Parkinson Disease Genetics: A "Continuum" from Mendelian to Multifactorial Inheritance

S. Petrucci^{1,2}, F. Consoli¹ and E.M. Valente^{*,1,3}

Abstract: Parkinson Disease (PD) is a common neurodegenerative disorder of intricate etiology, caused by progressive loss of aminergic neurons and accumulation of Lewy bodies. The predominant role of genetics in the etiology of the disease has emerged since the identification of the first pathogenetic mutation in SNCA (alpha-synuclein) gene, back in 1997. Mendelian parkinsonisms, a minority among all PD forms, have been deeply investigated, with 19 loci identified. More recently, genome wide association studies have provided convincing evidence that variants in some of these genes, as well as in other genes, may confer an increased risk for late onset, sporadic PD. Moreover, the finding that heterozygous mutations in the GBA gene (mutated in Gaucher disease) are among the strongest genetic susceptibility factors for PD, has widened the scenario of PD genetic background to enclose a number of genes previously associated to distinct disorders, such as genes causative of spinocerebellar ataxias, mitochondrial disorders and fragile X syndrome. At present, the genetic basis of PD defines a continuum from purely mendelian forms (such as those caused by autosomal recessive genes) to multifactorial inheritance, resulting from the variable interplay of many distinct genetic variants and environmental factors.

Keywords: Genetics, monogenic, multifactorial, Parkinson disease, parkinsonism, risk factor.

INTRODUCTION

Parkinson disease (PD) is the second most frequent neurodegenerative disorder after Alzheimer disease in aged populations, with a prevalence rate reaching up to 3% by age 75 years [1]. The main clinical features of PD are resting tremor, rigidity, bradykinesia and postural instability, but non-motor features such as cognitive decline, neuropsychiatric disturbances and autonomic failure often coexist with motor impairment. Pathological hallmarks of the disease are the degeneration of aminergic neurons in the substantia nigra pars compacta and other brain areas, as well as the deposition of alpha-synuclein and other proteins within intracytoplasmic inclusions known as Lewy bodies (LBs) [2, 3].

A genetic predisposition in PD has long been suspected, based on the detection of positive family history in up to 20% of patients [4]. However, only in recent years the role of genetics in PD has been deeply investigated, leading to major discoveries that have greatly improved knowledge of the disease basis. Through linkage studies, positional cloning strategies and high throughput techniques, 19 loci and 15 genes have been linked to monogenic autosomal dominant (AD) or autosomal recessive (AR) forms of PD (Table 1),

At present, the etiology of PD is thought to be multifactorial, resulting from the variable interplay of distinct genetic and environmental factors. The contribution of these factors seems to be inversely prevalent in the different forms of PD, spanning a broad spectrum where monogenic and idiopathic PD are at the opposite ends (Fig. 1). This review aims to discuss the contribution of genetics in determining PD phenotypes, from the highly penetrant autosomal recessive and dominant forms to the more complex scenario in which genetic variations in distinct genes variably influence the susceptibility to develop the disease.

MENDELIAN FORMS OF PD

Autosomal Recessive PD and Parkinsonisms

Among those parkinsonisms caused by recessively inherited mutations, three genes (PARK2/Parkin, PARK6/PINK1, PTEN-induced kinase 1 and PARK7/DJ-1, Daisuke-Junko-1) have been identified as causative of pure PD phenotypes [6-8]. The main

¹Neurogenetics Unit, CSS-Mendel Laboratory, IRCCS Casa Sollievo della Sofferenza, San Giovanni Rotondo, Italy

²Department of Experimental Medicine, "Sapienza" University of Rome, Rome, Italy; ³Department of Medicine and Surgery, University of Salerno, Salerno, Italy

that are collectively responsible only for less than 10% cases [5]. More recently, whole-genome association studies (GWAS) have identified polymorphic variants in several genes as susceptibility factors for the sporadic form of the disease, and mutations in genes apparently unrelated to PD, such as the *glucocerebrosidase A* (*GBA*) gene that is mutated in Gaucher's disease, have also been discovered as important risk factors.

^{*}Address correspondence to this author at the Neurogenetics Unit, CSS-Mendel Institute, viale Regina Margherita 261, 00198 Rome, Italy; Tel: +39 06 4416 0537; Fax: +39 06 4416 0548; E-mail: e.valente@css-mendel.it

Table 1. Mendelian PD genes and loci.

Locus	Chromosome	Gene	Inheritance	PD Phenotype	Mutation Types	MIM			
Typical PD									
PARK1/PARK4	4q21-22	SNCA	AD	Variable onset PD	point mutations, whole gene multiplications	163890			
PARK2	6q25.2-q27	PRKN	AR	Early onset PD	point mutations, exonic rearrangements	602544			
PARK6	1p36	PINK1	AR	Early onset PD	point mutations, exonic rearrangements	608309			
PARK7	1p36	DJ-1	AR	Early onset PD	point mutations, exonic rearrangements	602533			
PARK8	12q12	LRRK2	AD	Variable onset PD	point mutations	609007			
PARK17	16q11.2	VPS35	AD	Variable onset PD	point mutations	601501			
PARK18	3q27.1	EIF4G1	AD	Late onset PD	point mutations	600495			
Atypical Parkinsonisms									
PARK9	1p36	ATP13A2	AR	KRS	point mutations 6				
PARK14	22q13.1	PLA2G6	AR	Juvenile dystonia-parkinsonism	point mutations	603604			
PARK15	22q12-q13	FBXO7	AR	Pallido-pyramidal syndrome	point mutations	605648			
PARK19	1p31.3	DNAJC6	AR	Juvenile parkinsonism*	point mutations	608375			
PARK20	21q22.11	SYNJ1	AR	Juvenile parkinsonism*	point mutations	604297			
Putative PD-As	Putative PD-Associated Loci and Genes								
PARK3	2p13	SPR?	AD	Late onset PD	- 6				
PARK5	4p14	UCHL1	AD	Late onset PD	point mutations	191342			
PARK10	1p32	unknown	unclear	Late onset PD	-				
PARK11	2q37.1	GIGYF2	AD	Late onset PD	point mutations				
PARK12	Xq21-q25	unknown	unclear	Late onset PD	- 3				
PARK13	2p12	Omi/HtrA2	unclear	Late onset PD	point mutations	606441			
Non-PD Genes	Causative of PD) Phenotype	s						
not assigned	12q24.1	ATXN2	AD	Late onset PD	CAG expansions	601517			
not assigned	14q32.12	ATXN3	AD	Late onset PD	CAG expansions				
not assigned	Xq27.3	FMR1	X-linked	Late onset PD	CGG expansions (41-54, gray zone)				
not assigned	9p21.2	C9orf72	AD	Late onset PD	GGGGCC expansions				
not assigned	15q26.1	POLG1	AR/AD	Late onset parkinsonism	point mutations 17				
not assigned	10q24.31	Twinkle	AD	Variable onset parkinsonism	point mutations 60				
not assigned	1q21	GBA	risk factor/AD*	Variable onset PD	point mutations	606463			
direction of the contract of t									

AD, autosomal dominant; AR autosomal recessive; MIM: Mendelian Inheritance in Man online catalogue; PD, Parkinson disease; KRS, Kufor Rakeb Syndrome; considering the small number of reported cases and their clinical variability, the phenotypic spectrum related to these genes still has to be delineated (see text for details); "given the very high Odd's Ratio (>5), heterozygous mutations in the GBA gene can be considered as autosomal dominant with very low penetrance.

distinctive feature of these forms is represented by the early age at onset, slow progression and good response to Levodopa therapy. Other genes (PARK9/ATP13A2; PARK14/PLA2G6, phospholipase A2, group VI and PARK15/FBXO7, F-box only protein 7) are known to cause AR atypical parkinsonisms, that also present early age at onset but are characterized by a more rapidly progressive, complex phenotypes, in which parkinsonian signs are variably associated to other neurological features.

More recently, autosomal recessive mutations in two other genes (DNAJ/HSP40 homolog, subfamily C member 6, *DNAJC6*, and Synaptojanin 1, *SYNJ1*) have been described in members of consanguineous

families with juvenile parkinsonism, often associated to other neurological signs [9-12]. In all these forms, penetrance is usually complete and age-dependent.

Autosomal Recessive "Pure" Early Onset PD

Exonic rearrangements and/or point mutations in the *Parkin* gene are the commonest genetic alterations found in pure early onset PD (<40-45 years), with an overall mutation frequency of about 8-9%. Mutations in the *PINK1* and *DJ-1* genes are rarer, being identified in 3-4% and <1% of early onset PD, respectively [13].

Pathologically, a significant loss of dopaminergic neurons in the *substantia nigra* and *locus coeruleus*, without LBs in all but rare cases, have been described

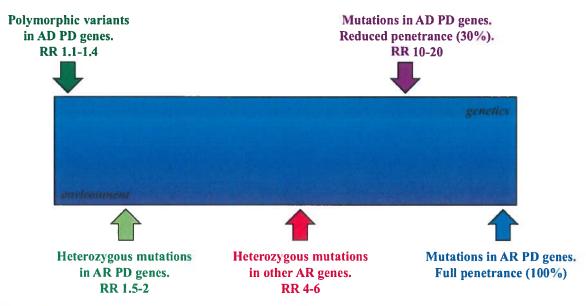


Fig. (1). Genetic factors associated to Parkinson disease and relative risks (RR).

in Parkin-mutated PD brains. At difference, the only PINK1 autoptic case reported so far was more similar to idiopathic PD, with nigral neuronal loss and LB pathology [14, 15]. No neuropathological data are yet available for DJ-1 mutated patients.

The occurrence of Parkin biallelic mutations is inversely correlated to the age at onset of parkinsonism: the earlier the onset, the higher the probability to detect mutations in this gene, with mean onset ages around 30 years. Other typical features of the Parkin-related phenotype are a slow and benign progression of the disease, usually without cognitive or vegetative impairment, and a good and long-lasting response to Levodopa and dopamine agonists. However, motor fluctuations and Levodopa-induced diskynesias may occur. A symmetrical presentation of symptoms, dystonia at onset, hyperreflexia, sleep benefit and psychiatric disturbances may also be present [16]. In PINK1-related parkinsonism, mean age at onset is in the fourth decade (usually later than Parkin); atypical features such as dystonia at onset, hyperreflexia and diurnal fluctuations are rarer, but psychiatric disturbances have been reported in a substantial subset of mutated patients [17]. Finally, only few patients with DJ-1 biallelic mutations have been identified to date: the phenotype seems to be characterized by a very early onset (in the twenties), with frequent occurrence of dystonia at onset that may have atypical distributions such as cervical dystonia and blepharospasm, and psychiatric disturbances [18, 19].

Autosomal Recessive Atypical Early Onset **Parkinsonisms**

These atypical early onset parkinsonisms are characterized by the association with other neurological

signs, such as dystonia, spasticity, dementia, and abnormal ocular movements.

Kufor Rakeb syndrome (KRS), caused by mutations in PARK9/ATP13A2 gene, is a rare pallido-pyramidal syndrome presenting with juvenile Levodoparesponsive parkinsonism (average onset age in the second decade), associated to supranuclear gaze palsy, pyramidal signs, mini-myoclonus of the face and fingers, dementia, and progressive brain atrophy [20, 21]. Although only few mutated patients have been reported to date, there is phenotypic variability, insofar patients have been described presenting with later ages at onset, subtle parkinsonism, ataxia and axonal neuropathy, or variable neuroimaging features including iron brain deposition, absence of atrophy, or degenerative cerebellar involvement [22]. Surprisingly, a ATP13A2 homozygous mutation was recently detected in a family with neuronal ceroid lipofuscinoses, a metabolic storage disease, further widening the phenotypic spectrum of this gene [23]. Heterozygous ATP13A2 mutations have been reported in rare patients with early onset pure PD, implicating this gene as a possible susceptibility factor for idiopathic PD [24]. Besides KRS, two other genes (PARK14/PLA2G6 and PARK15/FBXO7) cause atypical early onset forms of parkinsonism. Mutations in PLA2G6 are responsible for infantile neuroaxonal dystrophy and neurodegeneration with brain iron accumulation (NBIA) but, in rare cases, they can be found in patients with a Levodopa-responsive form of dystonia-parkinsonism that is rapidly complicated by the occurrence of cognitive impairment, psychiatric disturbances and pyramidal signs [25]. Conversely, the rare FBX07-related disease is primarily a pyramidal syndrome without cognitive impairment, in which levodopa-responsive parkinsonian features appear well after the onset of spasticity [26].

Novel AR PD Genes: PARK19/DNAJC6 and PARK20/SYNJ1

Using homozygosity mapping and whole exome sequencing, pathogenetic biallelic mutations in the DNAJC6 gene have been detected in patients with autosomal recessive juvenile parkinsonism, either isolated or associated to other neurological features such as mental retardation, pyramidal signs and epilepsy [9, 10]. More recently, homozygous mutations in the SYNJ1 gene have been identified in patients from a consanguineous Italian family parkinsonism, dystonia and cognitive deterioration and in an Iranian kindred with early onset PD and generalized epilepsy [11, 12]. Despite the limited number of cases reported to date, a wide clinical variability seems to characterize both these two novel forms of parkinsonism. More studies are required in order to better delineate the DNAJC6 and SYNJ1 related phenotypes and the role of these genes in the pathogenesis of PD.

Autosomal Dominant PD

To date, at least eight genes and loci have been linked to AD PD, but only few of them (PARK1-PARK4/SNCA, alpha synuclein; PARK8/LRRK2, leucine-rich repeat kinase 2; PARK17/VPS35, vacuolar protein sorting 35 and PARK18/EIF4G1, eukaryotic translation initiation factor 4-gamma 1) have been unequivocally proved to be causative of the disease; conversely, the pathogenic role of other genes (such as PARK5/UCHL1, ubiquitin carboxyl-terminal esterase L1; PARK11/GIGYF2, GRB10-interacting gyf protein 2 and PARK13/HTRA2, HTRA serine peptidase 2) still remains controversial. While SNCA and LRRK2 genes have been studied in depth, available data are still scarce for VPS35 and EIF4G1 genes, that were more recently identified.

Excluding rare exceptions, AD parkinsonisms share common features, such as a later age at onset compared with AR PD, the occurrence of cognitive impairment of variable degree, and incomplete penetrance [27].

In terms of neuropathology, LBs have been detected in brains from patients with AD PD, with the exception of *VPS35* mutated cases, for whom data are still lacking. However, pathology is highly variable in LRRK2-related PD, in which LBs can be absent or present with variable distribution in the brain, possibly associated to ubiquitine positive inclusions as well as Tau pathology [28, 29].

PARK1- PARK4 / SNCA (Alpha Synuclein)

Five distinct point mutations, as well as whole duplications or triplications of the SNCA gene, have been detected in a few patients with AD PD. The SNCA p.A53T change was the first genetic mutation to be identified in a large Italian PD kindred with dominant

inheritance back in 1997 (the Contursi kindred), opening up an entirely new avenue of research on PD pathophysiology [30]. The same mutation was subsequently found in several Greek/Italian families all sharing a common ancestor, in two other PD families of Korean and Swedish origin and in an apparently sporadic Polish case. The phenotype of mutated patients carrying the p.A53T mutation ranged from typical late-onset PD to atypical PD with more severe features, such as earlier age at onset, rapid progression, and high prevalence of cognitive, psychiatric and autonomic impairment [30-35]. The other four SNCA mutations have been reported: i) in a single German family with PD (p.A30P) [36]; ii) in several Basque families with a severe form of early onset parkinsonism or Lewy body dementia (p.E46K) [37]; iii) in a unique familial case of Caucasian origin (p.H50Q) [38, 39]; iv) in a three-generation French PD pedigree with early onset and rapidly progressive parkinsonism associated to frequent psychiatric disturbances and marked pyramidal signs (p.G51D)

At difference from point mutations, that are extremely rare, *SNCA* locus multiplications represent a more frequent cause of AD PD. Triplications of the whole gene have been described in few families presenting a severe form of early onset parkinsonism, while *SNCA* duplications are even commoner, being reported in several familial and sporadic PD cases worldwide [41-43].

In this genetic condition, the disease severity appears to correlate well with the dosage of the *SNCA* gene, rather than with the extension of the multiplicated genomic region. Indeed, the presence of four *SNCA* copies is always causative of a fully penetrant, aggressive and rapidly progressive phenotype, with early onset (usually in the third to fourth decade) of parkinsonian signs and precocious non motor features (dementia, psychiatric disturbances and dysautonomia) [44-46]. Conversely, in patients with *SNCA* duplications the disease presentation is highly variable, even within families: in some patients it resembles idiopathic lateonset PD while others are more similar to patients bearing *SNCA* triplications [43, 47, 48]. Penetrance is estimated to be up to 30%, as several healthy carriers of the *SNCA* duplication have been reported [49-52].

PARK8 / LRRK2

At difference from SNCA-related phenotypes, the parkinsonian phenotype caused by LRRK2 is usually more similar to idiopathic PD. The LRRK2 gene comprises 51 exons, and more than 100 missense been described variations have (http://www.molgen.ua.ac.be/PDmutDB). However, only seven nucleotide changes are considered surely pathogenetic (p.N1437H, p.R1441C, p.R1441G, p.R1441H, p.Y1699C, p.G2019S and p.I2020T), based on functional studies and segregation analysis in large families [53]. Among these mutations, the p.G2019S missense change is the most frequent due to a founder effect, with variable frequencies worldwide that seems to decrease with increasing distance from the

Mediterranean areas. In fact, its prevalence reaches up to 40% among the Ashkenazi Jewish and the North African Arab communities, while it has been detected in about 5-7% of familial and about 0,5- 2% of sporadic cases in the Caucasian populations [54, 55]. There is incomplete age-dependent penetrance, ranging from 28% at age 59 to 74% at age 79, as demonstrated by the detection of this mutation in many sporadic patients, in pedigrees with unconventional patterns of inheritance and in aged healthy subjects [56]. LRRK2related phenotype presents a variable age at onset (ranging from the fourth to the eight decade); unilateral tremor is often the initial sign of disease, and response to treatment is good and sustained, although phenotypic variability has been observed even within families.

Novel AD PD Genes: PARK17 /VPS35 and PARK18/ EIF41G

Recently, two novel PD-causing genes, VPS35 and EIF4G1, have been identified using high throughput strategies. The VPS35 p.D620N mutation has been recurrently detected in tremor-dominant PD familial cases from different ethnicities [57, 58], with age at onset ranging from 40 to 52 years and a relative frequent occurrence of cognitive impairment. Penetrance seems to be reduced and age dependent. Other missense variants have been identified in very few cases, but their pathogenetic significance still remains undetermined [58-60]. Since the estimated prevalence is 0,4%, VPS35-related parkinsonism can be considered a rare cause of AD PD [61].

Similarly to VPS35, the pathogenetic role of EIF4G1 mutations in PD is still not well defined. Among the genetic variations identified in PD patients, only the p.R1205H substitution was found to clearly cosegregate with the disease in nine families, manifesting as a late onset parkinsonism with slow progression, good response to Levodopa therapy and spared cognitive functions in all but a few cases [60, 62]. The frequency of the p.R1205H mutation was estimated to be about 0.2% in European and African PD cohorts. while mutation screening studies failed to find EIF4G1 mutations in other ethnicities [63, 64]. Asymptomatic carriers have been reported, some older than 80 years [60].

Other AD PD Genes Requiring Genetic Validation

Other AD loci and genes have been associated to PD (Table 1), but their pathogenicity was not confirmed in subsequent studies. In the PARK3 locus (mapped to chromosome 2p13 by linkage analysis performed in large AD PD families), the underlying causative gene has not been identified yet. A single gene mapping within the region, the sepiapterin reductase (SPR) gene (involved in dopamine synthesis) has been possibly implicated in PD, but the presence of other causative or susceptibility genes for late-onset PD within this region cannot be excluded [65-69]. The PARK5/UCHL1 (mapping to chromosome 4p14) and PARK11/GIGYF2 (chromosome 2q) genes initially seemed good PD candidates based on the identification of potentially

pathogenic mutations in few PD familial cases; however, further screenings in large cohorts of patients failed to identify additional pathogenetic variants [70-73]. Finally, the mitochondrial serine protease gene (PARK13/Omi/HTRA2) has been proposed as a possible candidate since a genetic variant (p.G399S) in this gene was found significantly over-represented in PD patients compared to controls; these findings were not confirmed in subsequent studies, the same variant being detected at a similar frequency in patients and controls [74-76].

Non-PD Related Genes Causative of Mendelian **Parkinsonisms**

Parkinsonism may be the phenotypic expression of mutations in genes that are usually related to other neurologic diseases. Trinucleotide repeat expansions in the ATXN2 (ataxin-2) or ATXN3 (ataxin-3) genes, that cause two forms of spinocerebellar ataxia, have been found in familial cases with pure Levodoparesponsive PD [77]. Other interesting examples are the trinucleotide repeat expansions in the FMR1 (fragile X mental retardation protein) gene on chromosome X, that are responsible of different phenotypes, such as mental retardation in males (>200 CGG repeats, full mutation) or FXTAS, the Fragile X tremor/ataxia syndrome, in both genders (45-54 CGG repeats, premutation). A form of parkinsonism, resembling idiopathic PD, may be the initial presentation of FXTAS in premutated subjects, but it can be the unique phenotype in female and male carriers of milder expansions (41-54 CGG, gray zone) [78]. Finally, pathogenetic expansions in the C9orf72 gene, recently identified as causative of familial frontotemporal dementia (FTD) and amyotrophic lateral sclerosis (ASL), have been reported in a few patients with typical PD and a positive family history of other neurodegenerative diseases, including FTD and ALS [79].

A Blurred Boundary Between Causative and Susceptibility Mutations in PD

The GBA Gene

Gaucher disease is an AR lysosomal storage disorder caused by biallelic mutations in the glucocerebrosidase gene (GBA, on chromosome 1q21), encoding the glucocerebrosidase enzyme (GCase). Almost 300 GBA mutations have been identified, but the most frequent cluster is in the catalytic domain of the enzyme [80]. Frequency and distribution of GBA mutations vary among populations, being more common in Ashkenazi Jews (in particular the p.N409S and p. L444P mutations) [81-83]. The intuition of a possible link between this gene and PD first arose from the observation of a frequent occurrence of parkinsonian symptoms in GD patients or in heterozygous relatives, leading to perform molecular screening of the GBA gene in large cohorts of PD patients [81, 84-89]. These studies unraveled an impressive high frequency of GBA heterozygous mutations in patients compared to healthy controls,

ranging from 7% to 15% in various populations. Based on these results, *GBA* heterozygous mutations are now considered to be the commonest genetic risk factor for PD, with an average odds ratio of 5.43 and an estimated cumulative risk for PD of 2.2% by age 65 and of 10.9% by age 85 [81, 90]. Indeed, based on these high relative risk values, heterozygous *GBA* mutations can be considered more than susceptibility factors, gaining the same genetic weight as autosomal dominant mutations with reduced penetrance [91]. Moreover, *GBA* mutations have been recently proposed also as modifiers of PD progression, increasing four fold the risk to reach a higher Hoehn and Yahr stage, and five-fold the risk to develop earlier cognitive impairment and eventually dementia [92].

Compared to idiopathic PD, patients with GBA mutations have on average an earlier age at onset, more symmetrical clinical signs, and an increased occurrence of non-motor symptoms, including cognitive impairment, neuropsychiatric disturbances automomic dysfunctions [93]. The parkinsonian phenotype observed in GBA heterozygous carriers can be heterogeneous, ranging from classical late-onset levodopa-responsive PD to more severe presentations consistent with the diagnosis of PD-dementia and Dementia with Lewy body [81]. Not surprisingly, abundant q-synuclein inclusions and prominent diffuse Lewy bodies-type pathology have been found in brains from GBA-mutated PD patients [94]. Recently, a widespread decrease of GCase catalytic activity and of related protein levels have been reported in brains of PD patients either with or without GBA mutations, leading to increased a-synuclein aggregation as a result of the compromised neuronal lysosomal activity. In turn, these increased levels of a-synuclein may inhibit intracellular trafficking and lysosomal function of normal GCase, suggesting a bidirectional, positive feedback loop between α-synuclein accumulation and GCase deficiency [95, 96].

Mitochondrial Parkinsonisms

There is a close tie between mitochondrial dysfunction and the risk to develop PD and parkinsonism: for instance, high levels of mitochondrial DNA (mtDNA) deletions were detected in the substantia nigra of PD patients [97] and, on the other hand, parkinsonian signs may frequently occur in mitochondrial diseases (MDs) caused by mutations or deletions in mitochondrial genes, such as ND1, ND2 and ND3, the mtDNA tRNA(Lys), the mtDNA tRNA(Gln), Cytb and Ng [98].

A parkinsonism partially responsive to Levodopa has been occasionally reported in patients with single or biallelic mutations in *POLG1* (*mitochondrial DNA polymerase gamma*), a nuclear gene that, when altered, causes multiple deletions in the mtDNA. However, additional signs suggestive of an underlying mitochondrial defect (such as external progressive ophthalmoplegia, ataxia, neuropathy and miopathy) often coexisted with PD in these patients [99]. PD features were also described in members of a family

with autosomal dominant external progressive opthalmoplegia caused by a missense mutation in the *Twinkle* (*C10ORF2*) gene, encoding a mitochondrial DNA helicase involved in the maintenance of the mtDNA stability [100].

Besides their pathogenetic role in complex forms of parkinsonism, these nuclear and mitochondrial genes may play as risk factors in the pathogenesis of PD. The POLG1 polymorphic poliglutamine tract (poly-Q), commonly encoded by 10 or 11 repetitions of the CAG triplet, has been associated to PD in the presence of non-10/11Q, although controversially in different studies [99]. Even mitochondrial haplogroups (characterized by common polymorphisms evolved from the same ancestor) have been deeply investigated in relation to PD pathogenesis, and the protective role of haplogroups J and T has been confirmed by a recent metanalysis study [101, 102].

Genetic Risk Factors for PD

We only have crumbs of knowledge about the complex environmental-genetic interactions that may cause the common form of "idiopathic" PD. Genetic research has long been attempting to unravel this intricate interplay, through case-control association studies and, more recently, GWAS. Moreover, recent evidence has pointed towards a potential role of heterozygous mutations in AR PD genes as risk factors, by detecting endophenotypes in healthy carriers.

Heterozygous Mutations in AR PD Genes

While biallelic mutations in genes such as Parkin. PINK1 and DJ-1 have been unequivocally linked to AR early onset parkinsonism, the role of single heterozygous mutations in these genes is still controversial and debated. Extensive mutation screenings of these genes have shown that a substantial proportion of patients only carried a single heterozygous variant, and such variants were occasionally detected also in healthy controls [103]. Bearing in mind that PD is a common condition, the presence of these single mutations might be accidental and unrelated to the disease. However, cumulative evidence from many studies and comparison of the frequencies of these mutations among cohorts of patients and controls, now suggests that they may represent minor susceptibility factors that could mildly contribute to the risk of sporadic PD (Parkin odds ratio 2,53; PINK1 odds ratio 1,65) [104, 105]. Interestingly, some healthy subjects carrying heterozygous Parkin or PINK1 mutations were found to present mild signs of parkinsonism not fulfilling the diagnostic criteria for clinically definite PD, or subclinical signs definable as endophenotypes (e.g. abnormal responses neurophysiological testing, hyperechogenicity of the substantia nigra, nigrostriatal dysfunction on functional neuroimaging, discrete abnormalities in voxel-based morphometric analyses, and so on). However, no evidence of progression to classical PD could be observed in the majority of these cases [106]. Further

studies are needed, in particular neurological follow up of healthy heterozygous carriers, in order to establish the real impact of such variants on the disease susceptibility [106].

Polymorphic Variants in Autosomal Dominant PD

Before the advent of whole genome techniques, casecontrol association studies of selected polymorphisms within candidate genes had been largely adopted to search for genetic susceptibility factors of PD. However, only a few of the proposed associations could be replicated and confirmed in other populations or in metaanalysis. Among these, several variants in the SNCA gene, including the NACP-Rep1 polymorphism and variants in the 3'UTR region, have been consistently associated to an increased PD risk [107, 108]. The H1 haplotype of the MAPT gene, encoding the microtubule associated protein tau, has been recognized as a PD risk factor with an odds ratio of 1.5, while two common variants in LRRK2 (p.G2385R and p.R1628P) increased the risk of PD about two-fold, particularly in Asian populations. Intriguingly, the most reliable risk factors resulted to be polymorphic variants within the same genes mutated in monogenic forms of PD or other neurodegenerative diseases, establishing a direct link between the pathogenesis of familial and sporadic forms of PD [109].

Other Genetic Risk Factors for PD

Thanks to GWAS, an increasingly popular approach

to identify genetic factors influencing complex traits, and the creation of large patients samples consortia from many research PD study groups worldwide, association studies have highly reinforced their capability in finding low risk variants. To date, multiple GWAS and three different meta-analyses have been published. Besides confirming the ascertained associations with SNCA, MAPT and LRRK2 variants, these studies have highlighted many other genes and loci implicated in genetic PD susceptibility (Table 2) [110-112]. However, the combined population-attributable risk across all identified loci was 60.3% and 25.6% for the MAPT and SNCA loci alone, confirming the strong influence of these two genes on PD susceptibility.

CONCLUSION

Over the last decade, impressive evidence has highlighted a central role for genetic factors not only in determining the probability to develop PD, but also influencing the disease onset, progression and phenotypic manifestation. Rare highly penetrant pathogenic mutations and more common susceptibility variants in several distinct genes variably interplay with still largely unknown environmental factors to eventually determine if, when and how a single individual will become affected. Despite this tremendous progress, our knowledge is still largely incomplete, and it is foreseeable that many additional genetic determinants will have to be identified. The advent of innovative next

Table 2. Genetic risk factors in sporadic Parkinson Disease (from published GWAS).

Chromosome	Gene	Risk Variants	Odds Ratios (OR)	References
12q12	LRRK2 (PARK8)	G2385R, R1628P	2-3*	[5]
1q22	SYT11	SNPs	1,43	[110]
17q21.31	MAPT	H1 haplotype	1,4	[5]
4p16.3	GAK	SNPs	1,35	[112]
4q21-22	SNCA (PARK1/PARK4)	Rep1; 5' and 3' UTR variants; SNPs	1,2-1,4	[107, 108, 112]
18q12.3	RIT2	SNPs	1,2	[112]
2q24.3	STK39	SNPs	1,19	[111]
12q24.31	CCDC62/HIP1R	SNPs	1,15	[111]
16p11.2	STX1B	SNPs	1,14	[111]
4p15	BST1	SNPs	1,1	[112]
2q21.3	ACMSD	SNPs	1,02	[110]
6p21.32	HLA-DRB5	SNPs	0,95 -0,98*	[110]
3q27.1	MCCC1/LAMP3	SNPs	0,9	[111]
4q21.1	STBD1	SNPs	0,9	[111]
7p15.3	GPNMB	SNPs	0,89	[111]
8p22	FGF20	SNPs	0,89	[111]
10p13	ITGA8	SNPs	0,88	[111]
1q32	PARK16 locus	SNPs	0,88	[111]

AD: autosomal dominant, GWAs Genome Wide Association Studies, SNP, single nucleotide polymorphism. European population, "Asiatic population

generation sequencing (NGS) techniques is expected to give new acceleration to this research path, with the possibility to sequence the entire exome or even genome of an individual with high efficiency and in a time- and cost-effective way. NGS techniques are now expected to unravel rare pathogenetic variants acting as genetic modifiers of PD risk or progression that, due to their "rare" nature, cannot be picked up by GWAS. A perfect example is the GBA gene, that had never emerged in GWA studies despite being one of the strongest genetic factors influencing PD susceptibility. On the other hand, this innovative approach is unraveling a complexity of the human genome that is much greater than previously thought, with the identification even in the genome of "healthy" individuals of several genetic variants significance remains difficult to decipher. As a consequence, the interpretation of NGS data is going to pose major challenges when trying to link specific genetic variants to the disease risk or to certain phenotypic manifestations. Studies on large cohorts of well-phenotyped patients and guidelines for the analysis and interpretation of sequencing data are sought in order to make the most of this innovative technique to advance our knowledge on pathogenesis and natural history.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

ACKNOWLEDGEMENTS

This work was supported by the Italian Ministry of Health (Ricerca Corrente 2013) and by the European Community FP7 Program (Project MeFoPa).

Financial Disclosures of all Authors for the **Preceding 12 Months**

Prof. Valente serves on the editorial boards of BMC Neurology, Movement Disorders and Pediatric Research. She is the recipient of a European Research Council Starting Grant, and of research grants from the Italian Ministry of Health, the Italian Ministry of University and Research, the Telethon Foundation Italy and the European Community FP7 Program.

Dr. Petrucci reports no disclosures.

REFERENCES

- De Lau LM, Breteler MM. Epidemiology of Parkinson's disease. Lancet Neurol 2006; 5(6): 525-35
- Dauer W, Przedborski S. Parkinson's disease: mechanisms and models. Neuron 2003; 39(6): 889-909.
- [3] Braak H. Del Tredici K. Rub U. de Vos RA, Jansen Steur EN. Braak E. Staging of brain pathology related to sporadic Parkinson's disease. Neurobiol Aging 2003; 24(2): 197-211.
- [4] Bonifati V, Fabrizio E, Vanacore N, De Mari M, Meco G. Familial Parkinson's disease: a clinical genetic analysis. Can
- J Neurol Sci 1995; 22(4): 272-9. Lesage S, Brice A. Role of mendelian genes in "sporadic" [5] Parkinson's disease. Parkinsonism Relat Disord 2012; 18 Suppl 1: S66-70.

- Kitada T. Asakawa S. Hattori N. et al. Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. Nature 1998; 392(6676): 605-8.
- Valente EM, Abou-Sleiman PM, Caputo V, et al. Hereditary early-onset Parkinson's disease caused by mutations in PINK1. Science 2004; 304(5674): 1158-60.
- Bonifati V, Rizzu P, van Baren MJ, et al. Mutations in the DJ-[8] 1 gene associated with autosomal recessive early-onset parkinsonism. Science 2003; 299(5604): 256-9.
- Edvardson S, Cinnamon Y, Ta-Shma A, et al. A deleterious mutation in DNAJC6 encoding the neuronal-specific clathrinuncoating co-chaperone auxilin, is associated with juvenile parkinsonism. PLoS One 2012; 7(5): e36458.
- Koroglu C, Baysal L, Cetinkaya M, Karasoy H, Tolun A. DNAJC6 is responsible for juvenile parkinsonism with phenotypic variability. Parkinsonism Relat Disord 2013; 19(3): 320-4.
- Quadri M, Fang M, Picillo M, et al. Mutation in the SYNJ1 Gene Associated with Autosomal Recessive, Early-Onset Parkinsonism. Hum Mutat 2013; 34(9): 1208-15
- Krebs CE, Karkheiran S, Powell JC, et al. The Sac1 Domain of SYNJ1 Identified Mutated in a Family with Early-Onset Progressive Parkinsonism with Generalized Seizures. Hum Mutat 2013; 34(9): 1200-7.
- Puschmann A. Monogenic Parkinson's disease and
- parkinsonism: clinical phenotypes and frequencies of known mutations. Parkinsonism Relat Disord 2013; 19(4): 407-15. Crosiers D, Theuns J, Cras P, Van Broeckhoven C. Parkinson disease: insights in clinical, genetic and [14] pathological features of monogenic disease subtypes. J Chem Neuroanat 2011; 42(2): 131-41.
- Samaranch L, Lorenzo-Betancor O, Arbelo JM, et al. PINK1-[15] linked parkinsonism is associated with Lewy body pathology. Brain 2010; 133(Pt 4): 1128-42.
- [16] Lohmann E, Thobois S, Lesage S, et al. A multidisciplinary study of patients with early-onset PD with and without parkin
- mutations. Neurology 2009; 72(2): 110-6. Ephraty L, Porat O, Israeli D, et al. Neuropsychiatric and f171 cognitive features in autosomal-recessive early parkinsonism due to PINK1 mutations. Mov Disord 2007; 22(4): 566-9.
- [18] Dekker M, Bonifati V, van Swieten J, et al. Clinical features and neuroimaging of PARK7-linked parkinsonism. Mov Disord 2003; 18(7): 751-7.
- [19] Abou-Sleiman PM, Healy DG, Wood NW. Causes of Parkinson's disease: genetics of DJ-1. Cell Tissue Res 2004; 318(1): 185-8.
- Williams DR, Hadeed A, al-Din AS, Wreikat AL, Lees AJ. Kufor Rakeb disease: autosomal recessive, levodoparesponsive parkinsonism with pyramidal degeneration, [20] supranuclear gaze palsy, and dementia. Mov Disord 2005; 20(10): 1264-71.
- Ramirez A, Heimbach A, Grundemann J, et al. Hereditary parkinsonism with dementia is caused by mutations in ATP13A2, encoding a lysosomal type 5 P-type ATPase. Nat Genet 2006; 38(10): 1184-91.
- Eiberg H, Hansen L, Korbo L, et al. Novel mutation in ATP13A2 widens the spectrum of Kufor-Rakeb syndrome (PARK9). Clin Genet 2012; 82(3): 256-63.
- Bras J, Verloes A, Schneider SA, Mole SE, Guerreiro RJ. [23] Mutation of the parkinsonism gene ATP13A2 causes neuronal ceroid-lipofuscinosis. Hum Mol Genet 2012; 21(12): 2646-50.
- Djarmati A, Hagenah J, Reetz K, et al. ATP13A2 variants in [24] early-onset Parkinson's disease patients and controls. Mov Disord 2009; 24(14): 2104-11.
- Yoshino H, Tomiyama H, Tachibana N, et al. Phenotypic spectrum of patients with PLA2G6 mutation and PARK14linked parkinsonism. Neurology 2010; 75(15): 1356-61.
- [26] Di Fonzo A, Dekker MC, Montagna P, et al. FBXO7 mutations cause autosomal recessive, early-onset early-onset parkinsonian-pyramidal syndrome. Neurology 2009; 72(3):
- Sundal C, Fujioka S, Uitti RJ, Wszolek ZK. Autosomal dominant Parkinson's disease. Parkinsonism Relat Disord [27] 2012; 18 Suppl 1: S7-10.

- [28] Houlden H, Singleton AB. The genetics and neuropathology of Parkinson's disease. Acta Neuropathol 2012; 124(3): 325-38.
- [29] Wider C, Skipper L, Solida A, et al. Autosomal dominant dopa-responsive parkinsonism in a multigenerational Swiss family. Parkinsonism Relat Disord 2008; 14(6): 465-70.
- [30] Polymeropoulos MH, Lavedan C, Leroy E, et al. Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. Science 1997; 276(5321): 2045-7.
- [31] Papadimitriou A, Veletza V, Hadjigeorgiou GM, Patrikiou A, Hirano M, Anastasopoulos I. Mutated alpha-synuclein gene in two Greek kindreds with familial PD: incomplete penetrance? Neurology 1999; 52(3): 651-4.
- [32] Spira PJ, Sharpe DM, Halliday G, Cavanagh J, Nicholson GA. Clinical and pathological features of a Parkinsonian syndrome in a family with an Ala53Thr alpha-synuclein mutation. Ann Neurol 2001; 49(3): 313-9.
- [33] Ki CS, Stavrou EF, Davanos N, et al. The Ala53Thr mutation in the alpha-synuclein gene in a Korean family with Parkinson disease. Clin Genet 2007; 71(5): 471-3.
- [34] Puschmann A, Ross OA, Vilarino-Guell C, et al. A Swedish family with de novo alpha-synuclein A53T mutation: evidence for early cortical dysfunction. Parkinsonism Relat Disord 2009; 15(9): 627-32.
- [35] Michell AW, Barker RA, Raha SK, Raha-Chowdhury R. A case of late onset sporadic Parkinson's disease with an A53T mutation in alpha-synuclein. J Neurol Neurosurg Psychiatry 2005; 76(4): 596-7.
- [36] Kruger R, Kuhn W, Muller T, et al. Ala30Pro mutation in the gene encoding alpha-synuclein in Parkinson's disease. Nat Genet 1998; 18(2): 106-8.
- [37] Zarranz JJ, Alegre J, Gomez-Esteban JC, et al. The new mutation, E46K, of alpha-synuclein causes Parkinson and Lewy body dementia. Ann Neurol 2004; 55(2): 164-73.
- [38] Proukakis C, Dudzik CG, Brier T, et al. A novel alphasynuclein missense mutation in Parkinson disease. Neurology 2013; 80(11): 1062-4.
- [39] Appel-Cresswell S, Vilarino-Guell C, Encarnacion M, et al. Alpha-synuclein p.H50Q, a novel pathogenic mutation for Parkinson's disease. Mov Disord 2013; 28(6): 811-3.
- [40] Lesage S, Anheim M, Letournel F, et al. G51D alphasynuclein mutation causes a novel parkinsonian-pyramidal syndrome. Ann Neurol 2013; 73(4): 459-71.
- [41] Devine MJ, Gwinn K, Singleton A, Hardy J. Parkinson's disease and alpha-synuclein expression. Mov Disord 2011; 26(12): 2160-8.
- [42] Singleton AB, Farrer M, Johnson J, et al. alpha-Synuclein locus triplication causes Parkinson's disease. Science 2003; 302(5646): 841.
- [43] Elia AE, Petrucci S, Fasano A, et al. Alpha-synuclein gene duplication: Marked intrafamilial variability in two novel pedigrees. Mov Disord 2013; 28(6): 813-7.
- [44] Fuchs J, Nilsson C, Kachergus J, et al. Phenotypic variation in a large Swedish pedigree due to SNCA duplication and triplication. Neurology 2007; 68(12): 916-22.
- [45] Ikeuchi T, Kakita A, Shiga A, et al. Patients homozygous and heterozygous for SNCA duplication in a family with parkinsonism and dementia. Arch Neurol 2008; 65(4): 514-9.
- [46] Gwinn K, Devine MJ, Jin LW, et al. Clinical features, with video documentation, of the original familial lewy body parkinsonism caused by alpha-synuclein triplication (lowa kindred). Mov Disord 2011; 26(11): 2134-6.
- [47] Chartier-Harlin MC, Kachergus J, Roumier C, et al. Alphasynuclein locus duplication as a cause of familial Parkinson's disease. Lancet 2004; 364(9440): 1167-9.
- [48] Ibanez P, Bonnet AM, Debarges B, et al. Causal relation between alpha-synuclein gene duplication and familial Parkinson's disease. Lancet 2004; 364(9440): 1169-71.
- [49] Nishioka K, Hayashi S, Farrer MJ, et al. Clinical heterogeneity of alpha-synuclein gene duplication in Parkinson's disease. Ann Neurol 2006; 59(2): 298-309.
- [50] Nishioka K, Ross OA, Ishii K, et al. Expanding the clinical phenotype of SNCA duplication carriers. Mov Disord 2009; 24(12): 1811-9.

- [51] Ahn TB, Kim SY, Kim JY, et al. alpha-Synuclein gene duplication is present in sporadic Parkinson disease. Neurology 2008; 70(1): 43-9.
- [52] Shin CW, Kim HJ, Park SS, Kim SY, Kim JY, Jeon BS. Two Parkinson's disease patients with alpha-synuclein gene duplication and rapid cognitive decline. Mov Disord 2010; 25(7): 957-9.
- [53] Ross OA, Soto-Ortolaza AI, Heckman MG, et al. Association of LRRK2 exonic variants with susceptibility to Parkinson's disease: a case-control study. Lancet Neurol 2011; 10(10): 898-908.
- [54] Gasser T, Hardy J, Mizuno Y. Milestones in PD genetics. Mov Disord 2011; 26(6): 1042-8.
- [55] Bardien S, Lesage S, Brice A, Carr J. Genetic characteristics of leucine-rich repeat kinase 2 (LRRK2) associated Parkinson's disease. Parkinsonism Relat Disord 2011; 17(7): 501-8.
- [56] Healy DG, Falchi M, O'Sullivan SS, et al. Phenotype, genotype, and worldwide genetic penetrance of LRRK2associated Parkinson's disease: a case-control study. Lancet Neurol 2008; 7(7): 583-90.
- [57] Vilarino-Guell C, Wider C, Ross OA, et al. VPS35 mutations in Parkinson disease. Am J Hum Genet 2011; 89(1): 162-7.
- [58] Zimprich A, Benet-Pages A, Struhal W, et al. A mutation in VPS35, encoding a subunit of the retromer complex, causes late-noset Parkinson disease. Am J Hum Genet 2011; 89(1): 168-75
- [59] Sheerin UM, Charlesworth G, Bras J, et al. Screening for VPS35 mutations in Parkinson's disease. Neurobiol Aging 2012; 33(4): 838 e1-5.
- [60] Nuytemans K, Bademci G, Inchausti V, et al. Whole exome sequencing of rare variants in EIF4G1 and VPS35 in Parkinson disease. Neurology 2013; 80(11): 982-9.
- [61] Sharma M, Ioannidis JP, Aasly JO, et al. A multi-centre clinico-genetic analysis of the VPS35 gene in Parkinson disease indicates reduced penetrance for disease-associated variants. J Med Genet 2012; 49(11): 721-6.
 [62] Chartier-Harlin MC, Dachsel JC, Vilarino-Guell C, et al.
- [62] Chartier-Harlin MC, Dachsel JC, Vilarino-Guell C, et al. Translation initiator EIF4G1 mutations in familial Parkinson disease. Am J Hum Genet 2011; 89(3): 398-406.
- [63] Schulte EC, Mollenhauer B, Zimprich A, et al. Variants in eukaryotic translation initiation factor 4G1 in sporadic Parkinson's disease. Neurogenetics 2012; 13(3): 281-5.
- [64] Zhao Y, Ho P, Prakash KM, et al. Analysis of EIF4G1 in Parkinson's disease among Asians. Neurobiol Aging 2013; 34(4): 1311 e5-6.
- [65] Gasser T, Muller-Myhsok B, Wszolek ZK, et al. A susceptibility locus for Parkinson's disease maps to chromosome 2p13. Nat Genet 1998; 18(3): 262-5.
- [66] DeStefano AL, Lew MF, Golbe LI, et al. PARK3 influences age at onset in Parkinson disease: a genome scan in the GenePD study. Am J Hum Genet 2002; 70(5): 1089-95.
 [67] Pankratz N, Uniacke SK, Halter CA, et al. Genes influencing
- [67] Pankratz N, Uniacke SK, Halter CA, et al. Genes influencing Parkinson disease onset: replication of PARK3 and identification of novel loci. Neurology 2004; 62(9): 1616-8.
- [68] Karamohamed S, DeStefano AL, Wilk JB, et al. A haplotype at the PARK3 locus influences onset age for Parkinson's disease: the GenePD study. Neurology 2003; 61(11): 1557-61
- [69] Sharma M, Mueller JC, Zimprich A, et al. The sepiapterin reductase gene region reveals association in the PARK3 locus: analysis of familial and sporadic Parkinson's disease in European populations. J Med Genet 2006: 43(7): 557-62.
- [70] Leroy E, Boyer R, Auburger G, et al. The ubiquitin pathway in Parkinson's disease. Nature 1998; 395(6701): 451-2.
- [71] Pankratz N, Nichols WC, Uniacke SK, et al. Significant linkage of Parkinson disease to chromosome 2q36-37. Am J Hum Genet 2003; 72(4): 1053-7.
- [72] Lautier C, Goldwurm S, Durr A, et al. Mutations in the GIGYF2 (TNRC15) gene at the PARK11 locus in familial Parkinson disease. Am J Hum Genet 2008; 82(4): 822-33.
- [73] Tan EK, Schapira AH. Summary of GIGYF2 studies in Parkinson's disease: the burden of proof. Eur J Neurol 2010; 17(2): 175-6.

- [74] Strauss KM, Martins LM, Plun-Favreau H, et al. Loss of function mutations in the gene encoding Omi/HtrA2 in Parkinson's disease. Hum Mol Genet 2005; 14(15): 2099-111.
- [75] Ross OA, Soto AI, Vilarino-Guell C, et al. Genetic variation of Omi/HtrA2 and Parkinson's disease. Parkinsonism Relat Disord 2008; 14(7): 539-43.
- [76] Simon-Sanchez J, Singleton AB. Sequencing analysis of OMI/HTRA2 shows previously reported pathogenic mutations in neurologically normal controls. Hum Mol Genet 2008; 17(13): 1988-93.
- [77] Socal MP, Emmel VE, Rieder CR, Hilbig A, Saraiva-Pereira ML, Jardim LB. Intrafamilial variability of Parkinson phenotype in SCAs: novel cases due to SCA2 and SCA3 expansions. Parkinsonism Relat Disord 2009; 15(5): 374-8.
- [78] Hall DA, O'Keefe JA. Fragile x-associated tremor ataxia syndrome: the expanding clinical picture, pathophysiology, epidemiology, and update on treatment. Tremor Other Hyperkinet Mov (N Y). 2012; 2. pii: tre-02-56-352-1.
- [79] Lesage S, Le Ber I, Condroyer C, et al. C9orf72 repeat expansions are a rare genetic cause of parkinsonism. Brain 2013; 136(Pt 2): 385-91.
- [80] Hruska KS, LaMarca ME, Scott CR, Sidransky E. Gaucher disease: mutation and polymorphism spectrum in the glucocerebrosidase gene (GBA). Hum Mutat 2008; 29(5): 567-83.
- [81] Sidransky E, Nalls MA, Aasly JO, et al. Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. N Engl J Med 2009; 361(17): 1651-61.
- [82] Wan L, Hsu CM, Tsai CH, Lee CC, Hwu WL, Tsai FJ. Mutation analysis of Gaucher disease patients in Taiwan: high prevalence of the RecNcil and L444P mutations. Blood Cells Mol Dis 2006; 36(3): 422-5.
- [83] Scott SA, Edelmann L, Liu L, Luo M, Desnick RJ, Kornreich R. Experience with carrier screening and prenatal diagnosis for 16 Ashkenazi Jewish genetic diseases. Hum Mutat 2010; 31(11): 1240-50.
- [84] Sidransky E. Gaucher disease: complexity in a "simple" disorder. Mol Genet Metab 2004; 83(1-2): 6-15.
- [85] Neudorfer O, Giladi N, Elstein D, et al. Occurrence of Parkinson's syndrome in type I Gaucher disease. QJM 1996; 89(9): 691-4.
- [86] Machaczka M, Rucinska M, Skotnicki AB, Jurczak W. Parkinson's syndrome preceding clinical manifestation of Gaucher's disease. Am J Hematol 1999; 61(3): 216-7.
- [87] Bembi B, Zambito Marsala S, Sidransky E, et al. Gaucher's disease with Parkinson's disease: clinical and pathological aspects. Neurology 2003; 61(1): 99-101.
- [88] Goker-Alpan O, Schiffmann R, LaMarca ME, Nussbaum RL, McInemey-Leo A, Sidransky E. Parkinsonism among Gaucher disease carriers. J Med Genet 2004; 41(12): 937-40
- [89] Halperin A, Elstein D, Zimran A. Increased incidence of Parkinson disease among relatives of patients with Gaucher disease. Blood Cells Mol Dis 2006; 36(3): 426-8.
- [90] Rana HQ, Balwani M, Bier L, Alcalay RN. Age-specific Parkinson disease risk in GBA mutation carriers: information for genetic counseling. Genet Med 2013; 15(2): 146-9.
- [91] Anheim M, Elbaz A, Lesage S, et al. Penetrance of Parkinson disease in glucocerebrosidase gene mutation carriers. Neurology 2012; 78(6): 417-20.
 [92] Winder-Rhodes SE, Evans JR, Ban M, et al.
- [92] Winder-Rhodes SE, Evans JR, Ban M, et al. Glucocerebrosidase mutations influence the natural history of Parkinson's disease in a community-based incident cohort. Brain 2013; 136(Pt 2): 392-9.

- [93] Brockmann K, Srulijes K, Hauser AK, et al. GBA-associated PD presents with nonmotor characteristics. Neurology 2011; 77(3): 276-80.
- [94] Murakami T, Shoji M, Imai Y, et al. Pael-R is accumulated in Lewy bodies of Parkinson's disease. Ann Neurol 2004; 55(3): 439-42.
- [95] Gegg ME, Burke D, Heales SJ, et al. Glucocerebrosidase deficiency in substantia nigra of parkinson disease brains. Ann Neurol 2012; 72(3): 455-63.
- [96] Mazzulli JR, Xu YH, Sun Y, et al. Gaucher disease glucocerebrosidase and alpha-synuclein form a bidirectional pathogenic loop in synucleinopathies. Cell 2011; 146(1): 37-52.
- [97] Bender A, Krishnan KJ, Morris CM, et al. High levels of mitochondrial DNA deletions in substantia nigra neurons in aging and Parkinson disease. Nat Genet 2006; 38(5): 515-7.
- [98] Finsterer J. Parkinson's syndrome and Parkinson's disease in mitochondrial disorders. Mov Disord 2011; 26(5): 784-91.
- in mitochondrial disorders. Mov Disord 2011; 26(5): 784-91.
 [99] Orsucci D, Caldarazzo lenco E, Mancuso M, Siciliano G. POLG1-related and other "mitochondrial Parkinsonisms": an overview. J Mol Neurosci 2011; 44(1): 17-24.
- [100] Baloh RH, Salavaggione E, Milbrandt J, Pestronk A. Familial parkinsonism and ophthalmoplegia from a mutation in the mitochondrial DNA helicase twinkle. Arch Neurol 2007; 64(7): 998-1000.
- [101] Mancuso M, Filosto M, Orsucci D, Siciliano G. Mitochondrial DNA sequence variation and neurodegeneration. Hum Genomics 2008; 3(1): 71-8.
- [102] Hudson G, Nalls M, Evans JR, et al. Two-stage association study and meta-analysis of mitochondrial DNA variants in Parkinson disease. Neurology 2013; 80(22): 2042-8.
 [103] Corti O, Lesage S, Brice A. What genetics tells us about the
- [103] Corti O, Lesage S, Brice A. What genetics tells us about the causes and mechanisms of Parkinson's disease. Physiol Rev 2011: 91(4): 1161-218.
- [104] Klein C, Lohmann-Hedrich K, Rogaeva E, Schlossmacher MG, Lang AE. Deciphering the role of heterozygous mutations in genes associated with parkinsonism. Lancet Neurol 2007; 6(7): 652-62.
- [105] Marongiu R, Ferraris A, Ialongo T, et al. PINK1 heterozygous rare variants: prevalence, significance and phenotypic spectrum. Hum Mutat 2008; 29(4): 565.
- [106] Bruggemann N, Mitterer M, Lanthaler AJ, et al. Frequency of heterozygous Parkin mutations in healthy subjects: need for careful prospective follow-up examination of mutation carriers. Parkinsonism Relat Disord 2009; 15(6): 425-9.
- [107] Maraganore DM, de Andrade M, Elbaz A, et al. Collaborative analysis of alpha-synuclein gene promoter variability and Parkinson disease. JAMA 2006; 296(6): 661-70.
- [108] Mata IF, Shi M, Agarwal P, et al. SNCA variant associated with Parkinson disease and plasma alpha-synuclein level. Arch Neurol 2010; 67(11): 1350-6.
- [109] Tan EK. The role of common genetic risk variants in Parkinson disease. Clin Genet 2007; 72(5): 387-93.
- [110] Sharma M, Ioannidis JP, Aasly JO, et al. Large-scale replication and heterogeneity in Parkinson disease genetic loci. Neurology 2012, 79(7): 659-67.
- [111] Lill CM, Roehr JT, McQueen MB, et al. Comprehensive research synopsis and systematic meta-analyses in Parkinson's disease genetics: The PDGene database. PLoS Genet 2012; 8(3): e1002548.
- [112] Pankratz N, Beecham GW, DeStefano AL, et al. Metaanalysis of Parkinson's disease: identification of a novel locus, RIT2. Ann Neurol 2012; 71(3): 370-84.