1	Elemental concentration and migratability in bioplastics derived from organic waste
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16	
17	Abstract
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19	In line with the Circular Economy approach, the production of polyhydroxyalkanoate (PHA) with organic
20	waste as the feedstock may a biotechnological application to reduce waste and recover high-value materials.
21	The potential contaminants that could transfer from bio-waste to a PHA include inorganic elements, such as
22	heavy metals. Hence, the total content and migratability of certain elements were evaluated in several PHA
23	samples produced from different origins and following different methods. The total content of certain
24	elements in PHA ranged between 0.0001 (Be) and 49500 mg kg <sup>-1</sup> (Na). The concentrations of some alkaline
25	(Na and K) and alkaline earth (Ca and Mg) metals were highest, which are of little environmental concern.
26	The feedstock type and PHA stabilisation and extraction procedures affected the element contents. Several
27	sets of experiments were conducted to evaluate the migration of elements from the PHA samples under

28 different storage times, temperatures, and pH levels. The total contents of some heavy metals (As, Cd, Fe,

36	Keywords
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34	to come into contact with food under frozen and refrigerated conditions.
33	Toy Safety Directive and by Commission Regulation (EU) 10/2011 on plastic materials and articles intended
32	phase extraction reagents and commercial PHA were below the migration limits stipulated by the current
31	wastewater treatment. Both the PHA obtained by extraction from wet biomass (acid storage) with aqueous
30	the PHA samples produced from the mixture of the organic fraction of municipal waste and sludge from
29	Hg, Ni, Pb, and Zn) in PHA produced from fruit waste or crops (commercial PHA) were lower than those in

- Biopolymers; polyhydroxyalkanoates; bioaccessibility; toxic elements; inductively coupled plasma mass
  spectrometry; cold vapour generation atomic fluorescence spectrometry.

### 41 1. Introduction

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43 Polyhydroxyalkanoates (PHAs) can be considered to be the sustainable polymers of the future due to their 44 physicochemical properties, biodegradability, and biocompatibility (Kourmentza et al., 2017; Tsang et al., 2019; Valentino et al., 2017). In line with the Circular Economy approach (European Commission, 2015), 45 the production of PHAs may be a biotechnological application to reduce waste through reuse and recovering 46 47 high-value substances. PHAs are naturally polymerised by bacteria using organic waste as a low-cost carbon 48 source (Khiewwijit et al., 2015; Kourmentza et al., 2017). In recent years, the organic fraction of municipal 49 solid waste has been used as a new feedstock for microorganism-based PHA production, which has become a 50 valuable strategy for reducing biopolymer production costs and increasing their sustainability (Valentino et 51 al., 2018; Valentino et al. 2019).

52 One of the main concerns relating to the use of waste as a raw material is the content of priority pollutants 53 that can migrate from the waste, through the technology chain, and into the end-products (i.e., PHA). The possible chemical contaminants include inorganic elements, such as heavy metals. In recent years, ecological 54 55 and public health concerns associated with environmental contamination by these elements have increased 56 (Astolfi et al., 2019a; He et al., 2005). Heavy metal (As, Cd, Hg, and Pb) toxicity can damage the functioning of the brain, lungs, kidney, liver, blood composition, and other important organs (Alina et al., 57 58 2012; Chowdhury et al., 2000; Henson and Chedrese, 2004; Martin and Griswold, 2009; Smith et al., 2000). 59 Repeated long-term exposure to some metals and their compounds may even cause cancer (Järup, 2003). 60 Toxic elements can originate from geogenic, industrial, agricultural, pharmaceutical, domestic, and 61 atmospheric sources (Astolfi et al., 2017, 2006; Canepari et al., 2018; Manigrasso, et al., 2019; Marconi et 62 al., 2011; Vitali et al., 2019). Sources of heavy metals from household municipal solid waste include 63 household dust, batteries, disposable household materials, paints and inks, body care products and medicines, 64 and household pesticides (Bardos, 2004). Significant concentrations of heavy metals are also present in industrial wastewater and municipal sludge (Altas, 2009). 65

66 Under waste law, Italy has a national end-of-waste system that regulates the procedure for the authorisation
67 of certain non-hazardous waste recovery options and establishes the conditions under which biodegradable
68 waste can cease to be considered waste (Italian Regulation, 1998).

69 The European Commission has issued several directives on plastic materials, including Directive 94/62/EC 70 Packaging and Packaging waste (European Parliament, Council of the European Union, 1994), Directive 71 2009/48/EC on the safety of toys (European Parliament, Council of the European Union, 2009), and Commission Regulation (EU) No 10/2011 on plastic materials and articles intended to come into contact 72 73 with food (European Commission, 2011). In particular, the presence of As, Cd, Cr, Hg, and Pb in plastics is 74 regulated by Annex XVII of the EU REACH Regulation on chemicals (European Parliament, Council of the 75 European Union, 2006). Moreover, Directive 2009/48/EC on the safety of toys specifies limits for the 76 migration of 19 elements (Al, Sb, As, Ba, B, Cd, Cr(III), Cr(VI), Co, Cu, Pb, Mn, Hg, Ni, Se, Sr, Sn, organic 77 Sn, and Zn) from toy materials and parts.

To our knowledge, the elements of PHA biopolymers have not yet been determined. This study describes the 78 79 first investigation of the occurrence and migratability of heavy metals and other elements in several PHA samples with different sources and produced following different process steps. The contents of 40 elements 80 81 (Al, As, B, Ba, Be, Bi, Ca, Cd, Ce, Co, Cr, Cs, Cu, Fe, Ga, K, La, Li, Mg, Mn, Mo, Na, Nb, Ni, P, Pb, Rb, 82 Sb, Se, Si, Sn, Sr, Te, Ti, Tl, U, V, W, Zn, and Zr) and Hg were quantified by inductively coupled plasma 83 mass spectroscopy (ICP-MS) and cold-vapour generation atomic fluorescence spectrometry (CV-AFS), 84 respectively. Although there is currently no specific PHA regulation, our results were compared with current 85 regulations and guidelines for similar materials and/or applications.

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#### 87 2. Materials and methods

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#### 89 2.1 PHA samples

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Two types of feedstock were considered, including a) a mixture of the organic fraction of municipal solid waste and biological sludge from urban wastewater treatment (collected from the Treviso pilot plant in Italy; PHA TV) and b) fruit waste (collected in from Lisbon pilot plant in Portugal; PHA L). Details regarding the production of PHA TV have been reported in previous studies (Valentino et al., 2019a, 2019b). The production of PHA L was similar to that for PHA TV. The main difference was the type of feedstock, which was organic waste originating from fruit processing. 97 The extraction step of the production technologies (especially PHA extraction from biomass) received the
98 most attention. Thus, both PHA-rich biomass before extraction and purified PHA after extraction from
99 biomass were considered. Three different types of extraction for the latter were considered.

100 After PHA accumulation, the PHA-rich biomass was stabilised for storage via overnight thermal drying 101 (biomass from Treviso only; PHA TV-d) or acidification with  $H_2SO_4$  (biomass from both Treviso and

102 Lisbon; PHA TV-a and PHA L-a, respectively).

For the type-a) feedstock samples, four different PHA samples were analysed: 1) raw PHA-rich biomass
(dried biomass, no extraction; PHA TV-d-ne); 2) PHA after extraction from dried biomass with CHCl<sub>3</sub>
(reference method; PHA TV-d-CHCl<sub>3</sub>); 3) PHA after extraction from dried biomass with NaClO (PHA TV-d-NaClO); and 4) PHA after extraction from wet biomass (acid storage) with aqueous-phase extraction (PHA

- 107 TV-a-ap). For each type, at least two different samples were collected from the pilot plant at different108 operation times.
- For the type-b) feedstock samples, one type of PHA sample was analysed, i.e., PHA after extraction fromwet biomass (acid storage) with aqueous-phase extraction reagents (PHA L-a-ap).
- Overall, 24 different PHA samples produced under six different conditions were analysed. For comparison, three types of commercial PHA obtained from different producers (PHA comm) that were produced from fruit waste or crops were analysed, and their results were expressed as the average.

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#### 115 2.2. Materials and reagents

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117 Analytical reagent-grade concentrated HNO<sub>3</sub> (67–70%; superpure) was obtained from Carlo Erba Reagents 118 S.r.l. (Milan, Italy), CH<sub>3</sub>COOH (100%; superpure) was purchased from Merck KGaA (Darmstadt, 119 Germany), while HCl (assay >36%; residue <3 mg L<sup>-1</sup>) and H<sub>2</sub>O<sub>2</sub> (assay >30%) were obtained from 120 Promochem, LGC Standards GmbH (Wesel, Germany).

121 The 5% HCl was used as a carrier, and 0.05% NaBH<sub>4</sub> (Sigma-Aldrich Chemie GmbH, St. Louis, USA) in

122 0.05% NaOH (assay >98%, anhydrous pellets, RPE for analysis, ACS – ISO; Carlo Erba Reagents, Milan,

123 Italy) was used as a reducing agent for CV-AFS.

Deionised water (resistivity ≤18.3 MΩ cm) was obtained using an Arioso Power I RO-UP Scholar UV system from Human Corporation (Songpa-Ku, Seoul, Korea). Graduated tubes (2.5-5–10 mL in polypropylene) were purchased from Artiglass S.r.l. (Due Carrare, PD, Italy). Syringe filters with cellulose nitrate membranes (GVS Filter Technology, Indianapolis, USA; pore size, 0.45 µm) were used to filter the samples prior to analysis.

All plastic containers, polypropylene tubes, pipette tips, quartz digestion tubes, and reagents that came intocontact with the samples or standards were checked for contamination.

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# 132 2.3. Standard solutions

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An ICP-MS multi-element standard solution  $(1.000 \pm 0.005 \text{ mg L}^{-1} \text{ As}, \text{ Al}, \text{ Ba}, \text{ Be}, \text{ Bi}, \text{ Cd}, \text{ Cr}, \text{ Cs}, \text{ Cu}, \text{ Ga},$ 134 La, Li, Mn, Mo, Nb, Ni, Pb, Rb, Sb, Se, Sn, Te, Ti, Tl, U, V, W, and Zr;  $5.00 \pm 0.03 \text{ mg L}^{-1}$  Ce and Co; 135  $10.00 \pm 0.05 \text{ mg L}^{-1}$  Fe and Zn;  $50.00 \pm 0.25 \text{ mg L}^{-1}$  P and Si;  $55.00 \pm 0.25 \text{ mg L}^{-1}$  B and Sr;  $500.0 \pm 2.5 \text{ mg}$ 136  $L^{-1}$  K, Mg, and Na; and 1000  $\pm$  5 mg  $L^{-1}$  Ca) was purchased from Ultra Scientific/Agilent Technologies 137 (North Kingstown, RI, USA), while a CV-AFS standard solution of Hg ( $1002 \pm 7 \text{ mg L}^{-1}$ ) was obtained from 138 139 SCP Science (Baie D'Urfé, Canada). All standard solutions were diluted with the same percentage of acid present in the samples [3% (v/v) HNO<sub>3</sub> for the digests, and 1% (v/v) HNO<sub>3</sub>, 3% (v/v) CH<sub>3</sub>COOH, or 0.07 M 140 141 HCl for the extracts obtained by the migration tests].

To maintain the daily ICP-MS performance, a multi-standard tuning solution with 0.005 mg L<sup>-1</sup> of Ba, Be, Ce, Co, In, Pb, Mg, Tl, and Th was prepared from a multi-standard stock solution  $(10.00 \pm 0.05 \text{ mg L}^{-1})$ obtained from Spectro Pure/Ricca Chemical Company (Arlington, TX, USA). A standard solution of Y (at 0.005 mg L<sup>-1</sup> from 1000 ± 2 mg L<sup>-1</sup>), purchased from Panreac Química (Barcelona, Spain), and a multielement standard solution of Sc, Rh, In, and Th (at 0.010 mg L<sup>-1</sup> from 1000 ± 5 mg L<sup>-1</sup>), obtained from Merck KGaA (Darmstadt, Germany), in 1% (v/v) HNO<sub>3</sub> were employed as the internal standards to control the nebuliser efficiency, as previously reported (Astolfi et al., 2020a, 2020b).

A solution of trace SRM 1643e elements in water (National Institute of Standards and Technology, NIST;Gaithersburg, MD, USA) was used for quality control.

152 **2.4. Instrumentation** 

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A quadrupole ICP-MS (820-MS Bruker; Bremen, Germany) equipped with a collision-reaction interface (CRI) and glass nebuliser (0.4 mL min<sup>-1</sup>; MicroMistTM; Analytik Jena AG, Jena, Germany) was used to analyse all elements, excluding Hg, which was analysed by CV-AFS (AFS 8220 Titan, FullTech Instruments; Rome, Italy). The operating parameters for the ICP-MS and CV-AFS analyses are summarised in Table S1 and are detailed in previous studies (Astolfi et al., 2020; 2019b). A Gibertini Elettronica (Milan, Italy) Europe 60 analytical balance (sensitivity, 0.1 mg) was used to weigh all samples.

A Milestone (FKV; Sorisole, Bergamo, Italy) Ethos1 Touch Control microwave system was used for acid
digestion, and a water bath (WB12; Argo Lab, Modena, Italy) with electronic temperature control was used
for the migration tests.

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#### 165 **2.5. Sample preparation**

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# 167 **2.5.1 Total acid digestion**

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169 Weighed amounts (~50 mg or ~200 mg for CV-AFS or ICP-MS analysis, respectively) of the PHA samples 170 were transferred into quartz vessels; to these, 1 mL 67% HNO<sub>3</sub>, 0.5 mL 30% H<sub>2</sub>O<sub>2</sub>, and 1.5 mL deionised water were added. The resulting solutions were subsequently heated with microwave energy for 40 min. The 171 following heating program was used: first step, heating to 50 °C for 5 min; second step, hold temperature at 172 50 °C for 5 min; third step, heating to 100 °C for 5 min; fourth step, hold temperature at 100 °C for 5 min; 173 174 fifth step, heating to 150 °C for 5 min; sixth step, hold temperature at 150 °C for 5 min; seventh step, heating 175 to 180 °C for 5 min; and eighth step, hold temperature at 180 °C for 15 min. Prior to use, the quartz vessels 176 were cleaned with the same reagents and digestion program used for the PHA samples, rinsed with deionised 177 water, and thoroughly dried with absorbent paper. The solutions obtained from digestion were diluted to 5 mL with 3% HCl or 10 mL with deionised water and filtered for CV-AFS or ICP-MS analysis, respectively. 178 Prior to ICP-MS analysis, the digests were further diluted 1:10 and 1:50 with deionised water. 179

Blanks (3% HNO<sub>3</sub>) were periodically analysed alongside the samples to check for any losses or cross-contamination.

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# 183 2.5.2 Migration tests

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Ten blank solutions were tested with each sample set extracted during the different migration tests to subtract the background signal from the reagents and trace and control the contributions from possible sample contamination. The blanks were deducted from all measurements. The procedure for each treatment is detailed in the following sections. At the end of each migration test, the solution was filtered and analysed by CV-AFS or ICP-MS. Prior to CV-FS analysis, the extracts were further diluted with 6% HCl in a ratio of 1:2.

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# 191 **2.5.2.1 H<sub>2</sub>O treatment experiments**

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Weighed amounts (~25 mg) of the PHA samples were transferred into polypropylene tubes, to which 5 mL of deionised  $H_2O$  (pH ~7) was added. The tubes were then covered with a cap and heated at 70 °C for 2 h or stored in the dark at 20 °C for 10 days.

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#### 197 **2.5.2.2 CH<sub>3</sub>COOH treatment experiments**

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Weighed amounts (~25 mg) of the PHA samples were extracted in dilute CH<sub>3</sub>COOH (food simulant B; pH ~4.5) according to Commission Regulation (EU) No 10/2011 (European Commission, 2011) on plastic materials and articles intended to come into contact with food. Five millilitres of a 3% CH<sub>3</sub>COOH solution, simulating food with a pH below 4.5, were pipetted into each tube with the PHA samples, which were then capped and placed in a water bath at 70 °C for 2 h or stored in the dark at 20 °C for 10 days.

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# 205 2.5.2.3 HCl treatment experiments

Weighed amounts (~25 mg) of the PHA samples were transferred into polypropylene tubes mixed with 5 mL
of 0.07-M HCl and then heated in a water bath for 2 h to 37 °C according to European standard EN 71-3
(British Standards Institution, 1994).

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#### 211 **2.6. Method performance**

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213 Method performance parameters, such as the trueness [per cent bias (%bias) or recovery (%R)], precision 214 (intraday and interday within-laboratory precision), linearity range, and detection and quantification limits 215 (LODs and LOQs, respectively), were assessed. Fortified PHA TV-a-ap samples were used for the quality control of the total digestion treatment, while a fortified reference water solution (SRM 1643e) was used for 216 the migration tests. Detailed information on the calibration graphs and quality control for elemental analysis 217 was shown in the Supplementary Material. Briefly, the instruments were calibrated externally using mixed 218 219 standards prepared by serial dilution in the same matrix as the final samples (3% HNO<sub>3</sub> for digests, and 1% HNO<sub>3</sub>, 3% CH<sub>3</sub>COOH, and 0.07 M HCl for extracts). The linear ranges of the 41 elements examined in this 220 221 study are listed in Table S2.

Owing to the lack of certified reference materials and a blank matrix for PHA biopolymers, the analytical quality control was verified by conducting recovery experiments. The obtained recoveries of the spiked PHA samples ranged from 90 to 102% (Table S2), confirming that there was no significant loss during digestion or contamination, or interference during analysis.

A reference water solution (SRM 1643) was fortified with 2  $\mu$ g L<sup>-1</sup> Hg; 25  $\mu$ g L<sup>-1</sup> Ce, Cs, Ga, La, Nb, Sn, Ti, U, W, and Zr; and 2500  $\mu$ g L<sup>-1</sup> P and Si to assess the quality of the analysis of extracts obtained by the migration tests. Fortified SRM 1643e was analysed after each calibration curve and revealed element concentrations that were within 10% of the certified values and ranged from 90 to 96% of spiked elements, excluding Al, As, Ba, Se, and Te (within 15%; Table S2).

The method was determined to be within-laboratory precision, both intra-day (repeatability) and inter-day (reproducibility), at the concentration used for the recovery study by calculating the per cent coefficient of variation (%CV). Three spiked samples were analysed on the same day to assess the repeatability and on three different days to assess the reproducibility. The results are listed in Table S2. The LODs and LOQs were calculated as three and ten times the standard deviation of the blank samples, respectively (Table S3). The method blanks were treated as samples to subtract the background signal from the reagents. The LODs and LOQs ranged from 0.0001 (Be and Nb) to 100 mg kg<sup>-1</sup> (Ca) and 0.0004 (Be) to 350 mg kg<sup>-1</sup> (Ca), respectively.

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# 240 **2.7. Statistical analysis**

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Statistical analyses were conducted using IBM SPSS Statistics 25 software (IBM Corp., Armonk, NY, USA).
For each element, values below the LOD were replaced with a value equal to half the LOD. When the
percentage of values <LOD exceeded 20%, the element was excluded from the statistical elaboration.</li>

The differences in the PHA concentration were tested by Kruskal-Wallis and pairwise post-hoc tests. The results (Table 1) were considered to be statistically significant with p-values of <0.05.

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#### 248 **3. Results and discussion**

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# 250 **3.1 Total elemental content**

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Table 1 shows the results of the Kruskal Wallis test, which was conducted to verify the presence and concentrations of differences elements among the six different types of PHA samples. Nb exhibited no significant differences between all of the PHA samples, while the levels of Bi, Se, Te, Tl, V, W, and Zr were very low or below the LODs, with an inadequate number of observations for multivariate analysis (Table 1). Thus, these elements were eliminated from the multivariate analysis data set.

Focusing on PHA-rich biomass with no extraction (PHA content around 50% w/w, after thermal drying), the concentrations of alkaline and alkaline earth metals were highest, with the levels of Na, Ca, Mg, and K ranging between 0.2% and 5% on a dry-weight basis. Furthermore, P was present at a concentration of approximately 0.4%, while Fe and Mn were present at concentrations of approximately 0.1% and 0.01%, respectively. However, all of these elements are of no or little concern from environmental or health viewpoints. Zn was the heavy metal present at the highest concentration of approximately 300 mg kg<sup>-1</sup> 263 (0.03%). The concentration of Cu was approximately 70 mg/kg (0.007%), while those of other toxic metals, 264 such as Ni and Pb, were below 10 mg kg<sup>-1</sup> (0.001%). Moreover, Cd is below 0.1 mg kg<sup>-1</sup> (0.0001%). The 265 contents of metalloids, such as and Se, were below 1 mg kg<sup>-1</sup> (0.0001%).

PHA extraction from dried biomass using hypochlorite destroys the organic matter, causing it to concentrate or dilute the metals, depending on how strongly sorbed to the remaining solid phase they are. For example, the contents of Al, As, Hg, K, Na, Mo, and U were strongly reduced, while those of Cd, Fe, Mn, Pb, and Zn were increased. Furthermore, chloroform dissolves the PHA, causing the metals to remain in the residual solid phase, i.e., chloroform extraction also acts as a method of purifying PHA from inorganic elements (Ca, Cd, Cu, Mn, Na, Rb, Sr, and Zn). The contents of all investigated elements decreased, excluding B, Cr, Nb, Sb, Se, Te, Tl, V, W, and Zr, which were already at a low concentration.

The metal concentrations of biomass that underwent extraction after acid stabilisation and aqueous-phase extraction were typically lower than those of biomass that underwent stabilisation by thermal drying. This could be expected as inorganic elements were retained in the solid phase during thermal drying, while they could be desorbed more easily in the liquid phase during acidic stabilisation.

The effects of different feedstock types (municipal waste/sludge vs fruit waste) under the same acid stabilisation and aqueous-phase extraction were compared. The comparison of PHA TV-a-ap and PHA L-aap indicates that more inorganic elements are present in the PHA derived from municipal waste/sludge than that derived from fruit waste (excluding As, Na, and Si, although the difference was not significant). The concentration of inorganic elements in commercial PHA, which is industrially obtained from crop substrates, is usually lower than that of waste-based PHA.

283 The contents of some environmentally relevant elements (Al, As, Cd, Hg, Ni, Pb, and Zn) were highest in the 284 PHA derived from municipal waste. This was not surprising, as this source also included a significant 285 fraction of sludge from wastewater treatment (approximately one-third). However, there are still 286 environmental benefits to recovering organic carbon from waste in the form of biodegradable plastics that can counteract the higher level of contaminants, if the content of contaminants is sufficiently low. The 287 contents of most relevant contaminants were at the mg kg<sup>-1</sup> or  $\mu$ g kg<sup>-1</sup> level, which can be considered 288 acceptable. The European Commission has issued several directives on requirements for plastic materials 289 290 intended to be used in different ways, such as Directive 94/62/EC on packaging and packaging waste 291 (European Parliament, Council of the European Union, 1994), Directive 2009/48/EC on the safety of toys 292 (European Parliament, Council of the European Union, 2009), and Commission Regulation (EU) No 10/2011 293 on plastic materials and articles intended to come into contact with food (European Commission, 2011). In particular, Annex II of Directive 94/62/EC (European Parliament, Council of the European Union, 1994) on 294 packaging and packaging waste determines the essential requirements that all packaging on the market 295 within the European Community should comply with. Article 11 of Directive 94/62/EC (European 296 297 Parliament, Council of the European Union, 1994) states that Member States shall ensure that the sum of the 298 concentrations of Pb, Cd, Hg, and Cr(VI) in packaging or packaging components shall not exceed 100 mg kg<sup>-1</sup>. The sum of these metals was well below this level in selected PHA samples. 299

According to regulation 1907/2006/EC (European Parliament, Council of the European Union, 2006), Cd 300 301 contamination receives particular attention, and the regulation states that mixtures and articles composed of plastic materials shall not be placed on the market if the concentration of Cd is equal to or greater than 0.01% 302 (100 mg kg<sup>-1</sup>) of the plastic material. The Cd contents of all waste-based PHA samples in our study are well 303 304 below this limit. Commission Regulation (EU) No 10/2011 establishes a set of specific requirements for 305 PHA obtained through microbial fermentation by using Alcaligenes eutrophus. According to this regulation, PHA should not contain more than 100, 5, 2, 1, and 1 mg kg<sup>-1</sup> of Zn, Cu, Pb, As, and Cr, respectively (Table 306 2). By comparing these values with Table 1, it can be seen that the contents of Cr, Cu, and Pb in PHA 307 308 produced from municipal waste/sludge exceeded these limits, while only the content of Cu exceeded the 309 limit in PHA from fruit waste. Additionally, the content of Cr in commercial PHA exceeded the limit. Hence, 310 in terms of heavy metals content, these samples did not comply with the regulations for plastics intended to 311 come into direct contact with food, and the extraction techniques need to be further improved.

Inorganic species may be present as residues from the catalysts or additives used in the production of polyethene terephthalate (PET), the most favourable packaging material for drinking water (Bach et al., 2012). Sb<sub>2</sub>O<sub>3</sub> is the preferred catalyst for the synthesis of PET due to its low cost and sufficient catalytic activity (Keresztes et al., 2009), and PET typically contains Sb levels in the range of 170-300 mg kg<sup>-1</sup> (Nishioka et al., 2002; Keresztes et al., 2009; Westerhoff et al., 2008). The maximum average content of Sb (0.131  $\pm$  0.085 mg kg<sup>-1</sup>; Table 1) of the studied PHA samples was approximately 1000 times lower than that in PET. Other metals, such as Co, Cr, Fe, and Mn, have also been found in PET bottles, with contents of 27, 0.1, 1.3, and 0.3 mg kg<sup>-1</sup>, respectively (Westerhoff et al., 2008). In this case, the content of Co in all the PHA
samples was at least 100 times lower, while that of Mn was lower than or similar to the values in literature
for PHA L-a-ap and PHA comm.

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#### **323 3.2 Migratable concentrations**

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Extraction tests were conducted on all PHA samples to assess compliance/noncompliance with the European
Directive migration limit (Table 2) and explore possible controls and variations in element migratability.

Table 3 summarises the results obtained for all analysed elements by the different migration tests in terms of concentration, and detailed data are provided in the Supplementary Material (Tables S4-S8). The migratable concentrations ranged from  $<0.2 \ \mu g \ kg^{-1}$  for Be, Cs, La, Nb, Tl, and U to  $>1000 \ m g \ kg^{-1}$  for Ca, K, Mg, Na, and P, and was highly variable, ranging from less than 1% to approximately 100%. The results for each treatment are discussed in detail in the following sections.

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# 333 **3.2.1 Element migratability in water**

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The elemental analysis results for the  $H_2O$  experiment are presented in Tables 3, S4, and S5. The concentrations of many of the minerals commonly found in  $H_2O$ , including Ca, K, Mg, Na, and P, were elevated, and the levels of many trace elements were lower. Heating can increase the migration speed and concentrations of migrated elements than the storage of solutions at room temperature (20 °C). The concentrations of Cu, Mn, and Sb in PHA TV-d-ne and Sb in PHA TV-a-ap after heating to 70 °C for 2 h were higher than those achieved 10 days at 20 °C. Small differences were detected for the other elements in all PHA samples.

The migration test for materials and articles intended for food contact was conducted using the time and temperature specified in Commission Regulation (EU) No 10/2011 (European Commission, 2011). We conducted the following overall migration tests: 1) test for 10 days at 20 °C, covering any contact with food under frozen and refrigerated conditions; and 2) tests for 2 h at 70 °C, covering all other contact conditions, including heating up to 70 °C for up to 2 h, or up to 100 °C for up to 15 min, which are not followed by long-

term storage at room or refrigerated temperatures. The contents of Ba, Co, and Cu in PHA TV-d-ne; Mn in 347 PHA TV-d-ne, PHA TV-d-CHCl<sub>3</sub>, and PHA TV-d-NaClO; and Pb in PHA TV-d-NaClO exceeded the 348 349 migration limits of Commission Regulation (EU) No 10/2011 for both tests. The contents of Ba in PHA TVd-CHCl<sub>3</sub> and Fe in PHA TV-d-ne exceeded the limits for test 1), and those of Ba in PHA TV-d-NaClO and 350 Sb in all PHA samples, excluding PHA TV-d-CHCl<sub>3</sub> and PHA comm, exceeded the limits for test 2). The 351 352 concentrations of all the selected elements in PHA comm were below the migration limits, thus, these PHA 353 samples are food (water)-contact safe under frozen and refrigerated conditions, at 70 °C for 2 h, or at 100 °C 354 for up to 15 min.

The lower levels of elements observed in polymeric materials than Sb explain the lack of studies conducted on elemental migration into bottled water (Bach et al., 2012). The specific migration limit for Sb is 0.04 mg kg<sup>-1</sup> (European Commission, 2011). Consistent with our results, temperature significantly affects the release of Sb (Bach et al., 2012; Cheng et al., 2010; Keresztes et al., 2009; Westerhoff et al., 2008). There were no significant traces of inorganic elements other than Sb in PET-bottled water subjected to different conditions (Cheng et al., 2010; Reimann et al., 2010), and more metals (Al, Ce, Pb, and Zr) can leach from glass into water than PET bottles (Reimann et al., 2010).

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#### **363 3.2.2 Element migratability in CH<sub>3</sub>COOH**

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365 Tables 3, S6, and S7 show the results of testing PHA samples with CH<sub>3</sub>COOH, i.e., food simulant B listed in 366 Commission Regulation (EU) No. 10/2011 (European Commission, 2011). Food simulants must be used to 367 test migration from materials and articles not yet in contact with food. Food simulant B is used for foods with hydrophilic characteristics, can extract hydrophilic substances, and have a pH below 4.5 (beverages, 368 369 such as clear fruit or vegetable juices of normal strength or concentrated, fruit nectars, lemonades; and milk 370 products, such as fermented milk including yoghurt, buttermilk and similar products, and processed and preserved cheese) The data show that the concentrations of elements such as Ba, Co, Cu, Fe, Mn, and Zn 371 increased under acidic conditions (Tables 3, S6, and S7). The maximum levels of the analysed elements in 372  $CH_3COOH$  increased by more than double the concentration obtained in  $H_2O$  (Table 3). The levels of 373 elements released from PHA L-a-ap and PHA comm were below the migration limits; thus, these materials 374

are safe for contact with food under frozen and refrigerated conditions, while PHA comm is safe for contact
with food at 70 °C for 2 h, or at 100 °C for up to 15 min.

In contrast with our results, for all analysed elements (Ag, Al, As, Ba, Cd, Co, Cr, Cu, Mn, Mo, Ni, Pb, Se, Tl, and V), excluding Sb, in all PET bottle samples, Cheng et al. (2010) detected no significant leaching due to low pH, and the levels of metals were well below the specific migration limit listed in Commission Regulation (EU) No 10/2011 (European Commission, 2011). Keresztes et al. (2009) also demonstrated that the Sb dissolution rate into water is higher into sparkling water than that into still water due to the lower pH of carbonated water.

383

# 384 **3.2.3 Element migratability in HCl**

385

The original European Council Directive 88/378/EEC (Council of the European Union, 1988) on toy safety 386 387 stipulated migratable limits for some hazardous elements based on the ingestion of a small amount of material and defined by 2 h of extraction under simulated gastric conditions (dilute HCl at 37 °C) according 388 389 to European standard EN 71-3, as listed in Table 2 (British Standards Institution, 1994; Turner, 2018). An 390 amended directive (European Parliament, Council of the European Union, 2009) provided revised limits on migration in dilute HCl that depended on the matrix being tested (dry, brittle, powder-like, or pliable toy 391 392 materials; liquid or sticky toy materials; and scraped-off toy materials), as well as limits for additional 393 elements and different oxidation states of Cr (Table 2). Furthermore, there is currently a proposal to reduce 394 the concentration limits of Pb (Council of the European Union, 2017). The results of this study (Tables 3 and 395 S8) reveal low concentrations of the elements listed by the original Toy Safety Directive (European Council 396 Directive 88/378/EEC). Instead, the extracted concentrations of Cd in PHA TV-d-NaClO and Pb in all PHA 397 samples excluding PHA L-a-ap and PHA comm exceeded the current or proposed European migratable 398 limits.

The contents of Cd and Pb in second-hand plastic toys also exceeded their migration limits stipulated by the current EU Toy Safety Directive, with the release of Cd from yellow and red Lego bricks exceeding the limit by one order of magnitude (Turner et al., 2018). Cd and Pb were used as stabilisers in polyvinyl chloride (PVC) or, in association with other elements (Cr, Mo, and S) as coloured pigments (Turner et al., 2018).

403

# 404 3.2.4 Bioaccessibilities of elements

405 Exposure to elemental concentrations and their health hazards are related to their migratable levels rather 406 than their total contents. The percentage migration relative to the total elemental content can be a measure of the bioaccessibility. The results shown in Figures 1 and S1-S4 indicate wide variations in the bioaccessibility 407 408 of the toxic elements listed in Table 2. Sb and Se were not considered because all data were <LOQs. The 409 standard deviation of the mean bioaccessibility percentage values in each PHA was very high as the 410 elemental composition of each PHA sample collected at different operation times from the pilot plant (batch) was highly variable. Therefore, we reported all the results for each batch of the different types of PHA. 411 Generally, the migration of elements is higher in acidic solutions and with heating (Figures 1 and S1-S4). For 412 all considered conditions, the bioaccessibility percentages of Al, Cr, and Fe were lower than 60% for all 413 types of PHA, and that of Sn was below 40% (Figure S1). As, Cd, and Hg appeared to be more bioaccessible 414 in PHA TV-d-ne; PHA TV-d-ne and PHA TV-NaClO; and PHA TV-NaClO and PHA TV-a-ap (Figure 1), 415 respectively. 416

417

# 418 **4.** Conclusions

419

In the context of the European Circular Economy strategy, an analytical method for determining the presence and migration of 41 elements, including alkaline metals, alkaline earth metals, heavy metals and metalloids, in PHA biopolymers was optimised. The obtained LODs were suitable for the identifying element concentrations well below the limits set for plastic materials by European regulations.

PHA samples produced from different origins and with different process steps were analysed. The elemental content of PHA was generally low, ranging from ppb to a few ppm, except for alkaline and alkaline earth metals (which are of little environmental concern). The feedstock type affected the contaminant levels, with the PHA from fruit waste having a lower element content than PHA obtained from the mixture of the organic fraction of municipal waste and sludge from wastewater treatment. The crop-derived commercial PHA was found to have lower concentrations of heavy metals than the waste-based PHA samples. The PHA stabilisation and extraction process also affected the contents of contaminants, where acid stabilisation and extraction with aqueous extractants resulted in lower heavy metal contents than thermal stabilisation andextraction with either hypochlorite or chloroform.

433 Although there is currently no specific regulation, the PHA samples were compared to the regulations and guidelines for similar materials and/or applications. The total content of the analysed elements for all tested 434 PHA types complied with the current regulations and guidelines, such as the limits for Cd in plastic materials 435 based on REACH regulation, including toys. According to Commission Regulation (EU) No 10/2011, PHA 436 437 TV-a-ap and PHA L-a-ap are food (water)-contact safe under frozen and refrigerated conditions and PHA comm in all tested conditions; PHA L-a-ap and PHA are safe for contact with food under frozen and 438 439 refrigerated conditions, and PHA comm is safe for contact with food at 70 °C for 2 h, or at 100 °C for up to 15 min. The levels of elements released from PHA L-a-ap and PHA comm were below the migration limits 440 441 as stipulated by the current and proposed EU Toy Safety Directive.

442

# 443 Declaration of competing interest

444

445 The authors declare that they have no known competing financial interests or personal relationships that 446 could have appeared to influence the work reported in this paper.

447

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449

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# **Table 1**

622 Number of valid data (N), descriptive statistics (mg kg<sup>-1</sup>) and Kruskal-Wallis test for each analysed element in the different PHA samples.

Types of feedstock		Municipal waste/sludge											Fruit waste			Crops			
Element		PHA	A TV-d-ne	a	PHA TV-d-CHCl <sub>3</sub> <sup>b</sup>		PHA TV-d-NaClO <sup>c</sup>		PHA TV-a-ap <sup>d</sup>		PHA L-a-ap <sup>e</sup>			PHA comm <sup>f</sup>					
	N	Median	Mean	SD	Median	Mean	SD	Median	Mean	SD	Median	Mean	SD	Median	Mean	SD	Median	Mean	SD
Al	23	548 <sup>f,c</sup>	1090	1500	52	49	37	<b>9.6</b> <sup>a</sup>	9.0	5.9	51 <sup>f</sup>	52	33	6.11	6.11	0.92	2.76 <sup>a,d</sup>	2.76	0.62
As	20	0.79 <sup>d,e,f</sup>	0.73	0.13	0.145	0.152	0.096	0.206 <sup>d</sup>	0.213	0.091	0.029 <sup>a,c</sup>	0.035	0.018	$0.072^{\mathrm{a}}$	0.072	0.070	0.028 <sup>a</sup>	0.036	0.019
В	19	4.2 <sup>f</sup>	4.5	1.1	5.0 <sup>f</sup>	10	11	<b>4.1</b> <sup>f</sup>	5.5	2.6	2.1	3.2	3.0	1.93	1.93	0.19	<1 <sup>a,b,c</sup>	<1	-
Ba	21	28 <sup>e,f</sup>	23	15	2.2	2.6	1.3	28 <sup>d,e,f</sup>	38	28	1.92 <sup>c</sup>	1.89	0.59	1.31 <sup>a,c</sup>	1.31	0.30	<1 <sup>a,c</sup>	<1	-
Be	18	0.031 <sup>b,f</sup>	0.027	0.018	<0.0001 <sup>a,c</sup>	< 0.0001	-	0.053 <sup>b,e,f</sup>	0.056	0.025	0.0032	0.0043	0.0032	0.000657 <sup>c</sup>	0.000657	0.000071	<0.0001 <sup>a,c</sup>	< 0.0001	-
Bi	12	0.16	< 0.02	-	< 0.02	< 0.02	-	0.52	0.41	0.27	< 0.02	< 0.02	-	< 0.02	< 0.02	-	< 0.02	< 0.02	-
Ca	24	20400 <sup>d,f</sup>	19200	7600	827 <sup>c</sup>	848	220	28900 <sup>c,d,e,f</sup>	28000	2500	393 <sup>a,c</sup>	719	850	<b>467</b> <sup>c</sup>	467	320	422 <sup>a,c</sup>	420	8.7
Cd	17	0.101 <sup>d,e,f</sup>	0.096	0.070	0.015 <sup>c</sup>	0.019	0.015	0.36 <sup>b,d,e,f</sup>	0.37	0.22	<0.005 <sup>a,c</sup>	0.0088	0.0085	<0.005 <sup>a,c</sup>	< 0.005	-	<0.005 <sup>a,c</sup>	0.006	0.003
Ce	21	0.66 <sup>f</sup>	0.57	0.42	0.062 <sup>c</sup>	0.073	0.042	1.29 <sup>b,d,e,f</sup>	1.10	0.80	0.052 <sup>c</sup>	0.067	0.041	0.046 <sup>c</sup>	0.046	0.032	<0.006 <sup>a,c</sup>	< 0.006	-
Со	21	0.46 <sup>b,d,f</sup>	0.46	0.13	0.03 <sup>a,c</sup>	< 0.01	-	0.36 <sup>b,d,f</sup>	0.53	0.35	0.051 <sup>a,c</sup>	0.062	0.046	0.052	0.052	0.020	<0.01 <sup>a,c</sup>	< 0.01	-
Cr	24	3.2 <sup>b</sup>	2.8	1.1	0.53 <sup>a,c</sup>	0.55	0.16	2.64 <sup>b</sup>	2.66	0.70	2.6	4.2	4.2	0.95	0.95	0.49	1.3	1.2	1.0
Cs	22	$0.081^{b,f}$	0.071	0.051	0.0013 <sup>a,c</sup>	0.0023	0.0021	0.076 <sup>b,f</sup>	0.085	0.064	0.0059	0.0065	0.0042	0.0036	0.0036	0.0016	<0.0004 <sup>a,c</sup>	< 0.0004	-
Cu	24	50 <sup>b,f</sup>	74	43	3.3 <sup>a,c</sup>	3.8	1.5	77 <sup>b,f</sup>	88	23	18	22	18	14	14	13	0.211 <sup>a,c</sup>	0.203	0.057
Fe	24	779 <sup>e,f</sup>	805	590	43.5 <sup>c</sup>	45.0	5.1	1430 <sup>b,d,e,f</sup>	1660	1200	82 <sup>c</sup>	77	35	<b>19.4</b> <sup>a</sup>	19.4	4.3	22 <sup>a,c</sup>	19	17
Ga	21	0.20 <sup>b,f</sup>	0.29	0.28	0.011 <sup>a,c</sup>	0.014	0.012	0.35 <sup>b,f</sup>	0.38	0.21	0.087	0.086	0.031	0.0490	0.0490	0.0092	<0.001 <sup>a,c</sup>	< 0.001	-
Hg	24	0.22 <sup>d,e,f</sup>	0.18	0.12	0.023	0.025	0.017	0.066 <sup>e,f</sup>	0.071	0.020	0.012 <sup>a</sup>	0.023	0.027	0.0062 <sup>a,c</sup>	0.0062	0.0053	0.0034 <sup>a,c</sup>	0.0033	0.0020
К	23	1810 <sup>b,c,d,e,f</sup>	2280	1700	59 <sup>a</sup>	63	22	150 <sup>a</sup>	129	56	41 <sup>a</sup>	42	24	29.3 <sup>a</sup>	29.3	9.6	<b>18.7</b> <sup>a</sup>	21.0	5.2
La	21	0.39 <sup>f</sup>	0.34	0.24	0.034 <sup>c</sup>	0.041	0.025	0.80 <sup>b,d,e,f</sup>	0.66	0.47	0.027 <sup>c</sup>	0.032	0.017	0.017 <sup>c</sup>	0.017	0.015	<0.004 <sup>a,c</sup>	< 0.004	-
Li	24	0.59 <sup>b,f</sup>	0.57	0.32	0.016 <sup>a,c</sup>	0.026	0.026	0.75 <sup>b,f</sup>	0.78	0.56	0.061	0.063	0.045	0.0230	0.0230	0.0045	0.0042 <sup>a,c</sup>	0.0078	0.0069
Mg	24	3490 <sup>d,e,f</sup>	4360	3300	514	548	350	3570 <sup>d,e,f</sup>	6470	5600	169 <sup>a,c</sup>	220	160	141 <sup>a,c</sup>	141	23	19 <sup>a,c</sup>	24	10
Mn	22	83 <sup>e,f</sup>	75	13	5.8 <sup>c</sup>	5.3	3.1	166 <sup>b,d,e,f</sup>	171	14	<b>1.26</b> <sup>c</sup>	1.38	0.42	0.18 <sup>a,c</sup>	0.18	0.15	0.24 <sup>a,c</sup>	0.32	0.30
Мо	24	0.56 <sup>b,f</sup>	0.60	0.19	0.0173 <sup>a,c,d</sup>	0.0196	0.0058	0.058 <sup>b</sup>	0.062	0.013	0.074 <sup>b</sup>	0.104	0.080	0.044	0.044	0.016	0.0262 <sup>a</sup>	0.0247	0.0068
Na	24	49500 <sup>b,d,f</sup>	51100	36000	205 <sup>a</sup>	202	80	753	1140	870	299 <sup>a</sup>	878	1200	1290	1290	190	65 <sup>a</sup>	231	290
Nb	16	0.056	0.052	0.038	0.00512	0.00538	0.00065	0.058	0.069	0.033	0.0105	0.0103	0.0014	0.00530	0.00530	0.00001	< 0.005	< 0.005	-

Ni	23	9.0 <sup>e,f</sup>	8.7	1.3	2.23 <sup>c</sup>	2.24	0.62	11.3 <sup>b,d,e,f</sup>	13.5	5.9	1.7 <sup>c</sup>	2.3	1.8	0.55 <sup>a,c</sup>	0.55	0.22	0.093 <sup>a,c</sup>	0.072	0.043
Р	24	4057 <sup>b,d,e,f</sup>	4300	790	341 <sup>a</sup>	330	160	3190 <sup>d,f</sup>	3020	1200	64 <sup>a,c</sup>	128	160	<b>108</b> <sup>a</sup>	108	13	24 <sup>a,c</sup>	29	15
Pb	21	5.9 <sup>f</sup>	7.4	4.2	0.73 <sup>c</sup>	0.87	0.58	26 <sup>b,e,f</sup>	59	89	2.4	3.8	4.1	0.260 <sup>c</sup>	0.260	0.084	<0.1 <sup>a,c</sup>	< 0.1	-
Rb	24	1.3 <sup>b,f</sup>	1.8	1.1	0.032 <sup>a,c</sup>	0.035	0.010	0.81 <sup>b,f</sup>	0.91	0.65	0.107	0.107	0.067	0.043	0.043	0.017	0.0112 <sup>a,c</sup>	0.0116	0.0041
Sb	22	$0.088^{\mathrm{b,f}}$	0.106	0.058	0.019 <sup>a,d</sup>	0.021	0.013	0.049 <sup>f</sup>	0.066	0.033	0.133 <sup>b,f</sup>	0.131	0.085	0.034	0.034	0.021	<0.002 <sup>a,c,d</sup>	< 0.002	-
Se	1	< 0.2	< 0.2	-	< 0.2	< 0.2	-	< 0.2	< 0.2	-	< 0.2	<0.2	-	< 0.2	< 0.2	-	<0.2	< 0.2	-
Si	24	855 <sup>b,f</sup>	790	320	181 <sup>a,c</sup>	188	63	686 <sup>b,f</sup>	620	220	329	332	52	564 <sup>f</sup>	564	170	125 <sup>a,c,e</sup>	113	23
Sn	24	1.06 <sup>f</sup>	1.01	0.79	0.101 <sup>c</sup>	0.104	0.026	<b>1.8</b> <sup>b,e,f</sup>	2.0	1.4	0.14	0.50	0.79	0.070 <sup>c</sup>	0.070	0.037	0.011 <sup>a,c</sup>	0.036	0.044
Sr	24	110 <sup>d,e,f</sup>	114	42	4.25	4.31	0.73	112 <sup>d,e,f</sup>	112	16	3.5 <sup>a,c</sup>	6.5	6.6	2.5 <sup>a,c</sup>	2.5	1.1	0.485 <sup>a,c</sup>	0.475	0.023
Те	2	< 0.005	< 0.005	-	< 0.005	< 0.005	-	< 0.005	< 0.005	-	< 0.005	< 0.005	-	< 0.005	< 0.005	-	< 0.005	< 0.005	-
Ti	24	10.3 <sup>b,e,f</sup>	11.5	6.8	0.88 <sup>a,c</sup>	0.83	0.36	17 <sup>b,e,f</sup>	21	13	2.18	2.17	0.61	<b>0.48</b> <sup>a,c</sup>	0.48	0.27	0.078 <sup>a,c</sup>	0.098	0.043
Tl	15	0.012	0.015	0.010	0.00165	0.00164	0.00085	0.023	0.029	0.019	< 0.0003	< 0.0003	-	< 0.0003	< 0.0003	-	< 0.0003	< 0.0003	-
U	19	0.130 <sup>b,d,e,f</sup>	0.131	0.077	0.0106 <sup>a</sup>	0.0110	0.0030	0.058 <sup>e,f</sup>	0.069	0.035	$0.0047^{a}$	0.0086	0.0092	<0.001 <sup>a,c</sup>	< 0.001	-	<0.001 <sup>a,c</sup>	< 0.001	-
V	14	1.00	0.86	0.35	< 0.03	< 0.03	-	0.83	0.75	0.45	0.062	0.072	0.051	< 0.03	< 0.03	-	< 0.03	< 0.03	-
W	11	0.078	0.066	0.021	< 0.02	< 0.02	-	< 0.02	< 0.02	-	0.09	0.11	0.10	0.0251	0.0251	0.0088	< 0.02	< 0.02	-
Zn	24	178 <sup>e,f</sup>	330	270	26 <sup>c</sup>	46	47	766 <sup>b,d,e,f</sup>	1070	870	39 <sup>c</sup>	48	41	12.0 <sup>a,c</sup>	12.0	2.9	2.3 <sup>a,c</sup>	3.6	2.4
Zr	14	0.76	0.61	0.44	0.108	0.123	0.048	1.14	1.36	0.68	0.18	0.20	0.16	0.24	0.24	0.25	<0.1	< 0.1	-

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 $625 \qquad a,b,c,d,e,f \text{ Different superscript letters within rows are significantly different (p < 0.05). a = PHA TV-d-ne; b = PHA TV-d-CHCl3; c = PHA TV-d-NaClO; d = PHA TV-a-ap; e = PHA L-ap; e = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-a-ap; e = PHA L-ap; e = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA TV-d-ne; b = PHA TV-d-NaClO; d = PHA TV-d-ne; b = PHA T$ 

626 a-ap; f = PHA comm.

# 627 **Table 2**

628 Migratable limits (mg kg<sup>-1</sup>) of elements in food and toys and as defined by the original and amended EC toy 629 safety directives.

630

			2009/48/EC <sup>c</sup>								
Element	CR (EU) No 10/2011ª	88/378/EEC <sup>b</sup>	in dry, brittle, powder-like or	in liquid or							
			pliable toy material	sticky toy material	in scraped-off toy material						
Al	-	-	5625	1406	70000						
As	$1^{e}$	25	3.8	0.9	47						
В	-		1200	300	15000						
Ba	1	1000	4500 (1500)	1125 (375)	56000 (18750)						
Be	-	-	-	-	-						
Bi	-	-	-	-	-						
Ca	-	-	-	-	-						
Cd		75	1.9 (1.3)	0.5 (0.3)	23 (17)						
Ce	-	-	-	-	-						
Со	0.05	-	10.5	2.6	130						
Cr	1 <sup>e</sup>	60	Cr(III) 37.5; Cr(VI) 0.02	Cr(III) 9.4; Cr(VI) 0.005	Cr(III) 460; Cr(VI) 0.2						
Cs	-	-	-	-	-						
Cu	5 <sup>e</sup>	-	622.5	156	7700						
Fe	48	-	-	-	-						
Ga	-	-	-	-	-						
Hg	-	-	7.5	1.9	94						
K	-	-	-	-	-						
La	-	-	-	-	-						
Li	0.6	-	-	-	-						
Mg	-	-	-	-	-						
Mn	0.6	-	1200	300	15000						
Мо	-	-	-	-	-						
Na	-	-	-	-	-						
Nb	-	-	-	-	-						
Ni	-	-	75	18.8	930						
Р	-	-	-	-	-						
Pb	$2^{e}$	-	13.5 (2.0)	3.4 (0.5)	160 (23)						
Rb	-	-	-	-	-						
Sb	0.04	60	45	11.3	560						
Se	-	-	37.5	9.4	460						
Si	-	-	-	-	-						
Sn	-	-	15000	3750	180000						
Sr	-	-	4500	1125	56000						
Te	-	-	-	-	-						
Ti	-	-	-	-	-						
Tl	-	-	-	-	-						
U	-	-	-	-	-						
V	-	-	-	-	-						
W	-	-	-	-	-						
Zn	25 (100 <sup>e</sup> )	-	3750	938	46000						
7r		_	_		_						

<sup>a</sup> Commission Regulation (CR) (EU) No 10/2011 (European Commission, 2011) on plastic materials and articles to come into
 contact with food establishes a set of specific requirements for PHA (which has been obtained through microbial fermentation by
 using Alcaligenes Eutrophus.

<sup>b</sup> The original European Council Directive 88/378/EEC (Council of the European Union, 1988) on toy safety stipulated migratable
 limits for eight hazardous elements.

636 <sup>c</sup> The amended Directive on toy safety (European Parliament, Council of the European Union, 2009) with revised values shown in

parentheses. There is currently a proposal to reduce the concentration limits of Pb (Council of the European Union, 2017).

#### Table 3

Summary of the migratable concentrations<sup>a</sup> ranges [minimum and maximum mean; mg kg<sup>-1</sup>] of each element detected in the extracts of different PHA samples. 

	Н	20	H <sub>2</sub>	0	CH <sub>3</sub> CO	ЮН	CH <sub>3</sub> C	СООН	HCl			
Element	(10 d;	20 °C)	(2 h; 7	(0 °C)	(10 d; 2	0 °C)	(2 h;	70 °C)	(2 h; 37 °C)			
	min	max	min	max	min	max	min	max	min	max		
Al	< 0.2	3.8	0.321	3.4	< 0.04	7.4	0.88	9	< 0.1	9		
As	< 0.1	0.16	< 0.02	0.107	< 0.05	0.210	< 0.06	0.46	< 0.2	0.87		
В	< 0.2	3.9	0.56	3.5	0.211	3.2	0.160	3.8	0.168	3.7		
Ba	0.39	1.01	0.401	1.4	< 0.1	18.8	0.364	18.7	0.285	25.6		
Be	< 0.0003	0.00044	< 0.0002	0.0024	< 0.0002	0.021	< 0.0002	0.0217	< 0.0003	0.037		
Bi	< 0.002	0.0058	< 0.001	0.00733	< 0.0007	0.17	0.00074	0.064	< 0.002	0.19		
Ca	28	6190	100	7510	69	15200	65	15400	244	21300		
Cd	< 0.01	0.08	< 0.003	0.09	< 0.01	0.27	0.00228	0.28	< 0.005	0.31		
Ce	< 0.001	0.0024	< 0.001	0.012	0.00061	0.22	< 0.001	0.2180	0.0014	0.42		
Со	0.0020	0.201	0.00236	0.244	0.0033	0.468	0.0058	0.48	0.0055	0.43		
Cr	< 0.02	0.15	0.0369	0.60	< 0.02	0.56	0.058	1.09	< 0.3	4.0		
Cs	< 0.001	0.0126	< 0.0002	0.0153	< 0.0006	0.0107	< 0.001	0.0230	< 0.0002	0.017		
Cu	0.26	21	0.28	35	0.26	56	0.22	65	0.24	67		
Fe	0.21	54	0.25	3.9	<0.3	156	1.29	171	0.7	520		
Ga	0.0081	0.027	0.00983	0.08	0.0107	0.47	< 0.01	0.34	< 0.01	0.43		
Hg	< 0.001	0.0028	0.0042	0.0118	0.00149	0.0239	0.0041	0.0272	0.0042	0.0396		
К	5.5	909	4.0	1245	7	896	12.4	1650	11.5	1400		
La	< 0.0002	0.0027	< 0.001	0.006	0.00077	0.17	0.0014	0.14	0.0012	0.35		
Li	< 0.006	0.104	0.0069	0.149	0.0034	0.118	0.00184	0.166	< 0.01	0.115		
Mg	14	2210	23	2860	21	4040	18	4110	26	6300		
Mn	< 0.1	21.7	< 0.1	17	< 0.1	102	0.16	103	0.219	168		
Mo	< 0.01	0.146	< 0.02	0.32	< 0.003	0.027	0.0058	0.076	0.0095	0.0581		
Na	100	22700	195	37800	93	26800	156	35000	105	22800		
Nb	< 0.0002	0.00113	< 0.0003	0.00059	0.000145	0.0064	< 0.0004	0.00606	< 0.0003	0.0074		
Ni	0.10	3.2	0.11	3.8	< 0.1	11.5	< 0.1	11.3	< 0.1	12.5		
Р	<2	1280	6.0	778	10.9	3020	8.7	1700	5.0	1840		
Pb	0.0180	19	< 0.01	18	0.036	12.5	0.064	11.4	0.040	35		
Rb	0.0053	0.62	0.0020	0.83	0.0073	0.59	0.012	1.06	0.0071	0.87		
Sb	< 0.04	< 0.04	< 0.04	0.177	< 0.04	< 0.04	< 0.04	0.062	< 0.04	0.083		
Se	< 0.2	0.308	< 0.1	0.35	< 0.04	0.127	< 0.08	0.18	< 0.4	0.55		
Si	5.0	48	<6	97	<30	102	26.7	112	<6	440		
Sn	0.0012	0.012	0.004	0.016	0.00105	0.12	0.00346	0.110	< 0.001	0.26		
Sr	0.17	31	0.33	25	0.19	62	0.17	62	0.30	72.0		
Te	< 0.01	< 0.01	< 0.003	0.0086	< 0.01	0.0158	< 0.01	< 0.01	< 0.03	0.038		
Ti	< 0.1	13	0.0517	13	< 0.03	8.3	0.0353	13.6	<1	28		
Tl	< 0.0003	0.0032	< 0.0002	0.0080	< 0.0003	0.0156	< 0.0006	0.0135	< 0.004	0.010		
U	< 0.0002	0.0011	< 0.0003	0.0045	< 0.0003	0.019	< 0.0003	0.039	< 0.0003	0.038		
V	< 0.03	0.138	< 0.04	0.55	< 0.02	0.25	< 0.04	0.62	<1	<1		
W	< 0.001	0.012	0.0056	0.08	< 0.001	0.011	< 0.002	0.017	< 0.001	0.0103		
Zn	0.43	19.5	0.471	11.6	1.2	607	1.01	620	1.4	715		
Zr	0.00040	0.018	< 0.001	0.019	0.00119	0.09	0.00040	0.0901	0.0014	0.13		

<sup>a</sup> Note that numbers in bold denote non-compliance according to the Commission Regulation (EU) No 10/2011 (European Commission, 2011) for migration test with H<sub>2</sub>O and CH<sub>3</sub>COOH; and the European Union Toy Safety Directive (Council of the European Union, 1988; European Parliament, Council of the European Union, 2009; Council of the European Union, 2017) for migration test with HCl. Contact time in days (d) or hours (h) at contact temperature in °C.



Figure 1 Percentage of bioaccessibility of As, Cd, Hg and Pb in the extracts of different PHA samples taken
at different operation times of the pilot plant (batch). The data not shown are lower than the quantification
limits.