

Thrombin regulates the ability of Schwann cells to support neuritogenesis and to maintain the integrity of the nodes of Ranvier

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ABSTRACT

Schwann cells (SC) are characterized by a remarkable plasticity that enables them to promptly respond to nerve injury promoting axonal regeneration. In peripheral nerves after damage SC convert to a repair-promoting phenotype activating a sequence of supportive functions that drive myelin clearance, prevent neuronal death, and help axon growth and guidance. Regeneration of peripheral nerves after damage correlates inversely with thrombin levels. Thrombin is not only the key regulator of the coagulation cascade but also a protease with hormone-like activities that affects various cells of the central and peripheral nervous system mainly through the protease-activated receptor 1 (PAR1). Aim of the present study was to investigate if and how thrombin could affect the axon supportive functions of SC. In particular, our results show that the activation of PAR1 in rat SC cultures with low levels of thrombin or PAR1 agonist peptides induces the release of molecules, which favor neuronal survival and neurite elongation. Conversely, the stimulation of SC with high levels of thrombin or PAR1 agonist peptides drives an opposite effect inducing SC to release factors that inhibit the extension of neurites. Moreover, high levels of thrombin administered to sciatic nerve *ex vivo* explants induce a dramatic change in SC morphology causing disappearance of the Cajal bands, enlargement of the Schmidt-Lanterman incisures and calcium-mediated demyelination of the paranodes. Our results indicate thrombin as a novel modulator of SC plasticity potentially able to favor or inhibit SC pro-regenerative properties according to its level at the site of lesion.

Key words: Protease-activated receptor 1; glia; rat; peripheral nervous system.

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Ethical Approval: All procedures involving animals were carried out in accordance with the guidelines of the European Communities Council Directive (86/609/EEC of 24 November 1986) and the Italian National law DL/116/92. All methods used were carried out in accordance with guidelines of the protocols n. 7FF2C.6.EXT.96 that has been approved by the Ministry of Health (AMT, Aut. N. 1184/2016-PR 16/12/2016).

Introduction

The strong regenerative potential of peripheral nerves is mainly due to the ability of Schwann cells (SC) to convert to a repair-promoting phenotype, which encourages axon growth (for a recent review see¹). Unfortunately, nerve stumps distal to transection maintain full or mildly reduced capacity for supporting regeneration only for about 1 month. Fading of the SC repair phenotype occurs in chronic denervation and is amplified in aging.²⁻⁴

The lesion of a peripheral nerve is accompanied by a local dramatic increase in thrombin activity respect to the uninjured control.^{5,6} Circulating concentration of prothrombin in normal subjects ranges from 700 nM to 1.7 μ M.⁷ It is also worth noting that both thrombin and the extrinsic pathway FX/FXa are locally generated in sciatic nerve.⁸ In the peripheral nervous system (PNS), low concentrations of thrombin were found to enhance the regeneration of mouse peripheral nerve after its crushing,⁹ while high concentrations had deleterious effects.¹⁰ Accordingly, in the central nervous system (CNS) thrombin is neuroprotective against ischemia at low concentration but causes degeneration at high concentration.¹¹ Thrombin is the key effector protease of the coagulation cascade mediating hemostasis, thrombosis, and inflammatory responses to vascular injury predominantly through the protease-activated receptor 1 (PAR1).¹² It is well known that SC primary cultures release factors that promote the viability and neurogenesis of neuronal cells.¹³ In this connection, our previous data indicate that the activation of PAR1 by low levels of specific agonist peptides in SC cultures enhances their ability to release molecules (such as decorin and macrophage migration inhibitory factor, MIF) that favor neuronal survival and neurite extension.¹⁴ PAR1 is the main thrombin receptor and acts as a biosensor translating the dynamic changes in the proteolytic microenvironment into cellular responses. It is activated when thrombin cleaves its N-terminal exodomain at a specific site. This cleavage event unmasks a new N-terminus that then serves as a tethered ligand that binds intramolecularly to the receptor effecting transmembrane signaling. A synthetic agonist peptide (PAR1 AP) with a sequence corresponding to the tethered ligand is able to stimulate PAR1 signalling without the need of proteolytic activation.¹ PAR1 is a G protein-coupled receptor (GPCR) and its signaling is transduced *via* activation of phospholipase C (PLC), generation of inositol trisphosphate (IP3) and diacylglycerol (DAG), consecutive Ca^{2+} release from intracellular stores, and protein kinase C (PKC) activation. Unlike classic GPCRs, which are internalized and recycled back to the cell surface after agonist stimulation, activated PAR1 is internalized and sorted from endosomes to lysosomes and degraded. Internalization and lysosomal sorting is critical for terminating activated PAR1 signaling.¹⁶ PAR1 is expressed within the blood vascular system and throughout the CNS, predominantly in neurons, microglia, astrocytes, and oligodendrocytes.^{11,17} PAR1 agonists mediate oligodendroglial pathology¹⁸ and targeting this receptor genetically or pharmacologically reduces its dysmyelinating effects.¹⁹ In the PNS, PAR1 is present on the SC plasma membrane and particularly concentrated at the level of the non-compacted SC microvilli of the nodes of Ranvier.^{14,20}

In the present study we compared the ability of low and high levels of thrombin in modulating the SC neurotrophic properties in cultures and the integrity of the nodes of Ranvier in sciatic nerve *ex vivo* explants. To note high levels of thrombin or PAR1 AP directly applied to rat sciatic nerve have been demonstrated to produce a rapid conduction block.²⁰ Our data show that high and low levels of thrombin displayed completely different actions inhibiting the former and favoring the latter the ability of SC of supporting neurogenesis. In addition, thrombin at high levels determined a Ca^{2+} -dependent paranodal demyelination of sciatic nerve fibers.

Materials and Methods

Materials

Collagenase type I, donkey serum, Hanks' Balanced Salt Solution (HBSS), hirudin, mouse monoclonal anti-S100b antibody (AB_477499), poly-D-lysine hydrobromide, poly-L-ornithine hydrobromide, Sudan black B, thapsigargin, thrombin, 0.25% trypsin were from Sigma-Aldrich, Milan, Italy.

Advanced Dulbecco's Modified Eagle Medium (Advanced DMEM), fetal bovine serum, Fluo-4 AM (stock solution 1 mM in DMSO), glycine, horse serum, N,N'-[1,2-ethanediybis(oxy-2,1-phenylene)]bis[N-[2-[(acetyloxy)methoxy]-2-oxoethyl]], bis[(acetyloxy)methyl] ester (BAPTA-AM; stock solution 5 mM in DMSO) were from Invitrogen, Monza, Italy.

Phosphate buffer saline (PBS) (0.1 g/L Ca^{2+} Chloride, 0.2 g/L Potassium Chloride, 0.2 g/L Potassium Dihydrogen Phosphate, 0.1 g/L Magnesium Chloride Hexahydrate, 8 g/L Sodium Chloride, 2.1716 g/L Disodium Phosphate Heptahydrate) was from Corning.

The PAR1 activating peptide (PAR1 AP) TFLLR-NH2 was from Bachem AG, Bubendorf, Switzerland.

Rabbit polyclonal antibody anti PAR1 (bs-0828R) was from Bioss Antibodies Inc., Woburn, MA, USA.

SCH79797 (PAR1 selective antagonist) (stock solution 10 mM in ethanol) was from Tocris Bioscience, Bristol, UK

Dylight 549-labelled anti-rabbit IgG and donkey AlexaFluor 488-labelled anti-mouse IgG were from Jackson ImmunoResearch Laboratories, West Grove, PA, USA.

Affinity-purified goat anti-mouse IgG, Vectashield mounting solution were from Vector Laboratories, Burlingame, CA, USA.

Preparation and treatments of primary Schwann cell cultures

Schwann cells were isolated from the sciatic nerves of male and female newborn Wistar rats and cultured as previously described,^{21,22} with some modifications. All procedures involving animals were carried out in accordance with the guidelines of the European Communities Council Directive (86/609/EEC of 24 November 1986) and the Italian National law DL/116/92. All methods used were carried out in accordance with guidelines of the protocols n. 7FF2C.6.EXT.96 that has been approved by the Ministry of Health (AMT, Aut. n. 1184/2016-PR 16/12/2016). Briefly, the bilateral sciatic nerves were removed from 2- to 3-day-old rats, minced with fine dissecting scissors into fragments, digested twice with 0.25% trypsin and 1 mg/mL collagenase type I in Advanced DMEM at 37°C for 30 min. Contaminating fibroblasts were removed by immunopanning. Briefly tissue culture plates were precoated with affinity-purified goat anti-mouse IgG and then incubated with OX-7 hybridoma culture supernatant containing anti-Thy1 IgG, which binds the cell surface molecule Thy1.1 expressed on fibroblasts. Non-adherent cells were plated at 500-1000 cells/mm² in petri dishes of 5 cm diameter (Falcon) precoated with poly-D-lysine hydrobromide. Cells were cultured in Advanced DMEM supplemented with 10% fetal bovine serum. Cells were incubated at 37°C in a humidified 5% CO₂ atmosphere and medium was changed every three days. The cell preparation consisted of 95% Schwann cells, as assessed by immunocytochemistry with anti-S100b antibody. Schwann cells were plated in 6-well plates (7 × 10⁵ cells/well) in 2 mL of Advanced DMEM supplemented with 10% fetal bovine serum. After 24 h cells were washed three times in HBSS and then treated for 48 h in advanced DMEM serum-free medium with thrombin (1, 10, 100 U/mL), the PAR1 activating peptide (PAR1 AP) (1, 10, 100 μ M), hirudin (10, 100 U/mL), SCH79797 (1 μ M) alone or in combination. The PAR1 specific inhibitor SCH79797 was added 5 min before PAR1 AP.

PC12 neurite outgrowth and survival assays

The conditioned medium (CM) from Schwann cells was recovered, spun, supplemented with 5% fetal bovine serum and 10% horse serum and added to rat pheochromocytoma PC12 cells (ECACC) for assessing neurite outgrowth. In fact, previously published data indicate that Schwann cells release factors promoting cell survival and neurite outgrowth of PC12 cells.^{13,14}

PC12 cells were grown on coverslips precoated with poly-L-ornithine hydrobromide in 24-well tissue culture plates. Cells were seeded at low density (5×10^3 cells/well) to avoid contact inhibition of neuritogenesis at higher densities. PC12 treated with nerve growth factor (NGF; 50 ng/mL) were used as positive controls. Medium was replaced every three days and after a week, PC12 cells were stained with hematoxylin and imaged with Nikon light microscope (Eclipse E600; Nikon Instruments SpA, Florence, Italy). The number of undifferentiated and neurite-extending cells was determined by visual examination of the field and expressed as the percentage of the total cells. Cells with neurites were defined as cells that possessed at least one neurite with a length equal or longer than the cell body diameter ($12 \mu\text{m}$ is the mean diameter of the control non-differentiated cells); 50-100 cells were assessed in one experiment, and data are the means \pm SE of three experiments.

For cell survival assays PC12 cells were seeded in 96-well plates (2×10^4 cells/well) and after 24 h from seeding they were cultured with the serum-free conditioned medium from Schwann cells treated with thrombin (10, 100 U/mL) or PAR1 AP (10, 100 μM) as mentioned above. Conditioned medium was replaced every three days and after a week MTT assay was used to evaluate cell viability. The MTT test measures the ability of living cells to reduce MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] into formazan crystals by the activity of mitochondrial dehydrogenases. MTT assay was performed essentially as described by Fabrizi *et al.*²³ Briefly, 10 μL of MTT solution (5 mg/mL) was added to each well and the incubation was continued for 3 h. Lysis buffer was prepared by dissolving 40% (w/v) SDS in deionized water, after adding an equal volume of N,N dimethylformamide, and the pH was adjusted to 4.7. After a 3-h incubation with MTT, 100 μL of the lysis buffer was added to each well and the absorbance read at 570 nm on a microplate reader (Model 550 Microplate Reader, BioRad Laboratories, Hercules, CA, USA). The relative cell viability (%) was expressed as a percentage relative to the untreated control cells.

In addition, cell viability was also assessed in the same samples by measuring the release of lactate dehydrogenase (LDH) in the culture medium by the Cytotoxicity Detection Kit (Roche, Mannheim, Germany) as previously described.²⁴

Measurement of $[\text{Ca}^{2+}]_i$ mobilization

Schwann cells seeded onto poly-L-lysine precoated chamber slides (Ibidi, Martinsried, Germany) were loaded with 1 μM of the cytosolic Ca^{2+} indicator Fluo-4 AM at 37°C, 5% CO_2 for 15 min in PBS pH 7.4 containing 25 mM glucose and 1% bovine albumin. After dye loading, cells were washed three times and then images were recorded using confocal microscope TCS-SP5 (Leica Microsystems, Heidelberg, Germany). Fluorescence intensity vs. time was quantified with LAS-X analysis software (Leica Microsystems) and expressed as $\Delta F/F_0$. The mean fluorescence value of all cells in one field measured at the start of the experiment is F_0 and F is the fluorescence intensity recorded in the same field as the experiment runs.

Ex vivo nerve explants

Sciatic nerves were rapidly removed from female adult Wistar rats (200-250 g) and cut in 4-mm long segments. The pieces of sciatic nerve were incubated free-floating in advanced DMEM at

37°C in 5% CO_2 /95% humidified air atmosphere and treated for 30 min, 1 h and 2 h with: 1, 10, 100 U/mL of thrombin; 1, 10, 100 μM PAR1 AP; 10, 100 U/mL hirudin; 1 μM SCH79797 (5 min pre-treatment); 1 μM BAPTA-AM (30 min pre-treatment); 100 nM thapsigargin alone or in combinations. At the end of the procedure samples were fixed for 30 min at room temperature (RT) with 4% paraformaldehyde in 0.12 M phosphate buffer, pH 7.4, washed in PBS and teased on positively charged glass slides. For immunofluorescence staining, teased fibers were blocked with 10% donkey serum containing 0.2% gelatin and 0.3% Triton X-100 for 1 h at RT and then incubated overnight at 4°C with rabbit anti-PAR1 (diluted 1:100) and mouse anti-S100b antibodies (diluted 1:1000). After washings with PBS, nerve preparations were incubated with donkey Dylight 549-labelled anti-rabbit IgG (diluted 1:400) and donkey AlexaFluor 488-labelled anti-mouse IgG (diluted 1:400). Negative controls were done by omission of the first antibody or by replacing the primary antibody with an equivalent amount of Igs purified from preimmune sera. After a final wash with PBS, the preparations were overlaid with coverslips using Vectashield mounting solution and imaged with confocal microscope TCS-SP5 (Leica Microsystems, Heidelberg, Germany). For Sudan black B staining, teased fibers were immersed in 0.1% Sudan black B diluted in 70% ethanol for 20 min at RT. Slides were then washed three times for 5 min each in PBS containing 0.02% Tween 20. After a final wash with PBS, the preparations were overlaid with coverslips using Vectashield mounting solution and imaged with Nikon light microscope (Eclipse E600; Nikon Instruments SpA).

Variable pressure-scanning electron microscope

Hydrated teased fibers from *ex vivo* nerve explants were observed using a variable pressure-scanning electron microscope (VP-SEM) (Hitachi SU-3500). VP-SEM combined with Peltier coolstage control optimally allowed high-resolution imaging of hydrated specimens under varying pressure conditions, limiting water vapour loss through control of stage temperature. Hydrated samples prepared as described in section above were directly observed on positively charged glass slides without conductive coating. A thin film of distilled water was deposited onto hydrated samples at RT to limit water-loss during low vacuum procedures. By appropriate control of chamber pressure and cooling temperature, particular attention has been paid to avoid the formation of collapsed surface of the Ranvier nodes. All samples were observed at an accelerating voltage of 6 kV with working distance between 5 and 6.2 mm and the correlative pressure/temperature were stabilized at 50 Pa/-25°C before.

By VP-SEM technique alterations in morphology and structure of the nodes were observed in their native hydrated state to assess minimal conformational changes of the nerves. Respect to conventional scanning electron microscope (SEM), VP-SEM is less time-consuming and less prone to create artifacts during processing.

Statistics

All experiments were performed in triple if not otherwise stated and reproduced three times. Statistical analyses were conducted using GraphPad Prism version 4.00 software. Comparisons were analysed using one-way ANOVA with Bonferroni-corrected *t*-test. A minimum statistical value $P < 0.05$ was considered significant.

Results

As mentioned above, SC in culture release factors that enhance neuronal survival and neurite extension¹³. Consistently with those previous reports, in neuronal PC12 cell line only $21 \pm 4\%$ of the untreated cells bore neurites at 7 days while this percentage

increased to $40 \pm 3\%$ after addition of the conditioned medium of SC primary cultures (CM-SC). NGF used as positive control led the percentage of PC12 cells bearing neuritis to $88 \pm 9\%$ (Figure 1A). We have previously shown that the neurotrophic properties of SC primary cultures are augmented when PAR1 is activated by low levels of specific agonist peptides (PAR1 AP).¹⁴ Here we broaden those previous results expanding the range of concentrations of PAR1 AP and using also thrombin which is the main PAR1 natural ligand. Thus, following the same procedure of our previous paper¹⁴, SC were treated with thrombin (1, 10, 100 U/mL) and PAR1 AP (1, 10, 100 μ M). After 48 h CM-SC was transferred to PC12 cells and neurite elongation was evaluated after 7 days. No difference in neurite extension respect to controls was observed using 1 μ M PAR1 AP or 1 U/mL thrombin (*not shown*). Conversely, CM obtained from SC treated with 10 μ M PAR1 AP or 10 U/mL of thrombin increased the percentage of PC12 cells bearing neurites respect to controls (Figure 1 B,C). These results are in line with our previous data.¹⁴ When we raised the level of the PAR1 agonists we observed a completely different result. In fact, CM obtained from SC treated with 100 U/mL of thrombin or 100 μ M PAR1 AP determined a reduction in the number of PC12 cells bearing neurites respect to controls (Figure 1 B,C). Thrombin was inhibited by hirudin which occludes both the thrombin active site and exosite I (fibrinogen and PAR1 recognition site) (Figure 1B). Similarly the PAR1 specific inhibitor SCH79797 abolished the effect of the PAR1 AP- treatment (Figure 1C).

The viability of PC12 cells was evaluated by MTT test in order to check if thrombin or PAR1 AP treatments could determine the

release from SC of cytotoxic molecules. No difference in cell survival was measured in PC12 cells receiving CM from SC treated with high levels of thrombin (100 U/mL) and PAR1 AP (100 μ M) (Figure 2). The same samples were tested for LDH release and no statistically significant differences were measured respect to controls (not shown). Conversely, CM from SC treated with low levels of thrombin (10 U/mL) and PAR1 AP (10 μ M) slightly increased PC12 cell viability consistently with our previous data.¹⁴

PAR1 is a GPCR and its activation is known to determine a rapid and transient rise in intracellular Ca^{2+} concentration.¹⁶ In SC by increasing the concentration of thrombin from 10 to 100 U/ml, we observed a parallel transient increase in intracellular Ca^{2+} levels (Figure 3 A,B). A rapid rise in cytosolic Ca^{2+} levels was also obtained using PAR1 AP (*not shown*). In order to check if the ability of thrombin in modifying SC neurotrophic properties could be calcium-mediated we used the intracellular Ca^{2+} chelator BAPTA-AM. When this compound was administered to SC it reverted the effect of both high and low levels of thrombin (Figure 3C). As mentioned above, the main thrombin receptor PAR1 is well expressed on the SC plasma membrane. While not being evenly distributed, it appears concentrated at the level of the nodes of Ranvier.^{14,20}

In order to explore whether the activation of the thrombin/PAR1 pathway could affect the SC/axon interaction in nerve fibers, we moved to *ex vivo* explants of sciatic nerve where the structure of the nodes of Ranvier are preserved.

Nerve explants were cultured with different concentrations of

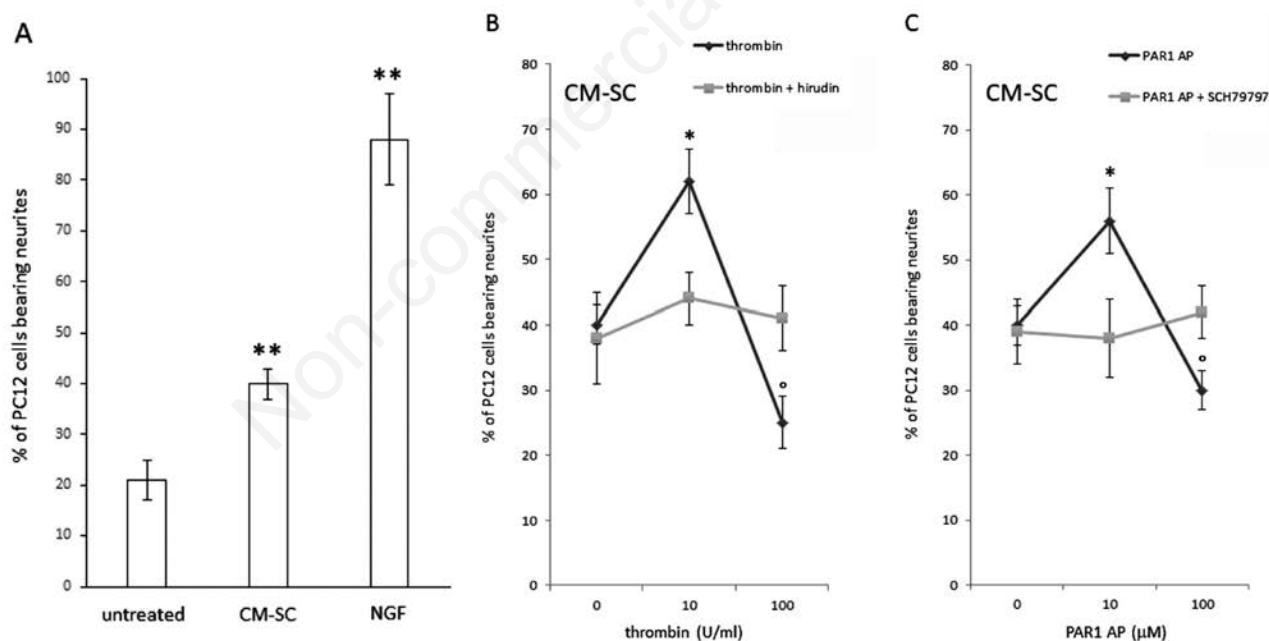


Figure 1. The conditioned medium from Schwann cells (CM-SC) stimulated with high or low levels of thrombin (or PAR1 AP) differently affects PC12 neurite outgrowth. A) PC12 cells were left untreated, treated with the culture medium from Schwann cells (CM-SC) or treated with NGF for 7 days. B) PC12 cells were exposed for 7 days to the conditioned medium (CM-SC) from untreated Schwann cells (ctrl) or from Schwann cells treated with: 10 U/mL thrombin; 100 U/mL thrombin; 10 U/mL hirudin + 10 U/mL thrombin; 100 U/mL hirudin + 100 U/mL thrombin; 100 U/mL hirudin alone; when SC were treated with 10 U/mL of thrombin their ability to promote neurite extension of PC12 was enhanced respect to the untreated SC while it was reduced with thrombin at 100 U/mL; thrombin effect was inhibited by hirudin. C) PC12 cells were exposed for 7 days to the conditioned medium (CM-SC) from untreated Schwann cells (ctrl) or from Schwann cells treated with: 10, 100 μ M PAR1 AP alone or in combination with 1 μ M SCH79797; When SC were treated with 10 μ M PAR1 AP their ability to promote neurite extension of PC12 was enhanced respect to the untreated SC while it was reduced with PAR1 AP at 100 μ M; PAR1 AP was blocked by the PAR1 inhibitor SCH79797; the percentage of neurite-bearing cells was calculated from 5 random fields per well, and the data are expressed as a mean percentage of 6 wells. Data are presented as mean \pm SEM. $^{\circ}P \leq 0.05$; $^*P \leq 0.01$ respect to ctrl; $^{**}P \leq 0.001$ respect to untreated.

thrombin (1, 10, 100 U/mL) or PAR1 AP (1, 10, 100 μ M) for 30 min, 1 h and 2 h. Then, nerve fibers were fixed, teased and stained for PAR1 and the SC cytoplasmic marker S100b. In untreated nerve explants PAR1 appeared mainly localized at the level of the nodes while after 2 h of treatment with 100 U/mL of thrombin it became more diffuse and scattered respect to the controls (Figure 4). To note, together with PAR1, also the distribution of S100b was considerably altered by thrombin treatment. In particular, in untreated nerve fibers S100b labeled the characteristic network of anastomosing, cytoplasmic channels known as Cajal bands (Figure 5 A,B) and the Schmidt-Lanterman incisures (SLI), another SC cytoplasmic compartment (Figure 5C). The treatment with 1 and 10 U/mL of thrombin (or 1 and 10 μ M of PAR1 AP) did not modify the morphology of the fibers respect to the untreated controls up to 2 h (*not shown*). Conversely, in fibers treated for 2 h with 100 U/mL of thrombin (or 100 μ M PAR1 AP, *not shown*) we observed a completely different distribution of the S100b staining characterized by the almost complete disappearance of the Cajal bands (Figure 5 D,E) and the marked dilation of the SLI (Figure 5F).

In order to analyze if these changes in the cytoplasmic compartments of SC corresponded to any alteration in myelin distribution, we labeled myelin by Sudan black staining. By this method we observed that nerve fibers treated with high concentrations of thrombin or PAR1 AP (100 U/mL and 100 μ M, respectively) were characterized by an increased paranodal demyelination respect to controls (Figure 6A, Table 1). Demyelination at the level of the paranodes could also be observed at low percentage in untreated controls and tended to increase with the time in culture ($5\pm 3\%$ at 30 min, $8\pm 2\%$ at 1 h and $12\pm 4\%$ at 2 h). Alterations in the morphology and structure of the nodes were analyzed in more detail by electron microscopy. In particular, nerve fibers from *ex vivo* sciatic nerve samples were directly observed using a VP-SEM, which operates in a wet mode with no need of desiccation and coating. Nerve fibers treated with high levels of thrombin or PAR1 AP and analyzed by electron microscopy showed an evident displacement of the SC at the nodes that consequently appeared elongated (Figure 6B). No alteration in the morphology of the nodes was observed with 1-10 U/mL thrombin or 1-10 μ M PAR1 AP (*not shown*). As shown in Table 1, the thrombin-induced paranodal demyelination could be inhibited by hirudin, which blocks both the

PAR recognition and thrombin active sites.

The thrombin-induced demyelinating effect was also impeded by the intracellular calcium chelator BAPTA-AM (Table 1). Thus, since the observed paranodal demyelination appeared to be calcium-related, we challenged nerve fibers with thapsigargin, a compound known to induce an increase in cytosolic Ca^{2+} with a mechanism different from the one elicited by the activation of the thrombin/PAR1 axis. In fact, thapsigargin raises the intracellular Ca^{2+} concentration by blocking the endoplasmic reticulum Ca^{2+} ATPase (SERCA) and activating plasma membrane Ca^{2+} channels²⁵.

Similarly to thrombin (100 U/mL), thapsigargin induced a pro-

Table 1. Percentage of paranodal demyelination in nerve fibers following PAR1 overactivation. *Ex vivo* nerve explants were treated with: thrombin (1, 10, 100 U/mL), 100 U/mL hirudin + 100 U/mL thrombin, 1 μ M BAPTA AM + 100 U/mL thrombin; PAR1 AP (1, 10, 100 μ M), 1 μ M BAPTA AM + 100 μ M PAR1 AP, 1 μ M SCH79797 + 100 μ M PAR1 AP. After 30 min, 1 h and 2 h of treatment samples were fixed, teased and stained by Sudan black. The percentage of nodes of Ranvier showing demyelination was calculated from 5 random fields per well, and the data are expressed as a mean percentage of 6 wells. Data are presented as mean \pm SEM.

	30 min	1 h	2 h
Thrombin (THR)			
0	5 \pm 3	8 \pm 2	12 \pm 4
1 U/mL THR	4 \pm 2	7 \pm 3	8 \pm 6
10 U/mL THR	6 \pm 3	7 \pm 5	12 \pm 5
100 U/mL THR	15 \pm 3*	30 \pm 5*	46 \pm 6*
BAPTA AM + 100 U/mL THR	6 \pm 2	5 \pm 3	11 \pm 4
Hirudin + 100 U/mL THR	7 \pm 2	10 \pm 4	15 \pm 6
PAR 1 AP			
0	5 \pm 3	8 \pm 2	12 \pm 4
1 μ M PAR 1 AP	6 \pm 4	11 \pm 5	10 \pm 3
10 μ M PAR 1 AP	3 \pm 1	9 \pm 5	10 \pm 5
100 μ M PAR 1 AP	13 \pm 3*	25 \pm 4*	32 \pm 7*
BAPTA AM + 100 μ M PAR 1 AP	4 \pm 2	7 \pm 4	9 \pm 3
SCH79797 + 100 μ M PAR 1 AP	8 \pm 4	14 \pm 6	18 \pm 7

* $P\leq 0.01$ respect to control.

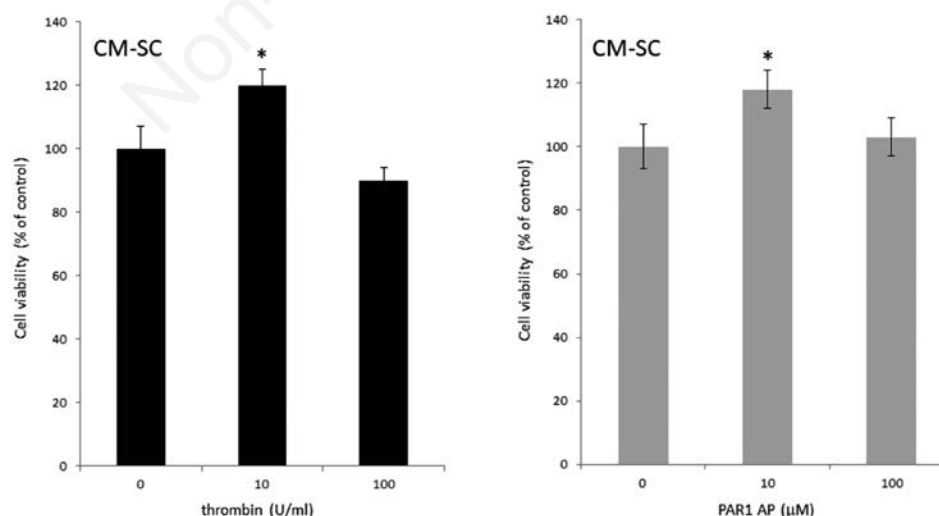


Figure 2. The conditioned medium (CM) from thrombin-treated Schwann cells (SC) does not affect PC12 cell viability. PC12 cells were exposed for 7 days to the CM of SC left untreated or treated with thrombin (10 and 100 U/mL) and PAR1 AP (10 and 100 μ M). Cell viability was evaluated by the MTT assay. Data are presented as mean \pm SEM of three independent experiments. * $P\leq 0.01$ respect to the untreated control.

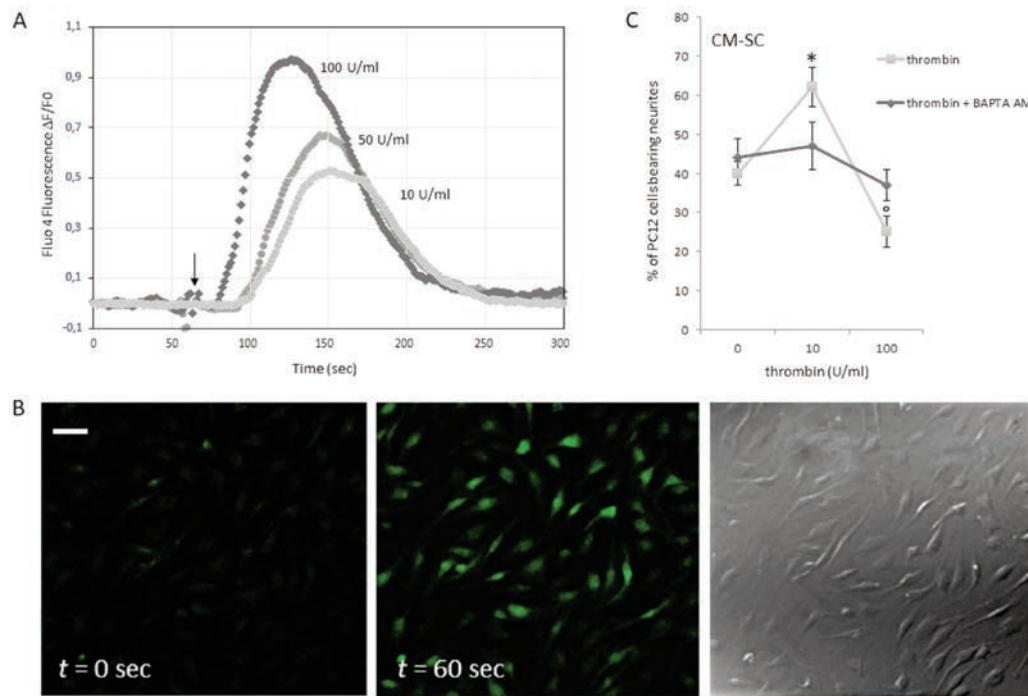


Figure 3. Thrombin dose-dependently induces an increase in intracellular Ca^{2+} levels in Schwann cells (SC). A) Representative tracings of the cytosolic Ca^{2+} indicator dye Fluo4 after application of different doses of thrombin (10, 50, 100 U/mL) to SC; arrow points at the stimulus administration; tracings indicate the mean fluorescence value of all cells in one field and are indicative of three independent experiments. B) SC loaded with Fluo4 at time 0 and after 60 s from the application of 100 U/mL of thrombin; time 0 corresponds to the stimulus administration (arrow in A). Phase contrast is shown on the right; scale bar: 50 μm . C) PC12 cells were exposed for 7 days to the conditioned medium (CM-SC) from SC treated with 10 and 100 U/mL of thrombin alone or in combination with 1 μM BAPTA AM. Data are presented as mean \pm SEM. $^{\circ}\text{P}\leq 0.05$; $^{*}\text{P}\leq 0.01$ respect to samples left untreated or treated with vehicle alone.

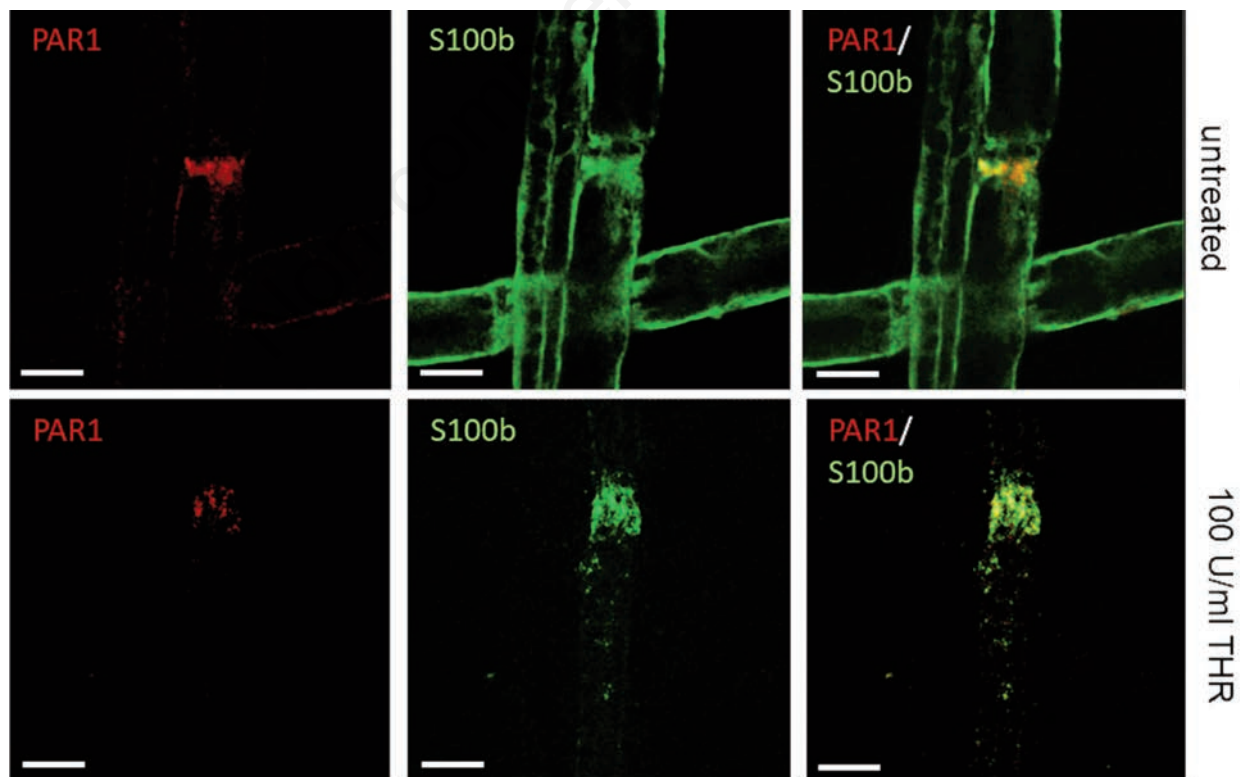


Figure 4. Double immunofluorescence staining for PAR1 and S100b of fibers from *ex vivo* nerve explants after 2-h culture. A) PAR1 appears highly expressed at the level of the nodes of Ranvier in untreated control. B) After treatment with 100 U/mL of thrombin both PAR1 and S100b are scattered and in clusters. Scale bar: 10 μm .

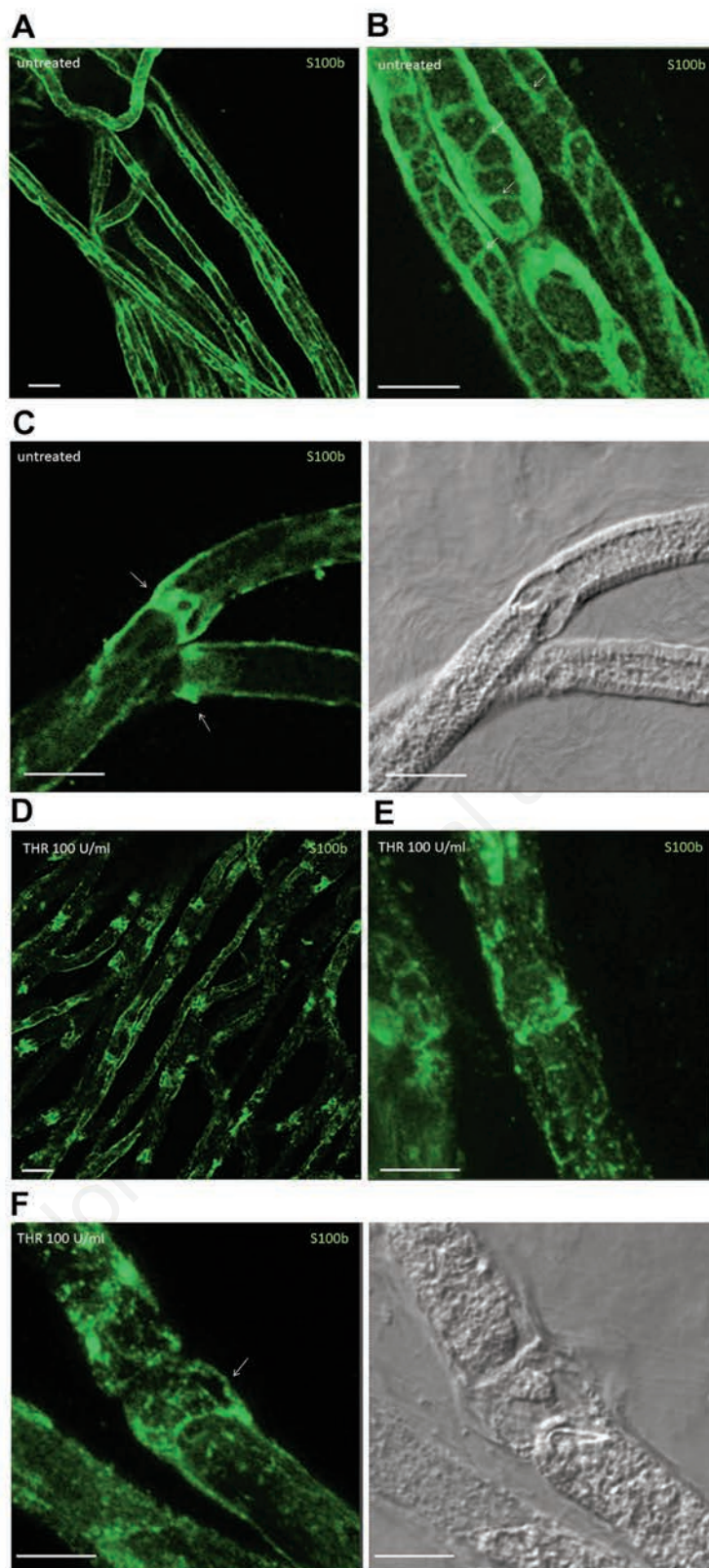


Figure 5. Immunofluorescence staining of fibers from *ex vivo* nerve explants for the SC cytosolic marker S100b. Thrombin-treatment alters the morphology of both the Cajal bands and the Schmidt-Lanterman incisures (SLI). A) Control fibers left untreated for 2 h; scale bar: 20 μm . B) Control fibers at higher magnification showing a node of Ranvier and the presence of Cajal bands (arrows); scale bar: 10 μm . C) Schmidt-Lanterman incisures (arrows) in control fibers left untreated for 2 h; phase contrast image on the right; scale bar: 10 μm . D) Nerve fibers treated for 2 h with 100 U/mL of thrombin (THR); scale bar: 20 μm . E) Nerve fibers treated for 2 h with 100 U/mL of thrombin (THR) at higher magnification showing the highly disorganized architecture of the Cajal bands; scale bar 10 μm . F) Dilation of a Schmidt-Lanterman incisure (arrow) in nerve fibers treated for 2 h with 100 U/mL of thrombin (THR); phase contrast image on the right; scale bar: 10 μm .

nounced alteration in S100b and myelin distribution in nerve fibers. When administered to *ex vivo* nerve explants this compound determined extensive paranodal demyelination of nerve fibers ($57\pm 6\%$ at 2 h) with concurrent enlargement of the SLI and disappearance of Cajal bands. In addition, in thapsigargin-treated samples demyelination appeared also in vast areas of the internodes (Figure 7).

Discussion

In this paper we show that the activation of the thrombin/PAR1 axis in SC profoundly affects two major properties of these cells that is their ability to promote neuritogenesis and their capacity to maintain the integrity of the nodes of Ranvier.

When treated with low levels of thrombin SC in culture show an enhanced ability to release factors promoting PC12 neurite extension. This effect can be blocked by the administration of hirudin, which inhibits the thrombin proteolytic activity, and it can

be reproduced by a specific PAR1 agonist peptide. These results are consistent with our previous data indicating that the activation of PAR1 induces the release from SC of molecules, which favor neuronal survival and neurite extension such as decorin and MIF.¹⁴ Although it would be improper to directly translate these findings *in vivo*, these data are in line with results obtained *in vivo* where thrombin at low concentration was demonstrated to favor nerve regeneration.⁹ As mentioned above, peripheral nerve damage is accompanied by the increase in thrombin activity at the site of injury.^{5,6} Thrombin activity was also found to be increased early in the clinical course of experimental autoimmune neuritis (EAN) in rat sciatic nerve.² In our model system SC treated with high levels of thrombin (100 U/mL) showed a reduced capacity of promoting PC12 neurite extension respect to controls. This effect appeared not related to the release of cytotoxic molecules from thrombin-treated SC primary cultures. The different effect exerted by thrombin in cultures at low and high doses is not surprising since in other systems this protease was reported to display different actions according to its concentration.¹¹

We observed divergent effects of low and high levels of

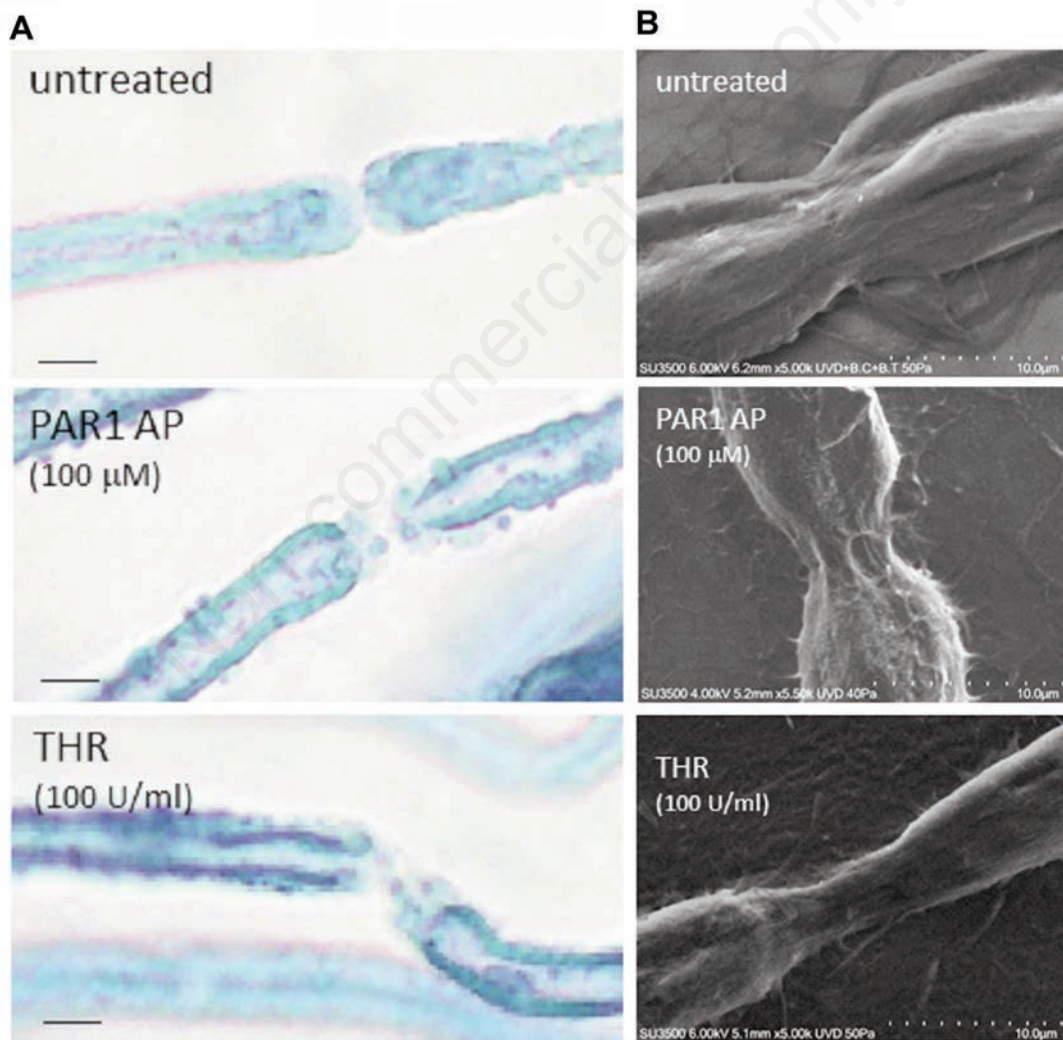


Figure 6. PAR1 overactivation determines in *ex vivo* nerve explants paranodal demyelination associated to a displacement of SC at the level of the nodes. A) Nerve fibers treated for 2 h with 100 U/mL of thrombin (THR) or 100 μM PAR1 AP observed by Sudan black staining are characterized by demyelination and myelin fragmentation at the level of the paranodes; scale bars: 10 μm. B) Nerve fibers treated for 2 h with 100 U/mL of thrombin (THR) or 100 μM PAR1 AP observed by VP-SEM show displacement of SC in the nodal region and elongated nodes; scale bars: 10 μm.

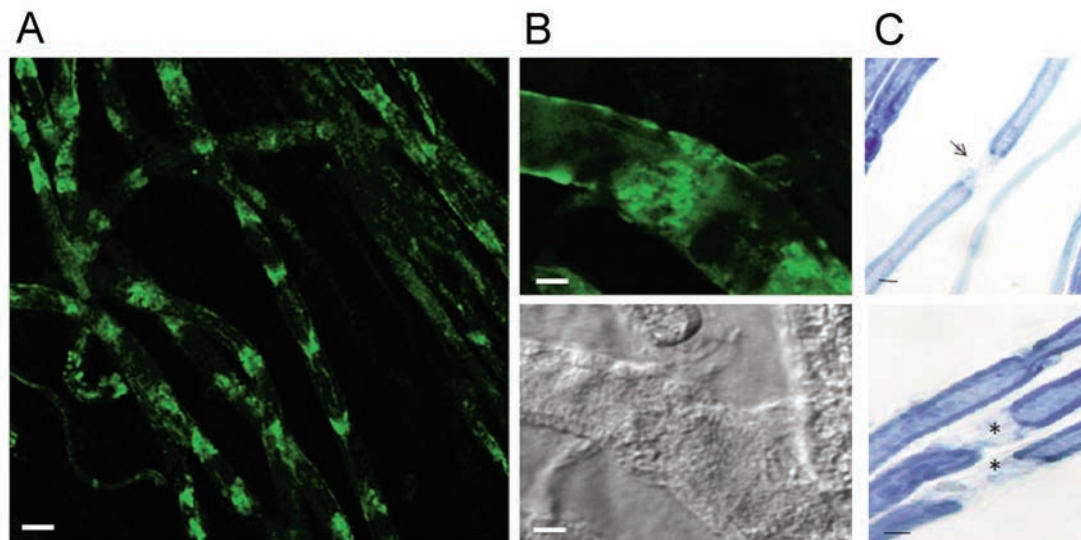


Figure 7. Thapsigargin promotes extensive demyelination in nerve fibers. *Ex vivo* nerve explants were treated for 2 h with thapsigargin and then stained for S100b (A,B) and with Sudan black (C). In nerve fibers thapsigargin, similarly to thrombin (100 U/mL), determines the disappearance of the Cajal bands and the dilation of the Schmidt-Lanterman incisures (A,B). Demyelination is observed not only at the paranodal region (arrow) but also in the internodal areas (asterisks) (C). Scale bars: A,C) 10 μm ; B) 20 μm .

thrombin on SC also in sciatic nerve *ex vivo* explants. In this model system although the nerve is transected the three-dimensional relationship between SC and the axon is mainly maintained. Low levels of thrombin in this model did not determine any evident modification in the morphology of SC and in particular of the nodes. Conversely, high levels of thrombin (100 U/mL) or PAR1 AP (100 μM) induced a profound rearrangement of SC and in particular the disappearance of the Cajal bands. Interestingly, a similar pattern of alteration of the Cajal bands is observed after chronic nerve compression.²⁷ Studies using periaxin-null mice indicate that the Cajal bands facilitate the microtubule-based transport of proteins and organelles in SC and aberrations in their architecture compromises myelin maintenance.²⁸ Actually, our data show that nerve fibers treated with high levels of PAR1 agonists present evident signs of demyelination at the level of the paranodes. It is notable that PAR1 is mostly expressed on the microvilli of the SC, that is in close proximity to the paranodes.^{14,20} In addition, thrombin (100-200 U/mL) and PAR1 AP (150-300 μM) cause a conduction failure in sciatic nerve within 30 min of application.²⁰

Our data indicate that the observed paranodal demyelination induced after PAR1 overactivation is calcium-mediated. In connection with this result, elevation of intracellular Ca^{2+} levels has been reported to induce myelin basic protein (MBP) network disassembly and myelin vesiculation in CNS.²⁹ Besides, in ischemia myelin is damaged in a calcium-dependent manner, abolishing action potential propagation,³⁰⁻³² and specific thrombin inhibitors were shown to reduce neurological deficit in models of middle cerebral artery occlusion.³³ The beneficial effect of thrombin inhibition was recently reported to occur in EAN,²⁵ and diabetic neuropathy³⁴ indicating the pathogenic importance of this pathway also in the PNS.

Our results obtained with high levels of thrombin in SC cultures and *ex vivo* nerve explants can be reproduced by PAR1 AP and inhibited by the specific PAR1 antagonist SCH79797 indicating the involvement of PAR1 in these processes. Nevertheless, the possibility of activation and transactivation of other members of the PAR family cannot be completely ruled out. In particular,

thrombin in adequate concentrations is known to activate the receptor for trypsin PAR2³⁵ and the low-affinity thrombin receptor PAR4.³⁶ However, in our SC cultures while thrombin and PAR1 AP dose-dependently induce a consistent rise in cytosolic Ca^{2+} , the specific agonist peptides for PAR2 and PAR4 determine only negligible modifications of the intracellular Ca^{2+} levels (*unpublished observations*). Our data contribute towards considering thrombin, which appears deregulated at sites of PNS injury a possible novel therapeutic target. Potentially a pharmacological reduction of its excessive activity at the level of nerve lesions could circumvent the demyelinating properties of this protease and restore the SC neurotrophic activities. Nevertheless, the complete pathways, which underlie thrombin deleterious or protective effect in PNS, need further investigation before considering possible therapeutic applications.

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