8] Dinarello CA, Goldin NP, Wolff SM. Demonstration and characterization of two distinct human leukocytic pyrogens. *J Exp Med* 1974;139:1369–81.
- [9] Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. *Annu Rev Immunol* 2009;27:519–50.
- [10] Kim B, Lee Y, Kim E, Kwak A, Ryo S, Bae SH, et al. The interleukin-1alpha precursor is biologically active and is likely a key alarmin in the IL-1 family of cytokines. *Front Immunol* 2013;4:391.
- [11] Zheng Y, Humphry M, Maguire JJ, Bennett MR, Clarke MC. Intracellular interleukin-1 receptor 2 binding prevents cleavage and activity of interleukin-1alpha, controlling necrosis-induced sterile inflammation. *Immunity*. 2013 Feb 21;38(2):285–95.
- [12] Molgora M, Supino D, Mantovani A, Garlanda C. Tuning inflammation and immunity by the negative regulators IL-1R2 and IL-1R8. *Immunol Rev* 2018 Jan;281(1):233–47.
- [13] Gross O, Thomas CJ, Guarda G, Tschopp J. The inflammasome: an integrated view. *Immunol Rev* 2011 Sep;243(1):136–51.
- [14] Bianchi ME, Crippa MP, Manfredi AA, Mezzapelle R, Rovere Querini P, Venereau E. High-mobility group box 1 protein orchestrates responses to tissue damage via inflammation, innate and adaptive immunity, and tissue repair. *Immunol Rev* 2017 Nov;280(1):74–82.
- [15] Bakouche O, Brown DC, Lachman LB. Subcellular localization of human monocyte interleukin 1: evidence for an inactive precursor molecule and a possible mechanism for IL-1 release. *J Immunol* 1987;138:4249–55.
- [16] Brody DT, Durum SK. Membrane IL-1: IL-1 α precursor binds to the plasma membrane via a lectin-like interaction. *J Immunol* 1989;143:1183–7.
- [17] Kurt-Jones EA, Beller DI, Mizel SB, Unanue ER. Identification of a membrane-associated interleukin-1 in macrophages. *Proc Natl Acad Sci U S A* 1985;82:1204–8.
- [18] Cohen I, Rider P, Carmi Y, Braiman A, Dotan S, White MR, et al. Differential release of chromatin-bound IL-1alpha discriminates between necrotic and apoptotic cell death by the ability to induce sterile inflammation. *Proc Natl Acad Sci U S A* 2010 Feb 9;107(6):2574–9.
- [19] Wessendorf JHM, Garfinkel S, Zhan X, Brown S, Maciag T. Identification of a nuclear localization sequence within the structure of the human interleukin-1 α precursor. *J Biol Chem* 1993;268:22100–4.
- [20] Cohen I, Rider P, Vornov E, Tomas M, Tudor C, Wegner M, et al. Corrigendum: IL-1 α is a DNA damage sensor linking genotoxic stress signaling to sterile inflammation and innate immunity. *Sci Rep* 2016 Jan 11;6:19100.
- [21] Bessa J, Meyer CA, de Vera Mudry MC, Schlicht S, Smith SH, Iglesias A, et al. Altered subcellular localization of IL-33 leads to non-resolving lethal inflammation. *J Autoimmun* 2014 Dec;55:33–41.
- [22] Li S, Amo-Aparicio J, Neff CP, Tengesdal IW, Azam T, Palmer BE, et al. Role for nuclear interleukin-37 in the suppression of innate immunity. *Proc Natl Acad Sci U S A* 2019 Mar 5;116(10):4456–61.
- [23] Cavalli G, Dinarello CA. Suppression of inflammation and acquired immunity by IL-37. *Immunol Rev* 2018 Jan;281(1):179–90.
- [24] Mantovani A, Barajon I, Garlanda C. IL-1 and IL-1 regulatory pathways in cancer progression and therapy. *Immunol Rev* 2018 Jan;281(1):57–61.
- [25] Di Paolo NG, Shafiani S, Day T, Papayannopoulou T, Russell DW, Iwakura Y, et al. Interdependence between Interleukin-1 and Tumor Necrosis Factor Regulates TNF-Dependent Control of Mycobacterium tuberculosis Infection. *Immunity*. 2015 Dec 15;43(6):1125–36.
- [26] Maffei CM, Mirels LF, Sobel RA, Clemons KV, Stevens DA. Cytokine and inducible nitric oxide synthase mRNA expression during experimental murine cryptococcal meningoencephalitis. *Infect Immun* 2004 Apr;72(4):2338–49.
- [27] Voronov E, Dotan S, Gayvoronsky L, White RM, Cohen I, Krelin Y, et al. IL-1-induced inflammation promotes development of leishmaniasis in susceptible BALB/c mice. *Int Immunol* 2010 Apr;22(4):245–57.
- [28] Orzalli MH, Smith A, Jurado KA, Iwasaki A, Garlick JA, Kagan JC. An Antiviral Branch of the IL-1 Signaling Pathway Restricts Immune-Evasive Virus Replication. *Mol Cell* 2018 Sep 6;71(5):825–40.
- [29] Hurgin V, Novick D, Werman A, Dinarello CA, Rubinstein M. Antiviral and immunoregulatory activities of IFN-gamma depend on constitutively expressed IL-1alpha. *Proc Natl Acad Sci U S A* 2007 Mar 20;104(12):5044–9.
- [30] Nicklin MJ, Barton JL, Nguyen M, FitzGerald MG, Duff GW, Kornman K. A sequence-based map of the nine genes of the human interleukin-1 cluster. *Genomics*. 2002 May;79(5):718–25.
- [31] Dinarello CA. The IL-1 family of cytokines and receptors in rheumatic diseases. *Nat Rev Rheumatol* 2019 Oct;15(10):612–32.
- [32] Beck G, Habicht GS. Isolation and characterization of a primitive interleukin-1-like protein from an invertebrate, *Asterias forbesi*. *Proc Natl Acad Sci U S A* 1986 Oct;83(19):7429–33.
- [33] Rivers-Auty J, Daniels MJD, Colliver I, Robertson DL, Brough D. Redefining the ancestral origins of the interleukin-1 superfamily. *Nat Commun* 2018 Mar 20;9(1):1156.
- [34] Klagsbrun M. The fibroblast growth factor family: structural and biological properties. *Prog Growth Fact Res* 1989;1(4):207–35.
- [35] Cavalli G, Cenci S. Autophagy and Protein Secretion. *J Mol Biol* 2020 Apr 3;432(8):2525–45.
- [36] Kluck V, van Deuren RC, Cavalli G, Shaikat A, Arts P, Cleophas MC, et al. Rare genetic variants in interleukin-37 link this anti-inflammatory cytokine to the pathogenesis and treatment of gout. *Ann Rheum Dis* 2020 Apr;79(4):536–44.
- [37] Dinarello CA. IL-1: discoveries, controversies and future directions. *Eur J Immunol* 2010 Mar;40(3):599–606.
- [38] Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. *Immunity*. 2013 Dec 12;39(6):1003–18.
- [39] Garlanda C, Riva F, Bonavita E, Mantovani A. Negative regulatory receptors of the IL-1 family. *Semin Immunol* 2013 Nov 13;25:4087–415.
- [40] Colotta F, Re F, Muzio M, Bertini R, Polentarutti N, Sironi M, et al. Interleukin-1 type II receptor: a decoy target for IL-1 that is regulated by IL-4. *Science* 1993 Jul 23;261(5120):472–5.
- [41] McMahan CJ, Slack JL, Mosley B, Cosman D, Lupton SD, Brunton LL, et al. A novel IL-1 receptor, cloned from B cells by mammalian expression, is expressed in many cell types. *EMBO J* 1991 Oct;10(10):2821–32.
- [42] Re F, Sironi M, Muzio M, Matteucci C, Introna M, Orlando S, et al. Inhibition of interleukin-1 responsiveness by type II receptor gene transfer: a surface “receptor” with anti-interleukin-1 function. *J Exp Med* 1996 Apr 1;183(4):1841–50.
- [43] Symons JA, Young PR, Duff GW. Soluble type II interleukin 1 (IL-1) receptor binds and blocks processing of IL-1 beta precursor and loses affinity for IL-1 receptor antagonist. *Proc Natl Acad Sci U S A* 1995 Feb 28;92(5):1714–8.
- [44] Banchereau J, Pascual V, O’Garra A. From IL-2 to IL-37: the expanding spectrum of anti-inflammatory cytokines. *Nat Immunol* 2012 Oct;13(10):925–31.
- [45] Ballak DB, Li S, Cavalli G, Stahl JL, Tengesdal IW, van Diepen JA, et al. Interleukin-37 treatment of mice with metabolic syndrome improves insulin sensitivity and reduces pro-inflammatory cytokine production in adipose tissue. *J Biol Chem* 2018 Sep 14;293(37):14224–36.
- [46] Cavalli G, Justice JN, Boyle KE, D’Alessandro A, Eisenmesser EZ, Herrera JJ, et al. Interleukin 37 reverses the metabolic cost of inflammation, increases oxidative respiration, and improves exercise tolerance. *Proc Natl Acad Sci U S A* 2017 Feb 28;114(9):2313–8.
- [47] Cavalli G, Koenders M, Kalabokis V, Kim J, Tan AC, Garlanda C, et al. Treating experimental arthritis with the innate immune inhibitor interleukin-37 reduces joint and systemic inflammation. *Rheumatology (Oxford)* 2016 Dec;55(12):2220–9.
- [48] Berda-Haddad Y, Robert S, Salers P, Zekraoui L, Farnarier C, Dinarello CA, et al. Sterile inflammation of endothelial cell-derived apoptotic bodies is mediated by interleukin-1alpha. *Proc Natl Acad Sci U S A* 2011 Dec 20;108(51):20684–9.
- [49] Aksejtijevich I, Masters SL, Ferguson PJ, Dancey P, Frenkel J, van Royen-Kerkhoff A, et al. An autoinflammatory disease with deficiency of the interleukin-1-receptor antagonist. *N Engl J Med* 2009 Jun 4;360(23):2426–37.
- [50] Cavalli G, Dinarello CA. Anakinra therapy for non-cancer inflammatory diseases. *Front Pharmacol* 2018;9:1157.
- [51] Berk DR, Bayliss SJ. Neutrophilic dermatoses in children. *Pediatr Dermatol* 2008 Sep-Oct;25(5):509–19.
- [52] Tzanetakou V, Kanni T, Giatrakou S, Katoulis A, Papadavid E, Netea MG, et al. Safety and efficacy of anakinra in severe hidradenitis suppurativa: a randomized clinical trial. *JAMA Dermatol* 2016 Jan;152(1):52–9.
- [53] Kanni T, Argyropoulou M, Spyridopoulos T, Pistiki A, Stecher M, Dinarello CA, et al. MABp1 targeting interleukin-1 alpha for moderate to severe hidradenitis suppurativa not eligible for adalimumab: a randomized study. *J Invest Dermatol* 2018 Apr;138(4):795–801.
- [54] Tarte S, Gurung P, Samir P, Burton A, Kanneganti TD. Cutting edge: dysregulated CARD9 signaling in neutrophils drives inflammation in a mouse model of neutrophilic dermatoses. *J Immunol* 2018 Sep 15;201(6):1639–44.
- [55] Satoh TK, Mellett M, Contassot E, French LE. Are neutrophilic dermatoses autoinflammatory disorders? *Br J Dermatol* 2018 Mar;178(3):603–13.
- [56] Navarini AA, Satoh TK, French LE. Neutrophilic dermatoses and autoinflammatory diseases with skin involvement—innate immune disorders. *Semin Immunopathol* 2016 Jan;38(1):45–56.
- [57] Kawaguchi Y, McCarthy SA, Watkins SC, Wright TM. Autocrine activation by interleukin 1alpha induces the fibrogenic phenotype of systemic sclerosis fibroblasts. *J Rheumatol* 2004 Oct;31(10):1946–54.
- [58] Kawaguchi Y, Hara M, Wright TM. Endogenous IL-1alpha from systemic sclerosis fibroblasts induces IL-6 and PDGF-A. *J Clin Invest* 1999 May;103(9):1253–60.
- [59] Raines EW, Dower SK, Ross R. Interleukin-1 mitogenic activity for fibroblasts and smooth muscle cells is due to PDGF-AA. *Science*. 1989 Jan 20;243(4889):393–6.
- [60] Rabe KF, Hurd S, Anzueto A, Barnes PJ, Buist SA, Calverley P, et al. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. *Am J Respir Crit Care Med* 2007 Sep 15;176(6):532–55.
- [61] Fabbri LM, Rabe KF. From COPD to chronic systemic inflammatory syndrome? *Lancet*. 2007 Sep 15;370(9589):797–9.
- [62] Caramori G, Adcock IM, Casolari P, Ito K, Jazrawi E, Tsaprouni L, et al. Unbalanced oxidant-induced DNA damage and repair in COPD: a link towards lung cancer. *Thorax* 2011 Jun;66(6):521–7.
- [63] Greenberg SB, Allen M, Wilson J, Atmar RL. Respiratory viral infections in adults with and without chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2000 Jul;162(1):167–73.
- [64] Mallia P, Message SD, Gielen V, Contoli M, Gray K, Kebabdz T, et al. Experimental rhinovirus infection as a human model of chronic obstructive pulmonary disease exacerbation. *Am J Respir Crit Care Med* 2011 Mar 15;183(6):734–42.
- [65] Suwara MI, Green NJ, Borthwick LA, Mann J, Mayer-Barber KD, Barron L, et al. IL-1 α released from damaged epithelial cells is sufficient and essential to trigger

- inflammatory responses in human lung fibroblasts. *Mucosal Immunol* 2014 May;7(3):684–93.
- [66] Hill AR, Donaldson JE, Blume C, Smithers N, Tezera L, Tariq K, et al. IL-1 α mediates cellular cross-talk in the airway epithelial mesenchymal trophic unit. *Tissue Barriers* 2016 Jun 28;4(3):e1206378.
- [67] Piper SC, Ferguson J, Kay L, Parker LC, Sabroe I, Sleeman MA, et al. The role of interleukin-1 and interleukin-18 in pro-inflammatory and anti-viral responses to rhinovirus in primary bronchial epithelial cells. *PLoS One* 2013 May 28;8(5):e63365.
- [68] Osei ET, Noordhoek JA, Hackett TL, Spanjer AIR, Postma DS, Timens W, et al. Interleukin-1 α drives the dysfunctional cross-talk of the airway epithelium and lung fibroblasts in COPD. *Eur Respir J* 2016 Aug;48(2):359–69.
- [69] Tracy EC, Bowman MJ, Henderson BW, Baumann H. Interleukin-1 α is the major alarmin of lung epithelial cells released during photodynamic therapy to induce inflammatory mediators in fibroblasts. *Br J Cancer* 2012 Oct 23;107(9):1534–46.
- [70] Calverley PMA, Sethi S, Dawson M, Ward CK, Finch DK, Penney M, et al. A randomised, placebo-controlled trial of anti-interleukin-1 receptor 1 monoclonal antibody MEDI8968 in chronic obstructive pulmonary disease. *Respir Res* 2017 Aug 09;18(1):153.
- [71] Richeldi L, Collard HR, Jones MG. Idiopathic pulmonary fibrosis. *Lancet*. 2017 May 13;389(10082):1941–52.
- [72] Ogushi F, Tani K, Endo T, Tada H, Kawano T, Asano T, et al. Autoantibodies to IL-1 alpha in sera from rapidly progressive idiopathic pulmonary fibrosis. *J Med Investig* 2001 Aug;48(3–4):181–9.
- [73] Schulte JJ, Husain AN. Connective tissue disease related interstitial lung disease. *Surg Pathol Clin* 2020 Mar;13(1):165–88.
- [74] Maniwa K, Ogushi F, Tani K, Ohmoto Y, Muraguchi M, Sone S. Increased incidence of autoantibodies to interleukin-1 α in rheumatoid arthritis with interstitial lung disease. *Respirology*. 2000 Dec;5(4):315–20.
- [75] Fleischmann RM, Tesser J, Schiff MH, Schechtman J, Burmester GR, Bennett R, et al. Safety of extended treatment with anakinra in patients with rheumatoid arthritis. *Ann Rheum Dis* 2006 Aug;65(8):1006–12.
- [76] Elhai M, Meune C, Boubya M, Avouac J, Hachulla E, et al. Mapping and predicting mortality from systemic sclerosis. *Ann Rheum Dis* 2017 Nov;76(11):1897–905.
- [77] Beretta L, Cappiello F, Barili M, Bertolotti F, Scorza R. T-889C IL-1 α promoter polymorphism influences the response to oral cyclophosphamide in scleroderma patients with alveolitis. *Clin Rheumatol* 2007 Jan;26(1):88–91.
- [78] Maekawa T, Jinnin M, Ohtsuki M, Ihn H. Serum levels of interleukin-1 α in patients with systemic sclerosis. *J Dermatol* 2013 Feb;40(2):98–101.
- [79] Duan H, Fleming J, Pritchard DK, Amon LM, Xue J, Arnett HA, et al. Combined analysis of monocyte and lymphocyte messenger RNA expression with serum protein profiles in patients with scleroderma. *Arthritis Rheum* 2008 May;58(5):1465–74.
- [80] Lin E, Vincent FB, Sahhar J, et al. Analysis of serum interleukin(IL)-1 α , IL-1 β and IL-18 in patients with systemic sclerosis. *Clin Transl Immunol* 2019 Apr 6;8(4):e1045.
- [81] Aden N, Nuttall A, Shiwen X, de Winter P, Leask A, et al. Epithelial cells promote fibroblast activation via IL-1 α in systemic sclerosis. *J Invest Dermatol* 2010 Sep;130(9):2191–200.
- [82] Birnhuber A, Crnkovic S, Biasin V, Marsh LM, et al. IL-1 receptor blockade skews inflammation towards Th2 in a mouse model of systemic sclerosis. *Eur Respir J* 2019 Sep 29;54(3):1900154.
- [83] Giacomelli R, Liakouli V, Berardicurti O, Ruscitti P, Di Benedetto P. Interstitial lung disease in systemic sclerosis: current and future treatment. *Rheumatol Int* 2017 Jun;37(6):853–63.
- [84] Montgomery ST, Dittrich AS, Garratt LW, Turkovic L, Frey DL, et al. Interleukin-1 is associated with inflammation and structural lung disease in young children with cystic fibrosis. *J Cyst Fibros* 2018 Nov;17(6):715–22.
- [85] Butler B, De Dios R, Nguyen L, McKenna S, Ghosh S, Wright CJ. Developmentally regulated innate immune NF κ B signaling mediates IL-1 α expression in the perinatal murine lung. *Front Immunol* 2019 Jul 10;10:1555.
- [86] Eldredge LC, Creasy RS, Presnell S, Debley JS, Juul SE, Mayock DE, et al. Infants with evolving bronchopulmonary dysplasia demonstrate monocyte-specific expression of IL-1 in tracheal aspirates. *Am J Physiol Lung Cell Mol Phys* 2019 Jul 1; 317(1):L49–56.
- [87] Berglund M, Melgar S, Kobayashi KS, Flavell RA, et al. IL-1 receptor-associated kinase M downregulates DSS-induced colitis. *Inflamm Bowel Dis* 2010;16(10): 1778–86.
- [88] Casini-Raggi V, Kam L, Chong YJ, Fiochi C, Pizarro TT, Cominelli F. Mucosal imbalance of IL-1 and IL-1 receptor antagonist in inflammatory bowel disease. A novel mechanism of chronic intestinal inflammation. *J Immunol* 1995;154(5): 2434–40.
- [89] Voronov E, Apte RN. IL-1 in colon inflammation, colon carcinogenesis and invasiveness of colon cancer. *Cancer Microenviron* 2015;8(3):187–200.
- [90] Bersudsky M, Luski L, Fishman D, White RM, et al. Non-redundant properties of IL-1 α and IL-1 β during acute colon inflammation in mice. *Gut*. 2014;63(4): 598–609.
- [91] Menghini P, Corridoni D, Buttò LF, Osme A, Shivaswamy S, et al. Neutralization of IL-1 α ameliorates Crohn's disease-like ileitis by functional alterations of the gut microbiome. *Proc Natl Acad Sci U S A* 2019 Dec 16;116(52):26717–26.
- [92] Baird GS, Montine TJ. Multiplex immunoassay analysis of cytokines in idiopathic inflammatory myopathy. *Arch Pathol Lab Med* 2008 Feb;132(2):232–8.
- [93] De Paepe B, Creus KK, De Bleecker JH. Role of cytokines and chemokines in idiopathic inflammatory myopathies. *Curr Opin Rheumatol* 2009 Nov;21(6): 610–6.
- [94] Lundberg IE, Nyberg P. New developments in the role of cytokines and chemokines in inflammatory myopathies. *Curr Opin Rheumatol* 1998 Nov;10(6): 521–9.
- [95] Authier FJ, Mhiri C, Chazaud B, Christov C, Cherin P, Barlovatz-Meimon G, et al. Interleukin-1 expression in inflammatory myopathies: evidence of marked immunoreactivity in sarcoid granulomas and muscle fibres showing ischaemic and regenerative changes. *Neuropathol Appl Neurobiol* 1997 Apr;23(2):132–40.
- [96] Englund P, Nennesmo I, Klareskog L, Lundberg IE. Interleukin-1 α expression in capillaries and major histocompatibility complex class I expression in type II muscle fibers from polymyositis and dermatomyositis patients: important pathogenic features independent of inflammatory cell clusters in muscle tissue. *Arthritis Rheum* 2002 Apr;46(4):1044–55.
- [97] Lundberg I, Kratz AK, Alexanderson H, Patarroyo M. Decreased expression of interleukin-1 α , interleukin-1 β , and cell adhesion molecules in muscle tissue following corticosteroid treatment in patients with polymyositis and dermatomyositis. *Arthritis Rheum* 2000 Feb;43(2):336–48.
- [98] Nyberg P, Wikman AL, Nennesmo I, Lundberg I. Increased expression of interleukin 1 α and MHC class I in muscle tissue of patients with chronic, inactive polymyositis and dermatomyositis. *J Rheumatol* 2000 Apr;27(4):940–8.
- [99] Grundtman C, Salomonsson S, Dorph C, Bruton J, Andersson U, Lundberg IE. Immunolocalization of interleukin-1 receptors in the sarcolemma and nuclei of skeletal muscle in patients with idiopathic inflammatory myopathies. *Arthritis Rheum* 2007 Feb;56(2):674–87.
- [100] Sugihara T, Sekine C, Nakae T, Kohyama K, Harigai M, Iwakura Y, et al. A new murine model to define the critical pathologic and therapeutic mediators of polymyositis. *Arthritis Rheum* 2007 Apr;56(4):1304–14.
- [101] Sugihara T, Okiyama N, Watanabe N, Miyasaka N, Kohsaka H. Interleukin-1 and tumor necrosis factor α blockade treatment of experimental polymyositis in mice. *Arthritis Rheum* 2012 Aug;64(8):2655–62.
- [102] Zong M, Dorph C, Dastmalchi M, Alexanderson H, Pieper J, Amoudruz P, et al. Anakinra treatment in patients with refractory inflammatory myopathies and possible predictive response biomarkers: a mechanistic study with 12 months follow-up. *Ann Rheum Dis* 2014 May;73(5):913–20.
- [103] Groh M, Rogowska K, Monsarrat O, Denoel A, Blanche P, Guillemin L. Interleukin-1 receptor antagonist for refractory anti-MDA5 clinically amyopathic dermatomyopathy. *Clin Exp Rheumatol* 2015 Nov-Dec;33(6):904–5.
- [104] Cooper Jr LT. Myocarditis. *N Engl J Med* 2009 Apr 9;360(15):1526–38.
- [105] Bertheloot D, Latz E. HMGB1, IL-1 α , IL-33 and S100 proteins: dual-function alarmins. *Cell Mol Immunol* 2017 Jan;14(1):43–64.
- [106] De Luca G, Cavalli G, Campochiaro C, Tresoldi M, Dagna L. Myocarditis: An Interleukin-1-Mediated Disease? *Front Immunol* 2018 June 13;9:1335.
- [107] Toldo S, Kannan H, Bussani R, Anzini M, Sannino C, Sinagra G, et al. Formation of the inflammasome in acute myocarditis. *Int J Cardiol* 2014 Feb 15;171(3): e119–21.
- [108] Wang Y, Gao B, Xiong S. Involvement of NLRP3 inflammasome in CVB3-induced viral myocarditis. *Am J Physiol Heart Circ Physiol* 2014 Nov 15;307(10): H1438–47.
- [109] Shah KB, Mauro AG, Flattery M, Toldo S, Abbate A. Formation of the inflammasome during cardiac allograft rejection. *Int J Cardiol* 2015 Dec 15;201: 328–30.
- [110] Abbate A. The heart on fire: inflammasome and cardiomyopathy. *Exp Physiol* 2013 Feb;98(2):385.
- [111] McTiernan CF, Lemster BH, Frye C, Brooks S, Combes A, Feldman AM. Interleukin-1 beta inhibits phospholamban gene expression in cultured cardiomyocytes. *Circ Res* 1997 Oct;81(4):493–503.
- [112] Tatsumi T, Matoba S, Kawahara A, Keira N, Shiraishi J, Akashi K, et al. Cytokine-induced nitric oxide production inhibits mitochondrial energy production and impairs contractile function in rat cardiac myocytes. *J Am Coll Cardiol* 2000 Apr; 35(5):1338–46.
- [113] Ikonomidis I, Lekakis JP, Nikolaou M, Paraskevaidis I, Andreadou I, Kaplanoglou T, et al. Inhibition of interleukin-1 by anakinra improves vascular and left ventricular function in patients with rheumatoid arthritis. *Circulation*. 2008 May 20;117(20):2662–9.
- [114] De Luca G, Campochiaro C, Dinarello CA, Dagna L, Cavalli G. Treatment of dilated cardiomyopathy with interleukin-1 inhibition. *Ann Intern Med* 2018 Dec 4;169(11):819–20.
- [115] Cavalli G, Foppoli M, Cabrini L, Dinarello CA, Tresoldi M, Dagna L. Interleukin-1 receptor blockade rescues myocarditis-associated end-stage heart failure. *Front Immunol* 2017 Feb 9;8:131.
- [116] Cavalli G, Pappalardo F, Mangieri A, Dinarello CA, Dagna L, Tresoldi M. Treating life-threatening myocarditis by blocking interleukin-1. *Crit Care Med* 2016 Aug; 44(8):e751–4.
- [117] Parisi F, Paglionico A, Varriano V, Ferraccioli G, Gremese E. Refractory adult-onset Still disease complicated by macrophage activation syndrome and acute myocarditis: a case report treated with high doses (8 mg/kg/d) of anakinra. *Medicine (Baltimore)* 2017 Jun;96(24):e6656.
- [118] Thygesen K, Alpert JS, Jaffe AS, Simoons ML, Chaitman BR, White HD, et al. Third universal definition of myocardial infarction. *J Am Coll Cardiol* 2012 Oct 16;60(16):1581–98.
- [119] Anderson JL, Morrow DA. Acute myocardial infarction. *N Engl J Med* 2015 May 25;376(21):2053–64.
- [120] Frangogiannis NG. The inflammatory response in myocardial injury, repair, and remodelling. *Nat Rev Cardiol* 2014 May;11(5):255–65.
- [121] Turner NA, Das A, Warburton P, O'Regan DJ, Ball SG, Porter KE. Interleukin-1 α stimulates proinflammatory cytokine expression in human cardiac myofibroblasts. *Am J Physiol Heart Circ Physiol* 2009 Sep;297(3):H1117–27.

- [122] Sun Y, Weber KT. Infarct scar: a dynamic tissue. *Cardiovasc Res* 2000 May;46(2):250–6.
- [123] Dinarello CA. Biological basis for interleukin-1 in disease. *Blood*. 1996;87:2095–147.
- [124] Suzuki K, Murtuza B, Smolenski RT, Sammut IA, Suzuki N, Kaneda Y, et al. Overexpression of interleukin-1 receptor antagonist provides cardioprotection against ischemia-reperfusion injury associated with reduction in apoptosis. *Circulation*. 2001 Sep 18;104(12 Suppl 1) (I308-I303).
- [125] Furuichi K, Wada T, Iwata Y, Kokubo S, Hara A, Yamahana J, et al. Interleukin-1-dependent sequential chemokine expression and inflammatory cell infiltration in ischemia-reperfusion injury. *Crit Care Med* 2006 Sep;34(9):2447–55.
- [126] Pinteaux E, Rothwell NJ, Boutin H. Neuroprotective actions of endogenous interleukin-1 receptor antagonist (IL-1ra) are mediated by glia. *Glia*. 2006 Apr 1;53(5):551–6.
- [127] Bujak M, Dobaczewski M, Chatila K, Mendoza LH, Li N, Reddy A, et al. Interleukin-1 receptor type 1 signaling critically regulates infarct healing and cardiac remodeling. *Am J Pathol* 2008 Jul;173(1):57–67.
- [128] Mauro AG, Mezzaroma E, Torrado J, Kundur P, Joshi P, Stroud K, et al. Reduction of myocardial ischemia-reperfusion injury by inhibiting interleukin-1 alpha. *J Cardiovasc Pharmacol* 2017 Mar;69(3):156–60.
- [129] Toldo S, Mezzaroma E, Van Tassel BW, Farkas D, Marchetti C, Voelkel NF, et al. Interleukin-1beta blockade improves cardiac remodeling after myocardial infarction without interrupting the inflammasome in the mouse. *Exp Physiol* 2013 Mar;98(3):734–45.
- [130] Chen CJ, Kono H, Golenbock D, Reed G, Akira S, Rock KL. Identification of a key pathway required for the sterile inflammatory response triggered by dying cells. *Nat Med* 2007 Jul;13(7):851–6.
- [131] Abbate A, Van Tassel BW, Biondi-Zoccai G, Kontos MC, Grizzard JD, Spillman DW, et al. Effects of interleukin-1 blockade with anakinra on adverse cardiac remodeling and heart failure after acute myocardial infarction [from the Virginia Commonwealth University-Anakinra Remodeling Trial (2) (VCU-ART2) pilot study]. *Am J Cardiol* 2013 May 15;111(10):1394–400.
- [132] Abbate A, Kontos MC, Grizzard JD, Biondi-Zoccai GG, Van Tassel BW, Robati R, et al. Interleukin-1 blockade with anakinra to prevent adverse cardiac remodeling after acute myocardial infarction (Virginia Commonwealth University Anakinra Remodeling Trial [VCU-ART] Pilot study). *Am J Cardiol* 2010 May 15;105(10):1371–7.
- [133] Morton AC, Rothman AM, Greenwood JP, Gunn J, Chase A, Clarke B, et al. The effect of interleukin-1 receptor antagonist therapy on markers of inflammation in non-ST elevation acute coronary syndromes: the MRC-ILA Heart Study. *Eur Heart J* 2015 Feb 7;36(6):377–84.
- [134] Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med* 2017 Sep 21;377(12):1119–31.
- [135] Buckley LF, Abbate A. Interleukin-1 blockade in cardiovascular diseases: a clinical update. *Eur Heart J* 2018 Jun 7;39(22):2063–9.
- [136] Imazio M, Gaita F. Acute and recurrent pericarditis. *Cardiol Clin* 2017;35(4):505–13.
- [137] Brucato A, Emmi G, Cantarini L, et al. Management of idiopathic recurrent pericarditis in adults and in children: a role for IL-1 receptor antagonism. *Intern Emerg Med* 2018;13(4):475–89.
- [138] Picco P, Brisca G, Traverso F, Loy A, Gattorno M, Martini A. Successful treatment of idiopathic recurrent pericarditis in children with interleukin-1β receptor antagonist (anakinra): an unrecognized autoinflammatory disease? *Arthritis Rheum* 2009;60(1):264–8.
- [139] Galluzzo A, Imazio M. Advances in medical therapy for pericardial diseases. *Expert Rev Cardiovasc Ther* 2018;16(9):635–43.
- [140] Brucato A, Imazio M, Gattorno M, et al. Effect of Anakinra on recurrent pericarditis among patients with colchicine resistance and corticosteroid dependence. *JAMA*. 2016 Nov 8;316(18):1906–12.
- [141] Imazio M, Andreis A, De Ferrari GM, et al. Anakinra for corticosteroid-dependent and colchicine-resistant pericarditis: the IRAP (international registry of Anakinra for pericarditis) study. *Eur J Prev Cardiol* 2020 Jun;27(9):956–64.
- [142] Adler Y, Charron P, Imazio M, Badano L, Barón-Esquivias G, Bogaert J, et al. 2015 ESC Guidelines for the diagnosis and management of pericardial diseases: The Task Force for the Diagnosis and Management of Pericardial Diseases of the European Society of Cardiology (ESC) Endorsed by: The European Association for Cardio-Thoracic Surgery (EACTS). *Eur Heart J* 2015 Nov 7;36(42):2921–64.
- [143] Caorsi R, Insalaco A, Longo C, Martini G, Cattalini M, Consolini R, et al. IL-1 blockade in pediatric recurrent pericarditis: a multicentric retrospective study on the Italian cohort. *Ann Rheum Dis* 2019 June. <https://doi.org/10.1136/annrheumdis-2019-eular.5854>.
- [144] Thornton P, McColl BW, Greenhalgh A, Denes A, Allan SM, Rothwell NJ. Platelet interleukin-1alpha drives cerebrovascular inflammation. *Blood*. 2010 Apr 29;115(17):3632–9.
- [145] Burzynski LC, Humphry M, Pырillou K, Wiggins KA, Chan JNE, Figg N, et al. The coagulation and immune systems are directly linked through the activation of interleukin-1alpha by Thrombin. *Immunity*. 2019 Apr 16;50(4):1033–42.
- [146] Bester J, Matshailwe C, Pretorius E. Simultaneous presence of hypercoagulation and increased clot lysis time due to IL-1beta, IL-6 and IL-8. *Cytokine*. 2018 Oct;110:237–42.
- [147] Tunjungputri RN, Li Y, de Groot PG, Dinarello CA, Smeekens SP, Jaeger M, et al. The inter-relationship of platelets with interleukin-1beta-mediated inflammation in humans. *Thromb Haemostasis* 2018 Dec;118(12):2112–25.
- [148] Brunn GJ, Saadi S, Platt JL. Constitutive repression of interleukin-1alpha in endothelial cells. *Circ Res* 2008 Apr 11;102(7):823–30.
- [149] Nishimura S, Nagasaki M, Kunishima S, Sawaguchi A, Sakata A, Sakaguchi H, et al. IL-1alpha induces thrombopoiesis through megakaryocyte rupture in response to acute platelet needs. *J Cell Biol* 2015 May 11;209(3):453–66.
- [150] Folco EJ, Mawson TL, Vromman A, Bernardes-Souza B, Franck G, Persson O, et al. Neutrophil extracellular traps induce endothelial cell activation and tissue factor production through interleukin-1alpha and Cathepsin G. *Arterioscler Thromb Vasc Biol* 2018 Aug;38(8):1901–12.
- [151] Bettiol A, Prisco D, Emmi G. Behcet: the syndrome. *Rheumatology (Oxford)* 2020 May 1;59(Supplement_3):iii101–7.
- [152] Emmi G, Silvestri E, Squatrito D, D'Elia MM, Ciucciarelli L, Prisco D, et al. Behcet's syndrome pathophysiology and potential therapeutic targets. *Intern Emerg Med* 2014 Apr;9(3):257–65.
- [153] Seyahi E. Phenotypes in Behcet's syndrome. *Intern Emerg Med* 2019 Aug;14(5):677–89.
- [154] Becatti M, Emmi G, Bettiol A, Silvestri E, Di Scala G, Taddei N, et al. Behcet's syndrome as a tool to dissect the mechanisms of thrombo-inflammation: clinical and pathogenetic aspects. *Clin Exp Immunol* 2019 Mar;195(3):322–33.
- [155] Emmi G, Bettiol A, Silvestri E, Di Scala G, Becatti M, Fiorillo C, et al. Vascular Behcet's syndrome: an update. *Intern Emerg Med* 2019 Aug;14(5):645–52.
- [156] Bettiol A, Hatemi G, Vannozzi L, Barilaro A, Prisco D, Emmi G. Treating the different phenotypes of Behcet's syndrome. *Front Immunol* 2019;10:2830.
- [157] Yazici H, Ugurlu S, Seyahi E. Behcet syndrome: is it one condition? *Clin Rev Allergy Immunol* 2012 Dec;43(3):275–80.
- [158] Khalid U, Saleem T. Hughes-Stovin syndrome. *Orphanet J Rare Dis* 2011 Apr 13;6:15.
- [159] Becatti M, Emmi G, Bettiol A, Silvestri E, Di Scala G, Taddei N, et al. Behcet's syndrome as a tool to dissect the mechanisms of thrombo-inflammation: clinical and pathogenetic aspects. *Clin Exp Immunol* 2019 Mar;195(3):322–33.
- [160] Emmi G, Becatti M, Bettiol A, Hatemi G, Prisco D, Fiorillo C. Behcet's syndrome as a model of thrombo-inflammation: The role of neutrophils. *Front Immunol* 2019 May 14;10:1085.
- [161] Silvestri E, Emmi G, Prisco D. Vascular Behcet's disease: new insights in the management of thrombosis. *Expert Rev Cardiovasc Ther* 2013 Dec;11(12):1583–5.
- [162] Karasneh J, Hajeer AH, Barrett J, Ollier WER, Thornhill M, Gul A. Association of specific interleukin 1 gene cluster polymorphisms with increased susceptibility for Behcet's disease. *Rheumatology (Oxford)* 2003 Jul;42(7):860–4.
- [163] Song GG, Kim J-H, Lee YH. Associations between interleukin-1 polymorphisms and susceptibility to vasculitis: a meta-analysis. *Z Rheumatol* 2016 May;75(4):406–15.
- [164] Keller M, Spanou Z, Schaerli P, Britschgi M, Yawalkar N, et al. T cell-regulated neutrophilic inflammation in autoinflammatory diseases. *J Immunol* 2005 Dec 1;175(11):7678–86.
- [165] Erkan F, Gül A, Tasali E. Pulmonary manifestations of Behcet's disease. *Thorax*. 2001 Jul;56(7):572–8.
- [166] Kawakami T, Ohashi S, Kawa Y, Takahama H, Ito M, Soma Y, et al. Elevated serum granulocyte colony-stimulating factor levels in patients with active phase of sweet syndrome and patients with active behcet disease: implication in neutrophil apoptosis dysfunction. *Arch Dermatol* 2004 May;140(5):570–4.
- [167] Mantovani A, Cassatella MA, Costantini C, Jaillon S. Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat Rev Immunol* 2011 Jul 25;11(8):519–31.
- [168] Clancy DM, Henry CM, Sullivan GP, Martin SJ. Neutrophil extracellular traps can serve as platforms for processing and activation of IL-1 family cytokines. *FEBS J* 2017 Jun;284(11):1712–25.
- [169] Becatti M, Emmi G, Silvestri E, Bruschi G, Ciucciarelli L, Squatrito D, et al. Neutrophil activation promotes fibrinogen oxidation and thrombus formation in Behcet disease. *Circulation*. 2016 Jan 19;133(3):302–11.
- [170] Lee Y, Wakita D, Dagvadorj J, Shimada K, Chen S, Huang G, et al. IL-1 Signaling Is critically required in stromal cells in Kawasaki disease vasculitis mouse model: role of both IL-1α and IL-1β. *Arterioscler Thromb Vasc Biol* 2015 Dec;35(12):2605–16.
- [171] Yates CM, Abdelhamid M, Adam DJ, Nash GB, Bradbury AW, Ed Rainger G. Endovascular aneurysm repair reverses the increased titer and the inflammatory activity of interleukin-1α in the serum of patients with abdominal aortic aneurysm. *J Vasc Surg* 2011 Aug;54(2):497–503.
- [172] Marchesi A, Tarissi de Jacobis I, Rigante D, Rimini A, Malorni W, Corsello G, et al. Kawasaki disease: guidelines of the Italian Society of Pediatrics, part I - definition, epidemiology, etiopathogenesis, clinical expression and management of the acute phase. *Ital J Pediatr* 2018 Aug 30;44(1):102.
- [173] Marchesi A, Tarissi de Jacobis I, Rigante D, Rimini A, Malorni W, Corsello G, et al. Kawasaki disease: guidelines of the Italian Society of Pediatrics, part II - treatment of resistant forms and cardiovascular complications, follow-up, lifestyle and prevention of cardiovascular risks. *Ital J Pediatr* 2018 Aug 30;44(1):103.
- [174] Esper F, et al. Association between a novel human coronavirus and Kawasaki disease. *J Infect Dis* 2005;191:499–502.
- [175] Maggio MC, Cimaz R, Alaimo A, Comparato C, Di Lisi D, Corsello G. Kawasaki disease triggered by parvovirus infection: an atypical case report of two siblings. *J Med Case Rep* 2019 Apr 24;13(1):104.
- [176] Alphonse MP, et al. Inositol- triphosphate 3- kinase C mediates inflammasome activation and treatment response in Kawasaki disease. *J Immunol* 2016;197:3481–9.
- [177] Maury CP, Salo E, Pelkonen P. Circulating interleukin-1 beta in patients with Kawasaki disease. *N Engl J Med* 1988;319(25):1670–1.
- [178] Senzaki H. The pathophysiology of coronary artery aneurysms in kawasaki disease: Role of matrix metalloproteinases. *Arch Dis Child* 2006;91:847–51.

- [179] Makata H, Ichiyama T, Uchi R, Takekawa T, Matsubara T, Furukawa S. Anti-inflammatory effect of intravenous immunoglobulin in comparison with dexamethasone in vitro: implication for treatment of Kawasaki disease. *Naunyn-Schmiedeberg's Arch Pharmacol* 2006;373:325–32.
- [180] Bhol KC, Desai A, Kumari S, Colon JE, Ahmed AE. Pemphigus vulgaris: The role of IL-1 and IL-1 receptor antagonist in pathogenesis and effects of intravenous immunoglobulin on their production. *Clin Immunol* 2001 Aug;100(2):172–80.
- [181] Suzuki H, Uemura S, Tone S, Iizuka T, Koike M, Hirayama K, et al. Effects of immunoglobulin and gamma-interferon on the production of tumour necrosis factor-alpha and interleukin-1 beta by peripheral blood monocytes in the acute phase of Kawasaki disease. *Eur J Pediatr* 1996;155:291–6.
- [182] Weng K-P, Hsieh K-S, Ho T-Y, Huang S-H, Lai C-R, Chiu Y-T, et al. IL-1beta polymorphism in association with initial intravenous immunoglobulin treatment failure in taiwanese children with kawasaki disease. *Circ J* 2010 Mar;74(3):544–51.
- [183] Tremoulet AH, Jain S, Kim S, Newburger J, Arditi M, Franco A, et al. Rationale and study design for a phase I/IIa trial of anakinra in children with Kawasaki disease and early coronary artery abnormalities (the ANAKID trial). *Contemp Clin Trials* 2016 May;48:70–5.
- [184] Kone-Paut I, Cimaz R, Herberg J, Bates O, Carbasse A, Saulnier JP, et al. The use of interleukin 1 receptor antagonist (anakinra) in Kawasaki disease: A retrospective cases series. *Autoimmun Rev* 2018 Aug;17(8):768–74.
- [185] Gambacorta A, Buonsenso D, De Rosa G, Lazzareschi I, Gatto A, Brancato F, et al. Resolution of giant coronary aneurysms in a child with refractory Kawasaki disease treated with Anakinra. *Front Pediatr* 2020 May 7;8:195.
- [186] Wu F, Zhao S, Yu B, Chen Y, Wang W, et al. A new coronavirus associated with human respiratory disease in China. *Nature*. 2020 Mar;579(7798):265–9.
- [187] Michaudel C, Maillet I, Fauconnier L, Quessiaux V, Chung KF, Wiegman C, et al. Interleukin-1 α mediates ozone-induced myeloid differentiation factor-88-dependent epithelial tissue injury and inflammation. *Front Immunol* 2018 May7;9:916.
- [188] Colafrancesco S, Alessandri C, Conti F, Priori R. COVID-19 gone bad: A new character in the spectrum of the hyperferritinemic syndrome? *Autoimmun Rev* 2020 Jul;19(7):102573.
- [189] McGonagle D, Sharif K, O'Regan A, Bridgewood C. The role of cytokines including interleukin-6 in COVID-19 induced pneumonia and macrophage activation syndrome-like disease. *Autoimmun Rev* 2020 Jun;19:6:102537.
- [190] McGonagle D, O'Donnell JS, Sharif K, Emery P, Bridgewood C. Immune mechanisms of pulmonary intravascular coagulopathy in COVID-19 pneumonia. *Lancet Rheumatol* 2020 Jul;2(7):e437–45.
- [191] Matthay MA, Zemans RL, Zimmerman GA, Arabi YM, et al. Acute respiratory distress syndrome. *Nat Rev Dis Primers* 2019 Mar 14;5(1):18.
- [192] Park WY, Goodman RB, Steinberg KP, Ruzinski JT, et al. Cytokine balance in the lungs of patients with acute respiratory distress syndrome. *Am J Respir Crit Care Med* 2001 Nov 15;164(10 Pt 1):1896–903.
- [193] Dahmer MK, Quasney MW, Sapru A, Gildengorin G, Curley MAQ, et al. Interleukin-1 receptor antagonist is associated with pediatric acute respiratory distress syndrome and worse outcomes in children with acute respiratory failure. *Pediatr Crit Care Med* 2018 Oct;19(10):930–8.
- [194] Awasthi S, Yadav KK, Pandey M, Mahdi AA, Awasthi N. Interleukin 1 receptor antagonist (IL1RA) gene polymorphism and levels associated with adverse outcome in severe community-acquired pneumonia in children: A hospital-based study in India. *Pediatr Pulmonol* 2018 Sep;53(9):1276–83.
- [195] Colafrancesco S, Scriver R, Barbati C, Conti F, Priori R. Targeting the immune system for pulmonary inflammation and cardiovascular complications in COVID-19 patients. *Front Immunol* 2020 Jun 23;11:1439.
- [196] Cavalli G, De Luca G, Campochiaro C, Della-Torre E, Ripa M, Canetti D, et al. Interleukin-1 blockade with high-dose anakinra in patients with COVID-19, acute respiratory distress syndrome, and hyperinflammation: a retrospective cohort study. *Lancet Rheumatol* 2020;2(6):e325–31.
- [197] Huet T, Beaussier H, Voisin O, Jouveshomme S, Dauriat G. Anakinra for severe forms of COVID-19: a cohort study. *Lancet Rheumatol* 2020 Jul;2(7):e393–400.
- [198] Navarro-Millán I, Sattui SE, Lakhanpal A, Zisa D, Siegel CH, Crow MK. Use of anakinra to prevent mechanical ventilation in severe COVID-19: A case series. *Arthritis Rheum* 2020 Jun 30. <https://doi.org/10.1002/art.41422>.
- [199] Aouba A, Baldolli A, Geffray L, Verdon R, Bergot E, Martin-Silva N, et al. Targeting the inflammatory cascade with anakinra in moderate to severe COVID-19 pneumonia: case series. *Ann Rheum Dis* 2020 Oct;79(10):1381–2.
- [200] González-García A, García-Sánchez I, Lopes V, Muñoz Moreno-Arrones O, et al. Successful treatment of severe COVID-19 with subcutaneous anakinra as a sole treatment. *Rheumatology (Oxford)* 2020 Aug 1;59(8):2171–3.
- [201] Dimopoulos G, de Mast Q, Markou N, Theodorakopoulou M, Komnos A. Favorable anakinra responses in severe Covid-19 Patients with secondary hemophagocytic lymphohistiocytosis. *Cell Host Microbe* 2020 Jul 8;28(1):117–123.e1.
- [202] Filocamo G, Mangioni D, Tagliabue P, Aliberti S, Costantino G, Minoia F, et al. Use of anakinra in severe COVID-19: A case report. *Int J Infect Dis* 2020 Jul;96:607–9.
- [203] Pontali E, Volpi S, Antonucci G, Castellaneta M, Buzzi D. Safety and efficacy of early high-dose IV anakinra in severe COVID-19 lung disease. *J Allergy Clin Immunol* 2020 Jul;146(1):213–5.
- [204] Franzetti M, Pozzetti U, Carugati M, Pandolfo A, Molteni C. Interleukin-1 receptor antagonist anakinra in association with remdesivir in severe COVID-19: A case report. *Int J Infect Dis* 2020 Aug;97:215–8.
- [205] Guaraldi G, Meschiari M, Cozzi-Lepri A, et al. Tocilizumab in patients with severe COVID-19: a retrospective cohort study. *Lancet Rheumatol* 2020. Aug;2(8):e474–84.
- [206] Campochiaro C, Della-Torre E, Cavalli G, et al. Efficacy and safety of tocilizumab in severe COVID-19 patients: a single-centre retrospective cohort study. *Eur J Intern Med* 2020 Jun;76:43–9.
- [207] Della-Torre E, Campochiaro C, Cavalli G, et al. Interleukin-6 blockade with sarilumab in severe COVID-19 pneumonia with systemic hyperinflammation: an open-label cohort study. *Ann Rheum Dis* 2020 Oct;79(10):1277–85.
- [208] Press release Roche - Roche provides an update on the phase III COVACTA trial of Actemra/RoActemra in hospitalised patients with severe COVID-19 associated pneumonia. <https://www.roche.com/investors/updates/inv-update-2020-07-29.htm>; 2020.
- [209] Press release - Sanofi and Regeneron provide update on Kevzara® (sarilumab) Phase 3 U.S. trial in COVID-19 patients - Sanofi. <https://www.sanofi.com/-/media/Project/One-Sanofi-Web/Websites/Global/Sanofi-COM/Home/media-room/press-releases/2020/2020-07-02-22-30-00-2057183-en.pdf>.
- [210] Shakoory B, Carrillo JA, Chatham WW, Amdur RL, et al. Interleukin-1 receptor blockade is associated with reduced mortality in sepsis patients with features of macrophage activation syndrome: reanalysis of a prior phase III trial. *Crit Care Med* 2016 Feb;44(2):275–81.
- [211] Lener T, Yao Q. Macrophage activation syndrome complicating adult onset Still's disease: A single center case series and comparison with literature. *Semin Arthritis Rheum* 2016 Jun;45(6):711–6.
- [212] Verdoni L, Mazza A, Gervasoni A, Martelli L, Ruggeri M, et al. An outbreak of severe Kawasaki-like disease at the Italian epicentre of the SARS-CoV-2 epidemic: an observational cohort study. *Lancet*. 2020 Jun 6;395(10239):1771–8.
- [213] Belhadjer Z, Méot M, Bajolle F, Khraiche D, Legendre A, et al. Acute heart failure in multisystem inflammatory syndrome in children (MIS-C) in the context of global SARS-CoV-2 pandemic. *Circulation*. 2020 May 17. <https://doi.org/10.1161/CIRCULATIONAHA.120.048360>.
- [214] Jones VG, Mills M, Suarez D, Hogan CA, Yeh D, Segal JB, et al. COVID-19 and Kawasaki Disease: Novel virus and novel case. *Hosp Pediatr* 2020 Jun;10(6):537–40.
- [215] Brito-Zerón P, Baldini C, Bootsma H, Bowman SJ, Jonsson R, et al. Sjögren syndrome. *Nat Rev Dis Primers* 2016 Jul 7;2:16047.
- [216] Barone F, Colafrancesco S. Sjögren's syndrome: from pathogenesis to novel therapeutic targets. *Clin Exp Rheumatol Jul-Aug* 2016;34(4 Suppl 98):58–62.
- [217] Bolstad AI, Jonsson R. The role of apoptosis in Sjögren's syndrome. *Ann Med Interne (Paris)* 1998 Feb;149(1):25–9.
- [218] Skopouli FN, Katsiogiannis S. How stress contributes to autoimmunity-lessons from Sjögren's syndrome. *FEBS Lett* 2018 Jan;592(1):5–14.
- [219] Nandula SR, Dey P, Corbin KL, Nunemaker CS, Bagavant H, Deshmukh US. Salivary gland hypofunction induced by activation of innate immunity is dependent on type I interferon signaling. *J Oral Pathol Med* 2013 Jan;42(1):66–72.
- [220] Deshmukh US, Nandula SR, Thimmalapara PR, Scindia YM, Bagavant H. Activation of innate immune responses through Toll-like receptor 3 causes a rapid loss of salivary gland function. *J Oral Pathol Med* 2009 Jan;38(1):42–7.
- [221] Ciccia F, Accardo-Palumbo A, Alessandro R, Alessandri C, Priori R, Guggino G, et al. Interleukin-36 α axis is modulated in patients with primary Sjögren's syndrome. *Clin Exp Immunol* 2015 Aug;181(2):230–8.
- [222] Niu L, Zhang S, Wu J, Chen L, Wang Y. Upregulation of NLRP3 inflammasome in the tears and ocular surface of dry eye patients. *PLoS One* 2015 May 11;10(5):e0126277.
- [223] Kiripolsky J, McCabe LG, Kramer JM. Innate immunity in Sjögren's syndrome. *Clin Immunol* 2017 Sep;182:4–13.
- [224] Dietrich J, Schlegel C, Roth M, Witt J, Geerling G, Mertsch S, et al. Comparative analysis on the dynamic of lacrimal gland damage and regeneration after Interleukin-1 α or duct ligation induced dry eye disease in mice. *Exp Eye Res* 2018 Jul;172:66–77.
- [225] Zoukhri D, Macari E, Kublin CL. A single injection of interleukin-1 induces reversible aqueous-tear deficiency, lacrimal gland inflammation, and acinar and ductal cell proliferation. *Exp Eye Res* 2007 May;84(5):894–904.
- [226] Chen YT, Nikulina K, Lazarev S, Bahrami AF, Noble LB, Gallup M, et al. Interleukin-1 as a phenotypic immunomodulator in keratinizing squamous metaplasia of the ocular surface in Sjögren's syndrome. *Am J Pathol* 2010 Sep;177(3):1333–43.
- [227] Pflugfelder SC, Jones D, Ji Z, Afonso A, Monroy D. Altered cytokine balance in the tear fluid and conjunctiva of patients with Sjögren's syndrome keratoconjunctivitis sicca. *Curr Eye Res* 1999 Sep;19(3):201–11.
- [228] Solomon A, Dursun D, Liu Z, Xie Y, Macri A, Pflugfelder SC. Pro- and anti-inflammatory forms of interleukin-1 in the tear fluid and conjunctiva of patients with dry-eye disease. *Invest Ophthalmol Vis Sci* 2001 Sep;42(10):2283–92.
- [229] López-Miguel A, Tesón M, Martín-Montañez V, Enríquez-de-Salamanca A, et al. Clinical and molecular inflammatory response in Sjögren syndrome-associated dry eye patients under desiccating stress. *Am J Ophthalmol* 2016 Jan;161:133–41.e1-2.
- [230] Dubost JJ, Perrier S, Afane M, Viallard JL, et al. IL-1 receptor antagonist in saliva: characterization in normal saliva and reduced concentration in Sjögren's syndrome (SS). *Clin Exp Immunol* 1996 Nov;106(2):237–42.
- [231] Vijmasi T, Chen FYT, Chen YT, Gallup M, McNamara N. Topical administration of interleukin-1 receptor antagonist as a therapy for aqueous-deficient dry eye in autoimmune disease. *Mol Vis* 2013 Sep 19;19:1957–65.
- [232] Goldstein MH, Martel JR, Sall K, Goldberg DF, Abrams M, et al. Multicenter study of a novel topical interleukin-1 receptor inhibitor, Isunakinra, in subjects with moderate to severe dry eye disease. *Eye Contact Lens* 2017 Sep;43(5):287–96.

- [233] Brække Norheim K, Harboe E, Gøransson LG, Omdal R. Interleukin-1 inhibition and fatigue in primary Sjögren's syndrome—a double blind, randomised clinical trial. *PLoS One* 2012;7(1):e30123.
- [234] Cavalli G, Dagna L, Biavasco R, Doglioni C, Ferrero E, Ferrarini M. Erdheim-Chester disease: an in vivo human model of macrophage activation at the crossroad between chronic inflammation and cancer. *J Leukoc Biol* 2020 Aug;108(2):591–9.
- [235] Campochiaro C, Tomelleri A, Cavalli G, Berti A, Dagna L. Erdheim-Chester disease. *Eur J Intern Med* 2015 May;26(4):223–9.
- [236] Chiapparini L, Cavalli G, Langella T, Venerando A, De Luca G, Raspante S, et al. Adult leukoencephalopathies with prominent infratentorial involvement can be caused by Erdheim-Chester disease. *J Neurol* 2018 Feb;265(2):273–84.
- [237] Ferrero E, Corti A, Haroche J, Belloni D, Colombo B, Berti A, et al. Plasma Chromogranin A as a marker of cardiovascular involvement in Erdheim-Chester disease. *Oncoimmunology*. 2016 Jul;5(7):e1181244.
- [238] Cavalli G, Berti A, Campochiaro C, Dagna L. Diagnosing Erdheim-Chester disease. *Ann Rheum Dis* 2013 Jul;72(7):e19.
- [239] Pacini G, Cavalli G, Tomelleri A, De Luca G, Pacini G, Ferrarini M, et al. The fibrogenic chemokine CCL18 is associated with disease severity in Erdheim-Chester disease. *Oncoimmunology*. 2018;7(7):e1440929.
- [240] Cavalli G, Guglielmi B, Berti A, Campochiaro C, Sabbadini MG, Dagna L. The multifaceted clinical presentations and manifestations of Erdheim-Chester disease: comprehensive review of the literature and of 10 new cases. *Ann Rheum Dis* 2013 Oct;72(10):1691–5.
- [241] Aouba A, Georgin-Lavialle S, Pagnoux C, Martin Silva N, Renand A, Galateau-Salle F, et al. Rationale and efficacy of interleukin-1 targeting in Erdheim-Chester disease. *Blood*. 2010 Nov 18;116(20):4070–6.
- [242] Tomelleri A, Cavalli G, De Luca G, Campochiaro C, D'Aliberti T, Tresoldi M, et al. Treating heart inflammation with interleukin-1 blockade in a case of Erdheim-Chester disease. *Front Immunol* 2018;9:1233.
- [243] Cohen-Aubart F, Maksud P, Saadoun D, Drier A, Charlotte F, Cluzel P, et al. Variability in the efficacy of the IL1 receptor antagonist anakinra for treating Erdheim-Chester disease. *Blood*. 2016 Mar 17;127(11):1509–12.
- [244] Diamond EL, Abdel-Wahab O, Durham BH, Dogan A, Ozkaya N, Brody L, et al. Anakinra as efficacious therapy for 2 cases of intracranial Erdheim-Chester disease. *Blood*. 2016 Oct 06;128(14):1896–8.
- [245] Stevenson FT, Turck J, Locksley RM, Lovett DH. The N-terminal propeptide of interleukin 1 alpha is a transforming nuclear oncoprotein. *Proc Natl Acad Sci U S A* 1997 Jan 21;94(2):508–13.
- [246] Sorrentino R, Terlizzi M, Di Crescenzo VG, Popolo A, Pecoraro M, Perillo G, et al. Human lung cancer-derived immunosuppressive plasmacytoid dendritic cells release IL-1alpha in an AIM2 inflammasome-dependent manner. *Am J Pathol* 2015 Nov;185(11):3115–24.
- [247] Terlizzi M, Colarusso C, Popolo A, Pinto A, Sorrentino R. IL-1alpha and IL-1beta-producing macrophages populate lung tumor lesions in mice. *Oncotarget*. 2016 Sep 6;7(36):58181–92.
- [248] Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature*. 2008 Jul 24;454(7203):436–44.
- [249] Dinarello CA. Interleukin-1alpha neutralisation in patients with cancer. *Lancet Oncol* 2014 May;15(6):552–3.
- [250] Hong DS, Janku F, Naing A, Falchook GS, Piha-Paul S, Wheler JJ, et al. Xilonix, a novel true human antibody targeting the inflammatory cytokine interleukin-1 alpha, in non-small cell lung cancer. *Investig New Drugs* 2015 Jun;33(3):621–31.
- [251] Hong DS, Hui D, Bruera E, Janku F, Naing A, Falchook GS, et al. MABp1, a first-in-class true human antibody targeting interleukin-1 α in refractory cancers: an open-label, phase 1 dose-escalation and expansion study. *Lancet Oncol* 2014 May;15(6):656–66.
- [252] Hickish T, Andre T, Wyrwicz L, Saunders M, Sarosiek T, Kocsis J, et al. MABp1 as a novel antibody treatment for advanced colorectal cancer: a randomised, double-blind, placebo-controlled, phase 3 study. *Lancet Oncol* 2017 Feb;18(2):192–201.
- [253] Isambert N, Hervieu A, Rébé C, Hennequin A, Borg C, Zanetta S, et al. Fluorouracil and bevacizumab plus anakinra for patients with metastatic colorectal cancer refractory to standard therapies (IRAFU): a single-arm phase 2 study. *Oncoimmunology*. 2018 Aug 1;7(9):e1474319.
- [254] Wu TC, Xu K, Martinek J, Young RR, Banchereau R, George J, et al. IL1 receptor antagonist controls transcriptional signature of inflammation in patients with metastatic breast cancer. *Cancer Res* 2018 Sep 15;78(18):5243–58.
- [255] Whiteley A, Becerra C, McCollum D, Paulson AS, Goel A. A pilot, non-randomized evaluation of the safety of anakinra plus FOLFIRINOX in metastatic pancreatic ductal adenocarcinoma patients. *J Clin Oncol* 2016;34(suppl):e165750.