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Maria Rosaria Boni, Alessandra Polettini, Raffaella Pomi

Effect of ultrasonic post-treatment on

anaerobic digestion of lignocellulosic waste

and Andreina Rossi

Abstract

This paper evaluates the effects of ultrasonication (US) applied, individually or in combination with a mechanical treatment, to the effluent of anaerobic digestion (AD) of lignocellulosic waste, on methane (CH₄) production. US of the substrate downstream of AD is a relatively novel concept aimed at improving the degradation of recalcitrant components in order to enhance the overall energy efficiency of the process. US tests were carried out on real digestate samples at different energies (500–50,000 kJ/kg total solids (TS), corresponding to sonication densities of 0.08–0.45 W/ml). AD tests were performed on mixtures of sonicated (S_{us}) and untreated (S) substrate at two different S_{us}: S ratios (25:75 and 75:25 w/w), simulating post-sonicated material recycling to the biological process. The US effect was estimated through the solubilization degree of organic matter, as well as the CH₄ production yield and kinetics, which were all found to be enhanced by the treatment. At S_{us}: S = 75:25 and $E_s \ge 20,000$ kJ/kg TS (0.25 W/ml), CH₄ production improved by 20% and the values of the kinetic parameters increased by 64–82%.

Keywords

Sonication, lignocellulosic waste, anaerobic digestion, post-treatment, biomethane, hydrolysis

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Introduction

Anaerobic digestion (AD) of organic waste has been looked at with renewed interest over the last decade, taking into account the principles of the circular economy in waste management. These principles involve achieving biological stabilization of the residues and at the same time pursuing materials and energy recovery targets. Interest in AD of organic waste is further fostered by the potential to integrate the AD process into decentralized energy production systems as well as into the biorefinery approach.

Despite having been applied for more than a century, AD requires further research efforts to address unresolved issues including the poor efficiency of the conversion of lignocellulosic materials into methane (CH₄) (Sawatdeenarunat et al., 2014). These materials are typical components of municipal solid waste, including agro-industrial as well as forest residues, which represent interesting streams for bioenergy generation that do not compete with food production. The complex structure of lignocellulosic components, mainly formed by lignin, hemicellulose and cellulose, makes them highly resistant and biologically recalcitrant (Hendriks and Zeeman, 2009; Molinuevo-Salces et al., 2013; Zheng et al., 2014). Consequently, the hydrolysis of lignocellulosic materials often becomes the rate-limiting step during AD and complex organic molecules may be found undegraded in the final digestate (Mbaye et al., 2014; Wojnowska-Baryła et al., 2018). Efforts to tackle the recalcitrance of lignocellulosic materials to biological degradation have been oriented towards the application of pre/post-treatments to promote the hydrolysis of complex molecules. These rely on various mechanisms, including physico-mechanical (milling, grinding, and ultrasonication (US)), physico-chemical (steam explosion and wet oxidation), chemical (alkaline or acidic treatment, chemical oxidation, and organic solvents treatment) or biological processes (Carlsson et al., 2012).

Such processes have been widely investigated mainly as a pre-treatment stage of lignocellulosic substrates prior to AD. More recently, the application of the mentioned processes downstream of AD has been suggested as a novel concept to improve selectivity towards recalcitrant components and the overall energy efficiency of the process (Boni et al., 2016; Cesaro et al., 2014; Garoma and Pappaterra, 2018; Lindner et al., 2015; Menardo et al., 2011; Sambusiti et al., 2015; Somers et al., 2018).

Several studies (Braeutigam et al., 2014; Cesaro et al., 2012; Elbeshbishy and Nakhla, 2011; Elbeshbishy et al., 2011; Khanal et al., 2007; Pilli et al., 2011; Rasapoor et al., 2016, 2018; Somers et al., 2018; Zeynali et al., 2017; Zou et al., 2016) have suggested

Corresponding author:

Andreina Rossi, Department of Civil and Environmental Engineering, Sapienza University of Rome, Via Eudossiana 18-00184, Rome, Italy. Email: andreina.rossi@uniroma1.it

Department of Civil and Environmental Engineering, Sapienza University of Rome, Rome, Italy

that US can positively affect the anaerobic biodegradability of organic residues including wastewater sludge, manure, food waste, agricultural wastes, distillery residues, and by-products from bio-ethanol production. The cavitation effects caused by the application of ultrasound promote the disintegration of particulate matter, the disruption of complex molecules, and the liberation of more easily degradable monomers, thus enhancing biodegradability (Iskalieva et al., 2012).

The effect of US downstream of AD on the biodegradability/ bioavailability of AD digestate has received only limited investigation (Boni et al., 2016; Cesaro et al., 2014; Garoma and Pappaterra, 2018; Somers et al., 2018). In this downstream case, the sonicated digestate can either be directed to a second AD stage or be recycled back to the digester. Application of US as a post-treatment would allow improved selectivity towards the poorly biodegradable and recalcitrant components, thus optimizing energy efficiency and costs.

The performance of US depends on several factors, acting either synergistically or antagonistically. These factors include the operating conditions (treatment duration, temperature, US frequency, and power), the physico-chemical properties of the substrate (solids and lignin content and viscosity of the slurry) and the design parameters (reactor configuration and diameter and position of the transducer) (Gogate et al., 2011).

It must also be emphasized that the factors affecting US may be interdependent, which also complicates the prediction of their overall effect on anaerobic biodegradability. The different experimental conditions adopted (e.g., substrate properties, power input, and transducer design) also explain why a comparison of the US effects on CH_4 production reported in the literature often shows inconsistencies and is difficult to generalize.

Although some studies on US as a pre-treatment method for wastewater sludge (Grönroos et al., 2005; Show et al., 2007) have demonstrated that the US density (D_s) represents a key operating parameter for process optimization, many authors (Boni et al., 2016; Cesaro et al., 2014; Elbeshbishy et al., 2011; Gadhe et al., 2014; Zeynali et al., 2017; Zou et al., 2016) have mainly focused on irradiation time and specific energy (E_s).

A systematic investigation of the individual and joint influence of the relevant parameters of the US process, including E_s and D_s , on the anaerobic biodegradability of digestate is still missing in the literature. This study makes an attempt to fill in the gaps by means of the application of US, individually and in combination with a mechanical treatment, as a post-treatment downstream of AD. We expand the perspective of our previous research, focusing on a more complex residual matrix, investigating the combined influence of the relevant sonication parameters, and combining sonication with enhanced particle size reduction. The influence of the operating variables of the US treatment on the evolution of the AD process was assessed through detailed investigation of the solubilization degree of organic matter upon US, the CH₄ production yield and kinetics of the AD stage, as well as the related carbon mass balance.

Table	1.	Digestate	properties.

Parameter	Unit of measure	Value*
рН	-	7.7±0.20
Total solids, TS	g/l	83 <u>+</u> 0.9
Volatile solids, VS	g/l	59 <u>+</u> 1.1
Total organic carbon, TOC	g/l	29 <u>+</u> 0.9
Dissolved organic carbon, DOC	g/l	4.9 <u>+</u> 0.02
Chemical oxygen demand, COD	g/l	85 <u>+</u> 1.9
Soluble COD, sCOD	g/l	12.1 <u>+</u> 0.1
Carbohydrates	g/l	13.0 <u>+</u> 1.2
Soluble carbohydrates	mg/l	1095 <u>+</u> 112
Kjeldahl nitrogen, TKN	mg N-NH4 ⁺ /l	3404 <u>+</u> 16
Soluble Kjeldahl nitrogen, sTKN (tot)	mg N-NH4 ⁺ /l	1436 <u>+</u> 18
Soluble N-NH4 ⁺	mg/l N-NH4 ⁺ /l	1088 <u>+</u> 17
Acetate	mg/l	403 <u>+</u> 93
Propionate	mg/l	30.5 <u>+</u> 0.75
Iron	mg/l	213 <u>+</u> 1
Manganese	mg/l	15.3 <u>+</u> 0.6
Cobalt	mg/l	0.91 <u>+</u> 0.09
Nicckel	mg/l	2.54±0.90

Notes: * average values and related standard deviations of three replicates; $N\text{-}NH_4^+\text{:}$ ammoniacal nitrogen.

Materials and methods

Materials

The digestate of an Italian full-scale AD plant, fed with a mixture of organic residues from a food production plant, silage energy crops, olive husks and manure, was collected and stored at 4°C and characterized (see Table 1) according to the methods reported in Boni et al. (2016).

Fourier-transform infrared spectroscopy (FT-IR) was also adopted using an Agilent 640 IR spectrometer operated in the mid-IR range with a 4 cm⁻¹ resolution and 4000 scans/sample in the absorbance mode with attenuated total reflectance correction.

Experimental set-up

A number of US tests were performed by applying pre-set E_s values to the digestate. Each sonicated sample was then characterized to assess changes in composition of the substrate and to evaluate the solubilization degree of the relevant compositional parameters (see 'US and mechanical treatments' subsection). Mixtures of non-sonicated (S) and sonicated (S_{us}) digestate were then prepared (see 'AD tests' subsection) to evaluate the effect of US on the CH₄ production rate and yield in a dedicated AD stage. For control purposes, a 100% non-sonicated digestate (S) sample was also included in the AD test campaign.

The effect of digestate milling prior to US was also estimated by using the milled and sonicated material in AD experiments under the conditions mentioned above (see 'AD tests' subsection).

US and mechanical treatments. The US treatment was performed by means of a laboratory-scale ultrasound generator described elsewhere (Boni et al., 2016). The E_s , that is, the amount of energy supplied per unit of initial mass of total solids (TS), was varied in the range 500–50,000 kJ/kg TS, adopting D_s values of 0.08, 0.25 and 0.45 W/ml, which in turn corresponded to US intensities of 2.3, 7.7 and 13.4 W/cm².

In order to limit the treatment duration, at a D_s of 0.08 W/ml the US tests were performed at E_s values < 20,000 kJ/kg TS.

The mechanical treatment was performed by a knife mill operated for three minutes at 22,000 revolutions per minute; the sample was then sonicated at an E_s of 20,000 kJ/kg TS at a D_s of 0.25 W/ml.

The effects of US and the combined milling and US treatment were assessed using equation (1) (see Boni et al., 2016):

$$I_{\rm p}(\%) = \frac{sX_{us} - sX_0}{X_0 - sX_0} \times 100 \quad I_s(\%) = \frac{sX_{us} - sX_0}{sX_0} \times 100 \quad (1)$$

where X is the concentration of the parameter of concern (total organic carbon (TOC), chemical oxygen demand (COD), carbohydrates or proteins) before (X_0) and after (X_{us}) the US process and the prefix s indicates the corresponding soluble fraction.

The numerical values of I_p and I_s are affected by the initial composition of the material, more specifically by the initial partitioning of organic matter between the particulate $(X_0 - sX_0)$ and soluble (sX_0) forms. The relevant content of the particulate fraction of the initial material makes I_p more meaningful than I_s for assessing the effect of US on the disintegration of organic matter.

AD tests. The AD tests were arranged to investigate the effect of E_s applied during the US treatment on the yield and kinetics of CH₄ production at a fixed value of D_s (0.25 W/ml). A subset of the treated samples (see 'US and mechanical treatments' subsection) was selected for the AD tests. In particular, the digestate sonicated at E_s values of 500 and 30,000 kJ/kg TS were not considered for the AD tests, since the former yielded a very low solubilization degree of organic matter and the behavior of the latter was similar to the sample treated at $E_s = 20,000$ kJ/kg TS.

The AD experiments were carried out on mixtures of S_{us} and S at different ratios, namely S_{us} : S = 25:75 and 75:25 w/w in order to simulate two different recycling ratios of the post-sonicated material to the biological process. Moreover, on the basis of the results obtained in the AD tests, for the sample treated at E_s = 20,000 kJ/kg TS an additional AD test was performed on the milled and sonicated digestate at a 75% recycling ratio (S_{m+us}: S = 75:25).

A control mixture containing the untreated digestate only (100% S) was also tested.

A summary of the experiments is reported in Table 2. Each test was run in duplicate.

The experiments were carried out in stirred batch glass reactors (working volume = 0.5 l) at $T = 37 \pm 2^{\circ}C$. Before the onset of the experiments, the reactors were flushed with nitrogen gas for a few minutes to drive away air from the reactor headspace. The volumetric production and composition of the produced biogas were periodically analyzed. The AD tests were operated until no further appreciable increase in CH₄ production was observed,

Table 2. Summary of the anaerobic digestion tests performed.

Run	Mixture composition % by weight (w/w)	E _s (kJ/kgTS)
Control	100% S	-
3000_25S _{us}	$25\%S_{us} + 75\%~S_{us}$	3000
3000_75S _{us}	$75\%{ m S}_{ m us}+25\%~{ m S}$	3000
6000_75S _{us}	$75\%S_{us} + 25\%~S_{us}$	6000
10000_25S _{us}	$25\%S_{us} + 75\%~S_{us}$	10,000
10000_75S _{us}	$75\%S_{us} + 25\%~S_{us}$	10,000
20000_25S _{us}	$25\%S_{us} + 75\%~S$	20,000
20000_75S _{us}	$75\%S_{us} + 25\%~S_{us}$	20,000
50000_25S _{us}	$25\%S_{us} + 75\% \ S$	50,000
50000_75S _{us}	$75\%S_{us} + 25\%~S_{us}$	50,000
$20000_{75S_{m+us}}$	$75\%S_{m+us}$ + 25% S	20,000

which typically occurred within 56 days from the start of the experiment. In order to assess the metabolic pathways of the biochemical process, individual volatile fatty acids (VFAs), namely acetate, propionate, butyrate, valerate, hexanoate, and heptanoate, were determined at selected time intervals (see Boni et al. (2016) for details on the analytical procedure).

Kinetic model

The CH₄ production over time was modeled through the modified Gompertz equation or the Richards bacterial growth model (Zwietering et al., 1990). Interpolation was performed using TableCurve $2D^{\circledast}$ v. 5.01, which showed for all experiments that the best degree of fitting was obtained using equation (2):

$$M = P\left\{1 + \exp(1 + \nu) \exp\left[\frac{R_m}{P}(1 + \nu)^{\left(1 + \frac{1}{\nu}\right)} (\lambda - t)\right]\right\}^{\binom{-1}{\nu}}$$
(2)

where M (Nl CH₄/kg volatile solids (VS)) is the CH₄ production at time t (d), P (Nl CH₄/kg VS) is the maximum CH₄ production yield, R_m (Nl CH₄/kg VS·d) is the maximum CH₄ production rate, λ is the lag phase duration (d) and ν is a curve shape parameter. The time required to attain 95% of P (t_{95}) was also estimated.

Results and discussion

Effect of US energy on digestate properties

Within the range of the tested US conditions, no significant changes in both the TS and VS concentrations were observed. This implied that no relevant organic matter volatilization nor mineralization effects were produced by US, as also observed in previous studies (Boni et al., 2016; Cesaro et al., 2014). Other authors (Elbeshbishy et al., 2011, 2012) found that US produced a slight decrease in the solids content when applied to organic wastes of different origin before AD. In particular, in food waste samples sonicated at an E_s of 5000 kJ/kg TS, the total suspended solids and volatile suspended solids content was



Figure 1. Effect of specific energy and ultrasonication density on: (a-c) organic carbon; and (d-f) soluble carbohydrates solubilization.

found to decrease by 9% and 7%, respectively. A more pronounced reduction in VS concentration ($\sim 24\%$ or higher) was observed in agricultural waste sonicated at E_s values in excess of 2500 kJ/kg TS.

To assess the solubilization degree of the substrate upon US, Figure 1 reports the evolution of dissolved organic carbon and soluble carbohydrates as a function of E_s and D_s . At the highest values of D_s (0.25 and 0.45 W/ml), the I_p values varied in the ranges 3.3–50.0% for TOC and 1.7–27% for carbohydrates. The corresponding ranges for I_s were 16.3–250% and 18–290%.

At the lowest D_s value (0.08 W/ml), the solubilization effect observed at increasing E_s values was less pronounced, with I_p varying in the range 1.4–16.3% for TOC and 4.3–7.3% for carbohydrates. Accordingly, the maximum values attained by I_s (83% for TOC and 80% for carbohydrates) were lower than those observed at 0.25 W/ml and 0.45 W/ml. At all D_s values, the relationship between the solubilization degree and E_s was roughly linear up to a threshold beyond which a plateau was reached (see Figure 1). This suggests that further increasing E_s is ineffective at promoting the disintegration of complex organic molecules. The mentioned threshold value for E_s was found to be affected by D_s . In particular, at a D_s value of 0.08 W/ml, the E_s threshold was 6000 kJ/kg TS, which increased to 30,000 kJ/kg TS at $D_s = 0.25$ and 0.45 W/ml.

Moreover, the effect of D_s on the solubilization yield was less pronounced at lower E_s values (< 6000 kJ/kg TS), as depicted in Figure 1. This can be explained considering that, at the greatest specific energies, higher densities produce a larger number of transient bubbles, which could exert stronger cavitation forces, in turn fostering the breakdown of complex molecules. This has also been reported in previous studies on sonicated sewage sludge (Grönroos et al., 2005; Show et al., 2007).

At the highest D_s investigated (30,000 kJ/kg TS), an $I_{p(TOC)}$ of 38.6% was observed, corresponding to an $I_{p(COD)}$ of 37.0, which was comparable with that observed by Elbeshbishy et al. (2011), equal to 27%. Under such conditions, $E_s = 3192$ kJ/kg TS and D_s of 0.6 W/ml, the $I_{p(COD)}$ value reported by Gianico et al. (2013) (~ 2%) was significantly lower than the value (5.6%) obtained in our study at $E_s = 3000$ kJ/kg TS. This is related to the initial composition of the substrate, which plays a key role in dictating the degree of solubilization. Stemming from this, the comparison among I_p values obtained by different authors should be carried out carefully, as the results depend on the specific experimental conditions adopted as well as the substrate characteristics.

An increase in the dissolved organic nitrogen from approximately 350 mg ammonium (N-NH₄) ⁺/l (raw digestate) up to ~1200 mg N-NH₄⁺/l (samples sonicated at 0.25 W/ml and E_s in the range 500÷50000 kJ/kg TS) was observed. Since the ammonia content was not significantly modified by US, it can be argued that, under the tested operating conditions, US was able to produce only a partial disintegration of the complex



Figure 2. Spectra band heights in the second-derivative mode as a function of ultrasonication energies.

N-containing molecules (i.e., proteins), while the liberation of simple monomers was negligible.

Changes in fibers structure of US substrates

The FT-IR spectroscopy was used to identify the molecular changes that occurred in the digestate as a result of US. Digestate samples sonicated at 3000, 10,000, 20,000, and 50,000 kJ/kg TS at a D_s of 0.25 W/ml were analyzed and compared to the unsonicated digestate. The complex mixture of different chemical components in the digestate causes peak overlapping and suppression in the FT-IR spectra. Thus, a mathematical approach based on the calculation of the second derivative of transmittance was used to enhance the resolution of bands that were not clearly identifiable in the original spectra. The relationships between the band heights of the spectra in the second-derivative mode and the US energies are reported in Figure 2. The decrease in the band heights observed in the 1558 and 1569 cm⁻¹ peaks, which is due to the aromatic ring vibration, was related to the modification of the lignin aromatic structure induced by US (Corredor et al., 2009; Koutsianitis et al., 2015; Sun et al., 2005). In particular, for such wave numbers the band heights decreased as E_s increased, with the exception of the substrate sonicated at $E_s =$ 20,000 kJ/kg TS. These findings show that a partial delignification of the original substrate occurred as a result of US (see Figure 2(a) and Figure 2(b)).

The band at 1637 cm⁻¹ was associated with the H–O–H stretching vibration of adsorbed water in carbohydrates (Chen et al., 2011). The decrease in band heights (see Figure 2(c)) was particularly significant for E_s values between 3000 and

10,000 kJ/kg TS, suggesting that US produced a modification of the holocellulose structure.

The band at 2985 cm⁻¹ in the FT-IR spectra was associated with the C–H vibration of CH₂ and CH₃ groups of cellulose and lignin components (Gastaldi et al., 1998; Koutsianitis et al., 2015). The data reported in Figure 2(d) show that a significant reduction in the content of such groups occurred for the substrate sonicated at 50,000 kJ/kg TS.

Finally, the results reported in Figure 2(e) and Figure 2(f) related to the broad band in the 3600–3000 cm⁻¹ provide information about the existence of hydrogen bonds in O–H groups (Taherdanak and Zilouei, 2014), which are commonly associated to the crystalline structure and the degree of intermolecular regularity (or the crystallinity) of cellulose (Harmsen and Huijgen, 2010; Koutsianitis et al., 2015; Oh et al., 2005). For the analyzed samples, a remarkable decrease in the intensity of the peak associated to O–H groups was observed as E_s increased, suggesting that US affected the proportion between crystalline and amorphous forms of cellulosic materials.

Effect of US on CH₄ yields and kinetics

The results of the different AD experiments are summarized in terms of percentage variation from the control sample of the CH₄ production yield (ΔP) and kinetic parameters (ΔR_m , Δt_{95}) as well as carbohydrates solubilization yield ($I_{p(carb_mix)}$) calculated according to equation (3):

$$I_{p(carb_mix)} (\%) = \frac{sX_{mix} - sX_0}{X_0 - sX_0} \times 100$$
(3)



Figure 3. Percentage variation of methane production yield and kinetics and $I_{p(carb_mix)}$ between the tested mixtures and the control samples ($D_s = 0.25$ W/ml).

where X is the concentration of carbohydrates for the control (X_0) and the S_{us}+S mixture (X_{mix}) , and the prefix s indicates the corresponding soluble fraction.

The results in Figure 3(a) and Figure 3(b) emphasize the combined effect of E_s and S_{us} : S ratio on the degree of substrate solubilization as well as on the yield (P) and kinetics (R_m) of CH₄ production. The inspection of the parameters of the Richards' bacterial growth model (equation (2)) indicated, for S_{us}: S = 75:25 (see Figure 3(a)), an appreciable increase in CH₄ production compared to the control run (19.3% at E_s = 20,000 kJ/kg TS and 21.3% at E_s = 50,000 kJ/kg TS). For lower E_s values the observed change in CH₄ production was either less significant or not appreciable, with ΔP values of between -3.9 and 6.8%. Similar results were also obtained for the rate of CH₄ production (Figure 3(a)), with R_m increasing by 82.0 and 63.7% compared to the control at $E_s \ge 20,000$ kJ/kg TS.

It is also interesting to note that the overall duration of CH_4 production (as conventionally represented by t_{95}) was not significantly affected by US. In fact, at higher US energies, despite R_m being larger than for the control run, t_{95} remained virtually unchanged. The results obtained for the kinetic parameters may be interpreted in the sense that US produced to some extent simpler structures from the original substrate, allowing a faster initial hydrolysis phase as mirrored by the increased R_m values. However, the final stages of the digestion process were likely to be dominated by the degradation of more complex components that had evidently been poorly affected by the US pre-treatment. This conclusion may also be supported by the evidence gained from the FT-IR analyses, which indicated that cellulose and lignin structures, although reduced as a consequence of US, were still present in the sonicated samples.

The results reported in Figure 3(b) show that at a S_{us} : S ratio of 25:75, in agreement with previous results (Boni et al., 2016), no gain in the maximum CH₄ production compared to the control test

was evident. Conversely, a slight reduction was observed for the mixtures containing the substrate sonicated at E_s of 3000–20,000 kJ/kg TS. Such a reduction may tentatively be ascribed to the abovementioned effect of biomass inactivation upon US coupled with an only moderate degree of carbohydrates solubilization (2.7-5.2%) produced by the treatment. At $E_s = 50,000$ kJ/kg TS, the CH₄ production yield was slightly increased ($\sim 3.7\%$) from that of the control run, possibly as a result of the fact that the higher degree of substrate solubilization obtained at high US energies was capable of counterbalancing the biomass inactivation effect caused by US. Notwithstanding this, for Sus: S =25:75 no appreciable effect of US on either the CH₄ production yield or the degradation kinetics could be identified (Figure 3(b)). On the basis of the results obtained, the hydrolysis stage of the AD process appeared to be positively affected by the US treatment applied when the content of soluble carbohydrates was at least 12% higher than that in the original substrate (see Figure 3(a)).

Table 3 provides a summary of the performance reported in previous studies on anaerobic degradability of solid waste as a result of US operated at low frequencies (< 50 kHz). However, the comparison of the results from different studies is limited by the different substrate characteristics, the experimental methods adopted (in some cases not fully detailed), as well as the way the results are reported. Often, the operating parameters of US are used with no normalization, but their influence on the process performance may be lower than that exerted by other factors. In the studies focusing on the role played by the substrate properties on the effect of US on the AD process, only TS concentrations > 50 g/l were considered.

It is evident from Table 3 that a limited number of studies has been carried out to evaluate the ability of US to increase the biodegradability/bioavailability of recalcitrant organic compounds remaining in the digestate downstream of AD, as it has been more widely regarded as a method for substrate pre-treatment.

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$ \label{eq:constraints} Nerses of a four function of the four fu$				GZ.U	000'01	Sus: 5 = 25:75 Sus: 5 - 75.25	1.4.1	24.1	1.7.1	-11.6 (CH4) 1 0 (CH.)	-33.1	
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Openity wates of a foot industry 230 20000 Sus; S = 75:35 232 243 133 -111 (bH) 2.8 0.45 20000 Sus; S = 75:35 22 2000 Sus; S = 75:35 22 22 20 20 0.45 20000 Sus; S = 75:35 21 73 2 23 2000 20 20 2 <td></td> <td></td> <td></td> <td>0.08</td> <td>20,000</td> <td></td> <td>I</td> <td>Ι</td> <td>6.4</td> <td>Ι</td> <td>Ι</td> <td></td>				0.08	20,000		I	Ι	6.4	Ι	Ι	
$ \label{eq:relations} \mbox{Networks} $				0.25	20,000	Sus: $S = 25:75$	32.8	26.3	18.3	-12.1 (CH4)	2.8	
						Sus: $S = 75.25$				19.3 (CH₄)	82.0	
$ \label{eq:relations} \begin{tabular}{l lllllllllllllllllllllllllllllllllll$				0.45	20,000		27.5	I	18.7	I	I	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$				0.08	30,000			I	.	I	I	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$				67.U 7.7	30,000		0.45 0.75		26.9 26.0			
				0.08	50,000		p:		0.07			
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$				0.25	50,000	Sus: $S = 25:75$	43.1	47.9	26.9	3.7 (CH₄)	10.2	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$						Sus: $S = 75.25$				21.3 (CH₄)	63.7	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$				0.45	50,000		33.6	I	20.8	I	I	(/100 - t- :)
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Urganic wastes of a food industry					$T = 37 + 2^{\circ}C$						(Boni et al., 2016)
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	×	$< 30^{\circ}$ C	23.7	0.05	500	I	2.8	Ι	3.0	Ι	Ι	
$ \begin{array}{llllllllllllllllllllllllllllllllllll$					1000		3.7	I	4.0	1.	I	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$					0009	*Sus: S = 25:75 Suc: S = 75.25	9.1	I	8.9	8.6 [CH4]	23.0 2 E	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$					10,000	Sus: 5 = 75:25	12.5	Ι	10.8	18.5 (CH ₄)	56.0	
$ \begin{array}{llllllllllllllllllllllllllllllllllll$					15,000		12.5	Ι	12.0	. 1	I	
Organic Fraction of MunicipalNone50-100 50.372 20.372 $20.5.75$ 50.000 $5us: S = 75:75$ 22.1 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 14.7 -1.7 25.4 (CH_a) 42.1					20,000	Sus: S = 25:75	16.3	I	13.7	-1.4 (CH ₄)	10.2	
Organic Fraction of MunicipalNone $50-100$ 0.372 $Su: S = 75:75$ 22.1 $ 21.3$ 6.5 (CH_{4}) 4.4 Organic Fraction of MunicipalNone $50-100$ 0.372 2102 Unspecified $ 4.6$ (biogas) $ -$ Solid Waste digestate 4219 Unspecified $ 4.6$ (biogas) $ 6.6$ (biogas) $ -$ If S=25 g/l) $50-100$ 0.372 2102 Unspecified $ 4.6$ (biogas) $ 4.6$ (biogas) $ -$ <					30.000	c7:c/ = c :snc	14 9	I	14.7	19.1 (CH₄) 	/ .7c	
Organic Fraction of MunicipalNone $50-100$ 0.372 2102 Unspecified 25.4 (DHz) 48.1 Solid Waste digestate $50-100$ 0.372 2102 Unspecified $ 46$ (biogas) $ -$ Solid Waste digestate 6291 $ 46$ (biogas) $ -$ Lignocellulosic material and $50-100$ 2264 Unspecified $ -$ Lignocellulosic material and $50-100$ 2264 Unspecified $ -$ Lignocellulosic material and $50-100$ 2264 Unspecified $ -$ Lignocellulosic material and $50-100$ 2264 Unspecified $ -$ Lignocellulosic material and $50-100$ 2264 Unspecified $ -$					50,000	Sils: $S = 25.75$	22.1		21.3	-6.5 (CH/)	77	
Organic Fraction of Municipal None $50-100$ 0.372 2102 Unspecified - - 46 (biogas) - Cesaro et al., 2014) Solid Waste digestate $50-100$ 0.372 2102 Unspecified - - 46 (biogas) - - (Cesaro et al., 2014) Solid Waste digestate 6291 4219 6291 71 (biogas) - - 60 (biogas) - - - 60 (biogas) - - - 60 (biogas) -						Sus: S = 75:25			2	25.4 (CH4)	48.1	
Solid Waste digestate 4219 60 (biogas) – (TS=25 g/l) 60 (biogas) – 71 (biogas) – Lignocellulosic material and 50–100 2264 Unspecified – None – dried distilled grain with 6858 – None – – None – solubles digestate 6858 10.0 2264 Unspecified – None – – TS=16.9 g/l TS=16.9 g/l None – – – – –	Organic Fraction of Municipal	None	50-100	0.372	2102	Unspecified	Ι	I	Ι	46 (biogas)	I	(Cesaro et al., 2014)
10=20 g/u 0.271 0.2264 Unspecified $ -$	Solid Waste digestate				4219					60 (biogas)	I	
TS=16.9 g/l	l innocellulocic material and		50_100		1 6 2 9 1	l henerified	I	I	I	/ I (blogas) None		
solubles digestate 6858 – None – TS=16.9 g/l	dried distilled arain with				4581					None		
l5=16.9 g/l	solubles digestate				6858					None	Ι	
	15=16.9 g/l											

Table 3. Summary of the performance of anaerobic digestion (AD) of solid waste as result of ultrasonication (US), as reported in previous studies.

			0	herating conditions			-					
		ns	treatment		AD		for subs	trate solubil.	ncement ization)	AD yield	AD kinetics	
Substrate	(°C)	TS (g/l)	D _s [W/ml]	E _s (kJ/kgTS)			l _{pCOD} (%)	І _{рТКN} [%]	_{pcarb.} [%]	Variation from control [%]	Variation from control (%)	Reference
Digestate from a domestic wastewater treatment plant TS=12.6 g/l	Ambient	12.6	0.60 0.60 0.64 0.63	14,286 28,571 45,714 60,000	Batch reactor $T = 37^{\circ}C$		1 1 1 1	1 1 1 1	1 1 1 1	60 (CH₄) 74 (CH₄) 78 (CH₄) 133	1 1 1 1	(Garoma and Pappaterra, 2018)
Manure digestate [MAN] T5=98.5 g/l	< 20°C	64.3	0.13	3000 9000 15,000	Batch reactor $T = 37 \pm 1^{\circ}C$ F: M [on VS-basis] = 0.8	Kecycle ratio 0.2 0.5 0.5 0.2 0.2	3.9 10.8 16.2			-0.7 (biogas) 1.6 (biogas) 3.0 (biogas) -10.3 (biogas) 4.4 (biogas)	-10.7 -35.2 -10.2 -12.8	(Somers et al., 2018)
Potato waste digestate (POT) TS=140 g/l Mixed organic waste digestate (000)		33.5 83.9		3000 9000 15,000 3000	Unspecified Unspecified	2	13.6 19.5 12.9 25.2 25.2		1111		2 2 2 1	
15=1/3.8 g/t Pre-treatment Organic Fraction of Municipal Solid Waste and sewage sludge	None	90 ±30	7.0	90,692±4.534	Semi-continuous reactor T = 35 ÷ 40°C					24 [biogas]		(Cesaro et al., 2012)
Lignocellulosic material and dried distilled grain with solubles	None	50	0.372	2835 5630 8493	Unspecified				111	3 (biogas) 11 (biogas) 23 (biogas)		(Cesaro et al., 2014)
15=930 g/t Hog maure TS: 93 g/t sCOD/TCOD=0,38	 < 30° C < < 	93	Unspecified	250 500 2500 10,000 21,000	Batch reactor T = 37°C F/M= 4 gCOD/gVSS		7.0 11.0 16.0 19.0 24.0 26.0	0.7 16.7 19.5 33.5 30.6		11.7 (CH ₄) 28.0 (CH ₄) 10.9 (CH ₄) 16.3 (CH ₄) 19.9 (CH ₄) 18.7 (CH ₄)	33.7 61.3 35.5 46.6 75.4	(Etbeshbishy et al., 2011)
Chicken manure TS: 50 g/l	Room temperature	50	0.22 0.26 0.23 0.22 0.22	30,000 288 360 468 612 720	Batch reactor T = 38°C F/M = 0.1 [w/w]		27.0 	33.3 	1111	20.7 (CH4) 26.9 (CH4) 30.6 (CH4) 35.5 (CH4) 45.7 (CH4) 45.7 (CH4)	9. 008	(Braeutigam et al., 2014)
Food wastes TS: 60 g/l	None	60	0.2	0009	Hybrid plug flow digester T=27÷30°C	0LR = 500 gVS/m3d 0LR = 1500			11 1	36.U (CH4J 59 (biogas) 80 (biogas)	11 1	(Rasapoor et al., 2018)
Organic fraction of municipal solid waste	None	60 80	0.2 0.6 0.4	6000 18,000 3000 6000	Batch reactor $T = 37 \pm 1^{\circ}C$ F/M = 0.78 (VS basis)	gv»/cyg	1 1 1 1			24 (biogas) 24 (biogas) 20 (biogas) 9 (biogas)	1111	(Rasapoor et al., 2016)
												(continued)

(continued)
з.
Table

			Ō	erating conditions		i					
		US t	reatment		AD	Blodegra (or sub;	dability enha strate solubil	incement .ization)	AD yield	AD kinetics	
Substrate	T [°C]	TS (g/l)	D _s (W/ml)	E _s (kJ/kgTS)		l _{pcoD} (%)	І _{рТКN} [%]	l _{pcarb} . [%]	Variation from control (%)	Variation from control (%)	Reference
			9.0	13,500		I	Ι	I	13 (biogas)	I	
Fruit and vegetable wastes	None	50	0.110	1175	Batch reactor	I	I	I	29 (biogas)	I	(Zeynali et al., 2017)
				2380	$T = 35^{\circ}C$	Ι	Ι	Ι	80 (biogas)	I	
				3560		Ι	Ι	Ι	63 (biogas)	Ι	
Food waste	< 30°C	50	1.5	9000	Unspecified	5.05	5.12	Ι	I	I	(Gadhe et al., 2014)
TS: 72.5 g/l		50		18,000		10.76	11.57	Ι	I	Ι	
		50		27,000		15.02	17.88	I	I	Ι	
		80		5625		13.75	21.95	Ι	I	Ι	
		80		11,250		28.00	20.79	Ι	Ι	Ι	
		80		16,875		42.37	46.86	Ι	I	Ι	
Food waste	< 30°C	65.5	0.8	350	Unspecified	0.2	I	5.7	I	I	(Elbeshbishy
TS: 65.5 g/l			0.5	1200		5.1	I	9.1	I	I	et al., 2012)
			0.7	3000		8.2	Ι	18.0	I	I	
			0.6	5500		22.1	Ι	22.0	Ι	Ι	
			0.8	15,000		24.2	Ι	22.4	Ι	I	
			0.8	23,000		26.3	I	22.9	I	I	
					Batch reactor						
					$T = 37 \pm 2^{\circ}C$						
Maize straw (MS) and dairy	None	220	0.25	189.39 (DM)	DMus: $MS = 1:1 \ [w/w]$	I	I	I	43.9 (biogas)	I	(Zou et al., 2016)
manure (DM)				284.09 (DM)	DMus: $MS = 1:1 [w/w]$	I	Ι	I	15.1 (biogas)	I	
TS _{MS} : 893 g/l				378.79 (DM)	DMus: $MS = 1:1 [w/w]$	I	I	Ι	-38.7 (biogas)	I	
TS _{MS} : 222 g/l				189.39 (MS)	MSus: DM =1:1 [w/w]	I	I	I	51.5 (biogas)	I	
				284.09 (MS)	MSus: $DM = 1:1 [w/w]$	I	Ι	Ι	69.7 (biogas)	Ι	
				378.79 (MS)	MSus: $DM = 1:1 \ [w/w]$	Ι	Ι	I	27.8 (biogas)	I	

The observed gain in CH₄ production is in agreement with the results of our previous study on a different substrate (Boni et al., 2016), while a larger increase in biogas production was attained by Cesaro et al. (2014) and Garoma and Pappaterra (2018), who used digestate as the substrate. The results here are comparable with those observed by other authors (Elbeshbishy et al., 2011; Rasapoor et al., 2016) when applying US as a pre-treatment, at approximately the same E_s .

In order to evaluate the fate of the original substrate during the process, the carbon mass balance at the end of the experiments was calculated. To this aim, the contributions of the following forms of carbon were accounted for (see Figure 4): (a) carbon (C) in the form of the analyzed metabolic products (C2–C7 VFAs); (b) residual C in both the soluble and the particulate forms (including C present as non-degraded organic compounds and/or additional metabolic products, as well as microbial cells); and (c) gasified C, in the form of CH₄ and carbon dioxide (CO₂) (for CO₂, the



Figure 4. Carbon mass balance for the experimental runs.

amount dissolved in the liquid was also included). The term "balance" in Figure 4 represents the C mass that was apparently lost due either to inaccuracies in the analytical measurements or sample inhomogeneity and was thus required to close the materials balance. All contributions to the mass balance were calculated from direct measurements in the liquid and gaseous phases, with the exception of dissolved inorganic C. This was indirectly estimated using the chemical equilibrium model Visual Minteq based on CO_2 solubility as a function of pH and temperature.

It was evident that most of the initial C mass was retained in the digestate as residual C (44–60% in particulate forms and 17– 33% as soluble species). The amount of C gasified as CH₄ was always found to account for a low fraction of the initial TOC (7– 9%), corresponding to 138–174 Nl CH₄/kg TOC. It is worth mentioning that 3–4% of initial TOC was associated with the measured metabolic products, which clearly suggests that the VFAs in the system were completely degraded at the end of the process; in other words, although a high organic load was adopted, no inhibitory phenomena related to the potential accumulation of VFAs were found to occur during the experiments.

Effect of milling and US on anaerobic degradability of digestate

In order to further exploit US as a feasible treatment option for complex substrates to enhance their anaerobic degradability, a combined treatment based on milling and US was investigated. This stemmed from the consideration that the size of complex organic materials is expected to influence US efficiency. On the basis of the results discussed in the previous subsection that indicated that only E_s values in excess of 10,000 kJ/kg TS positively affected the AD process, the combined treatment was performed at an E_s of 20,000 kJ/kg TS.

The milled and subsequently sonicated substrate displayed a higher degree of TOC dissolution compared with the sonicatedonly substrate under the same US conditions. Conversely, the proteins and carbohydrates contents were not found to be affected by the treatment to any statistically significant degree (Figure 5(a)).



Figure 5. Influence of milling and ultrasonication combination on: (a) $I_{p(TOC)}$, $I_{p(Carbohydrates)}$ and $I_{p(proteins)}$ (a) and $I_{p(carb_mix)}$ as well as percentage variation of methane CH₄ production yield; and (b) kinetics between the tested mixtures and the control samples at E_s = 20,000 kJ/kgTS and D_s = 0.25 W/ml.

Starting from the results shown so far, the combined treatment appeared unable to produce the full disintegration of the lignocellulosic structure, since the solubilization yield of the mixture $20000_{75S_{(m+us)}}$ (expressed as $I_{p(carbohydrates)}$) was similar to that obtained for the corresponding mixture $20000_{75S_{us}}$. At the same time (results not reported graphically), the change in the total process duration (t_{95}) upon milling and US was not found to be statistically significant. This finding may explain why no significant gain in CH₄ yields is produced when US is applied on particles of smaller size. However, from Figure 5(b), we note that the application of milling prior to US positively affected the process kinetics. This result suggests that additional efforts should be devoted to improve the mechanical processing method prior to US, with a view to improving the overall degree of substrate solubilization.

Conclusions

The application of post-US of AD digestate produced a significant increase in CH₄ yield (~20%) and maximum production rate (64–82%) at $E_s \ge 20,000$ kJ/kgTS and $D_s = 0.25$ W/ml. Although the outcomes of the study indicated an improvement in the digestibility of the substrate, the processing conditions adopted suggested significant energy requirements that would in fact impair the full-scale applicability of the process. The results appear to indicate that, in order to improve the energetic profile of the combined process and therefore the associated overall environmental sustainability, efforts should be directed to reducing the US energy applied. Thus, further insights into the effects of different combinations of D_s and E_s should be obtained, more specifically by evaluating the effect of increasing D_s as a strategy for reducing E_s .

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ORCID iD

Andreina Rossi D https://orcid.org/0000-0001-7448-611X

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