

1 **Ultra-high performance separation of basic compounds on**
2 **reversed phase columns packed with fully/superficially porous**
3 **silica and hybrid particles by using UV transparent**
4 **hydrophobic cationic additives.**

5

6

7 Simone Manetto, Giulia Mazzocanti, Alessia Ciogli*, Claudio Villani, Francesco Gasparri

8 *Dipartimento di Chimica e Tecnologie del Farmaco, "Sapienza" Università di Roma, P. le Aldo*

9 *Moro 5, 00185 Roma (Italy)*

10

11 **Non-standard abbreviations:** UHPLC, ultra-high performance liquid chromatography; **ILs**, ionic
12 liquids; **ER-RPLC**, electrostatic repulsion reversed phase liquid chromatography; **CSH**, charged
13 surface hybrid, **TBAOH**, tetrabutylammonium hydroxide; **TBAHSO₄**, tetrabutylammonium
14 hydrogen sulfate; **TBA**, tetrabutylammonium; **SPP**, superficially porous particles; **FPP**, fully porous
15 particles; **RP^{plusTBA}**, reversed phase added of TBA.

16

17 **Keywords:** basic analytes, hydrophobic cationic additives, peak tailing, reversed phase columns.

18

19 *Corresponding author: Alessia Ciogli

20 E-mail address: alessia.ciogli@uniroma1.it

21 **Abstract**

22 The use of the tetrabutylammonium additive was investigated in the ultra-high performance reversed
23 phase liquid chromatographic elution of basic molecules of pharmaceutical interest. When added to
24 the mobile phase at low pH, the hydrophobic tetrabutylammonium cation interacts with the octadecyl
25 chains and with the residual silanols, thus imparting a positive charge to the stationary phase,
26 modulating retention and improving peak shape of protonated basic solutes. Two sources of additive
27 were tested: a mixture of tetrabutylammonium hydroxide/trifluoroacetic acid and
28 tetrabutylammonium hydrogen sulfate. Retention and peak shape of eleven basic pharmaceutical
29 compounds were evaluated on commercially available ultra-fast columns packed with octadecyl
30 stationary phases (Ascentis Express C18 2.0 μm , Acquity BEH C18 1.7 μm , Titan C18 1.9 μm). All
31 columns benefit from the use of additive, especially tetrabutylammonium hydrogen sulfate, providing
32 very symmetric peaks with reasonable retention times. Focusing on the probe compounds
33 amitriptyline and sertraline, efficiency and asymmetry values were investigated at increasing
34 retention factor. The trend is very different to that obtained in reversed phase conditions and the effect
35 lies in the complex molecular interaction mechanisms based on hydrophobic and ion exchange
36 interactions as well as electrostatic repulsion.

37 **Introduction**

38 Today, more than 70% of small molecules of pharmaceutical interest are bases, about 20% are acids,
39 and only 10% are neutral compounds [1]. However, despite the high number of basic compounds,
40 their analytical control by reversed phase liquid chromatography (RP-LC) poses still particular
41 challenges. Two critical factors should be taken into account in the elution of bases by RP-LC based
42 on silica-based stationary phases: control of retention and minimization of peak broadening and
43 tailing [1-3]. Basic compounds are ionizable and their retention is mainly affected by the ionic
44 behaviors of residual silanols present on the surface of packing materials [4]. Non-negligible amount
45 of underivatized, deprotonated silanols act as cation exchange sites and provide an interaction

46 mechanism that add to the hydrophobic effect typical of reversed phase stationary phase. In this case,
47 the stationary phase works as a mixed-mode support, the positively charged analytes at the pH
48 condition of the mobile phase are retained longer, and their peak shape is negatively affected (e.g.,
49 tailed and broad peaks) [5-7]. Different approaches have been developed to suppress the ion exchange
50 mechanism of silanols. They are gathered in two main groups: i) modification of the mobile phase
51 and ii) use of alternative solid supports. Lowering the pH of mobile phase, addition of tertiary amines
52 at acidic pH, and more recently, the introduction of room temperature ionic liquids (ILs) in mobile
53 phase represent some methods belonging to the first group [8-16]. At pH values between 2.5 and 3.5
54 silanols are mainly undissociated, the interactions with the ionized basic samples are reduced and
55 peak shape improves [17, 18]. However, pH reduction alone does not seem to be a wide-applicability
56 solution. Tertiary amines are the first choice and the most used mobile phase additives employed to
57 shield silanols [10]. In fact, positively charged amines interact electrostatically with the residual
58 anionic silanols making them no longer available for the interaction with solutes. Moreover,
59 hydrophobic protonated amines can also be applied in combination with chiral mobile phase additives
60 to obtain enantiomeric purity profile of chiral bases [19]. Concerning the ILs, they represents a valid
61 alternative as additives to shield free silanols but the mechanism of action seems to be concentration
62 dependent [20] and not easy to preview. In addition, the main drawback of using ILs as additives in
63 mobile phase lies in their absorption properties in UV region [21].

64 In the second group, innovative solid supports have been primarily focused on ultra-high performance
65 LC (UHPLC) applications. Among them, some companies produced silica bulk, named high purity
66 silica, with a reduced amount of metals and free silanols; others have commercialized hybrid silica
67 particles, where siliceous skeleton contains organic moieties, typically ethyl groups that provide
68 mechanical resistance and reduction of silanols [22-24]. From the silica derivatization point of view,
69 embedded polar or ionic functional groups were introduced in the long alkyl chain of hydrophobic
70 stationary phase in addition to the standard end-capping procedure [25-27]. Ihara et al. presented an
71 unconventional embedded RP stationary phase [28]. They prepared a long chain

72 octadecylimidazolium ILs-modified silica stationary where the cationic moiety is located close to the
73 silica network. Compared to the standard C-18 stationary phase, this support can interact with solutes
74 according to a mixed mechanism comprising hydrophobic, anion-exchange, electrostatic, and π - π
75 interactions. In 2010, a new concept of mixed mode stationary phase based on hybrid support was
76 introduced in the market as charged surface hybrid (CSH) materials, which involves positively
77 charged groups chemically bonded to hybrid silica surface [29, 30]. Columns packed with CSH silica
78 provide a wide range of selectivity for ionizable compounds and allows for their rapid equilibration
79 in pH switching protocols. This makes them ideal to be included in the routine method development
80 and to be used in analytical platforms where the pH of mobile phase is frequently changed [28]. Gritti
81 and Guichon have published many papers concerning the use of CSH supports and the theoretical
82 investigation of their interaction mechanism [31-33]. In particular, in a paper dated 2014, the authors
83 introduced the term electrostatic repulsion interactions RP-LC (ER-RPLC) to describe this new
84 version of mixed-mode mechanism [34-38]. The advantages of ER-RPLC lie in the shorter retention
85 times and the reduction of peak tailing when employed at acidic elution conditions.

86 However, despite all progress in particle technology, peak deformation of basic compounds can be
87 recorded under particular experimental conditions. Taking into account the routine operations in
88 analytical laboratories, low pH in combination with the positively charged additives is often a first
89 favored choice for the RP-LC analysis of basic molecules, like small molecules, peptides, and small
90 proteins, in order to suppress silanol activity effectively. Herein we report on the use of positively
91 charged hydrophobic additives for efficient UHPLC elution of basic molecules of pharmaceutical
92 interest. Commercially available UHPLC columns packed with octadecyl stationary phases, namely
93 Ascentis Express C18 2.0 μ m, Acquity BEH C18 1.7 μ m and Titan C18 1.9 μ m, were evaluated and
94 results compared with the Acquity CSH C18 1.7 μ m column.

95 **2. Materials and methods**

96 **2.1 Materials**

97 All solvents were LC grade and were purchased from Sigma-Aldrich (St. Louis, MO, USA) as well
98 as trifluoroacetic acid (TFA), tetrabutylammonium hydroxide 30-hydrate (TBAOH) ($\geq 99.0\%$ w/w),
99 and tetrabutylammonium hydrogen sulfate (TBAHSO₄) ($\geq 99.0\%$ w/w). All basic compounds, in
100 active pharmaceutical ingredient form, were available in house from previous studies. Solutions of
101 samples were prepared in mobile phases at a concentration of 1.0 mg/ml.

102 Four UHPLC columns, of 100 mm L x 3.0 mm I.D., were employed: Acquity BEH C18 1.7 μm and
103 Acquity CSH C18 1.7 μm from Waters Corporation (Milford, MA, USA), Ascentis Express C18 2.0
104 μm and Titan C18 1.9 μm from Supelco Sigma-Aldrich (Merck KGaA, Darmstadt, Germany).

105 **2.2 Instrumentation**

106 An UltiMate 3000 RSLC LC (Thermo Fisher Dionex, Sunnyvale, California) equipped with dual
107 gradient pumps was employed. The complete configuration of the system includes an in-line split-
108 loop well plate sampler, a thermostated column ventilated compartment (temperature range: 5–
109 110°C), and a diode array detector (UltiMate 3000 RSLC DAD) with a low dispersion 2.5 μl flow
110 cell. The UV detector was set at a time constant of 0.10 s and data collection rate of 100 Hz. UV
111 detection was performed at 220 nm. Data acquisition and processing were performed with
112 Chromeleon 7.2 (ThermoFisher).

113 **2.3 Chromatographic conditions**

114 The mobile phases contained different portions of water (eluent A) and acetonitrile (eluent B), both
115 with added TBAOH ion pair reagent (5 mM) and TFA (0.1% v/v). In addition, the eluent A was
116 buffered at pH 2.5 with TFA. Conversely, when TBAHSO₄ (5 mM) was added to water, the pH was
117 at once 2.5. For comparative purpose, the elution conditions of water/acetonitrile 90/10 and 0.1%
118 TFA v/v were used. Mobile phases were filtered through 0.2 μm Omnipore filters (Merck Millipore,
119 Darmstadt, Germany). Injected volumes were 0.5 μL . Hold up volume was estimated by injection of
120 uracil.

121 For data evaluation, the values of peak asymmetry at 5% of peak height were calculated according to
122 the European Pharmacopeia. The efficiency (N/m) was calculated by the second moment, as reported
123 in Eq. 1 [39].

$$124 \quad TP = \frac{t_r^2}{\mu_2} \quad \text{Eq.1}$$

125

126 Where TP is the number of theoretical plates, t_r the retention time of analyte, and μ_2 the second
127 statistical moment. The μ_2 is defined as reported in Eq. 2:

$$128 \quad \mu_2 = \frac{\int (t - \mu_1)^2 * f(t) dt}{\mu_0} \quad \text{Eq.2}$$

129 where t corresponds to the peak centroid, the μ_1 is the first statistical moment ($\mu_1 = \frac{\int t * f(t) dt}{\mu_0}$) and
130 the μ_0 is the zeroth statistical moment ($\mu_0 = \int f(t) dt$).

131

132 3. Results and Discussion

133 In the first part of the study, a collection of basic compounds of pharmaceutical interest has been
134 eluted on widely used C18 UHPLC columns in reversed phase mode by using only TFA as additive.

135 **Table 1** reports the characteristics of tested columns. Briefly, Ascentis Express and Titan C18 columns
136 are based on superficially and totally porous silica particles, respectively (SPP and FPP). They have
137 very comparable pore (90 Å vs. 80 Å) and particles sizes (2.0 µm vs. 1.9 µm) with different loading
138 of organic portion (8% vs. 13%). Acquity BEH C18 column is packed with hybrid carbon/siliceous
139 1.7 µm particles (130 Å pore size). The total carbon content is 18% but the portion of octadecyl
140 moiety is not specified. Acquity CSH C18 column has been used as comparative support that was
141 developed to improve both stability at low pH and peak symmetry of basic analytes.

142 **Figure 1** reports structures of investigated compounds grouped for their pharmaceutical activity:
143 samples **1-5** are antidepressant drugs, lidocaine (**6**), and ketamine (**7**) are employed as anaesthetics
144 while compounds **8-11** belong to the beta-blocker family. In this sample selection, amitriptyline was
145 chosen as a model compound having only an amino group far from the partially aromatic polycyclic
146 portion, so in principle, the effect of cationic additives should be easily evaluated (see later). An

147 organic/aqueous mobile phase, optimized for each compound, with a constant 0.1% TFA afforded
148 long retention times and broad peaks (**Table S1**). Keeping constant the organic portion in the mobile
149 phase for each compound, the Acquity BEH C18 is the most retentive stationary phase in RP
150 conditions. The Ascentis Express C18 shows capacity factors comparable to those of the Titan C18
151 although the stationary phase is based on superficially porous particles and has a low carbon content.
152 Besides, focusing on peak distortion and using $As_{5\%}$ as a measure of matrix activity (mainly correlated
153 to silanol activity), significant peak asymmetries were recorded on Ascentis Express C18 column
154 where $As_{5\%}$ values were often > 3.0 . On the other hand, lower $As_{5\%}$ values were found on the Acquity
155 BEH C18, while the Titan C18 showed intermediate results. Furthermore, for compounds **1, 6, 7, 9-**
156 **11**, peak symmetries on the Acquity BEH C18 are well comparable or superior to those recorded on
157 Acquity CSH C18 in the same analytical conditions, attesting the better surface inertness of the hybrid
158 organic/siliceous support.

159 As expected, the last two columns gave higher efficiencies as they are packed with smaller particles.
160 In particular, the plate number of Acquity CSH C18 column, 40-50% higher than those of the Acquity
161 BEH C18, reflects the mixed mode (hydrophobic and charge repulsion) retention mechanism
162 specifically developed for the elution of basic compounds. The fixed positive charge on CSH
163 stationary phase allows electrostatic repulsion with the positively charged analytes reducing capacity
164 factor and increasing the number of plates.

165 In this contest, taking into account the Acquity CSH C18 column embodies a solution for improving
166 peak shape of basic compounds, we demonstrate how the addition of cationic and hydrophobic
167 additive at low pH, in the mobile phase, represents a valid and flexible additional approach in the
168 elution of the same compounds on conventional UHPLC C18 supports, providing excellent peak
169 shape with asymmetry values close to 1.0. The widely known, not expensive, and UV transparent,
170 tetrabutylammonium (TBA) was chosen as a hydrophobic cationic additive. At acidic pH of the
171 mobile phase, two sources of TBA were evaluated: TBAOH and TBAHSO₄. To work at low pH,

172 TBAOH, as opposed to TBAHSO₄, must be added with a TFA. The screening has been done on six
173 samples randomly chosen and data are reported in **Table S2**.

174 As expected, these additives prevent unwanted interactions with the stationary phase ensuring
175 reasonable elution times and symmetries. In detail, asymmetry values are well comparable in both
176 cases, some slight differences in retention could be ascribed to the nature of counter ion as a
177 kosmotropic effect [40] but this aspect is actually under investigation in our laboratory. When
178 hydrogen sulfate anion is employed, the eluent A reaches a pH of 2.5 directly without any additional
179 pH correction. Therefore, for this facilitating aspect we decided to use only TBAHSO₄, namely from
180 now RP^{plusTBA} approach. The elution profiles of amitriptyline **1** on the three investigated columns are
181 given in **Figure 2** together with the chromatographic profile obtained on the Acquity CSH C18. The
182 effect of TBA is clearly shown in the figure. Nice peak shapes with asymmetry values close to 1.00
183 were recorded in addition to a decrease in retention time.

184 Comparing the column performances in the absence of additive (blue traces in **Figure 2**), the Ascentis
185 Express C18 seems to be the most sensible to the presence of cationic additive moving the *As5%* from
186 3.48 to 1.24. **Table S3** completes the investigation of all compounds with RP^{plusTBA} elution. In some
187 cases, two different elution conditions were evaluated, aiming to observe the peak distortion at high
188 values of retention (sertraline, paroxetine and citalopram in **Table S3**). Interestingly, the *As5%* values
189 remain almost constant. A deeper investigation of retention factor and plate number as a function of
190 organic modifier concentration has been performed for compound **1** on the Ascentis Express C18.

191 Logarithmic retention of amitriptyline in RP and RP^{plusTBA} modes decreases linearly with % ACN
192 (**Figure 3**). In order to obtain comparable ranges of log *k* with the two elution modes, lower contents
193 of ACN in the mobile phase were used in the RP^{plusTBA} mode. As an example, the right part of **Figure**
194 **3** shows that similar *k* values (6.17 and 8.51 in RP^{plusTBA} and RP, respectively) are recorded by using
195 20% and 30% of acetonitrile, with a fourfold reduction of peak asymmetry in RP^{plusTBA} mode.

196 This chromatographic behaviour indicates a shift in the prevailing retention mechanism from the
197 typical reversed phase to the electrostatic repulsion supported by the TBA additive.

198 Moreover, the higher slope of RP^{plusTBA} straight line provides a reduction of retention time with a tiny
199 increase of acetonitrile. At 35% of organic modifier, as reported at the bottom of **Figure 3**, the
200 retention factor is close to zero, compared to $k = 3.53$ in RP mode.

201 Different trends were observed for both efficiency and asymmetry as a function of retention factor.
202 **Figure 4 A** shows the variation of plate number per column when retention factor increases with the
203 content of water in the mobile phase. While in acidic RP conditions, we observe a van Deemter like
204 curve, in RP^{plusTBA} conditions experimental data fit with an inverted van Deemter like curve. Peak
205 asymmetry increases rapidly with retention factor in RP, while ~~and~~ it is smaller and remains almost
206 constant in RP^{plusTBA} mode (**Figure 4 B**).

207 Although it is difficult to provide a model detailing the molecular interaction mechanisms operative
208 in our chromatographic systems (due to the coexisting of hydrophobic, electrostatic repulsion, ion
209 exchange interactions), we believe the use of cationic, hydrophobic mobile phase additives is a
210 convenient solution for the elution of basic compounds. Similar results were in fact obtained with
211 sertraline (**Figure S1**) whose structural similarity with amitriptyline is limited to the exocyclic-alkyl-
212 amino portion, thus corroborating the assumption that electrostatic repulsion plays an important role
213 in the elution mechanism. With increased complexity of molecular structure, especially in the
214 presence of additional polar functional groups, the control of chromatographic properties in RP^{plusTBA}
215 requires a case-by-case investigation respect to the previous models. However, RP^{plusTBA} elution mode
216 results effective in any case as shown by the radar plots in **Figure 5**. In these plots, investigated
217 compounds were placed at each vertexes of radars while the axes report the asymmetry values or the
218 efficiency. Data points represent the best elution conditions obtained for each compound and the
219 connecting line gives an idea of the total score for each column. Detailed data of single elution
220 conditions are listed in **Tables S1-S3**. The Ascentis Express C18 column is mainly sensitive to the
221 presence of TBAHSO₄ and asymmetry values have been shifted from 3-4 to about 1 in all cases
222 (**Figure 5 A**). Comparing the results achieved on the three UHPLC columns, in **Figure 5 B**, the
223 Acquity BEH C18 column shows better $As_{5\%}$ and N/m values in line with the particle proprieties. The

224 only compound excluded from data analysis was atenolol that in all tested conditions was eluted with
225 the void volume. A selection of chromatographic traces of different probes, with and without the TBA
226 cationic additives, is presented in **Figure 6**. The chromatograms emphasize the quality of peak shape,
227 like a perfect Gaussian profile, in the elution conditions providing medium to high retentions.
228 Moreover, in two examples related to paroxetine and citalopram, the chromatograms prove that highly
229 symmetric peaks are obtained even with high retention factors, like in the amitriptyline case. The best
230 elution conditions for all tested compounds are given in **Table 2**.

231 A small sub-class of investigated compounds is represented by the beta-blockers. The effect of
232 electrostatic repulsion for these samples is very large, resulting in small retention factors even in the
233 presence of minor amounts of organic modifier (2% ACN, **Figure 7**). Moreover, the beta-blocker
234 nadolol has more than one stereocenters and exist as a pair of diastereomers that can be in principle
235 separated by achiral chromatography. Indeed, all UHPLC columns can separate the two diastereomers
236 (data in **Table S2**) by using TBAHSO₄ additive demonstrating a diastereoselectivity and efficiency
237 higher than that recorded by using TBAOH/TFA mixture in the mobile phase or recorded on the
238 Acquity CSH C18 column. As shown in **Figure 7**, two well separated peaks were observed at
239 reasonable retention time with a resolution of 2.34 ($\alpha = 1.12$), while the charged surface hybrid
240 stationary phase missed this target and showed a chromatographic performance similar to classical
241 C18 stationary phase (no diastereomer separation) except for the better peak symmetry under the
242 same RP conditions.

243

244 **4. Conclusions**

245 Recent years have seen a great research effort to finding solutions for the UHPLC separation of basic
246 compounds in reversed phase conditions. One advantageous approach exploits mobile phase cationic
247 hydrophobic additives. In this study, the TBAHSO₄ additive has been employed to improve the
248 elution of basic molecules of pharmaceutical interest on several UHPLC columns packed with C18

249 stationary phases. TBAHSO₄ resulted convenient for the analysis of basic compounds providing
250 highly symmetric peaks in a wide range of retention factors. All tested UHPLC columns, based on
251 either pure silica or hybrid organic-silica, benefit on the use of this mobile phase additive. In addition,
252 the mixed retention mechanism based on electrostatic repulsion and hydrophobic effect, results in
253 high efficiencies that usually compensate for low selectivities. Finally, the obtained results showed
254 the TBAHSO₄/C18 stationary phase combination represents a valid alternative to the use of CSH
255 materials.

256 **Acknowledgments**

257 This work was supported by Scientific Research “Ateneo 2018” funds (“Sapienza” Università di
258 Roma).

259 **Conflict of interest statement**

260 The authors declare no commercial or associative conflict of interest.

261 **References**

262 [1] Mc Calley D. V., The challenges of the analysis of basic compounds by high performance liquid
263 chromatography: Some possible approaches for improved separations. *J. Chromatogr. A* 2010, *1217*,
264 858-880.

265 [2] Petersson P., Forssen P., Edstrom L., Samie F., Tatterton S., Clarke A., Fornstedt T., Why ultra-
266 high performance liquid chromatography produces more tailing peaks than high performance liquid
267 chromatography, why it does not matter and how it can be addressed. *J. Chromatogr. A* 2011, *1218*
268 6914-6921.

269 [3] Gritti F., Guiochon G., Effect of the surface heterogeneity of the stationary phase on the range of
270 concentrations for linear chromatography. *Anal. Chem.* 2005, *77*, 1020-1030.

271 [4] Nawrocki J., The silanol group and its role in liquid chromatography. *J. Chromatogr A* 1997, *779*,
272 29-71.

- 273 [5] McCalley D. V., Rationalization of retention and overloading behavior of basic compounds in
274 reversed-phase HPLC using low ionic strength buffers suitable for mass spectrometric detection.
275 *Anal. Chem.* 2003, 75, 3404-3410.
- 276 [6] Neue U. D., Tran K., Méndez A., Carr P. W., The combined effect of silanols and the reversed-
277 phase ligand on the retention of positively charged analytes. *J. Chromatogr. A* 2005, 1063, 35-45.
- 278 [7] Bocian S., Buszewski B., Residual silanols at reversed-phase silica in HPLC – a contribution for
279 a better understanding. *J. Sep. Sci.* 2012, 35, 1191-1200.
- 280 [8] Bartha A., Stihlberg J., Electrostatic retention model of reversed-phase ion-pair chromatography.
281 *J. Chromatogr. A* 1994, 668, 255-284.
- 282 [9] Ruiz-Angel M. J., Torres-Lapasió J. R., Carda-Broch S., García-Alvarez-Coque M. C.,
283 Improvement of peak shape and separation performance of b-blockers in conventional reversed-
284 phase columns using solvent modifiers. *J. Chromatogr. Sci.* 2003, 41(7), 350-358.
- 285 [10] Kaliszan R., Marszałł M. P., Markuszewski M. J., Baczek T., Pernak J., Suppression of
286 deleterious effects of free silanols in liquid chromatography by imidazolium tetrafluoroborate ionic
287 liquids. *J. Chromatogr. A* 2004, 1030, 263-271.
- 288 [11] Pan L., LoBrutto R., Kazakevich Y. V., Thompson R., Influence of inorganic mobile phase
289 additives on the retention, efficiency and peak symmetry of protonated basic compounds in reversed-
290 phase liquid chromatography. *J. Chromatogr. A* 2004, 1049, 63-73.
- 291 [12] Ubeda-Torres M. T., Ortiz-Bolsico C., García-Alvarez-Coque M. C., Ruiz-Angel M.J., Gaining
292 insight in the behaviour of imidazolium-based ionic liquids as additives in reversed-phase liquid
293 chromatography for the analysis of basic compounds. *J. Chromatogr. A* 2015, 1380, 96-103.
- 294 [13] Calabuig-Hernández S., García-Alvarez-Coque M. C., Ruiz-Angel M. J., Performance of amines
295 as silanol suppressors in reversed-phase liquid chromatography. *J. Chromatogr. A* 2016, 1465, 98-
296 106.

- 297 [14] Caban M., Stepnowski P., The antagonistic role of chaotropic hexafluorophosphate anions and
298 imidazolium cations composing ionic liquids applied as phase additives in the separation of tri-cyclic
299 antidepressants. *Anal. Chim. Acta* 2017, 967, 102-110.
- 300 [15] Buszewska-Forajta M., Markuszewski M. J., Kaliszan R., Free silanols and ionic liquids as their
301 suppressors in liquid chromatography. *J. Chromatogr. A* 2018, 1559, 17-43.
- 302 [16] Peris-García E., García-Alvarez-Coque M.C., Carda-Broch S., Ruiz-Angel M.J., Effect of buffer
303 nature and concentration on the chromatographic performance of basic compounds in the absence
304 and presence of 1-hexyl-3-methylimidazolium chloride. *J. Chromatogr. A* 2019, 1602, 397-408.
- 305 [17] Tossell J. A., Sahai N., Calculating the acidity of silanols and related oxyacids in aqueous
306 solution. *Geochem. Cosmochim. Acta* 2000, 64, 4097-4113.
- 307 [18] Rosenholm J. M., Czuryzkiewicz T., Kleitz F., Rosenholm J. B., Lindèn M., On the nature of
308 the brønsted acidic groups on native and functionalized mesoporous siliceous SBA-15 as studied by
309 benzylamine adsorption from solution. *Langmuir* 2007, 23, 4315-4323.
- 310 [19] Reyes-Reyes M. L., Melgar-Fernández R., Balderas-Hernández P., Roa-Morales G., UHPLC
311 determination of enantiomeric purity of sertraline in the presence of its production impurities.
312 *Chromatographia* 2014, 77, 19-20.
- 313 [20] Zhang W., He L., Gu Y., Liu X., Jiang S., Effect of ionic liquids as mobile phase additives on
314 retention of catecholamines in reversed-phase high-performance liquid chromatography. *Anal. Lett.*
315 2003, 36, 827-838.
- 316 [21] Paul A., Mandal P. K., Samanta A., On the optical properties of the imidazolium ionic liquids.
317 *J. Phys. Chem. B* 2005, 109, 9148-9153.
- 318 [22] Wyndham K. D., Gara J. O', Walter T., Glose K., Lawrence N., Alden B., Izzo G., Hudalla C.,
319 Iraneta P., Characterization and evaluation of C18 HPLC stationary phases based on ethyl-bridged
320 hybrid organic/inorganic particles. *Anal. Chem.* 2003, 75, 6781-6788.

321 [23] Wyndham K., Walter T., Iraneta P., Alden B., Bouvier E., Hudalla C., Lawrence N., Walsh D.,
322 Synthesis and applications of BEH particles in liquid chromatography. *LCGC N. Am.* 2012, *30*, 20-
323 29.

324 [24] Pesek J. J., Matyska M. T., Hydride-based silica stationary phases for HPLC: Fundamental
325 properties and applications. *J. Sep. Sci.* 2005, *28*, 1845-1854.

326 [25] O'Sullivan G. P., Scully N. M., Glennon J. D., Polar-embedded and polar-encapped stationary
327 phases for LC. *Anal. Letters* 2010, *43*, 1609-1629.

328 [26] Long Z., Wang C., Guo Z., Zhang X., Nordahl L., Liang X., Strong cation exchange column
329 allow for symmetrical peak shape and increased sample loading in the separation of basic compounds.
330 *J. Chromatogr A* 2012, *125*, 67-71.

331 [27] Davies N. H., Euerby M. R., McCalley D.V., A study of retention and overloading of basic
332 compounds with mixed-mode reversed-phase/cation-exchange columns in high performance liquid
333 chromatography. *J. Chromatogr A* 2007, *1138*, 65-72.

334 [28] Qiua H., Mallik A. K., Takafujia M., Liu X., Jiang S., Ihara H., A new imidazolium-embedded
335 C18 stationary phase with enhanced performance in reversed-phase liquid chromatography. *Anal.*
336 *Chim. Acta* 2012, *738*, 95-101.

337 [29] Iraneta P. C., Wyndham K. D., McCabe D. R., Walter T. H., A review of Waters hybrid particle
338 technology- Part 3. Waters Corporation, Milford, USA, 2010.

339 [30] Lucie N., Hana V., Solich P., Evaluation of new mixed-mode UHPLC stationary phases and the
340 importance of stationary phase choice when using low ionic-strength mobile phase additives. *Talanta*
341 2012, *93*, 99-105.

342 [31] Gritti F., Guiochon G., Adsorption behaviors of neutral and ionizable compounds on hybrid
343 stationary phases in the absence (BEH-C18) and the presence (CSH-C18) of immobile surface
344 charges. *J. Chromatogr. A* 2013, *1282*, 58-71.

- 345 [32] Gritti F., Guiochon G., Effect of the ionic strength on the adsorption process of an ionic surfactant
346 onto a C18-bonded charged surface hybrid stationary phase at low pH. *J. Chromatogr. A* 2013, *1282*
347 46-57.
- 348 [33] Gritti F., Guiochon G., Hydrophilic interaction chromatography: A promising alternative to
349 reversed-phase liquid chromatography systems for the purification of small protonated bases. *J. Sep.*
350 *Sci.* 2015, *38*, 1633-1641.
- 351 [34] Gritti F., Guiochon G., Separation of peptides and intact proteins by electrostatic repulsion
352 reversed phase liquid chromatography. *J. Chromatogr A* 2014, *1374*, 112-121.
- 353 [35] Gritti F., Guiochon G., Comparison between the adsorption behaviors of an organic cation and
354 an organic anion on several reversed-phase liquid chromatography adsorbents. *J. Chromatogr. A*
355 2004, *1048*, 1-15.
- 356 [36] Gritti F., Guiochon G., Effect of the ionic strength of salts on retention and overloading behavior
357 of ionizable compounds in reversed-phase liquid chromatography I. Xterra-C18. *J. Chromatogr. A*
358 2004, *1033*, 43-55.
- 359 [37] Gritti F., Guiochon G., Critical contribution of nonlinear chromatography to the understanding
360 of retention mechanism in reversed-phase liquid chromatography. *J. Chromatogr. A* 2005, *1099*, 1-
361 42.
- 362 [38] Gritti F., Guiochon G., Determination of the adsorption energy distribution of neutral and
363 charged compounds onto endcapped silica-C18 adsorbent from polar liquid phases. *J. Colloid* 2009,
364 *71*, 480-486.
- 365 [39] Stevenson P.G., Gritti F., Guiochon G., Automated methods for the location of the boundaries
366 of chromatographic peaks. *J. Chromatogr. A* 2011, *1218*, 8255-8263.
- 367 [40] Cecchi T., Passamonti P., Retention mechanism for ion-pair chromatography with chaotropic
368 reagents. *J. Chromatogr. A* 2009, *1216*, 1789-1797.

369

370

371 **Figure captions.**

372 **Figure 1.** Structures of investigated compounds.

373 **Figure 2.** Amitriptyline eluted on the four investigated columns. Analytical conditions: in RP
374 H₂O/ACN 65/35 + 0.1% TFA (blue traces); in RP^{plusTBA} H₂O/ACN 75/25 + 5mM TBAHSO₄ (orange
375 traces). Flow rate 0.5 mL/min, T = 35°C, Sample: 1µg on column. Geometry columns: 100 mm L x
376 3.0 mm I.D.

377 **Figure 3.** Log k vs %ACN plot in RP and RP^{plusTBA} and chromatograms relating to the two areas
378 highlighted in the graph (right: similar k, bottom: similar %ACN). Sample: amitriptyline. Column
379 Ascentis Express C18, 100 mm L x 3.0 mm I.D. Analytical conditions: in RP H₂O/ACN + 0.1% TFA
380 (blue dots and trace); in RP^{plusTBA} H₂O/ACN + 5mM TBAHSO₄ (orange dots and trace). Flow rate
381 0.5 ml/min. T = 35°C.

382 **Figure 4. A)** Plates and **B)** As_{5%} vs k plots in RP and RP^{plusTBA}. See details in **Figure 3.**

383 **Figure 5. A)** Radar chart corresponds to the Ascentis Express C18 for both RP (blue trace) and
384 RP^{plusTBA}(orange trace) conditions. Vertexes are investigated compounds, axes the As_{5%}.. **B)** Radar
385 charts correspond to the three UHPLC columns (Titan C18 (green), Acquity BEH C18 (black) and
386 Ascentis Express C18 (gray)) in only RP^{plusTBA} conditions. Vertexes are investigated compounds,
387 axes are As_{5%} (left) and N/m (right). Eluents composition is shown in Supporting Information, **tables**
388 **S1-S2-S3.**

389 **Figure 6.** RP^{plusTBA} (orange traces) vs RP (blue traces) approaches. Columns and samples are reported
390 in the graphs. Eluents as reported in **Figure 3.** Detailed elution conditions: Sertraline: A/B 35/65 and
391 A/B 75/25 in RP and RP^{plusTBA} respectively. Citalopram: A/B 65/35 and A/B 80/20. Ketamine: A/B
392 85/15 and A/B 98/2. Paroxetine: A/B 65/35 and A/B 80/20. Lidocaine: A/B 85/15 and A/B 98/2.
393 Mirtazapine: A/B 90/10 and A/B 90/10.

394 **Figure 7.** Elution profiles of beta-blockers. Columns and samples are reported in the graphs. Eluents
395 as reported in **Figure 3**. Detailed elution conditions: Pindolol: A/B 90/10 and A/B 98/2, Sotalol: A/B
396 95/5 and A/B 98/2, Nadolol: A/B 90/10 and A/B 98/2 in RP (blue traces) and RP^{plusTBA} (orange trace)
397 respectively.