

DOTTORATO DI RICERCA IN BIOCHIMICA CICLO XXXII (A.A. 2016-2019)

"Endocrine disrupting effects of the pollutant β -hexachlorocyclohexane in cancer cells: possible chemopreventing role of the natural sesquiterpene β -caryophyllene"

Tutor
Prof. Margherita Eufemi

Ph.D. Coordinator Prof. Stefano Gianni

Ph.D. Student **Stefania Carissimi**

"Non temete i momenti difficili. Il meglio viene da lì"

Rita Levi Montalcini

CHAPTER 1: INTRODUCTION	7
1.1 Endocrin Disruptors compounds	7
1.2 Organochlorine pollutants	12
1.2.1 OCPs as endocrine and metabolic disruptors	16
1.2.2 OCPs and oncogenic modulation	18
1.3 β-Hexachlorocyclohexane	19
1.4 STAT3 protein	20
1.4.1 STAT3 Activation and Regulation	22
1.5 STAT3 Post-Translational Modifications (PTMs)	24
1.5.1 Phosphorylation	24
1.5.2 Acetylation	25
1.5.3 Methylation	25
1.5.4 Oxidation and Glutathionylation	26
1.6 STAT3 in tumorigenesis	27
1.6.1 STAT3 Inhibitors	28
1.7 The sesquiterpene β-caryophyllene	29
1.7.1 β-CRY as anticancer agent	30
CHAPTER 2: AIM OF THE RESEARCH	33
CHAPTER 3: MATERIALS AND METHODS	37
3.1 Reagents	37
3.2 Cell cultures	37
3.3 Cell viability assay	38
3.4 Cell lysate preparation	38
3.5 Western blotting	39
3.6 Total RNA Extraction	40
3.7 cDNA synthesis and Quantitative Real Time-PCR	40
3.8 Cell Cycle Assay	41

3.9 Scratch test Assay	41
3.10 Cell Proliferation Assay	42
3.11 Statistical analyses	42
4. RESULTS	43
4.1 β-Hexachlorocyclohexane triggers STAT3 pathways	43
4.2 Investigation of β-caryophyllene protective role against β-	
hexachlorocyclohexane carcinogenic property	48
4.2.1 β-caryophyllene inhibition of STAT 3 activation	48
4.2.2 β-caryophyllene inhibition of proliferation and migration	49
4.3 β-hexachlorocyclohexane effect on placenta	55
CHAPTER 5: DISCUSSION	59
REFERENCES	61
CHAPTER 6: APPENDIX	71

CHAPTER 1: INTRODUCTION

1.1 Endocrin Disruptors compounds

The U.S. Environmental Protection Agency (EPA) defines an endocrine-disrupting compound (EDC) as "an exogenous agent that interferes with synthesis, secretion, transport, metabolism, binding action, or elimination of natural blood-borne hormones that are present in the body and are responsible for homeostasis, reproduction, and developmental process." From a physiological perspective, an endocrine-disrupting substance is a compound, either natural or synthetic, which, through environmental or inappropriate developmental exposures, alters the homeostasis of hormonal system that enable the organism to communicate with and respond to its environment (**Figure 1**).

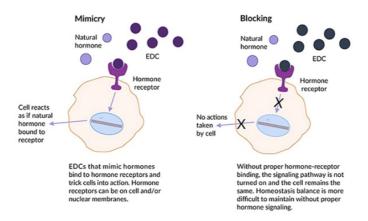


Fig. 1: Schematic representation of EDCs interference with physiological hormones pathways.

Today, there are nearly 1000 chemicals reported to have endocrine effects ("Home," n.d.). The prevalence of EDCs in our environment and their persistence in our bodies represents a significant health challenge since the endocrine system plays a central role in all vertebrates and regulates such critical biological functions as metabolism, development, reproduction, and behaviour.

Epidemiological studies link EDCs with alterations of the reproductive system, such as malformations, infertility and increased risk of endometriosis, neurobehavioral and neurodevelopmental changes, metabolic syndrome, bone disorders, immune disorders, and cancers in humans ("WHO | State of the science of endocrine disrupting chemicals - 2012," n.d.).

The group of molecules identified as endocrine disruptors is highly heterogeneous and includes synthetic chemicals used as industrial solvents and their by-products (polychlorinated biphenyls, polybrominated biphenyls, dioxins), plastics (bisphenol A), plasticizers (phthalates), pesticides (methoxychlor, chlorpyrifos, dichlorodiphenyltrichloroethane, hexachlorocyclohexane), fungicides (vinclozolin), and pharmaceutical agents (diethylstilbestrol).

Listed below, the main EDCs, their mechanism of action, hormones affected (**Table 1**) and effects on the physiological endocrine activity (**Table 2**).

Pesticide	Uses	Hormones affected	Mechanism	References
Organochlorines	Insecticides	Androgens, oestrogens, prolactin	Competitive inhibitor of androgen receptors, inhibits oestrogen-sensitive reporter binding to androgen receptors. Some induce the production of aromatase, an enzyme that converts androgen to oestrogen	Daxenberger (2002), Lemaire et al. (2004), Scippo (2004), Sonnenschein and Soto (1998), Storrs and Kiesecker (2004)
Organophosphates	Mostly insecticides, some herbicides	Oestrogens, thyroid hormones	Prevents thyroid hormone-receptor binding. Increases the expression of oestrogen responsive genes.	Gwinn et al. (2005), Jeong et al. (2006), Kang et al. (2004)
Carbamates	Herbicides and fungicides	Androgens, oestrogens, steroids.	Still largely unknown. Thought to affect androgen- and androgen-receptor dependant mechanisms. Shown to interfere with cellular microtubule formation in oestrogen-sensitive cells.	Daxenberger (2002), Goad et al. (2004), Lu et al. (2004), Morinaga et al. (2004)
Triazines	Herbicides	Androgens	Inhibition of natural ligands that bind to androgen receptors and androgen-binding proteins. Some induce or inhibit the production of aromatase, an enzyme that converts androgen to oestrogen	Hayes et al. (2002a), Ishihara et al. (2003), Meulenberg (2002)
Pyrethroids	Insecticides	Oestrogens, progesterone	Different compounds antagonise or potentiate the action of oestrogen by acting on the oestrogen receptor or possibly an alternative signalling pathway. Some Inhibit the action of progesterone by affecting the hormone itself.	Garey and Wolff (1998), Kim et al. (2004)

Table 1: List of common pesticides with Endocrine Disruptors effects, hormones affected and their mechanism of action.

The sources of exposure to EDCs are diverse and vary widely around the world. People who work with pesticides, fungicides, and industrial chemicals are at particularly high risk for exposure and thus for developing a reproductive or endocrine abnormality (McKinlay et al., 2008). Some EDCs were banned decades ago and others more recently, with significant differences between countries (**Table 3**). These complex mixtures enter the food chain and accumulate in animals and humans.

Pesticide	Description	ED Effects	References
Carbendazim	Fungicide	Increases oestrogen production by increasing aromatase activity.	Morinaga et al. (2004)
Carbofuran	Insecticide	Acute doses increase levels of progesterone, cortisol and	Goad et al. (2004)
Chlorothalonil	Eugainida	oestradiol whilst decreasing testosterone levels.	Tassian and Mataumura (2001)
Chlordane	Fungicide Insecticide	Triggers the proliferation of androgen-sensitive cells. Antagonises the action of androgens <i>via</i> binding competitively to their receptors and inhibiting the genetic transcription they induce.	Tessier and Matsumura (2001) Lemaire et al. (2004)
Chlordecone	Insecticide	Bids to both oestrogen and androgen receptors.	Okubo et al. (2004)
Chlorfenvinphos	Insecticide	Weak oestrogen mimic.	Vinggaard et al. (1999)
Chlorpyrifos methyl	Insecticide	Antagonises androgen activity.	Kang et al. (2004)
Cyhalothrin	Insecticide, acaricide	Decreases the secretion of thyroid hormones.	Akhtar et al. (1996)
Cypermethrin	Insecticide	Mimics the action of oestrogen. Metabolites also have oestrogenic effects.	Chen et al. (2002), McCarthy et al. (2006)
Cyproconazole	Fungicide	Inhibits the enzyme aromatase, decreasing the production of oestrogens and increasing the available androgens.	Trosken et al. (2004)
DDT and metabolites	Insecticide	Mimics the action of oestrogen, antagonises the action of androgens via binding competitively to their receptors and inhibiting the genetic transcription they induce. Promotes the proliferation of androgen-sensitive cells. Mimics the actions of oestrogens indirectly by stimulating the production of their receptors.	Bulayeva and Watson (2004), Lemaire et al. (2004), Tapiero (2002) Tessier and Matsumura (2001)
Deltamethrin	Insecticide	Shows weak oestrogenic activity.	Andersen (2002)
Diazinon	Insecticide, acaricide	Mimics the action of oestrogen.	Manabea et al. (in press)
Dibutyltin dichloride	Antifouling agent	Competitively inhibits the conversion of androgens to oestrogens. Inhibits androgen synthesis.	Heidrich (2001), Thibaut and Porte (2004)
Dichlorvos	Insecticide, acaricide	Weak androgen-receptor antagonist.	Andersen (2002)
Dicofol	Acaricide	Inhibits androgen synthesis, increases the synthesis of oestrogens. Also binds to the oestrogen receptor.	Okubo et al. (2004), Thibaut and Porte (2004)
НСВ	Fungicide	Severely disrupts thyroid hormone production. Enhances androgen action at low doses, but inhibits it at high ones.	Ralph et al. (2003), Verreault et al. (2004)
HCH (lindane)	Insecticide	Shortens oestrous cycles and lowers luteal progesterone concentrations. Increases the blood serum concentrations of insulin and oestradiol, decreases thyroxine concentrations.	Beard and Rawling (1999), Rawlings et al. (1998)
Heptachlor	Non-systemic insecticide	Binds to cellular oestrogen and androgen receptors.	Fang et al. (2003), Oduma et al. (1995)
Hexaconazole	Fungicide	Inhibits the enzyme aromatase, decreasing the production of oestrogens and increasing the available androgens.	Trosken et al. (2004)
Ioxynil	Herbicide	Antagonises the action of thyroid hormones and the expression of the genes coding for their cellular receptors.	Sugiyama et al. (2005)
Iprodione	Fungicide	Weakly promotes aromatase activity, increasing oestrogen production.	Andersen et al. (2002)
Kepone	Insecticide	Mimics the action of oestrogen weakly, binds strongly to androgen and progesterone receptors.	Fang et al. (2003), Tapiero et al. (2002)
Linuron	Herbicide	Competitively inhibits the binding of androgen to its receptor, inhibits androgen-inducing gene expression. Alters androgen-dependant ventral prostate gene expression.	Fang et al. (2003)
Malathion	Insecticide, acaricide	Inhibits catecholamine secretion, binds to thyroid hormone receptors.	Cocco (2002), Ishihara et al. (2003)
Mancozeb	Fungicide	Inhibits the production of thyroid hormones.	Cocco (2002)
Maneb	Fungicide	Inhibits the production of thyroid hormones.	Cocco (2002)
Metriam	Fungicide	Inhibits the production of thyroid hormones.	Cocco (2002)
Methiocarb	Herbicide, insecticide	Inhibits androgen activity whilst promoting oestrogen activity.	Andersen et al. (2002)

 Table 2: This table reports endocrine disrupting activity of each pesticide.

EDCs	Type or source	Legal status	NRs
Organochlorines (e.g., DDT)	Pesticides and plasticizers	1970s: DDT banned in most developed countries 2000s: restricted by the Stockholm Convention	ERα, AR
Dioxins (e.g., PCB, TCDD)	Environmental pollutants in food	2000s: PCB banned and other dioxins restricted by the Stockholm Convention	Via AhR: PPAR ₇ , ERs
Organotins (e.g., TBT, TPTO)	Environmental pollutants in food	Banned worldwide by the International Maritime Organization	RXR: PPARγ
PFCs (e.g., PFOA, PFOS)	Plasticizers	2009: restricted by the Stockholm Convention and the EU	ERs, AR, PPARs
BFRs (e.g., PBDE)	Flame retardants	PBDE banned in the EU and some U.S. states 2009: some BFRs banned by the Stockholm Convention	PXR, ERs, TR
Alkylphenols (e.g., octylphenol)	Surfactants	Restricted for some uses in the EU	ERs, AR, CAR
ВРА	Plasticizers	2009: Canada becomes the first country to ban BPA in baby bottles; WHO begins assessing BPA safety	ERs, AR, TR, GR
Phthalates (e.g., DEHP, DBP, DEP)	Plasticizers	Restricted in children's toys in the E.U. (1999) and the United States (2009) 2010: Australia bans products with > 1% of DEHP	PPARs, CAR/PXR, GR

Table3: It is reported, for each EDC, the type of source, the NR involved and the legal status.

Research conducted so far allow to identify some key issues necessary to a full understanding of mechanism of action and consequences of exposure to EDCs:

1) Age of exposure – The field of endocrine disruption has embraced the terminology "the fetal basis of adult disease" (Barker, 2003), since developing organisms are extremely sensitive to perturbation and fetal development is commonly known to be a period of increased susceptibility to chemical insult. Thus, it is crucial to determine whether or what kind of consequence the exposure to EDCs will lead to, taking into account when the exposure occurs;

- 2) *Latency from Exposure* Effects of EDCs may not be immediately apparent early in life but may be manifested in adulthood or during aging;
- 3) *Importance of mixtures* Contamination of environments are rarely due to a single compounds and effects of different classes of EDCs may be additive or even synergistic (Crews et al., 2003).
- 4) Non traditional dose-response dynamics even infinitesimally low levels of exposure may cause endocrine or reproductive abnormalities, particularly if exposure occurs during a critical developmental window. Moreover, low doses may even exert more potent effects than higher doses.

 5) Transgenerational, epigenetic effects Recent evidence suggests that the mechanism of transmission may involve the germ line and may be non genomic. One of the hypothesis leans toward epigenetic inheritance patterns, which involve chemical modifications to the DNA (DNA methylation), histone modifications, and non coding RNAs, rather than mutations of the DNA sequence itself.

1.2 Organochlorine pollutants

Organochlorine pesticides (OCPs) are a structurally heterogeneous class of organic compounds composed primarily of carbon, hydrogen and several chlorine atoms per molecule. They have been employed in agriculture and in public health as insecticides and biocides for several decades but were mostly banned between 1970 and 1990. Only a few active substances are still used in tropical countries. So, most exposures derive from past uses and, in certain cases, are still significant because of the long persistence and

biomagnifications of these compounds. Once released in the environment, they break down very slowly in air, water, soil and in living organisms and are thus subject to long trans-boundary air pollution transport where they are carried over long distances via the atmospheric transport and human activities (Jones and de Voogt, 1999).

The main OCPs are:

- Dichlorodiphenyl-trichloroethane (DDT), classified as "probably carcinogenic to humans (group 2A)" was widely used in agriculture and to eradicate malaria. It is readily absorbed and distributed in the body by lymphatic and blood circulation, with a preference for lipid tissue. Associations between cancer and exposure to DDT have been investigated in more than 100 cohort and case-control studies. A few of these, conducted on Chinese population, reported dose related association between liver cancer and blood DDT level (McGlynn et al., 2006), (Persson et al., 2012);
- **Hexachlorobenzene** (HCB), at first produced as a fungicide, is still formed as a major by-product and contaminant in the manufacture of chemicals such as solvents, chlorine-containing compounds and pesticides (ATSDR 1996)It has a strong impact on the human androgen axis and it is shown to cause liver tumors in animals (Randi et al., 2003);
- Polychlorinated biphenyls (PCBs), industrial chemicals consisting of paired phenyl rings with various degrees of chlorination which are able to bind to TRs. Some authors showed that PCBs can reduce circulating levels of T4 in animals (Goldey et al., 1995), others proposed they can exert neurotoxic effects on the developing brain by causing a state of hypothyroidism (Crofton, 2004);
- **Endosulfan**, mainly used to control pests in vegetables, fruits, cereal grains and cotton, ornamental shrubs, trees, vines and ornamental plants. In vitro studies show that Endosulfan acts as antagonist of androgen receptor (Li et al., 2008) and significantly increases cell

proliferation and estrogen receptor (ER) transactivation gene response in MCF-7 cells (Andersen et al., 2002). A cohort study conducted in 2003, among male school children (10–19 years old) showed that endosulfan exposure was associated with delayed male sexual maturity and interfered with the male sex-hormone synthesis (Saiyed Habibullah et al., 2003);

Hexachlorocyclohexane, (HCH) belongs to a family of eight isomers of 1,2,3,4,5,6-hexachlorocyclohexane that differ in their equatorial substitution pattern around the ring. They are denoted by Greek letters (α, β, γ, δ, ε, η and θ) (Figure 2) (Willett et al., 1998). The physical and chemical properties of the HCH isomers are quite different from one another as illustrated in Table 4.

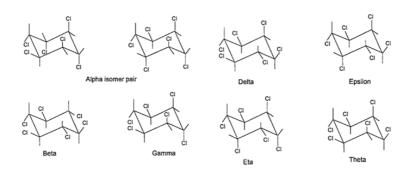


Fig.2: Structure of Hexachlorocyclohexane (HCH) isomers.

property	α	P	γ	o	€
melting point (°C) ^a	159-160	309-310	112-113	138-139	219-220
vapor pressure ^b	$(1.6 \pm 0.9) \times 10^{-2}$	$(4.2 \pm 0.3) \times 10^{-6}$	$(5.3 \pm 1.4) \times 10^{-3}$	2.1×10^{-3}	
log K _{ow} c	3.9 ± 0.2	3.9 ± 0.1	3.7 ± 0.5	4.1 ± 0.02	
BCF in human fatd	20 ± 8	527 ± 140	19 ± 9	8.5	
BCF in aquatic animals ^e	2.6 ± 0.5	2.9 ± 0.3	2.5 ± 0.4		

Table 4: Selected Physical Properties of HCH isomers.

The γ -isomer (γ -HCH), called lindane, has been widely used as a pesticide, due to its effectiveness, low cost and acute toxicity. The estimation of its global production between 1950 and 2000 was 600,000 tonnes and the majority was used in agriculture, as an insecticide on fruits and vegetables, rice paddies, Christmas trees, and animals and as a seed treatment. In medicine, γ -HCH has been applied topically to people for the treatment of lice and scabies. Although its employment has been restricted since the 1970's, because of its toxicity, the problem of residues of all isomers of HCH remains because of the high persistence and interconversion of these isomers in soil (Chessells et al., 1988), (Concha-Graña et al., 2006).

During the production process of γ -HCH others three isomers, α , β , and δ , are formed, however only the γ -HCH is known to be effective as an insecticide, despite its content ranges between 12% and 15% of the isomer mixture. Consequently, the α , β and δ -isomer mixture represents a big amount of byproducts, which are often dumped under unsuitable circumstances.

As OCPs bio-magnify through the food chain, they accumulate in the body. e.g., DDT can remain in the body for 50 years, lifelong sequestrated into the soft tissue compartments (mainly the adipose tissues), from which they partition to the bloodstream into plasma or serum lipids and are actively secreted into breast milk as the main elimination pathway in mammals (Mrema et al., 2013). Human exposure to OCPs begins during early prenatal

and continues during the breast-feeding neonatal periods, which are critical stages for the development and differentiation of sensitive body organs and systems. In fact, these chemicals cross the placenta to the fetus and are secreted into breast milk (Perera et al., 2005). In adults the dietary exposure route accounts for more than 90% of total organochlorine compounds burden for the general population, while workers are exposed mainly through inhalation and skin contact. OCPs are rapidly absorbed in the small intestine and enter circulatory system where they are distributed throughout the body and accumulate in body tissues with high lipid contents, with a continuous exchange between blood and tissues. As such these chemicals have been detected in human tissues such as blood (whole blood, cord blood, serum and plasma), adipose tissues (in autopsies and living subjects), breast milk, muscles and hair.

1.2.1 OCPs as endocrine and metabolic disruptors

There is growing concern in the scientific community that OCPs may be contributing to the rapid increased rates of diabetes and metabolic syndrome and a lot of literature links this pathological conditions to the exposure to compounds such as BPA, dioxins, organochlorine and organophosphate pesticides. The chlorine atoms are poorly reactive towards nucleophilic elimination reactions and thus their biodegradation reactions are limited to anaerobic environment of sludges. As a consequence, their interaction with biological systems is mostly limited to agonistic or antagonistic binding to the intracellular receptors for which natural hydrophobic substances, such as steroid derivatives, are the endogenous ligands. Agonistic binding leads to recruitment of coactivators and increase of transcriptional activity while antagonistic binding prevents coactivator recruitment and/or attracts corepressors, leading to decreased transcriptional activity of the receptors.

There are three main pathways through which EDCs cause metabolic disruption (Casals-Casas and Desvergne, 2011), shown in **Figure 3**:

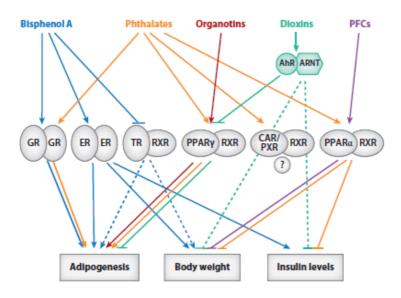


Fig. 3: Schematic representation of EDCs interactions with Nuclear Receptors, Xenosensors and PPARs and consequent activation and cross-talk between involved pathways.

- 1) Interaction with hormone receptors, such as ER (Estrogen Receptors), TR (thyroid Hormone receptors) and GR (Glucocorticoid Receptors): These receptors are important for the control of adipogenesis, weight gain, and insulin levels, although the underlying mechanisms are not yet well understood
- 2) Interaction with Xenosensors, such as PXR (pregnane X receptor), CAR (constitutive androstane receptor) and AhR (aryl hydrocarbon receptor): these receptors are mainly involved in regulating the metabolism of xenobiotics, yet their contribution to fatty acid, lipid, and glucose metabolism has been only recently appreciated

3) Interaction with PPARs (Peroxisome Proliferator–Activated Receptors): PPARγ is a member of the nuclear receptor superfamily and it is one of the major regulators of adipogenesis. It is primarily expressed in adipose tissue, and its activation promotes adipocyte differentiation as well as the induction of lipogenic enzymes. Additionally, it contributes to maintenance of metabolic homeostasis through transcriptional activation of genes implicated in energy balance.

Altogether, these many examples of EDCs interaction with receptors highlight the fact that a given compound can interfere with different NRs and different pathways. The final result of these interactions is given by cross-talk between these pathways, rather than to a linear causation chain and are much more complex to decipher in vivo.

1.2.2 OCPs and oncogenic modulation

It has been demonstrated that several OCPs exposure cause cancer in humans. Taking into account DNA toxicity OCPs fall into two classes of carcinogens: genotoxic and non-genotoxic. The latter one includes compounds which are able to affect the expression of epigenetic enzymes, in turn regulating the transcription of genes involved in pathological processes.

Hexachlorobenzene, an environmental pollutant, is known to cause liver tumours in animals through activation of c-myc, c-fos, c-jun proto-oncogenic proteins and PKC activity induction (Randi et al., 2003); the β isomer of Lindane is able to increase the mRNA expression level of metalloproteinase MMP-13 and the expression of a number of proto-oncogenes such as c-Neu, cyclin D1 and p27 (Wong and Matsumura, 2007).

Genome hypomethylation has been found in tumours (Das and Singal, 2004), whereas a recent study reported a strong correlation between increasing levels

of POPs, such as DDT, and global DNA hypomethylation (an aberrant epigenetic pattern of malignant cells) in a sample size of 70 subjects in the Greenlandic Innuit (Rusiecki et al., 2008). In a recent review it has been examined the association between work-related exposures and prostate cancer and the evidence for a possible relationship has been assessed (Krstev and Knutsson, 2019). The highest risk estimates for specific organochlorine pesticides were estimated for chlordane in a Swedish study (Hardell et al., 2006) and Lindane in the USA studies (Mills and Yang, 2003).

1.3 β-Hexachlorocyclohexane

The β -isomer (β -HCH) of Hexachlorocyclohexane is characterized by high lipid solubility, great stability and high persistence in the environment, due to its chemical structure. As it is shown in **Figure 4** this compound has all chlorine atoms in an equatorial position, and it lacks aromatic character and any axial chlorine atom, which can be the site for 1,2-elimination.

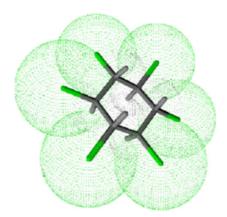


Fig.4: Structure of the β isomer of HCH.

Direct, non/genomic mechanisms of toxicity involve perturbation of the homeostatic redox levels of biological compartments and consequent cellular death through apoptosis. Indirect, genomic mechanisms of toxicity involve permanent modification of the transcription of gene elements by interference at the epigenetic level of DNA functioning.

Coosen et al. showed that 1μM β-HCH has mitogenic activity in human breast cancer cell line MCF-7 (Coosen and van Velsen, 1989) while the same phenomenon is not evident in human triple negative breast cancer cell line MDA-MB231. Moreover, β-HCH increased, in MCF-7 cells, pS2 gene mRNA level. The pS2 gene product is a polypeptide that is expressed by one-half of all breast tumors, and although its function has not been determined, its presence identifies tumors that are sensitive to anti-hormonal therapy (Steinmetz et al., 1996). In vivo assay with mouse xenograft model showed that 10-8M β-HCH significantly increased MCF-7 cell number achieving maximal responses at 10-5M (Steinmetz et al., 1996). β-HCH has been shown to induce activation of caspase-8, that plays the role in transduction of death signal (Said et al., 2004) and caspase-3, that initiates cell apoptosis (Khan et al., 2000). Moreover, Shi et al. demonstrated that β-HCH enhances reactive oxygen species (ROS) production in rat Sertoli cells and induces activation of JNKs and NFK-B and expression of FasL. Upon engagement of FasL to Fas receptor, the intrinsic program of apoptotic death is stimulated in cells leading to the activation of caspase-8. Finally, apoptosis of Sertoli cells is mediated by caspase-3, thereby disturbing the spermatogenic process (Shi et al., 2011).

1.4 STAT3 protein

Signal Transducer and Activator of Transcription 3 (STAT) is a member of the STAT family. STAT3 is a transcriptional factor that transmits signals from

cells surface into the nucleus by binding to specific DNA promoter sequences and regulating gene expression involved in many cellular processes.

STAT3 exists in two isoforms originating from alternative splicing: the full-length, called STAT3 α and the C-terminal truncated form, called STAT3 β (Figure 5) (Dewilde et al., 2008), (Ng et al., 2012). The β isoform lacks the C-terminal Transactivation domain, including the S727 residue. For this reason it has long been considered as a dominant negative regulator of STAT3 α transcriptional activity (Caldenhoven et al., 1996). However, several recent studies showed that STAT3 β plays unique functions, activating sets of specific genes. Its expression levels are lower compared to STAT3 α but when STAT3 β is over expressed can play different and opposite roles: it induces apoptosis and cell-cycle arrest, in several STAT3-dependent tumor cell lines with persistent STAT3 tyrosine (705) phosphorylation and plays an oncogenes function by triggering aggressive T cell leukemia in transplanted bone marrow cells (Ecker et al., 2009). In contrast however, STAT3 β seems to have a potent anti-inflammatory role in the initiation phases of skin and colon tumorogenesis (Marino et al., 2014).

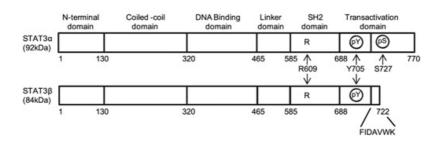


Fig 5: Schematic structure of STAT3 proteins. Comparison between $Stat3\alpha$ and $Stat3\beta$ splice variant.

Despite STAT3 transcriptional activity is mainly regulated by Y705 phosphorilation, there is an increasing consensus that unphosphorylated STAT3 (U-STAT3) is also transcriptionally active and continuously shuttling between the cytoplasm and the nucleus (Cimica et al., 2011) and it is shown to be able to interact with other factors and to directly or indirectly bind to DNA on binding sites distinct from the canonical gamma interferon activation site (GAS) sequences, thus exerting its control on a different subset of target genes, including several oncogenes (Yang et al., 2007), (Timofeeva et al., 2012), (Nishimoto et al., 2013). Ng D.C. et al showed that U-STAT3 can inhibit the microtubule-destabilizing protein stathmin, resulting in enhanced microtubules polymerization and cell migration (Ng et al., 2006).

STAT3 can be also found in the mitochondrion, phosphorylated on S727 residue. Activated Mito-STAT3 is shown to interact with Electron Transporter Chain (ETC) I and II, to preserve optimal ETC activity, increasing membrane polarization and ATP production, and enhancing the activity of lactate dehydrogenase. As a consequence, aerobic glycolysis is induced and ROS production decreased (Wegrzyn et al., 2009), (Szczepanek et al., 2011), (Tammineni et al., 2013).

1.4.1 STAT3 Activation and Regulation

In physiological conditions, the STAT3 protein is temporary activated and usually this activation lasts from a few minutes to several hours. However, numerous studies demonstrated the constitutive activation of STAT3 in a large number of human tumor cell lines. As reported in the Human Protein ATLAS, (**Figure 6**), the majority of cancer tissues display weak to moderate cytoplasmic and nuclear positivity to STAT3 protein.

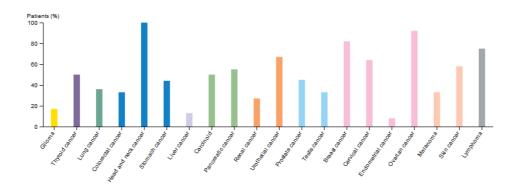


Fig. 6: Color-coded bars indicate the percentage of patients (maximum 12 patients) with high and medium STAT3 protein expression level (ATLAS).

This protein is canonically activated by Y705 phosphorilation downstream of cytokines (IL-6 family, leptin, IL-12, IL-17, and IL-10, which activate gp130/JAK/STAT3 pathway) growth factors (granulocyte colony-stimulating factor G-CSF-, epidermal growth factor EGF) and oncogenes (Src, Abl, Sis, Fps, Ros, Met and ErbB2 (Bowman et al., 2000)), (Figure 7) to mediate their functions under both physiological and pathological conditions (Yu et al., 2014), (Yuan et al., 2015). Under physiological conditions, STAT3 activation is tightly controlled by negative regulators, which mainly fall into three groups: Phosphatases (SHP-1, SHP-2, PTP1B) (Xu and Qu, 2008); Suppressor Of Cytokine Signaling (SOCS) proteins (SOCS3 act as a negative feedback regulator to inhibit JAK activity) (Krebs and Hilton, 2001); Protein Inhibitor of Activated STAT (PIAS) proteins (PIAS3, prevents the binding of STAT3 to its target DNA sequence) (C. D. Chung et al., 1997).

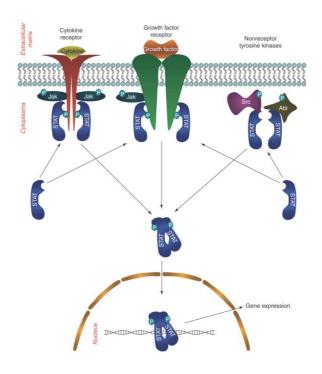


Fig 7: Main pathways responsible of STAT3 activation.

1.5 STAT3 Post-Translational Modifications (PTMs)

STAT3 can be post-translationally modified in a number of alternative ways, variably affecting its activity and cellular localization.

The main PTMs of STAT3 are Phosphorylation, Acetylation, Methylation, and Glutathionylation (**Figure 8**).

1.5.1 Phosphorylation

As already mentioned, STAT3 canonical activity mainly occurs due to Y705 phosphorylation, which enable STAT3 to form functional dimers, translocate in the nucleus and bind DNA. This is positively regulated by many activatory

signals such as cytokines (IL-6 family, leptin, IL-12, IL-17, and IL-10, as well as interferons) growth factors (granulocyte colony-stimulating factor -G-CSF-, epidermal growth factor -EGF-) and a number of oncogenes, the prototype of which are Src family kinases but also including Abl, Sis, Fps, Ros, Met and ErbB2. Phosphorylation can also occur on S727 residue and be carried out by kinases including MAPK and mTOR. While S727 phosphorylation is required for the optimal transcriptional induction of a subset of target genes (Aznar et al., 2001), it can also modify STAT3 activation dynamics by inhibiting the subsequent Y705 phosphorylation (J. Chung et al., 1997). It is now clear that Serine phosphorylation is responsible of STAT3 mitochondrial localization.

1.5.2 Acetylation

Under citokynes and grow factors stimuli STAT3 can also be acetylated on Lysine residues (K685 and K87) by the CBP/p300 histone acetyltransferase while its deacetylation is carried out by the nicotinamide adenine dinucleotide (NAD)-dependent silent information regulator protein (SIRT)1 (Nie et al., 2009). K685 acetylation leads to an enhanced STAT3 transcriptional activity and increased Y705 phosphorylation and dimer stability (Yuan et al., 2005), while K87 acetylation, promoted by insulin stimulation, can increase mitochondrial translocation and function (Xu et al., 2016).

1.5.3 Methylation

STAT3 methylation can occur in the nucleus on Lysine residues (K140 or K180). K140 is methylated by the histone methyltransferase SET9 when STAT3 is bound to its promoter, leading to impaired transcriptional activation of the target genes (Yang et al., 2010). In contrast, K180 is methylated by the lysine methyltransferase of the polycomb complex 2 (EZH2) operates. Kim et al. demonstrated that EZH2 methylation of STAT3 K180 residue is essential,

in glioblastoma cells, to enhance STAT3 activation and promote its transcriptional activity (Kim et al., 2013).

1.5.4 Oxidation and Glutathionylation

Finally, STAT3 can be both oxidized and glutathionylated on multiple cysteine residues, under conditions of oxidative stress and cytokine signalling, impairing its transcriptional activity (Xie et al., 2009), (Li et al., 2010). STAT3 oxidation can occur downstream of the cytoplasmic thiol peroxidase peroxiredoxin-2 (Prx2), one of the major H₂O₂ scavengers within the cell (Sobotta et al., 2015). This achieves ROS detoxification while at the same time impairing IL-6-induced, STAT3-mediated transcription, suggesting that STAT3 is part of a redox relay controlling redox homeostasis, and ROS and cytokine signalling. Therefore, this cross-talk between oxidative and non-oxidative STAT3 modifications can affect the activities of YP-STAT3 as well as cell proliferation and survival.

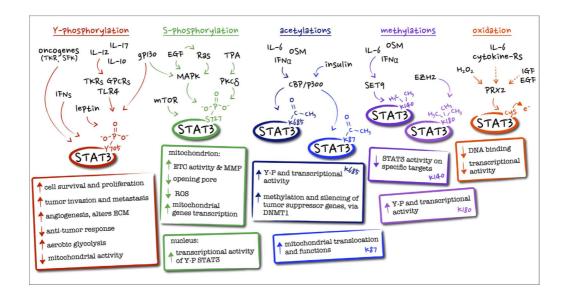


Fig 8: main STAT3 post-translational modifications, together with their known biological effects and upstream regulator.

1.6 STAT3 in tumorigenesis

STAT3 is constitutively activated in many tumours and most of its activators/regulator are deregulated in cancer. As a consequence it is mostly considered as an oncogenes (Yuan et al., 2015), (Yu et al., 2014), (Avalle et al., 2017).

Different mechanisms are correlated to aberrant STAT3 activation:

- 1) loss of the negative regulation of STAT3;
- 2) excessive stimulation of STAT3;
- 3) positive feedback loops that sustain persistent STAT3 activation;
- 4) somatic mutations that confer a hyperactive property to STAT3.

The direct effect of STAT3 activation consists in an increased expression of its target genes which are able to mediate all the crucial steps of cancer progression such as proliferation and survival of cells, their migration and invasion properties and their potential to induce angiogenesis and to evade the host immune response. In particular, STAT3 regulates:

- a) cell cycle, promotion of G1/S and G2/M transitions through cyclin D1, cyclin B and CDK1 stimulation (Ma et al., 2015);
- b) cellular survival, by controlling the anti-apoptotic genes Mcl-1, Bcl-2, Bcl-xl, Survivin (Dokduang et al., 2016) and by down-regulating the apoptotic gene p53 (Nicholas and B, 2011);
- c) tumor invasion and metastasis, by the expression of matrix metalloproteinase (MMPs);
- d) Angiogenesis, by promoting vascular endothelial growth factor (VEGF) (Zhu et al., 2011).

1.6.1 STAT3 Inhibitors

STAT3 pivotal role in cancer initiation, progression and metastasis makes it a great target for anticancer therapy. A lot of STAT3 pharmacological inhibitors are available and they mainly act at two levels: 1) indirectly blocking abnormally activated upstream proteins (Cetuximab and Panitumumab, monoclonal antibodies against EGFR; Gefitinib, Erlotinib, Lapatinib and PD153035, small molecules designed to block EGFR tyrosine kinase activity; AG490, and AZD1480, JAK inhibitors; Dasatinib and Indirubin, Src inhibitors); 2) directly suppressing the STAT3 activation (STA-21, S3I-201 and Stattic, small non peptidic molecules which are able to directly bind to STAT3 SH2 domain, preventing its dimerization; novel platinum (IV) compounds, CPA-1, CPA-7 able to cross link DNA and inhibit STAT3 DNA binding (Turkson et al., 2004)).

Besides synthetic molecules, several natural compounds display a STAT3 inhibitory activity. Among these we can find polyphenols such as Quercitin, Silibinin, and Epigallocatechin gallate; Curcumin, Caffeic acid and Celastrol, and the Sesquisterpene β-Caryophyllene.

1.7 The sesquiterpene β-caryophyllene

β-caryophyllene (CRY) is a plant compound which belongs to the bicycle sesquiterpene family. In nature, it mainly occurs as trans-βcaryophyllene ((E)-CRY) mixed with small amounts of its isomers, cis-βcaryophyllene (Z)-β-caryophyllene) and α-caryophyllene (α-humulene), together with its oxidation derivative β-caryophyllene oxide (CRYO) (**Figure 9**)

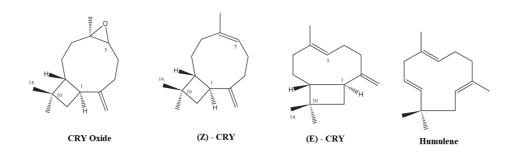


Fig. 9: Structures of trans-CRY, its isomers and oxidative product.

β-CRY has a low water solubility but is able to interact with artificial lipid bilayer, which strongly suggests its high affinity to the cell membranes. It is one of the major active components of essential oils derived from large number of spice and food plants. It is commonly found in basil, cinnamon, black pepper, cloves, cannabis, lavender, oregano and rosemary and it is used as a flavouring agent (approved by the Food and Drug Administration (FDA) and by the European Food Safety Authority (EFSA) with identification number FL no: 01.007).

Its biological effects include anti-inflammatory (Medeiros et al., 2007), anticarcinogenic (Langhasova et al., 2014), antimicrobial (Sabulal et al., 2006), antioxidative (Singh et al., 2006), and analgesic activities (Klauke et al., 2014). The latter is typical of β -CRY which belongs to a class of cannabinoids (CBs),

specifically phyto-cannabinoids (pCBs), that were identified as plant derivatives of Cannabis sativa L.

Gertsch et al. demonstrated that the (E)-CRY is the first Cannabis-derived functional CB receptor ligand which displays higher biding affinity to CB2 than its isomer Z-CRY, whereas CRYO and α -humulene possess no CB2 binding properties (Gertsch et al., 2008).

1.7.1 β-CRY as anticancer agent

Dahham et al. reported the antiproliferative effect of β-CRY on several cancer cell lines (Dahham et al., 2015). B-CRY isolated from essential oils of Aquilaria crassna stem bark led to strong growth inhibition in two colon cancer cell lines, HCT-116 and HT-29, as well as in pancreatic cancer cells, PANC-1, whereas it had no effects on other tested cancer cell lines. In contrast, Ambrož et al. studies revealed that β-CRY obtained from Myrica rubra did not affect CaCo-2 intestinal cancer cell viability (Ambrož et al., 2015). Jung et al. described the anticancer effects of \beta-CRY in in vivo model of high-fat dietinduced obese C57BL/6N mice. (Jung et al., 2015). They observed that a highfat diet (HFD) induces larger and more aggressive tumors in animals injected with B16F10 melanoma cells. β-CRY treatment abolished the HFD pro-cancer effects. The anticancer activity of β-CRY in vivo was also presented at the Euro Global Summit on Cancer Therapy in Valencia, 2015. In this report, a growth and vascularization of tumors developed from orthotopically grafted colon cancer cells into nude mice were reduced significantly after administration of β-CRY isolated from agar wood.

Furthermore β -CRY is able to enhance the efficacy of classical anticancer drugs, such as paclitaxel or doxorubicin (Ambrož et al., 2015), (Legault and Pichette, 2007), (Kim et al., 2014).

In the last few years, Di Sotto et al. deeply investigated the role of β -CRY, focusing their attention on the protective effects against DNA-damage caused by different toxic compounds. These authors demonstrated that β -CRY was able to protect cultured human lymphocytes from the genotoxic damage induced by ethyl methanesulfonate and colcemid (Di Sotto et al., 2010).

Another study shows the capability of β -CRY to inhibit the genotoxicity of a condensate of cigarette smoke both in bacterial and mammalian cells (Giacomo et al., 2016). At last, Giacomo et al. demonstrated that β -CRY protective role, against damage caused by Condensed Smoke Cigarette in mammalian cells, is exerted by inhibition of CSC-mediated STAT3 phosphorylation (Di Giacomo et al., 2018).

CHAPTER 2: AIM OF THE RESEARCH

OCPs have been deeply studied in the last decades and cytotoxic mechanisms of compounds such as dioxin and polychlorinated biphenyls have been widely described in literature (Yang et al., 2015). Still, the \u03b3-isomer of hexaclorocyclohexane has not been largely investigated, despite its relatively common geographic distribution (Fagnocchi et al., 2018) and its physicochemical properties make it extremely dangerous for human health. During the first part of my research we started investigating the cellular mechanisms triggered by β-HCH, as it has been found to be a major contaminant of the "Sacco River Valley" area, in the south of Rome. In 2005, an environmental emergency state was declared in the aforementioned area and a bio-monitoring study started in 2006 (Porta et al., 2013). Results of the epidemiological study, conducted on 1000 people residing in this area (Sorveglianza Sanitaria Ed Epidemiologica Della Popolazione Residente In Prossimità Del Fiume Sacco Rapporto tecnico attività 2013-2015 Dipartimento di Epidemiologia del Servizio Sanitario Regionale - Regione Lazio) showed above the average emetic concentrations of β -HCH; thus we decided to test, in vitro, a concentration that was extrapolated by those results. Taking into account the pleiotropic role of the protein signal transducer and activator of transcription 3 (STAT3), its function as a hub protein in cellular signalling pathways triggered by β-HCH was investigated in different cancer cell lines corresponding to tissues that are especially vulnerable to damage caused by environmental pollutants. Data obtained were recently published (Rubini et al., 2018) and suggest that β-HCH is able to exert its toxicity triggering STAT3 cellular pathways.

The second part of my research project was founded by the Sovena Foundation which gave me the chance to carry on a collaboration with Antonella Di Sotto

research group, at the department of Physiology and Pharmacology of La Sapienza, Rome. In the context of this collaboration it has been previously demonstrated that the sesquiterpene β -Caryophyllene is able to exert anticancer activities decrising Y705-STAT3 phosphorilation induced by Condensed Smoke Cigarette (Di Giacomo et al., 2018). Therefore we decided to evaluate β -CRY protective role against the deregulation of STAT3 pathways induced by β -HCH in human breast cancer cell lines MCF-7 and MDA-MB-468. The obtained data confirm that β -CRY exerts a chemopreventing effect by inhibiting β -HCH-induced STAT3 activation.

The last part of my PhD project has been carried out at University of Copenhagen, Department of Biomedical Science, where I worked in the laboratory of the Professor Ole Hartvig Mortensen. Here, I started investigating the effect of β –HCH on placental cells. We selected this model since there are increasing evidences that the placenta acts as a partial barrier for OCPs (Perera et al., 2005). A recent publication shows that β –HCH, together with many others OCPs, is ubiquitous in umbilical cord sera and placentas, indicating that the adverse effects of OCPs on foetus and infants should be of concern. Moreover, the concentrations of OCPs in umbilical cord sera and placentas are significantly associated with those in maternal sera, which suggests a direct influence of the maternal burden (Zhang et al., 2018). Moreover, a study published this year on Nature shows that black carbon particles, derived from particulate matter combustion, accumulate on the fetal side of the placenta, probably affecting the foetus from early life onwards (Bové et al., 2019).

For this last set of experiments, we selected a trophoblast cancer model, therefore using BeWo cell line. Since optimal and appropriate biomarkers of placental toxicity are still being researched, we mainly focused our attention on STAT3 pathways, as this protein seems to be a good marker for this model.

We investigated whether β -HCH molecular mechanism is exerted through STAT3 pathways activation. Moreover, we measured the mRNA expression level of several genes encoding transport proteins which are necessary for nutrients transport across placenta and their absorption. Preliminary data show that STAT3 is ones more involved in the cellular response to β -HCH and that the organochlorine compound also affects the expression level of nutrient transports such as GLUT1 and TauT.

CHAPTER 3: MATERIALS AND METHODS

3.1 Reagents

β-caryophyllene (CRY; ≥ 98.5% purity, Sigma-Aldrich Co, St. Louis, MO, USA), β-hexaclorocyclohexane (β-HCH) (Sigma-Aldrich), protease inhibitors cocktail (Sigma-Aldrich); BSA (Sigma-Aldrich); I-block (Thermo Fisher Scientific, Rodano, Italy); RPMI-1640 and DMEM (InVitrogen, MI); fetal bovine serum (FBS- Sigma-Aldirich); MTT (3-(4,5-dimethylthiazol-2-yl)-2,5diphenyl-2H-tetrazolium bromide) (Sigma-Aldrich,); Peroxidase and Alkaline phosphatase-conjugated secondary antibody (Jackson ImmunoResearch, Pero, Italy); ECL Fast Femto reagent (Immunological Science, Roma, Italy); BCIP/NBT reagents (Carl Roth, Milano, Italy, CAS No. 298-83-9 and 6578-06-9); TRIzol® Reagent (Immunological Science); Super Script II R-Transcriptase (FS-RT-3022, Fisher Molecular Biology, Rodano, Italy). βhexaclorocyclohexane (β-HCH) (Sigma-Aldrich, 33376), at a final concentration of 10 µM, was tested on each cell line pre-treated or not with specific inhibitors: 6 µM AZD1480 (Sigma-Aldrich, SML1505), 100 µM S3I-201 (Sigma-Aldrich, SML0330), 70 nM Dasatinib (Selleckchem, Roma, Italy, Cat. No. S1021), 0.8 µM Lapatinib (Sigma-Aldrich, CDS022971), and 15 µM Gefitinib (Sigma-Aldrich, SLM1657).

β-hexaclorocyclohexane was dissolved in sterile 100% DMSO while β-caryophyllene in sterile 100% Ethanol and stored at -20°C.

3.2 Cell cultures

Cell culture was carried out according to standard procedures in a humidified 5% CO₂ incubator at 37°C. Human HER2+ breast cancer cell line MCF-7 and human prostate cancer cell line LNCaP, were maintained in RPMI-1640 (Roswell Park Memorial Institute) medium; human triple negative breast

cancer cell line MDA-MB-468 and human hepatoma cell line HepG2 were grown in DMEM (InVitrogen, MI). All media were supplemented with 10% v/v fetal bovine serum (FBS- Sigma-Aldrich), 100 units/mL penicillin and 100 μg/mL streptomycin, 1% w/v sodium pyruvate, 2mM glutamine.

Human choriocarcinoma cell line BeWo, was grown in DMEM High Glucose (Thermo Fisher Scientific). The medium was supplemented with 10% v/v fetal bovine serum (FBS- Sigma-Aldrich), 1% Penicillin-Streptomycin 1000 units/mL (Invitrogen) and 2mM glutamine.

β-hexaclorocyclohexane (β-HCH), at a final concentration of 10 μM, was tested on each cell line, overnight pre-treated or not with 10 μg/mLβ-caryophyllene (β-CRY).

3.3 Cell viability assay

The β -HCH cytotoxicity was evaluated by seeding cells in 96-well plates and measuring cell viability after 24 and 48 hours of incubation in the presence of different concentrations of β -HCH (5, 10, 25, 50, 75, 100, 125, 150, 175, and 200 μ M). Cell viability was measured using MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide) (Sigma-Aldrich M2128). Briefly, the culture medium was removed and 125 μ L of MTT solution (0.5 mg/mL MTT in culture medium) was added to each well. After 2 hours of incubation, the solution was removed and the insoluble formazan dye resulting from the conversion of tetrazolium salt by metabolically active cells was dissolved by adding 125 μ L/well of DMSO and measured at 570 nm using the MultiskanTM FC Microplate Photometer (ThermoFisher).

3.4 Cell lysate preparation

Adherent cells were washed and scraped with cold PSB. Total protein extracts were obtained using a lysis buffer containing 2% SDS, 20 mM Tris-

hydrocloride Ph = 7.4, 2 M urea, 10% glycerol added with 2 mM sodium orthovanadate, 10 mM DTT, and a protease inhibitors cocktail diluted 1:100 (Sigma-Aldrich). After a 30" sonication with Labsonic M (Sartorius), and centrifugation (20min, 14.000 RPM, 4°C), the total protein extracted were quantified by measuring spectrophotometrically the absorbance at 260 nm, and re-suspended in 4X Laemmli Buffer (2% SDS, 62,5 mM Tris-HCl pH 6.8, 10% v/v glycerol, 5 mM DTT, 0.02% bromophenol blue) for the electrophoresis.

3.5 Western blotting

The lysate samples in 4X Laemmli buffer proteins were resolved by SDS-PAGE 10% TGX FastCastTM Acrylamide gel (BioRad, Segrate, Italy) (200V, 45 min) and transferred on PVDF membranes (BioRad) using Trans-Blot® TurboTM Transfer System (BioRad). Protein bands were electrotransferred to a PVDF membrane. The membranes were blocked with 3% w/v non-fat dried milk or 0.2% w/v I-block (Thermo Fisher Scientific, Rodano, Italy) in Trisbuffered saline containing 0.05% Tween-20 (TBS-T) and incubated with a specific primary antibody for 1 h. Subsequently, membranes were washed three times in TBS-T, and then incubated for an additional hour with appropriate horseradish peroxidase- or alkaline phosphatase-conjugated secondary antibody (Jackson ImmunoResearch, Pero, Italy). The peroxidase signal was detected with ECL Fast Femto reagent (Immunological Science, Roma, Italy), acquired by Molecular Imager® ChemiDocTM MP System (Bio-Rad), and the intensity of protein bands was quantified using the ImageLab Software. The alkaline phosphatase signal was detected with BCIP/NBT reagents (Carl Roth, Milano, Italy). β-actin was used as normalization protein. Immunodetection was carried out using primary anti-pY705-STAT3 (Cell Signaling D3A7, antibody dilution 1:2000), anti Tot-STAT3 (Cell Signaling D3Z2G, antibody dilution 1:1000), anti-JAK2 (Cell Signaling D2E12, antibody dilution 1:1000), anti-PY1007/1008JAK2 (Cell Signaling C80C3, antibody dilution 1:1000), anti-EGFR (Cell Signaling D38B1, antibody dilution 1:1000), anti-pY1173EGFR (Cell Signaling 53A5, antibody dilution 1:1000), anti-Src (Cell Signaling 32G6, antibody dilution 1:1000), and anti-pY416Src (Cell Signaling 6943S, antibody dilution 1:1000) primary antibodies.

At least three experimental replicates were performed for each biological sample. All results are expressed as mean \pm SD. Differences between experimental groups were determined by Student's t-test. p-value of <0.01 was considered statistically significant

3.6 Total RNA Extraction

Cells were harvested and total RNA was isolated with TRIzol® Reagent (Immunological Science) following the manufacturer's instructions. RNA was precipitated by adding isopropanol and subsequently centrifuged at 12,500 g for 10 minutes at 4°C. The precipitated RNA was washed with 75% ethanol and air dried. Total RNA was than resuspended in RNase-free water and quantified spectrophotometrically. Its quality was assessed by 1% agarose gel electrophoresis and staining with ethidium bromide.

3.7 cDNA synthesis and Quantitative Real Time-PCR

The reverse transcription was carried out with Super Script II R-Transcriptase (FS-RT-3022, Fisher Molecular Biology, Rodano, Italy). Gene expression was evaluated with specific primers for BIRC-5, c-MYC, CRP, p21, CCND1, MMP2 and S18 (housekeeping) (all from Qiagen S.r.l., Milano, Italy), Taut, GLUT1 and GLUT3 (TAG Copenhagen A/S), using CFX Connect™ Real-Time PCR Detection System (BioRad) with a SYBR green fluorophore based

real-time reaction (Brilliant SYBR Green QPCR Master Mix, Thermo Fisher Scientific). Expression data were analyzed using CFX ManagerTM Real Time PCR Detection System Software, Version 3.1 (BioRad).

3.8 Cell Cycle Assay

Cell cycle analysis was performed by flow cytometry using Propidium Iodide (PI) staining. MCF-7 cells were seeded on 6 well plates (150x10³/well), and grown for 24 hours. Then, cells were treated for 48 hours with 10 μM β-HCH alone of after an over-night pre-treatment with 10μg/ml β-CRY. Cells were scraped, collected and washed with PBS. Subsequently, cells were fixed with 70% ethanol solution, drop-wise added during vortex agitation and incubated for 24 hours at 4°C. Cells were collected by centrifugation, washed in PBS, resuspended in 200 μl of PBS containing PI (30 μg/ml), Triton-X100 (0,1 % w/v), RNase (0.2 mg/ml) and then incubated for 30 minutes at room temperature. Fluorescence intensity of PI (FL2) was recorded by a cytoflorimeter (Accuri C6, Becton Dickinson) and cell cycle analysis was performed using a specific software (Modfit LT, Verity Software House). Results were expressed, according to PI intensity, as the percentage of cells in each cell cycle phase.

3.9 Scratch test Assay

The effects of β -HCH and β -CRY on cell migration was evaluated. MCF7 and MDA-MB-468 cells (1 × 10⁴/well) were plated in 6 well plates (VWR®, tissue culture plate), and incubated for 24 hours to allow them to adhere. Afterwards, the cell monolayer was scraped with a pipette tip in order to create a "scratch", dividing the well in two distinctive chambers. Cells were washed with PBS to remove the debris, then fresh medium was added. Cells were incubated with 10 μ M β -HCH for 12, 24 and 48 hours, with or without an over-night pre-

treatment with 10 μ g/ml β -CRY. Cells that had invaded the scratch were detected via microscopy (Leica Microsystems).

3.10 Cell Proliferation Assay

A cell proliferation assay was performed to investigate the antiproliferative effect of β -CRY. Briefly, 10×10^5 cells/well of both MCF-7 and MDA-MB-468 cell lines were seeded into 6-well plates and treated for 12, 24 and 48 hours with $10 \mu M \beta$ -HCH alone or after an over-night pre-treatment with β -CRY 10 μ g/ml. Then, at each time point, cells were trypsinized, resuspended in fresh medium and viable cells number was evaluated using Trypan Blu dye and the Thoma cell counting chamber. Counts were carried out from 3 to 5 times.

3.11 Statistical analyses

The repeatability of results was confirmed by performing all experiments at least three times.

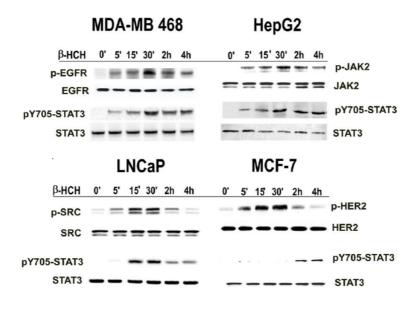
The obtained values are presented as mean and standard deviation. Statistical analysis was performer with GraphPad Prisma software using Student's t-test.

4. RESULTS

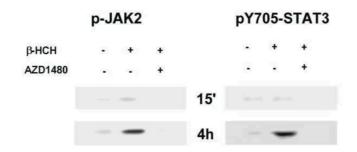
4.1 β-Hexachlorocyclohexane triggers STAT3 pathways

Data we recently published suggest that β -HCH is able to exert its toxicity triggering STAT3 cellular pathways. We chose a panel of cell lines representing different human tumor types associated with the expression and activation of specific receptors potentially involved in STAT3 activation: membrane and membrane associated tyrosine kinase receptors (EGFR in MDA-MB 468, JAK2 in HepG2, and HER2 in MCF-7 cells) and cytoplasmic non-receptor tyrosine kinases (SRC in LNCaP). The β -HCH tested concentration (10 μ M) was extrapolated both from environmental–epidemiological studies carried out on the exposed population living throughout the "Valle del Sacco" area (Sorveglianza Sanitaria Ed Epidemiologica Della Popolazione Residente In Prossimità Del Fiume Sacco Rapporto tecnico attività 2013-2015 Dipartimento di Epidemiologia del Servizio Sanitario Regionale - Regione Lazio) and from previous in vitro studies (Sharma et al., 2010), (Briz et al., 2011).

A time course assay was at first performed. The activation of STAT3 upon β-HCH treatment was followed by immunoblotting analysis, together with the activation of the specific receptors taken into account. Results indicate that STAT3 phosphorylation occurs within the same time as the receptors activation with the exception of MCF-7 cells (**Figure 10 A**). The delayed activation of pY705–STAT3 observed in MCF-7 cells is not directly related to the HER2 receptor but seems to be mediated by the JAK2 receptor (**Figure 10 B**)



B

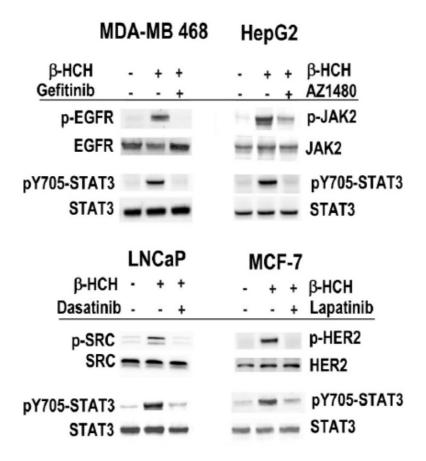


Int. J. Mol. Sci 19 (7), 2108. "STAT3, a Hub Protein of Cellular Signaling Pathways, Is Triggered by β-Hexaclorocyclohexane"

Fig. 10: (A) Immunoblot analysis of the time course assay performed on different cell lines treated with 10 μM β -HCH. Samples were analyzed for STAT3 and each cell line for a specific membrane or membrane associated tyrosine kinase receptor. Both unmodified and phosphorylated form were detected for each protein using specific antibodies. (B) Evaluation of JAK2 and STAT3 phosphorylation level in MCF-7 cell line treated with 10 μM β -HCH. Cells were incubated for 15 minutes and 4 hours in the absence or presence of JAK2 inhibitor (AZD1480), then subjected to Immunoblot analysis.

In order to verify if the cell-specific action of β -HCH is dependent on STAT3-mediated pathways, all cell lines were exposed to 10 μ M β -HCH, pre-treated or not with specific inhibitors of the different receptors/cytoplasmic tyrosine kinases analysed (Dasatinib as Src inhibitor, AZD1480 as JAK2 inhibitor, Gefitinib as EGFR inhibitor, and Lapatinib as HER2 inhibitor). For this experiment, a single incubation time with β -HCH was selected for each cell line, according to results of Y705–STAT3 phosphorylation obtained from the time course assay. In particular, we chose 15 minutes of β -HCH treatment for MDA-MB 468, HepG2, and LNCaP cells, and 2 hours for MCF-7 cells. Western blot assay was performed to analyze both unphosphorylated and phosphorylated receptors and STAT3. Immunoblot results show the absence of STAT3 and receptors phosphorylation in cells pre-treated with the specific inhibitors (**Figure 11**).

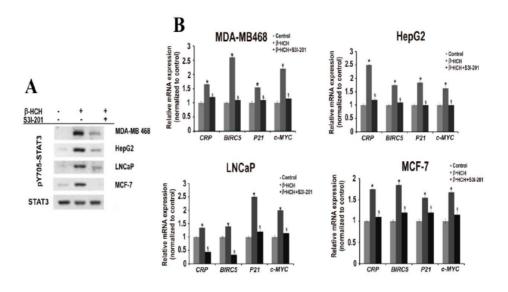
To further confirm the role of STAT3 in the cellular response to β -HCH, the expression profile of STAT3 specific target genes, which represents a different phase of the carcinogenesis process (P21 for cell cycle, CRP for inflammation, BIRC5-Survivin for apoptosis, and c-MYC for proliferation) was evaluated. The same analysis was performed in the presence of a specific STAT3 inhibitor, S3I-201. All cell lines were treated with β -HCH in the presence or absence of S3I-201 and the time points of treatment were selected taking into account the rapid or delayed STAT3 activation, previously observed. Results shown in **Figure 12 B** seem to confirm that STAT3 is involved in the cellular response to β -HCH and can probably mediate its potential tumor activity. Indeed, in all cell lines tested, β -HCH treatment leads to an increase in the expression level of STAT3-specific genes analyzed.



Int. J. Mol. Sci 19 (7), 2108. "STAT3, a Hub Protein of Cellular Signaling Pathways, Is Triggered by β-Hexaclorocyclohexane"

Fig. 11: Inhibition of specific signalling pathways activated by β-HCH. Each cell line was treated with 10 μM β-HCH in the presence or absence of specific inhibitors: Gefitinib as EGFR inhibitor in MDA-MB 468 cells; AZD1480 as JAK2 inhibitor in HepG2 cells; Dasatinib as SRC inhibitor in LNCaP cells; and Lapatinib as HER2 inhibitor in MCF-7 cells. Cells were incubated (15 min for MDA-MB 468, HepG2, and LNCaP cells and 2 h for MCF-7 cells) and cellular extracts were subjected to Immunoblot analysis. Detection of the unmodified and corresponding phosphorylated form was carried out using specific antibodies

As expected, this increase was not observed when cells were co-treated with the STAT3 inhibitor. As control, STAT3 activation was checked by Western Blotting in all cell lines and the pre-treatment with S3I-201 significantly reduced STAT3 phosphorylation (**Figure 12 A**). The overall results support the hypothesis that STAT3 is involved in pathways triggered by β -HCH and that it can mediate the inflammatory, anti-apoptotic and proliferative activity of this organochlorine compound.



Int. J. Mol. Sci 19 (7), 2108. "STAT3, a Hub Protein of Cellular Signaling Pathways, Is Triggered by β-Hexaclorocyclohexane"

Fig. 12: (A) Immunoblot analysis of pY705–STAT3 in cells treated for 4 hours with 10 μM β-HCH, in the presence or absence of a specific STAT3 inhibitor (S3I-201). Detection of phosphorylated and unphosphorylated STAT3 was carried out using specific antibodies. (**B**) RT-qPCR analysis of STAT3 target genes (CRP, BIRC5, p21, and c-MYC) performed on β-HCH treated or untreated cells, as well as on cells pre-incubated with a specific STAT3 inhibitor (S3I-201). Statistically significant differences (p < 0.05) between β-HCH treated and β-HCH untreated cells are marked by *, while statistically significant differences (p < 0.05) between β-HCH treated and pre-incubated or not with a specific STAT3 inhibitor are marked by §.

4.2 Investigation of β -caryophyllene protective role against β -hexachlorocyclohexane carcinogenic property

4.2.1 β-caryophyllene inhibition of STAT 3 activation

We decided to focus our attention on breast cancer cell lines MCF-7 and MDA-MB-468 and we investigated the protective role of the sesquiterpene β -caryophyllene against the β -HCH-induced phosphorylation of Y705-STAT3. Cells were over-night pre-treated with 10 μ g/mL of the natural compound. Afterwards, MDA-MB468 and MCF7 cells were incubated with 10 μ M β -HCH for 15 minutes or 2 hours, respectively. As shown in **Figure 13**, β -CRY was able to down-modulate phosphorylation of Y705-STAT3 in both cell lines.

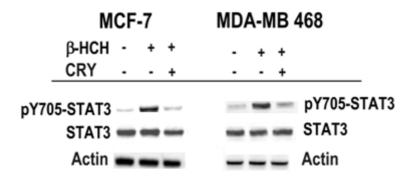
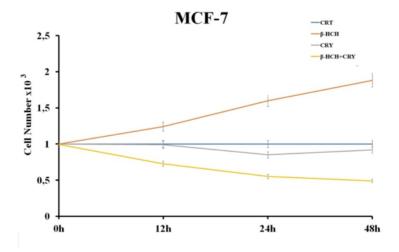


Fig. 13: Immunoblot analysis of human breast cancer cell lines (MCF-7 and MDA-MB 468) treated with 10 μM β-HCH, in presence or absence of β-Caryophyllene 10 μg/mL (over-night pre-treatment). Cells were incubated (15 min and 2h of β-HCH treatment for MDA-MB 468 and MCF-7 respectively), then cellular extracts were subjected to Immunoblot analysis of unmodified-STAT3, pY705 STAT3 and Actin.

4.2.2 β-caryophyllene inhibition of proliferation and migration

A cell proliferation and a scratch test assays were performed on both MCF-7 and MDA-MB-468 cell lines, to investigate the protective role of β -caryophyllene against β -HCH potential carcinogenic effect. The same experimental conditions were used for the two assays: cells were exposed for 12, 24 and 48 hours to 10 μ M β -HCH, alone or after an over-night pretreatment with 10 μ g/mL β -CRY.

As shown in **Figure 14**, β -HCH is able to induce, compared to untreated cells, a 2-fold increase of cells number while β -CRY is able to decrease the cell proliferation. This down-modulation is much more evident in cells over-night pre-treated with β -CRY and then exposed to β -HCH. The Scratch Test Assay confirmed data obtained from the proliferation assay. It is clear that cell invasion is greatly enhanced by β -HCH and decreased by β -CRY, especially when cells are pre-treated with the sesquiterpene and afterwards exposed to β -HCH (**Figure 15**). These results show that β -Caryophyllene is able to protect cells against β -HCH effects.



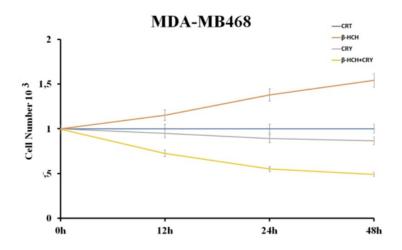


Fig. 14: Proliferation assay performed on MCF-7 and MDA-MB-468 cell lines. Cells were treated for 12h, 24h and 48h with 10 μ M β -HCH, alone or after an over-night pre-treatment with 10 μ g/mL β -CRY. Trypan Blue was used to detect viable cells after 12h, 24h and 48h. Data represent the means of three independent experiments

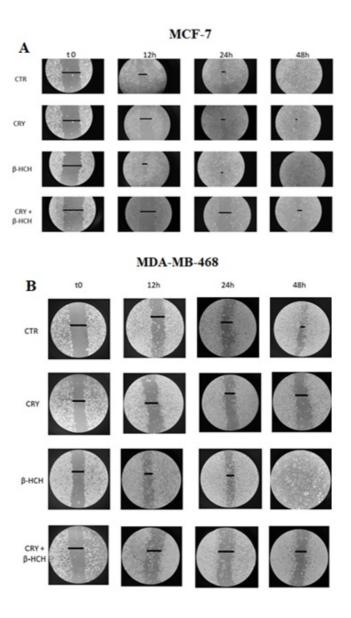
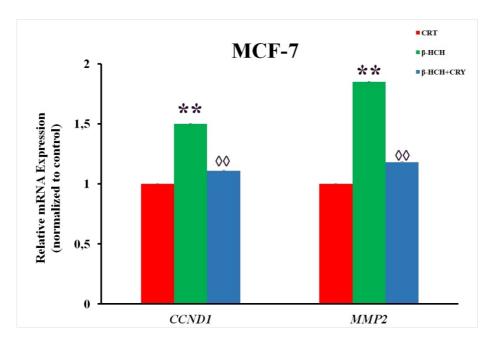


Fig 15 Cells migration was detected performing a Scratch Test Assay on A) MCF-7 cell line and B) MDA-MB-468 cell line. Cells were treated for 12h, 24h and 48h with 10 μM β-HCH, alone or after over-night pre-treatment with 10 μg/mL β-CRY. Images are representative of three different experiments.

To further investigate if β -CRY protective role against carcinogenic effect of β -HCH is due to the modulation of STAT3 protein activity, the expression profile of two genes under STAT3 control (CCND1, for cell cycle and MMP2, for cell invasion) was analyzed. Cells were over-night pre-treated with 10 μ g/mL β -CRY and then exposed to 10 μ M β -HCH for 48 hours. After RNA extraction, RT-PCR was performed.

β-HCH treatment was able to increase the expression of CCND1 and MMP2 in both cell lines. β-CRY pre-treatment down-modulated the expression of these genes (**Figure 16 A and B**). These data are in line with previous results, shown in Figure 10 and Figure 11. Indeed, CCND1 positive modulation can explain increased cells proliferation while MMP2 level increase confirm that cells ability to close the scratch depends on the effect on cell motility, rather than cell proliferation.





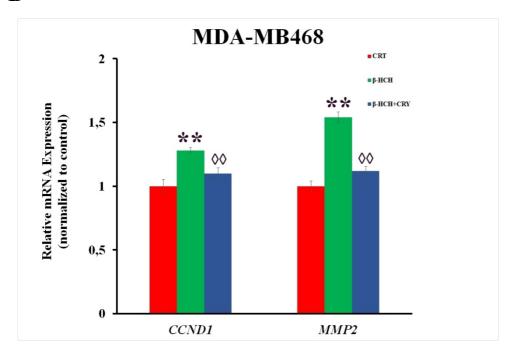


Fig 16: RT-qPCR analysis of STAT3 target genes (CCND1 and MMP2). Analysis was performed on A) MCF-7 cell line and B) MDA-MB-468 cell line. Cells were treated for 48 hours with 10 μM β -HCH, alone or after over-night pre-treatment with 10 μg/mL β -CRY. Statistically significant differences (p < 0.05) between β -HCH treated cells and control are marked with stars, while statistically significant differences (p < 0.05) between β -CRY pre-treated cells and β -HCH treated cell are marked with rhombus.

The mechanism underlying this inhibitory activity was further investigated with cell cycle analysis. MCF-7 cells were incubated for 48 hours with 10 μ M β -HCH, alone or after over-night pre-treatment with 10 μ g/mL β -CRY, then analysed by flow cytometer. **Figure 17** indicates a significant block of cell

cycle in G2/M phase when cells are exposed to both β -HCH and β -CRY. This checkpoint blocks the entry into mitosis when DNA is damaged. There is evidence in literature of another natural compound able to inhibit cancer cells proliferation arresting the cell cycle at this checkpoint. (Luk et al., 2005). Our results could explain the capability of β -CRY to contain β -Hexachlorocyclohexane caused damage, inducing both cell cycle arrest and decrease of cell cycle protein (CCND1) expression levels.

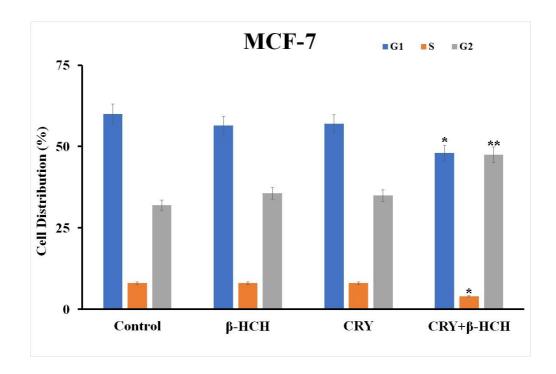


Fig 17: Analysis of cell cycle progression. MCF-7 cells were treated with 10 μ M β -HCH, alone or after over-night pre-treatment with 10 μ g/mL β -CRY. Cells were harvested after 48 hours, then they were fixed, stained and analyzed for DNA content. The percentage off cell distribution in G1, S and G2/M phase of the cell cycle are indicated.

4.3 β-hexachlorocyclohexane effect on placenta

Despite the evidence that β -HCH can accumulate in the placental tissue and be related to many foetal and infant diseases, we lack information about the molecular mechanism involved. The most interesting trophoblast cells feature is their physiological capability of invading the surrounding tissues. This phenomenon is temporally and locally controlled and it starts immediately after embryo implantation into the endometrium; still, this invasive capability is typically correlated to tumor progression and must be strictly regulated to avoid the cancerogenic development of placenta cells. This process is strictly regulated by numerous signalling mediators and the JAK/STAT3 is one of the most important pathways involved during the development of placenta. (Fitzgerald et al., 2008). We decided to investigate, using an in vitro model of cariocarcinoma (BeWo cell line), β-HCH activity on STAT3 pathways. First of all, the effect of β-HCH on cell viability was evaluated. As is it shown in Figure 18, after 24 hours of exposure β -HCH induced null cytotoxic effects up to the concentration of 125 µM, while it is evident, after 48 hours of exposure and only at the higher concentrations, up to 50% of cytotoxicity. No appreciable reduction in cell viability was observed when exposing cells to 10 μ M β -HCH so we used this concentration further on.

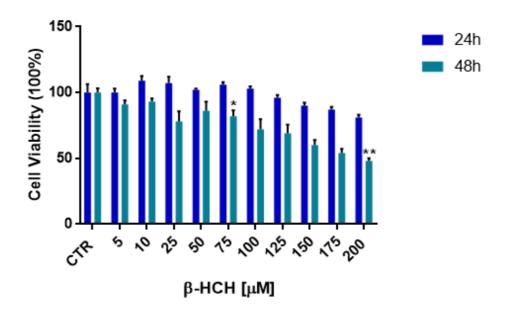
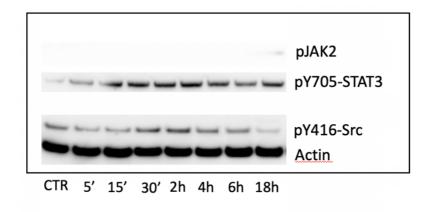


Fig. 18: BeWo cells were treated with increasing concentrations of β-HCH, for 24 and 48 hours. Cell viability was assessed by MTT assay. Cell viability is expressed as percentage of measured data of treated cells compared to control. Data represent the mean of at least three independent measurements and error bars indicate SEM. Data were subjected to statistical analysis and significant differences (p < 0.05) are marked with asterisks.

A time course assay on BeWo cell line treated with 10 μ M β -HCH was at first performed. The activation of STAT3 pathways, upon β -HCH treatment was followed by immunoblotting analysis. Preliminary results show that STAT3 phosphorylation occurs within the same time as the pY416-Src activation, already after 30 minutes of exposure to β -HCH, indicating that in this model the activation of pY705–STAT3 could be mediated by the cytoplasmatic non-receptor tyrosine kinase Src (**Figure 19**)



B

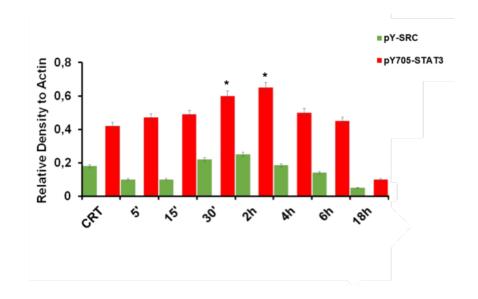


Fig 19: (A) Immunoblot analysis of the time course assay performed on BeWo cell line treated with 10 μM β-HCH. Samples were analysed for pJAK2, pY705-STAT3, pY416-Src and Actin proteins using specific antibodies (B) Densitometry of pY416-Src and pY705-STAT3 proteins, normalized to Actin, is shown. Statistically significant differences (p < 0.05) between β-HCH treated and β-HCH untreated cells are marked with asterisks.

The last experiment performed is a Real Time qPCR of genes encoding transport proteins expressed in the placental tissue, such as GLUT1, GLUT3 and TauT. **Figure 20** shows that 10 μ M β -HCH treatment is able to decrease the mRNA expression level of both GLUT1 and TauT genes, after 12 and 24 hours of treatment respectively. The correct expression of these genes is fundamental to guarantee the foetus nutrients supply during the pregnancy; so these preliminary results let us suppose that in uterus β -HCH exposure could lead to severe adverse effects on fetal development.

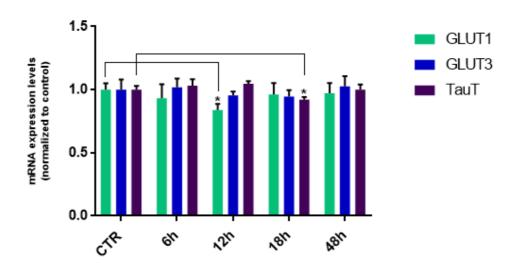


Fig 20: RT-qPCR analysis was performed on 10 μM β-HCH treated or untreated BeWo cells. The expression level of genes encoding glucose and Taurine transport (GLUT1/3 and TauT) was analyzed at different time points (6h, 12h, 18h, 48h). Statistically significant differences (p < 0.05) are marked with asterisks.

CHAPTER 5: DISCUSSION

My PhD project firstly aimed to clarify some of the molecular mechanisms involved in the alteration of key signalling pathways triggered by high levels of β -HCH, such as those revealed in human blood of people residing in the Valle del Sacco area, chronically exposed to this and many other organochlorine compounds. The obtained results support the hypothesis that STAT3 is involved in several cellular pathways affected by β -HCH.

STAT3 phosphorylation on residue Y705 was observed in all evaluated cell lines and it can mediate both a rapid and a delayed cellular response to β -HCH through different cellular pathways, distinctive of each cell line.

RT-PCR results show that this protein can also mediate the inflammatory, anti-apoptotic and proliferative activity of β -HCH (Rubini et al., 2018).

The natural sesquiterpene β -caryophyllene was evaluated for its beneficial properties against the cell injury induced by β -HCH. Particularly, the ability of the test substance to inhibit the STAT3 pathway, which is known to be involved in the β -HCH tumorigenic mechanism and in the neoplastic progression of damaged cells, has been demonstrated. β -CRY was also shown to be active against β -HCH-induced proliferation and migration. This inhibitory property seems to be related to β -CRY ability to interfere with STAT3 transcriptional activity. Indeed, gene such as CCDN1 and MMP2, under STAT3 control, were analyzed and their mRNA expression level was significally decreased in cells pre-treated with the sesquiterpene β -CRY, compared to those exposed to β -HCH alone.

Even though results we obtained are extremely interesting and promising, nothing is known yet about the nature of the interaction between β -CRY and STAT3 protein. It would be interesting to deeply investigate the molecular mechanism underlying the inhibition of STAT3 activation exerted by this

sesquiterpene. Moreover, the effect of β -HCH on placenta needs to be deeper investigated, together with the molecular mechanism involved. The very preliminary data shown here would need to be validated and the effect of this organochlorine in uterus exposure could be studied *in vivo*, to truly understand the effect of this compound on the fetal development. Finally the protective role of β -Caryophyllene could be investigated in placenta and *in vivo* too, in order to understand if this natural compound could be one day used as a chemopreventive agent.

REFERENCES

- Ambrož, M., Boušová, I., Skarka, A., Hanušová, V., Králová, V., Matoušková, P., Szotáková, B., Skálová, L., 2015. The Influence of Sesquiterpenes from Myrica rubra on the Antiproliferative and Pro-Oxidative Effects of Doxorubicin and Its Accumulation in Cancer Cells. Mol. Basel Switz. 20, 15343–15358. https://doi.org/10.3390/molecules200815343
- Andersen, H.R., Vinggaard, A.M., Rasmussen, T.H., Gjermandsen, I.M., Bonefeld-Jørgensen, E.C., 2002. Effects of currently used pesticides in assays for estrogenicity, androgenicity, and aromatase activity in vitro. Toxicol. Appl. Pharmacol. 179, 1–12. https://doi.org/10.1006/taap.2001.9347
- Avalle, L., Camporeale, A., Camperi, A., Poli, V., 2017. STAT3 in cancer: A double edged sword. Cytokine 98, 42–50. https://doi.org/10.1016/j.cyto.2017.03.018
- Aznar, S., Valerón, P.F., del Rincon, S.V., Pérez, L.F., Perona, R., Lacal, J.C., 2001. Simultaneous tyrosine and serine phosphorylation of STAT3 transcription factor is involved in Rho A GTPase oncogenic transformation. Mol. Biol. Cell 12, 3282–3294. https://doi.org/10.1091/mbc.12.10.3282
- Barker, D.J.P., 2003. Editorial: The Developmental Origins of Adult Disease. Eur. J. Epidemiol. 18, 733–736.
- Bové, H., Bongaerts, E., Slenders, E., Bijnens, E.M., Saenen, N.D., Gyselaers, W., Eyken, P.V., Plusquin, M., Roeffaers, M.B.J., Ameloot, M., Nawrot, T.S., 2019. Ambient black carbon particles reach the fetal side of human placenta. Nat. Commun. 10, 1–7. https://doi.org/10.1038/s41467-019-11654-3
- Bowman, T., Garcia, R., Turkson, J., Jove, R., 2000. STATs in oncogenesis. Oncogene 19, 2474–2488. https://doi.org/10.1038/sj.onc.1203527
- Briz, V., Molina-Molina, J.-M., Sánchez-Redondo, S., Fernández, M.F., Grimalt, J.O., Olea, N., Rodríguez-Farré, E., Suñol, C., 2011. Differential estrogenic effects of the persistent organochlorine pesticides dieldrin, endosulfan, and lindane in primary neuronal cultures. Toxicol. Sci. Off. J. Soc. Toxicol. 120, 413–427. https://doi.org/10.1093/toxsci/kfr019
- Caldenhoven, E., Dijk, T.B. van, Solari, R., Armstrong, J., Raaijmakers, J.A.M., Lammers, J.-W.J., Koenderman, L., Groot, R.P. de, 1996. STAT3β, a Splice Variant of Transcription Factor STAT3, Is a Dominant Negative Regulator of Transcription. J. Biol. Chem. 271, 13221–13227. https://doi.org/10.1074/jbc.271.22.13221
- Casals-Casas, C., Desvergne, B., 2011. Endocrine disruptors: from endocrine to metabolic disruption. Annu. Rev. Physiol. 73, 135–162. https://doi.org/10.1146/annurev-physiol-012110-142200

- Chessells, M.J., Hawker, D.W., Connell, D.W., Papajcsik, I.A., 1988. Factors influencing the distribution of lindane and isomers in soil of an agricultural environment. Chemosphere 17, 1741–1749. https://doi.org/10.1016/0045-6535(88)90102-6
- Chung, C.D., Liao, J., Liu, B., Rao, X., Jay, P., Berta, P., Shuai, K., 1997. Specific inhibition of Stat3 signal transduction by PIAS3. Science 278, 1803–1805. https://doi.org/10.1126/science.278.5344.1803
- Chung, J., Uchida, E., Grammer, T.C., Blenis, J., 1997. STAT3 serine phosphorylation by ERK-dependent and -independent pathways negatively modulates its tyrosine phosphorylation. Mol. Cell. Biol. 17, 6508–6516. https://doi.org/10.1128/mcb.17.11.6508
- Cimica, V., Chen, H.-C., Iyer, J.K., Reich, N.C., 2011. Dynamics of the STAT3 transcription factor: nuclear import dependent on Ran and importin-β1. PloS One 6, e20188. https://doi.org/10.1371/journal.pone.0020188
- Concha-Graña, E., Turnes-Carou, M.I., Muniategui-Lorenzo, S., López-Mahía, P., Prada-Rodríguez, D., Fernández-Fernández, E., 2006. Evaluation of HCH isomers and metabolites in soils, leachates, river water and sediments of a highly contaminated area. Chemosphere 64, 588–595. https://doi.org/10.1016/j.chemosphere.2005.11.011
- Coosen, R., van Velsen, F.L., 1989. Effects of the beta-isomer of hexachlorocyclohexane on estrogen-sensitive human mammary tumor cells. Toxicol. Appl. Pharmacol. 101, 310–318. https://doi.org/10.1016/0041-008x(89)90279-2
- Crews, D.P., Putz, O., Thomas, P., Hayes, T., Howdeshell, K., 2003. Wildlife as models for the study of how mixtures, low doses, and the embryonic environment modulate the action of endocrine-disrupting chemicals. Pure Appl. Chem. 75, 2305–2320.
- Crofton, K.M., 2004. Developmental disruption of thyroid hormone: correlations with hearing dysfunction in rats. Risk Anal. Off. Publ. Soc. Risk Anal. 24, 1665–1671. https://doi.org/10.1111/j.0272-4332.2004.00557.x
- Dahham, S.S., Tabana, Y.M., Iqbal, M.A., Ahamed, M.B.K., Ezzat, M.O., Majid, A.S.A., Majid, A.M.S.A., 2015. The Anticancer, Antioxidant and Antimicrobial Properties of the Sesquiterpene β -Caryophyllene from the Essential Oil of Aquilaria crassna. Mol. Basel Switz. 20, 11808–11829. https://doi.org/10.3390/molecules200711808
- Das, P.M., Singal, R., 2004. DNA methylation and cancer. J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol. 22, 4632–4642. https://doi.org/10.1200/JCO.2004.07.151
- Dewilde, S., Vercelli, A., Chiarle, R., Poli, V., 2008. Of alphas and betas: distinct and overlapping functions of STAT3 isoforms. Front. Biosci. J. Virtual Libr. 13, 6501–6514.
- Di Giacomo, S., Abete, L., Cocchiola, R., Mazzanti, G., Eufemi, M., Di Sotto, A., 2018. Caryophyllane sesquiterpenes inhibit DNA-damage by tobacco smoke in

- bacterial and mammalian cells. Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc. 111, 393–404. https://doi.org/10.1016/j.fct.2017.11.018
- Di Sotto, A., Mazzanti, G., Carbone, F., Hrelia, P., Maffei, F., 2010. Inhibition by beta-caryophyllene of ethyl methanesulfonate-induced clastogenicity in cultured human lymphocytes. Mutat. Res. 699, 23–28. https://doi.org/10.1016/j.mrgentox.2010.04.008
- Dokduang, H., Yongvanit, P., Namwat, N., Pairojkul, C., Sangkhamanon, S., Yageta, M.S., Murakami, Y., Loilome, W., 2016. Xanthohumol inhibits STAT3 activation pathway leading to growth suppression and apoptosis induction in human cholangiocarcinoma cells. Oncol. Rep. 35, 2065–2072. https://doi.org/10.3892/or.2016.4584
- Ecker, A., Simma, O., Hoelbl, A., Kenner, L., Beug, H., Moriggl, R., Sexl, V., 2009. The dark and the bright side of Stat3: proto-oncogene and tumor-suppressor. Front. Biosci. Landmark Ed. 14, 2944–2958.
- Fagnocchi, L., Poli, V., Zippo, A., 2018. Enhancer reprogramming in tumor progression: a new route towards cancer cell plasticity. Cell. Mol. Life Sci. CMLS 75, 2537–2555. https://doi.org/10.1007/s00018-018-2820-1
- Fitzgerald, J.S., Poehlmann, T.G., Schleussner, E., Markert, U.R., 2008. Trophoblast invasion: the role of intracellular cytokine signalling via signal transducer and activator of transcription 3 (STAT3). Hum. Reprod. Update 14, 335—344. https://doi.org/10.1093/humupd/dmn010
- Gertsch, J., Leonti, M., Raduner, S., Racz, I., Chen, J.-Z., Xie, X.-Q., Altmann, K.-H., Karsak, M., Zimmer, A., 2008. Beta-caryophyllene is a dietary cannabinoid. Proc. Natl. Acad. Sci. U. S. A. 105, 9099–9104. https://doi.org/10.1073/pnas.0803601105
- Giacomo, S.D., Mazzanti, G., Sotto, A.D., 2016. Mutagenicity of cigarette butt waste in the bacterial reverse mutation assay: The protective effects of β -caryophyllene and β -caryophyllene oxide. Environ. Toxicol. 31, 1319–1328. https://doi.org/10.1002/tox.22136
- Goldey, E.S., Kehn, L.S., Lau, C., Rehnberg, G.L., Crofton, K.M., 1995. Developmental exposure to polychlorinated biphenyls (Aroclor 1254) reduces circulating thyroid hormone concentrations and causes hearing deficits in rats. Toxicol. Appl. Pharmacol. 135, 77–88. https://doi.org/10.1006/taap.1995.1210
- Hardell, L., Andersson, S.-O., Carlberg, M., Bohr, L., Bavel, B. van, Lindström, G., Björnfoth, H., Ginman, C., 2006. Adipose Tissue Concentrations of Persistent Organic Pollutants and the Risk of Prostate Cancer. J. Occup. Environ. Med. 48, 700–707. https://doi.org/10.1097/01.jom.0000205989.46603.43
- Home [WWW Document], n.d. . TEDX Endocr. Disrupt. Exch. URL https://endocrinedisruption.org/home/ (accessed 9.27.19).
- Jones, K.C., de Voogt, P., 1999. Persistent organic pollutants (POPs): state of the science. Environ. Pollut. Barking Essex 1987 100, 209–221.

- Jung, J.I., Kim, E.J., Kwon, G.T., Jung, Y.J., Park, T., Kim, Y., Yu, R., Choi, M.-S., Chun, H.S., Kwon, S.-H., Her, S., Lee, K.W., Park, J.H.Y., 2015. β-Caryophyllene potently inhibits solid tumor growth and lymph node metastasis of B16F10 melanoma cells in high-fat diet-induced obese C57BL/6N mice. Carcinogenesis 36, 1028–1039. https://doi.org/10.1093/carcin/bgv076
- Khan, S.M., Dauffenbach, L.M., Yeh, J., 2000. Mitochondria and caspases in induced apoptosis in human luteinized granulosa cells. Biochem. Biophys. Res. Commun. 269, 542–545. https://doi.org/10.1006/bbrc.2000.2321
- Kim, C., Cho, S.K., Kim, K.-D., Nam, D., Chung, W.-S., Jang, H.-J., Lee, S.-G., Shim, B.S., Sethi, G., Ahn, K.S., 2014. β-Caryophyllene oxide potentiates TNFα-induced apoptosis and inhibits invasion through down-modulation of NF-κβ-regulated gene products. Apoptosis Int. J. Program. Cell Death 19, 708–718. https://doi.org/10.1007/s10495-013-0957-9
- Kim, E., Kim, M., Woo, D.-H., Shin, Y., Shin, J., Chang, N., Oh, Y.T., Kim, H., Rheey, J., Nakano, I., Lee, C., Joo, K.M., Rich, J.N., Nam, D.-H., Lee, J., 2013. Phosphorylation of EZH2 activates STAT3 signaling via STAT3 methylation and promotes tumorigenicity of glioblastoma stem-like cells. Cancer Cell 23, 839–852. https://doi.org/10.1016/j.ccr.2013.04.008
- Klauke, A.-L., Racz, I., Pradier, B., Markert, A., Zimmer, A.M., Gertsch, J., Zimmer, A., 2014. The cannabinoid CB₂ receptor-selective phytocannabinoid beta-caryophyllene exerts analgesic effects in mouse models of inflammatory and neuropathic pain. Eur. Neuropsychopharmacol. J. Eur. Coll. Neuropsychopharmacol. 24, 608–620. https://doi.org/10.1016/j.euroneuro.2013.10.008
- Krebs, D.L., Hilton, D.J., 2001. SOCS proteins: negative regulators of cytokine signaling. Stem Cells Dayt. Ohio 19, 378–387. https://doi.org/10.1634/stemcells.19-5-378
- Krstev, S., Knutsson, A., 2019. Occupational Risk Factors for Prostate Cancer: A Meta-analysis. J. Cancer Prev. 24, 91–111. https://doi.org/10.15430/JCP.2019.24.2.91
- Langhasova, L., Hanusova, V., Rezek, J., Stohanslova, B., Ambroz, M., Kralova, V., Vanek, T., Lou, J.D., Yun, Z.L., Yang, J., Skalova, L., 2014. Essential oil from Myrica rubra leaves inhibits cancer cell proliferation and induces apoptosis in several human intestinal lines. Ind. Crops Prod. Complete, 20–26. https://doi.org/10.1016/j.indcrop.2014.04.018
- Legault, J., Pichette, A., 2007. Potentiating effect of beta-caryophyllene on anticancer activity of alpha-humulene, isocaryophyllene and paclitaxel. J. Pharm. Pharmacol. 59, 1643–1647. https://doi.org/10.1211/jpp.59.12.0005
- Li, J., Li, N., Ma, M., Giesy, J.P., Wang, Z., 2008. In vitro profiling of the endocrine disrupting potency of organochlorine pesticides. Toxicol. Lett. 183, 65–71. https://doi.org/10.1016/j.toxlet.2008.10.002

- Li, L., Cheung, S.-H., Evans, E.L., Shaw, P.E., 2010. Modulation of gene expression and tumor cell growth by redox modification of STAT3. Cancer Res. 70, 8222–8232. https://doi.org/10.1158/0008-5472.CAN-10-0894
- Luk, S.C.-W., Siu, S.W.-F., Lai, C.-K., Wu, Y.-J., Pang, S.-F., 2005. Cell Cycle Arrest by a Natural Product via G2/M Checkpoint. Int. J. Med. Sci. 2, 64–69.
- Ma, J., Cui, B., Ding, X., Wei, J., Cui, L., 2015. Over-Expression of Cyclin D1 Promotes NSCs Proliferation and Induces the Differentiation into Astrocytes Via Jak-STAT3 Pathways. Neurochem. Res. 40, 1681–1690. https://doi.org/10.1007/s11064-015-1635-9
- Marino, F., Orecchia, V., Regis, G., Musteanu, M., Tassone, B., Jon, C., Forni, M., Calautti, E., Chiarle, R., Eferl, R., Poli, V., 2014. STAT3β controls inflammatory responses and early tumor onset in skin and colon experimental cancer models. Am. J. Cancer Res. 4, 484–494.
- McGlynn, K.A., Abnet, C.C., Zhang, M., Sun, X.-D., Fan, J.-H., O'Brien, T.R., Wei, W.-Q., Ortiz-Conde, B.A., Dawsey, S.M., Weber, J.-P., Taylor, P.R., Katki, H., Mark, S.D., Qiao, Y.-L., 2006. Serum concentrations of 1,1,1-trichloro-2,2-bis(p-chlorophenyl)ethane (DDT) and 1,1-dichloro-2,2-bis(p-chlorophenyl)ethylene (DDE) and risk of primary liver cancer. J. Natl. Cancer Inst. 98, 1005–1010. https://doi.org/10.1093/jnci/djj266
- McKinlay, R., Plant, J.A., Bell, J.N.B., Voulvoulis, N., 2008. Endocrine disrupting pesticides: implications for risk assessment. Environ. Int. 34, 168–183. https://doi.org/10.1016/j.envint.2007.07.013
- Medeiros, R., Passos, G.F., Vitor, C.E., Koepp, J., Mazzuco, T.L., Pianowski, L.F., Campos, M.M., Calixto, J.B., 2007. Effect of two active compounds obtained from the essential oil of Cordia verbenacea on the acute inflammatory responses elicited by LPS in the rat paw. Br. J. Pharmacol. 151, 618–627. https://doi.org/10.1038/sj.bjp.0707270
- Mills, P.K., Yang, R., 2003. Prostate cancer risk in California farm workers. J. Occup. Environ. Med. 45, 249–258. https://doi.org/10.1097/01.jom.0000058339.05741.0c
- Mrema, E.J., Rubino, F.M., Brambilla, G., Moretto, A., Tsatsakis, A.M., Colosio, C., 2013. Persistent organochlorinated pesticides and mechanisms of their toxicity. Toxicology 307, 74–88. https://doi.org/10.1016/j.tox.2012.11.015
- Ng, D.C.H., Lin, B.H., Lim, C.P., Huang, G., Zhang, T., Poli, V., Cao, X., 2006. Stat3 regulates microtubules by antagonizing the depolymerization activity of stathmin. J. Cell Biol. 172, 245–257. https://doi.org/10.1083/jcb.200503021
- Ng, I.H.W., Ng, D.C.H., Jans, D.A., Bogoyevitch, M.A., 2012. Selective STAT3-α or -β expression reveals spliceform-specific phosphorylation kinetics, nuclear retention and distinct gene expression outcomes. Biochem. J. 447, 125–136. https://doi.org/10.1042/BJ20120941
- Nicholas, C., B, G., 2011. The Jak-STAT Signal Transduction Pathway in Melanoma. Breakthr. Melanoma Res. undefined-undefined. https://doi.org/10.5772/18876

- Nie, Y., Erion, D.M., Yuan, Z., Dietrich, M., Shulman, G.I., Horvath, T.L., Gao, Q., 2009. STAT3 inhibition of gluconeogenesis is downregulated by SirT1. Nat. Cell Biol. 11, 492–500. https://doi.org/10.1038/ncb1857
- Nishimoto, A., Kugimiya, N., Hosoyama, T., Enoki, T., Li, T.-S., Hamano, K., 2013. JAB1 regulates unphosphorylated STAT3 DNA-binding activity through protein-protein interaction in human colon cancer cells. Biochem. Biophys. Res. Commun. 438, 513–518. https://doi.org/10.1016/j.bbrc.2013.07.105
- Perera, F.P., Rauh, V., Whyatt, R.M., Tang, D., Tsai, W.Y., Bernert, J.T., Tu, Y.H., Andrews, H., Barr, D.B., Camann, D.E., Diaz, D., Dietrich, J., Reyes, A., Kinney, P.L., 2005. A summary of recent findings on birth outcomes and developmental effects of prenatal ETS, PAH, and pesticide exposures. Neurotoxicology 26, 573–587. https://doi.org/10.1016/j.neuro.2004.07.007
- Persson, E.C., Graubard, B.I., Evans, A.A., London, W.T., Weber, J.-P., LeBlanc, A., Chen, G., Lin, W., McGlynn, K.A., 2012. Dichlorodiphenyltrichloroethane (DDT) and risk of hepatocellular carcinoma. Int. J. Cancer J. Int. Cancer 131, 2078–2084. https://doi.org/10.1002/ijc.27459
- Porta, D., Fantini, F., De Felip, E., Blasetti, F., Abballe, A., Dell'Orco, V., Fano, V., Ingelido, A.M., Narduzzi, S., Forastiere, F., 2013. A biomonitoring study on blood levels of beta-hexachlorocyclohexane among people living close to an industrial area. Environ. Health 12, 57. https://doi.org/10.1186/1476-069X-12-57
- Randi, A.S., Hernandez, S., Alvarez, L., Sanchez, M., Schwarcz, M., Kleiman d Pisarev, D.L., 2003. Hexachlorobenzene-induced early changes in ornithine decarboxylase and protein tyrosine kinase activities, polyamines and c-Myc, c-Fos and c-Jun proto-oncogenes in rat liver. Toxicol. Sci. Off. J. Soc. Toxicol. 76, 291–298. https://doi.org/10.1093/toxsci/kfg253
- Rubini, E., Altieri, F., Chichiarelli, S., Giamogante, F., Carissimi, S., Paglia, G., Macone, A., Eufemi, M., 2018. STAT3, a Hub Protein of Cellular Signaling Pathways, Is Triggered by β-Hexaclorocyclohexane. Int. J. Mol. Sci. 19, 2108. https://doi.org/10.3390/ijms19072108
- Rusiecki, J.A., Baccarelli, A., Bollati, V., Tarantini, L., Moore, L.E., Bonefeld-Jorgensen, E.C., 2008. Global DNA hypomethylation is associated with high serum-persistent organic pollutants in Greenlandic Inuit. Environ. Health Perspect. 116, 1547–1552. https://doi.org/10.1289/ehp.11338
- Sabulal, B., Dan, M., J, A.J., Kurup, R., Pradeep, N.S., Valsamma, R.K., George, V., 2006. Caryophyllene-rich rhizome oil of Zingiber nimmonii from South India: Chemical characterization and antimicrobial activity. Phytochemistry 67, 2469–2473. https://doi.org/10.1016/j.phytochem.2006.08.003
- Said, T.M., Paasch, U., Glander, H.-J., Agarwal, A., 2004. Role of caspases in male infertility. Hum. Reprod. Update 10, 39–51. https://doi.org/10.1093/humupd/dmh003

- Saiyed Habibullah, Dewan Aruna, Bhatnagar Vijay, Shenoy Udyavar, Shenoy Rathika, Rajmohan Hirehall, Patel Kumud, Kashyap Rekha, Kulkarni Pradip, Rajan Bagalur, Lakkad Bhadabhai, 2003. Effect of endosulfan on male reproductive development. Environ. Health Perspect. 111, 1958–1962. https://doi.org/10.1289/ehp.6271
- Sharma, H., Zhang, P., Barber, D.S., Liu, B., 2010. Organochlorine pesticides dieldrin and lindane induce cooperative toxicity in dopaminergic neurons: role of oxidative stress. Neurotoxicology 31, 215–222. https://doi.org/10.1016/j.neuro.2009.12.007
- Shi, Y., Song, Y., Wang, Yuping, Wang, Yinan, Liang, X., Hu, Y., Yu, H., Guan, X., Cheng, J., Yang, K., 2011. β-Benzene hexachloride induces apoptosis of rat Sertoli cells through generation of reactive oxygen species and activation of JNKs and FasL. Environ. Toxicol. 26, 124–135. https://doi.org/10.1002/tox.20536
- Singh, G., Marimuthu, P., de Heluani, C.S., Catalan, C.A.N., 2006. Antioxidant and biocidal activities of Carum nigrum (seed) essential oil, oleoresin, and their selected components. J. Agric. Food Chem. 54, 174–181. https://doi.org/10.1021/jf0518610
- Sobotta, M.C., Liou, W., Stöcker, S., Talwar, D., Oehler, M., Ruppert, T., Scharf, A.N.D., Dick, T.P., 2015. Peroxiredoxin-2 and STAT3 form a redox relay for H2O2 signaling. Nat. Chem. Biol. 11, 64–70. https://doi.org/10.1038/nchembio.1695
- Steinmetz, R., Young, P.C., Caperell-Grant, A., Gize, E.A., Madhukar, B.V., Ben-Jonathan, N., Bigsby, R.M., 1996. Novel estrogenic action of the pesticide residue beta-hexachlorocyclohexane in human breast cancer cells. Cancer Res. 56, 5403–5409.
- Szczepanek, K., Chen, Q., Derecka, M., Salloum, F.N., Zhang, Q., Szelag, M., Cichy, J., Kukreja, R.C., Dulak, J., Lesnefsky, E.J., Larner, A.C., 2011. Mitochondrial-targeted Signal transducer and activator of transcription 3 (STAT3) protects against ischemia-induced changes in the electron transport chain and the generation of reactive oxygen species. J. Biol. Chem. 286, 29610–29620. https://doi.org/10.1074/jbc.M111.226209
- Tammineni, P., Anugula, C., Mohammed, F., Anjaneyulu, M., Larner, A.C., Sepuri, N.B.V., 2013. The import of the transcription factor STAT3 into mitochondria depends on GRIM-19, a component of the electron transport chain. J. Biol. Chem. 288, 4723–4732. https://doi.org/10.1074/jbc.M112.378984
- Timofeeva, O.A., Chasovskikh, S., Lonskaya, I., Tarasova, N.I., Khavrutskii, L., Tarasov, S.G., Zhang, X., Korostyshevskiy, V.R., Cheema, A., Zhang, L., Dakshanamurthy, S., Brown, M.L., Dritschilo, A., 2012. Mechanisms of unphosphorylated STAT3 transcription factor binding to DNA. J. Biol. Chem. 287, 14192–14200. https://doi.org/10.1074/jbc.M111.323899

- Turkson, J., Zhang, S., Palmer, J., Kay, H., Stanko, J., Mora, L.B., Sebti, S., Yu, H., Jove, R., 2004. Inhibition of constitutive signal transducer and activator of transcription 3 activation by novel platinum complexes with potent antitumor activity. Mol. Cancer Ther. 3, 1533–1542.
- Wegrzyn, J., Potla, R., Chwae, Y.-J., Sepuri, N.B.V., Zhang, Q., Koeck, T., Derecka, M., Szczepanek, K., Szelag, M., Gornicka, A., Moh, A., Moghaddas, S., Chen, Q., Bobbili, S., Cichy, J., Dulak, J., Baker, D.P., Wolfman, A., Stuehr, D., Hassan, M.O., Fu, X.-Y., Avadhani, N., Drake, J.I., Fawcett, P., Lesnefsky, E.J., Larner, A.C., 2009. Function of Mitochondrial Stat3 in Cellular Respiration. Science 323, 793–797. https://doi.org/10.1126/science.1164551
- WHO | State of the science of endocrine disrupting chemicals 2012 [WWW Document], n.d. . WHO. URL http://www.who.int/ceh/publications/endocrine/en/ (accessed 9.27.19).
- Willett, K.L., Ulrich, E.M., Hites, R.A., 1998. Differential Toxicity and Environmental Fates of Hexachlorocyclohexane Isomers. Environ. Sci. Technol. 32, 2197–2207. https://doi.org/10.1021/es9708530
- Wong, P.S., Matsumura, F., 2007. Promotion of breast cancer by betahexachlorocyclohexane in MCF10AT1 cells and MMTV-neu mice. BMC Cancer 7, 130. https://doi.org/10.1186/1471-2407-7-130
- Xie, Y., Kole, S., Precht, P., Pazin, M.J., Bernier, M., 2009. S-Glutathionylation Impairs Signal Transducer and Activator of Transcription 3 Activation and Signaling. Endocrinology 150, 1122–1131. https://doi.org/10.1210/en.2008-1241
- Xu, D., Qu, C.-K., 2008. Protein tyrosine phosphatases in the JAK/STAT pathway. Front. Biosci. J. Virtual Libr. 13, 4925–4932. https://doi.org/10.2741/3051
- Xu, Y.S., Liang, J.J., Wang, Y., Zhao, X.-Z.J., Xu, L., Xu, Y.-Y., Zou, Q.C., Zhang, J.M., Tu, C.-E., Cui, Y.-G., Sun, W.-H., Huang, C., Yang, J.-H., Chin, Y.E., 2016. STAT3 Undergoes Acetylation-dependent Mitochondrial Translocation to Regulate Pyruvate Metabolism. Sci. Rep. 6, 39517. https://doi.org/10.1038/srep39517
- Yang, J., Huang, J., Dasgupta, M., Sears, N., Miyagi, M., Wang, B., Chance, M.R., Chen, X., Du, Y., Wang, Y., An, L., Wang, Q., Lu, T., Zhang, X., Wang, Z., Stark, G.R., 2010. Reversible methylation of promoter-bound STAT3 by histone-modifying enzymes. Proc. Natl. Acad. Sci. U. S. A. 107, 21499–21504. https://doi.org/10.1073/pnas.1016147107
- Yang, J., Liao, X., Agarwal, M.K., Barnes, L., Auron, P.E., Stark, G.R., 2007.
 Unphosphorylated STAT3 accumulates in response to IL-6 and activates transcription by binding to NFkappaB. Genes Dev. 21, 1396–1408. https://doi.org/10.1101/gad.1553707
- Yang, O., Kim, H.L., Weon, J.-I., Seo, Y.R., 2015. Endocrine-disrupting Chemicals:
 Review of Toxicological Mechanisms Using Molecular Pathway Analysis. J.
 Cancer Prev. 20, 12–24. https://doi.org/10.15430/JCP.2015.20.1.12

- Yu, H., Lee, H., Herrmann, A., Buettner, R., Jove, R., 2014. Revisiting STAT3 signalling in cancer: new and unexpected biological functions. Nat. Rev. Cancer 14, 736–746. https://doi.org/10.1038/nrc3818
- Yuan, J., Zhang, F., Niu, R., 2015. Multiple regulation pathways and pivotal biological functions of STAT3 in cancer. Sci. Rep. 5. https://doi.org/10.1038/srep17663
- Yuan, Z.-L., Guan, Y.-J., Chatterjee, D., Chin, Y.E., 2005. Stat3 dimerization regulated by reversible acetylation of a single lysine residue. Science 307, 269–273. https://doi.org/10.1126/science.1105166
- Zhang, Xiaolan, Wu, X., Lei, B., Jing, Y., Jiang, Z., Zhang, Xinyu, Fang, X., Yu, Y., 2018. Transplacental transfer characteristics of organochlorine pesticides in paired maternal and cord sera, and placentas and possible influencing factors. Environ. Pollut. 233, 446–454. https://doi.org/10.1016/j.envpol.2017.10.075
- Zhu, B.-H., Chen, H.-Y., Zhan, W.-H., Wang, C.-Y., Cai, S.-R., Wang, Z., Zhang, C.-H., He, Y.-L., 2011. (-)-Epigallocatechin-3-gallate inhibits VEGF expression induced by IL-6 via Stat3 in gastric cancer. World J. Gastroenterol. WJG 17, 2315–2325. https://doi.org/10.3748/wjg.v17.i18.2315

CHAPTER 6: APPENDIX

The Appendix collects the front page of my publications.





Articl

STAT3, a Hub Protein of Cellular Signaling Pathways, Is Triggered by β-Hexaclorocyclohexane

Elisabetta Rubini 1 $^{\odot}$, Fabio Altieri 1,2 $^{\odot}$, Silvia Chichiarelli 1 $^{\odot}$, Flavia Giamogante 1 $^{\odot}$, Stefania Carissimi 1 , Giuliano Paglia 1 $^{\odot}$, Alberto Macone 1 $^{\odot}$ and Margherita Eufemi 1,2,*

- Department of Biochemical Sciences, A. Rossi Fanelli, Sapienza University, P.le A. Moro 5, 00185 Rome, Italy; elisabetta.rubini@uniroma1.it (E.R.); fabio.altieri@uniroma1.it (F.A.); silvia.chichiarelli@uniroma1.it (S.C.); flavia.giamogante@uniroma1.it (F.G.); stefania.carissimi@uniroma1.it (S.C.); giuliano.pag@gmail.com (G.P.); alberto.macone@uniroma1.it (A.M.)
- ² Istituto Pasteur-Fondazione Cenci Bolognetti, Sapienza University, P.le A. Moro 5, 00185 Rome, Italy
- * Correspondence: margherita.eufemi@uniroma1.it; Tel.: +39-06-49910598

Received: 26 June 2018; Accepted: 17 July 2018; Published: 20 July 2018



Abstract: Background: Organochlorine pesticides (OCPs) are widely distributed in the environment and their toxicity is mostly associated with the molecular mechanisms of endocrine disruption Among OCPs, particular attention was focused on the effects of β -hexaclorocyclohexane (β -HCH), a widely common pollutant. A detailed epidemiological study carried out on exposed population in the "Valle del Sacco" found correlations between the incidence of a wide range of diseases and the occurrence of $\beta\textsc{-HCH}$ contamination. Taking into account the pleiotropic role of the protein signal transducer and activator of transcription 3 (STAT3), its function as a hub protein in cellular signaling pathways triggered by β-HCH was investigated in different cell lines corresponding to tissues that are especially vulnerable to damage by environmental pollutants. Materials and Methods: Human prostate cancer (LNCaP), human breast cancer (MCF-7 and MDA-MB 468), and human hepatoma (HepG2) cell lines were treated with 10 μ M β -HCH in the presence or absence of specific inhibitors for different receptors. All samples were subjected to analysis by immunoblotting and RT-qPCR. Results and Conclusions: The preliminary results allow us to hypothesize the involvement of STAT3, through both its canonical and non-canonical pathways, in response to β -HCH. Moreover, we ascertained the role of STAT3 as a master regulator of energy metabolism via the altered expression and localization of HIF-1 α and PKM2, respectively, resulting in a Warburg-like effect.

 $\textbf{Keywords:} \ STAT3; \ \beta\text{-}hexaclorocyclohexane} \ (\beta\text{-}HCH); signal \ transduction; energy \ metabolism$

1. Introduction

Organochlorine compounds are widely distributed in the environment and several studies link their basic molecular mechanism of endocrine disruption [1,2] with the onset of many pathological conditions such as chronic inflammatory processes, cardiovascular diseases, neurological and metabolic disorders, and oncogenesis [3–5]. The toxicity of organochlorine compounds is related to their physicochemical properties, because these pollutants belong to a group of organic compounds, known as "persistent organic pollutants" (POPs), that are resistant to degradation or biodegradation and that can be bioaccumulated into adipose tissue because of their lipotropic properties and great stability [6]. Observational epidemiological studies, carried out on population at high risk of exposure, revealed that POPs may play an important role in the development of a chronic inflammatory state by interfering with pathways associated with essential cellular processes and homeostasis [7]. Acute inflammation represents one of the early responses to injury but, if the causal insult becomes persistent, it may progress with a long chronic phase; although acute inflammation is necessary to rid the organism of

Hindawi Oxidative Medicine and Cellular Longevity Volume 2018, Article ID 5919237, 14 pages https://doi.org/10.1155/2018/5919237



Research Article

Antiviral and Antioxidant Activity of a Hydroalcoholic Extract from Humulus lupulus L.

Antonella Di Sotto 0,1 Paola Checconi 0,2,3 Ignacio Celestino,2,3 Marcello Locatelli,4 Stefania Carissimi, Marta De Angelis, Valeria Rossi, Dolores Limongi, Chiara Toniolo 0,6 Lucia Martinoli, Silvia Di Giacomo, Anna Teresa Palamara (), 23 and Lucia Nencioni (5)

ondence should be addressed to Antonella Di Sotto; antonella disotto@uniromal.it and Paola Checconi; paola.checconi@uniromal.it

Received 10 January 2018; Accepted 16 May 2018; Published 24 July 2018

Academic Editor: Ralf Haus

Copyright © 2018 Antonella Di Sotto et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

A hydroalcoholic extract from female inflorescences of Humulus Lupulus L. (HOP extract) was evaluated for its anti-influenza activity. The ability of the extract to interfere with different phases of viral replication was assessed, as well as its effect on the intracellular redox state, being unbalanced versus the oxidative state in infected cells. The radical scavenging power, inhibition of lipoperoxidation, and ferric reducing activity were assayed as antioxidant mechanisms. A phytochemical characterization of the extract was also performed. We found that HOP extract significantly inhibited replication of various viral strains, at different time from infection. Viral replication was partly inhibited when virus was incubated with extract before infection, suggesting a direct effect on the virions. Since HOP extract was able to restore the reducing conditions of infected cells, by increasing glutathione content, its antiviral activity might be also due to an interference with redox-sensitive pathways required for viral replication. Accordingly, the extract exerted radical scavenging and reducing effects and inhibited lipoperoxidation and the tBOOH-induced cytotoxicity. At phytochemical analysis, different phenolics were identified, which altogether might contribute to HOP antiviral effect. In conclusion, our results highlighted anti-influenza and antioxidant properties of HOP extract, which encourage further in vivo studies to evaluate its possible application.

Department of Physiology and Pharmacology V. Erspamer, Sapienza University of Rome, P.le Aldo Moro 5, 00185 Rome, Italy

²Department of Public Health and Infectious Diseases, Laboratory Affiliated to Istituto Pasteur Italia-Fondazione Cenci Bolognetti, Sapienza University of Rome, P. le Aldo Moro 5, 00185 Rome, Italy

³IRCCS San Raffaele Pisana, Department of Human Sciences and Promotion of the Quality of Life, San Raffaele Roma Open University, Via di Val Cannuta 247, 00166 Rome, Italy

⁴Department of Pharmacy, University "G. D'Annunzio" of Chieti-Pescara, Via dei Vestini 31, 66100 Chieti, Italy
⁵Department of Biochemical Sciences "A. Rossi Fanelli", Sapienza University of Rome, P.le Aldo Moro 5, 00185 Rome, Italy

⁶ Department of Environmental Biology, Sapienza University of Rome, P.le Aldo Moro 5, 00185 Rome, Italy





Article

STAT3 Post-Translational Modifications Drive Cellular Signaling Pathways in Prostate Cancer Cells

Rossana Cocchiola ¹0, Elisabetta Rubini ¹0, Fabio Altieri ¹0, Silvia Chichiarelli ¹0, Giuliano Paglia ¹0, Donatella Romaniello ²0, Stefania Carissimi ¹0, Alessandra Giorgi ¹, Flavia Giamogante ¹, Alberto Macone ¹0, Giacomo Perugia ³, Aymone Gurtner ⁴ and Margherita Eufemi ¹,*

- Department of Biochemical Sciences "A. Rossi Fanelli" and Istituto Pasteur-Fondazione Cenci Bolognetti, Sapienza University, Ple A. Moro 5, 00185 Rome, Italy; rossana.cocchiola@uniroma1.it (R.C.); elisabetta.rubini@uniroma1.it (E.R.); fabio.altie ri@uniroma1.it (F.A.); silvia.chichiarelli@uniroma1.it (S.C.); giuliano.paglia@uniroma1.it (G.P.); stefania.carissimi@uniroma1.it (S.C.); alessandra.giorgi@uniroma1.it (A.G.); flavia.giamogante@uniroma1.it (F.G.); alberto.macone@uniroma1.it (A.M.)
- Department of Biological Regulation, Weizmann Institute of Science, 234 Herzl Street, 7610001 Rehovot, Israel; donatella.romaniello@weizmann.ac.il
- Department of Gynecological-Obstretic Science and Urologic Sciences, Sapienza University, V.le Dell'Università, 00185 Rome, Italy; giacomo.perugia@uniroma1.it
- Department of Research, Advanced Diagnostics, and Technological Innovation, Translational Research Area, Regina Elena National Cancer Institute; via Elio Chianesi, 53, 00144 Rome, Italy; aymone.gurtner@ifo.gov.it
- Correspondence: margherita.eufemi@uniroma1.it; Tel.: +39-064-991-0598; Fax: +39-064-440-062

Received: 5 March 2019; Accepted: 9 April 2019; Published: 12 April 2019



Abstract: STAT3 is an oncoprotein overexpressed in different types of tumors, including prostate cancer (PCa), and its activity is modulated by a variety of post-translational modifications (PTMs). Prostate cancer represents the most common cancer diagnosed in men, and each phase of tumor progression displays specific cellular conditions: inflammation is predominant in tumor's early stage, whereas oxidative stress is typical of clinically advanced PCa. The aim of this research is to assess the correspondence between the stimulus-specificity of STAT3 PTMs and definite STAT3-mediated transcriptional programs, in order to identify new suitable pharmacological targets for PCa treatment. Experiments were performed on less-aggressive LNCaP and more aggressive DU-145 cell lines, simulating inflammatory and oxidative-stress conditions. Cellular studies confirmed pY705-STAT3 as common denominator of all STAT3-mediated signaling. In addition, acK685-STAT3 was found in response to IL-6, whereas glutC328/542-STAT3 and pS727-STAT3 occurred upon tert-butyl hydroperoxyde (tBHP) treatment. Obtained results also provided evidence of an interplay between STAT3 PTMs and specific protein interactors such as P300 and APE1/Ref-1. In accordance with these outcomes, mRNA levels of STAT3-target genes seemed to follow the differing STAT3 PTMs. These results highlighted the role of STAT3 and its PTMs as drivers in the progression of PCa.

RINGRAZIAMENTI

Ho desiderato questo momento più di ogni altra cosa al mondo, ho commesso errori, ho rifiutato consigli e ne ho accettati altri per poter essere qui oggi.

Ho avuti alti e bassi in questi anni, alternando momenti di smisurato entusiasmo ad altri di profondo odio nei confronti di questo percorso. In più di qualche occasione ho temuto di non farcela e spesso avrei voluto mollare la presa, eppure eccomi qui oggi a cercare le parole adatte a chiunque abbia contribuito a rendere questi tre anni l'esperienza più forte e significativa della mia ancora breve vita.

Dedico questa tesi ai miei genitori. Grazie per aver sempre creduto in me e per avermi permesso di inseguire il mio sogno. Grazie per avermi dato la possibilità di scegliere, di sbagliare e poi di ritornare sui miei passi. Vi voglio bene, so che non lo dico spesso, ma questo traguardo è nostro, perché sono la donna che sono grazie a voi.

Grazie a nonna per esserci sempre, anche se questo a volte significa trovare 15 telefonate perse sul telefono. Quelle telefonate ti hanno resa famosa anche all'estero e sono l'esempio perfetto di tutto l'amore che hai per me.

A mia sorella, la mia migliore amica, la donna che mi ha dimostrato cosa significhi avere il coraggio di inseguire i propri sogni sempre, a qualsiasi età e nonostante le parole di sconforto. So che sei la mia più grande sostenitrice e io sarò sempre la tua. Non smettere mai di lottare e segui sempre l'istinto perché quello non sbaglia mai.

Alle mie sorelle acquisite. Rossana, grazie per essere presente e attenta, un po' mamma e un po' amica. Giorgia, grazie per la follia contagiosa e per la grandissima forza con cui affronti la vita ogni giorno. Siete le sorelle Bella e

Che Mi Capisce da ormai più di dieci anni e non avrei potuto desiderare niente di meglio.

Alla Professoressa Eufemi, grazie per avermi dato la possibilità di essere qui oggi a presentare questo progetto, per avermi permesso di farne parte sin dal principio. Aver contribuito col mio lavoro a portare avanti questa grande battaglia è stato e sarà sempre un grande onore per me.

Al professore Altieri e alla professoressa Chichiarelli, grazie per i consigli, per la disponibilità ogni volta che ho avuto bisogno di un aiuto e grazie per i caffè e le chiacchierate rigeneranti.

Grazie a tutti i ragazzi che ho incontrato in questi tre anni. Non dimenticherò mai quello che ho imparato tra queste quattro mura. Porterò con me ogni momento di questo viaggio, i sorrisi e i momenti bui perché entrambi mi hanno reso la persona che sono oggi.

Ad Antonella, semplicemente grazie per esserci stata quando più ne avevo bisogno. Mi hai mostrato cosa significhi fare questo lavoro con passione smisurata ed estrema dedizione. Ti ringrazio per essere stata un uragano di positività e gentilezza, in perfetto equilibrio tra rigore e dolcezza. Grazie perché in quel laboratorio ho ritrovato la passione che avevo perduto e ho capito che tipo di ricercatrice voglio diventare. E grazie ovviamente alla Professoressa Mazzanti per avermi dato la possibilità concreta di portare a termine questo percorso.

A Maddalena, grazie per esserci stata sempre, per esserti aperta con me e per essere diventata la meravigliosa amica che sei oggi. Sono fiera di te, del coraggio e della forza disumana che hai, della tenacia con cui hai intrapreso

questo percorso nonostante chi ti abbia remato contro e scoraggiata. Continua a crescere e a far vedere a tutti di che pasta sei fatta.

A Flavia, la più grande conquista di questi tre anni di dottorato. Grazie per la tua intelligenza intellettuale e umana. Grazie perché mi hai avvicinata e pian piano conquistata. Ti sei lasciata scoprire poco per volta e ti sei dimostrata per quella che sei realmente. Una persona genuina e con un cuore d'oro e una grande, grandissima amica. Senza la tua presenza e il tuo sostegno, senza i tuoi consigli e le tue tirate d'orecchie, questo percorso non sarebbe stato lo stesso; e diciamocelo pure, in più di qualche occasione sarebbe stato un vero inferno. La vita però ha deciso di far girare la mia ruota e dopo avermi portato via degli amici me ne ha fatti incontrare altri, meravigliosi e autentici.

Valeria, tu sei una di questi (pochi ma buoni) nuovi amici che la vita mi ha dato. Una volta mi hai detto che con te sono riuscita ad abbattere i muri che ho alzato per proteggermi dalle persone. Questo mi ha permesso di scoprirti e di riscoprire la nostra amicizia. Non c'è cosa che mi renda più felice. Quello che abbiamo è speciale perché l'abbiamo costruito dal nulla e l'abbiamo consolidato nonostante centinaia di chilometri di distanza a separarci, con periodiche ed eterne videochiamate che in un secondo annullavano ogni distanza.

Caterina, l'ultima sopravvissuta a questo covo di matti, la ragazza cazzuta che ha ascoltato ogni mio sfogo e mi ha accompagnata negli ultimi mesi, forse i più faticosi di questo percorso. Grazie per esserci ancora e per essere mia amica. Sono sicura che questo per noi sia solo l'inizio.

Dedico a voi, amiche mie, i ricordi più belli di questi tre anni. In fondo sono gli unici momenti di cui valga la pena fare tesoro.

Ai miei amici Poratci dico grazie perché dopo quasi nove anni siamo ancora qui gli uni per gli altri. Sono rimasti i migliori, quelli che ad oggi posso davvero chiamare amici. A voi devo forse il ringraziamento più grande perché è grazie ad alcuni di voi se sono tornata in carreggiata due anni fa. Non dimenticherò mai le parole dure e sincere che mi avete urlato in faccia quella sera perché sono state la doccia fredda di cui avevo bisogno per sbloccarmi e per capire che la passione per questo lavoro è la sola cosa necessaria per andare avanti. So che non sono un'amica infallibile ma voglio che sappiate che faccio del mio meglio e che il bene che provo per ognuno di voi non riuscirei mai a raccontarlo a parole.

Grazie infinite a tutti quelli che hanno incrociato la mia strada nell'ultimo anno. Copenaghen è diventata la mia seconda casa, il mio porto sicuro, la città che mi ha accolta a braccia aperte e mi ha scaldato il cuore, nonostante le temperature rigide. Grazie a questo meraviglioso viaggio sono nata la seconda volta e sono diventata la versione migliore di me stessa, almeno fino a questo momento. Grazie alla mia famiglia danese (e per un quinto tedesca), le mie coinquiline, per essere state calorose e affettuose e per avermi fatta sentire a casa sempre. A Vesterbrogade ho trovato delle sorelle e per questo non smetterò mai di esservi riconoscente.

Grazie a Valeria, che è da sempre un tassello fondamentale della mia vita ma che lo è diventata ancora di più durante questa folle avventura. Grazie per avermi ricordato che si può fare sempre meglio e che la nostra amicizia può essere molto più di quanto non fosse già prima. Ti voglio bene e ti sono grata e non sarò mai in grado di spiegarti quanto.

Ole Hartvig Mortensen. Un semplice grazie non sarebbe sufficiente e tutte le parole del mondo suonerebbero scontate e banali. Dovrei dedicarti un intero capitolo di questa tesi per spiegare cosa sia significato per me lavorare con un professionista come te. Mi limiterò a ringraziarti per la smisurata fiducia che

hai riposto in me fin dal primo giorno di lavoro e per aver capito, in così poco tempo, chi sono veramente e quanto io sia disposta a fare per realizzare questo piccolo grande sogno. Grazie per aver silenziosamente e spesso inconsapevolmente raccolto i pezzi di me che lasciavo per strada e per avermi restituito fiducia, in me stessa e nel mio sogno. Sei stato l'ultimo e fondamentale pezzo del puzzle senza il quale questa tesi non sarebbe stata possibile e con lei la mia rivincita.

Spero la vita non separi mai del tutto le nostre strade perché so che ho ancora tanto da imparare da te.

Alle mie amiche di sempre, come sempre, grazie per essere costantemente presenti e sempre al 100% voi.

Infine, ma non per importanza, grazie a Giacomo. Sei il regalo più bello che potessi ricevere, la persona che dovevo incontrare per capire che c'è ancora speranza e che a volte semplicemente ne vale pena. Grazie per non esserti arreso con me e per essere l'uomo meraviglioso che sei.

Che questo traguardo sia solo l'inizio della prossima grande avventura.

Vi voglio bene,

Stefania.