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# Mechanical Characterization of Methanol Plasma Treated Fluorocarbon Ultrathin Films Through Atomic Force Microscopy

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Methanol plasma has been proposed as an effective way to improve the performances of fluorocarbon (CF<sub>x</sub>) ultrathin films as stent coatings as it can successfully modulate fluorine content and wettability of the films. Nevertheless, plasma treatment may affect mechanical properties of the films, which therefore need comprehensively characterizing to verify the suitability of treated films for application as stent coating materials. In this work we investigate mechanical properties of methanol plasma treated CF<sub>x</sub> ultrathin films on stainless steel. In particular, cohesion of the films and their adhesion to the substrate is investigated using small punch test combined with atomic force microscopy (AFM) imaging. Also, elastic and viscoelastic properties are investigated at the nanometer scale using two different AFM based advanced technique for nanomechanical characterization, i.e., HarmoniX<sup>TM</sup> and contact resonance AFM (CR-AFM). Overall, methanol plasma treated CF<sub>x</sub> films have been demonstrated to be suitable for application as stent coating also on the basis of their nanomechanical properties.

**Keywords:** fluorocarbon films, adhesion, elastic properties, viscoelastic properties, contact resonance atomic force microscopy, HarmoniX, methanol plasma, biomaterials

## 1. INTRODUCTION

The mechanical characterization of thin coatings is increasingly emerging as a need in the development and optimization of biomaterials. Indeed, biomaterials for medical devices are often subjected to surface modification processes, such as functionalization and deposition of thin coatings, with the aim of improving their functionality and biocompatibility, without sacrificing the high structural properties given by the bulk materials. Besides the opportune chemical, physical and biocompatibility properties, a coating for medical devices must exhibit appropriate mechanical properties in order to maintain its integrity, cohesion and adhesion to the underlying substrate, even after significant elastic and plastic deformations to which the device can undergo. Together with cohesion and adhesion to the substrate, the stiffness of the coating, i.e., the part of the biomaterial in contact with the biological medium, is emerging as key factor, in addition to chemical composition, surface energy and roughness, in the regulation of the biological response. Indeed, the stiffness of biomaterials has been demonstrated to affect adhesion, proliferation, differentiation and migration of several kind of cells (Lo et al., 2000; Guo et al., 2006; Chowdhury et al., 2010).

For instance, neurons have been shown to proliferate better on soft substrates (Flanagan et al., 2002) while fibroblasts and chondrocytes exhibited maximum spread and proliferation on stiffer substrates (Subramanian and Lin, 2005; Hopp et al., 2013). Native mesenchymal stem cells undergo different differentiation behavior when interacting with surfaces with different elastic modulus (Engler et al., 2006). Endothelial cells have been observed to exhibit tube-like structures on soft substrates and high spread and proliferation on stiffer substrates (Califano and Reinhart-King, 2008, 2010); smooth muscle cells and myoblasts have been shown to have higher projected area on stiff substrates (Engler et al., 2004a,b). Moreover, substrate stiffness has been demonstrated to affect adhesion, spreading and activation of platelets, suggesting that the mechanical properties at the surface of cardiovascular devices could also have an effect on blood contact behavior and clot formation (Qiu et al., 2014). Therefore, the capability to characterize the mechanical properties of thin coatings for medical devices is fundamental, not only from the point of view of the integrity and mechanical behavior of the device, but also for the comprehension of the effect of surface stiffness on the biological events occurring at the interface of the device. Indeed, the capability of measuring and modulating the elastic modulus at the very surface of an implant might allow improved body response and implant performance. In previous studies, fluorocarbon (CF<sub>x</sub>) thin films deposited by plasma enhanced chemical vapor deposition (PECVD) with thickness of about 35 nm have been demonstrated to be effective as coatings for cardiovascular stents, thanks to their chemical inertness, high uniformity, flexibility and adhesion to the substrate (Haïdopoulos et al., 2005). These characteristics make them resistant to corrosion (Touzin et al., 2010) and able to overcome the high deformation occurring during stent deployment (about 25%) (Lewis et al., 2008). The higher hemocompatibility of CF<sub>x</sub> coatings in respect of bare metallic surfaces has been shown (Montaño-Machado et al., 2017) and can mainly attributed to the presence of fluorine content. Nevertheless, the hydrophobic nature of these coatings could limit protein adsorption, hemocompatibility and endothelialization (Arima and Iwata, 2007; Tang et al., 2008). Recently, a novel oxidative methanol plasma treatment has been developed and demonstrated to be effective in tuning the hydrophobicity of CF<sub>x</sub> coatings without affecting their integrity, uniformity and morphology (Montaño-Machado et al., 2019). By analyzing coatings with different wettability and oxygen and fluorine content, it has been shown that the blood contact behavior of CF<sub>x</sub> coatings can be improved by modulating surface energy and fluorine species content (Montaño-Machado et al., 2019). Nevertheless, the effect of the treatment on the mechanical properties of the modified films, which could affect the cohesion and adhesion to the substrate as well as the blood contact behavior, has not been evaluated yet. The mechanical characterization of these ultrathin coatings is challenging. The capability of the coating to maintain its integrity after a determined deformation can be tested by the method developed by Lewis et al. (2008), called small punch test, consisting in the application of a known deformation and the subsequent morphological analysis of the coating.

As for the quantitative evaluation of the elastic modulus, it cannot be obtained by conventional techniques, such as micro- and nanoindentation due to the limited thickness which makes the results of the measurements strongly affected by the mechanical properties of the underlying substrate (Fischer-Cripps, 2006; Reggente et al., 2017). Moreover, micro- and nanoindentation do not allow one to visualize the distribution of these surface properties because of its poor spatial resolution. Alternatives methods, based on atomic force microscopy (AFM), can be used to obtain quantitative maps of mechanical properties, such as indentation modulus, damping, adhesion and energy dissipation (Passeri et al., 2013a) as well as viscoelastic moduli, i.e., storage and loss modulus, and loss tangent (Killgore and DelRio, 2018).

In this work, we perform an exhaustive mechanical characterization of methanol plasma treated CF<sub>x</sub> ultrathin films, with thickness in the range 30–65 nm, deposited on stainless steel in order to verify the effect of plasma treatment on the mechanical properties of the films. Cohesion of the films and their adhesion to the substrate after deformation was analyzed with small punch test method combined with AFM morphological characterization. The challenging analysis of elastic and viscoelastic response of ultrathin soft films on stiff substrates was performed with two different AFM based techniques for nanomechanical characterizations, i.e., HarmoniX<sup>TM</sup> and contact resonance AFM (CR-AFM), for the latter using two different data analysis procedures, in order to cross-validate these methods.

## 2. MATERIALS AND METHODS

### 2.1. Materials

#### 2.1.1. Pre-treatment of 316L Stainless Steel Substrates

316 L stainless steel disks of 12.7 mm diameter and 0.5 mm thickness (Goodfellow, Devon, PA, USA) are used as substrates. First, they have been cleaned in an ultrasonic bath with acetone, deionized water, and methanol for 10 min for each solvent and then dried with particle-free compressed air. Then an electropolishing and an acid dipping treatment have been performed in order to reduce the surface roughness. Electropolishing has been carried out in 100 mL of solution containing glycerol, phosphoric acid and deionized water for 3 min at 90°C. Acid dipping was performed for 30 s at 50°C in a solution consisted in nitric acid, hydrofluoric acid, and deionized water (Montaño-Machado et al., 2019).

#### 2.1.2. Plasma-Enhanced Chemical Vapor Deposition of Fluorocarbon Coating

CF<sub>x</sub> coatings with different wettability and oxygen and fluorine content have been obtained by PECVD, following the protocols previously developed (Montaño-Machado et al., 2019). Specimens were introduced into the previously described radio frequency (RF) plasma reactor (Lewis et al., 2008) at 6 cm below the electrode for a pulsed H<sub>2</sub> plasma etching at 100 W for 100 s ( $t_{\text{on}} = 100$  ms;  $t_{\text{off}} = 300$  ms). Etching was carried out to (i) remove the layer of organic contaminants and (ii) reduce

Q8

the oxide layer on the sample surface. After the plasma etching of the substrates, a fluorocarbon coating has been deposited by pulsed afterglow plasma polymerization using  $C_2F_6$  and  $H_2$  as precursors. The sample was placed at 11 cm below the electrode and the coating was realized using a peak power input of 150 W, a duty cycle of 20%, a pressure of 700 mTorr and a total gas flow rate of 20 sccm for a deposition time of 5 min. An oxidation treatment with methanol plasma was then carried out with the sample at 6 cm below the electrode, using a continuous power of 50 W and a flow of 10 sccm during 30 and 90 s. In this way, three different coatings with decreasing hydrophobicity (static contact angle of  $117^\circ$  for the unmodified  $CF_x$  coating,  $87^\circ$  for the coating after 30 s methanol treatment and  $68^\circ$  for the coating after 90 s methanol treatment) and increasing O/F ratio (0.01 for the unmodified  $CF_x$  coating, 0.44 for the coating after 30 s methanol treatment and 1.08 for the coating after 90 s methanol treatment) were obtained (Montaño-Machado et al., 2019). For the sake of clarity, from herein  $CF_x$ -A refers to the sample which has not been processed with methanol plasma treatment, while  $CF_x$ -B and  $CF_x$ -C indicate the samples modified by 30 and 90 s methanol plasma treatment, respectively.

## 2.2. Instrumentation and Equipment

### 2.2.1. X-Ray Photoelectron Spectroscopy Analysis

Chemical composition of the coating was analyzed by X-ray photoelectron spectroscopy (XPS), using an X-ray photoelectron spectrometer (XPS-PHI 5600-ci Spectrometer-Physical Electronics, Eden Prairie, MN, USA), with a base pressure below  $5 \times 10^{-7}$  Pa. Survey and high resolution spectra were acquired using the  $K\alpha$  line of a standard aluminum ( $K\alpha = 1486.6$  eV) and magnesium ( $K\alpha = 1253.6$  eV) X-ray sources, respectively, operated at 300 W, without charge compensation. Three spots on three samples were analyzed for each experiment.

### 2.2.2. Small Punch Test

A plastic deformation of 25%, i.e., the estimated maximum deformation to which a stent undergo during deployment (Migliavacca et al., 2005), was applied to the coated samples using a custom-made small punch test device mounted on a SATEC T20000 testing machine (Instron, Norwood, MA, USA) as previously described (Lewis et al., 2007). All deformations were performed at room temperature at a displacement rate of  $0.05 \text{ mm s}^{-1}$  and a maximal load of 2200 N. The state of the coating, i.e., the morphology and the eventual presence of delamination or cracks, after deformation was analyzed by AFM. A Dimension<sup>TM</sup> 3100 AFM (Veeco, Woodbury, NY, USA) was used in tapping mode with a silicon tip (OTESPA, Bruker).  $20 \times 20 \mu\text{m}$  images in correspondence of the topmost part of the samples, where the maximum deformation (25%) occurs, were acquired and analyzed. Three specimens for each condition ( $CF_x$ -A,  $CF_x$ -B and  $CF_x$ -C) were tested.

### 2.2.3. Nanomechanical Characterizations by Atomic Force Microscopy

Mechanical characterizations of the films have been performed through both CR-AFM and HarmoniX<sup>TM</sup>.

HarmoniX<sup>TM</sup> has been carried out using a standard AFM apparatus (ICON, Bruker Inc.), equipped with a standard T-shape Si cantilever (HMX10, Bruker Inc.) having  $k_c = 4 \text{ N/m}$  (as reported by the manufacturer) and first flexural and torsional resonance frequencies in air equal to  $f_{0,1} = 52.9 \text{ kHz}$  and  $t_{0,1} = 949 \text{ kHz}$ , respectively. Cantilever force sensitivity has been measured from force-deflection curves performed on Si (100) single crystal. Calibration procedure has been performed using a blend of polystyrene (PS) and low-density polyethylene (LDPE) film deposited on a Si substrate (PS/LDPE, Bruker Inc.), being  $M_{PS} = 1.6 \text{ GPa}$  and  $M_{LDPE} = 100 \text{ MPa}$  the indentation modulus of PS and LDPE, respectively, as supplied by the producer.

CR-AFM has been carried out using a standard AFM apparatus (Solver, NT-MDT, Russia). The AFM setup was equipped with a Si cantilever (CSG10, NT-MDT, Russia) with spring constant  $k_c = 0.116 \text{ N/m}$ , determined through the method described by Sader et al. (1999). In order to evaluate the instrumental parameters required to analyze CR-AFM data, a well-established experimental procedure was followed (Reggente et al., 2015): standard force-deflection curves have been acquired on Si (100) single crystal to calibrate the cantilever force sensitivity; tip radius  $R_t$  has been evaluated by reconstructing the tip shape through the analysis of the images collected on an array of inverted tips used as reference sample (TGZ1, NT-MDT, Russia); the exact position of the tip along the cantilever axis has been determined through scanning electron microscopy (SEM) analysis. Being the tip in contact with the sample surface, contact resonance frequencies (CRFs) were detected for each sample and CRFs values and the corresponding uncertainties have been evaluated from statistics performed on not <512 points of CR-AFM images. Calibration of CR-AFM has been performed using as the reference the PS regions of the above described PS/LDPE sample. Numerical simulation of the entire experimental session was performed using Matlab (version 7.1.0.246, 2005).

## 3. TECHNIQUES

### 3.1. HarmoniX<sup>TM</sup>

HarmoniX<sup>TM</sup> is tapping mode based AFM technique in which a T-shaped cantilever with an out-of-axis tip is used (Sahin et al., 2007; Sahin and Erina, 2008). The first flexural mode of the cantilever, characterized by the first flexural resonance frequency  $f_{0,1}$ , is used to reconstruct sample morphology, as in standard AFM tapping mode. During tapping performed at frequency  $f_{0,1}$ , the tip periodically (with period  $T_0 = 1/f_{0,1}$ ) interacts with the sample surface. The actual tip-sample interaction force depends on the instantaneous value of tip-sample separation  $d$ , i.e., is given by the van der Waals force if  $d > a_0$ , and by the sum of van der Waals and repulsive (elastic) forces if  $d < a_0$ , where  $a_0$  is an intermolecular distance. More explicitly, by modeling the repulsive force using the Derjaguin-Muller-Toporov (DMT) (Derjaguin et al., 1975) and assuming that the lateral extension of the sample is much bigger than the tip size (Santos et al., 2011), the tip-sample interaction force  $F_{ts}$  is given by



$$F_{ts} = \begin{cases} -\frac{HR_t}{6d^2} & \text{if } d > a_0 \\ -\frac{HR_t}{6a_0^2} + \frac{4}{3}E^*\sqrt{R_t(a_0 - d)^3} & \text{if } d < a_0 \end{cases} \quad (1)$$

where  $H$  is the Hamaker constant,  $R_t$  is the tip radius,  $a_0$  an intermolecular distance, and  $E^*$  is the reduced modulus given by  $E^* = (M_s^{-1} + M_t^{-1})^{-1}$  being  $M_s$  and  $M_t$  the indentation modulus of the sample and of the tip, respectively (Derjaguin et al., 1975; García and San Paulo, 1999). When  $M_s \ll M_t$  as in the case of the materials investigated in this work,  $E^* \approx M_s$ . This force is measured by recording the cantilever deflection  $\Delta z$  which is proportional to the tip-sample force through the (normal) spring constant  $k_c$ . Being periodic with period  $T_0$ , the force-separation curve could be, in principle, reconstructed through inverse Fourier transform after acquiring the harmonic components (multiple of  $f_{0,1}$ ) of the deflection signal. Nevertheless, the presence of higher deflection modes of the cantilever distorts the harmonic components of the force-separation curve. A smart solution consists in using a T-shaped cantilever with an out-of-axis tip. Indeed, during tapping, the tip-sample interaction produces a torque  $T_{ts} = F_{ts}l_t$  where  $l_t$  is the distance between the tip location and the cantilever axis. This results in the torsion of the cantilever of an angle  $\Delta\phi = T_{ts}/k_\phi$  where  $k_\phi$  is the cantilever torsional spring constant (Green et al., 2004; Pettersson et al., 2007). Thus, the cantilever torsional signal is proportional to  $F_{ts}$  and is periodic with period  $T_0$ . Therefore, the periodic  $F_{ts}$  can be reconstructed by inverse Fourier transform of the components of the torsional signal at frequencies multiple of  $f_{0,1}$ . Being the value of the first free torsional resonance  $t_{0,1}$  much higher than  $f_{0,1}$ , this allows the analysis of harmonic components of the torsional signal in an undistorted spectral region and leads signal-to-noise ratio higher than the one relative to the deflection signal. Therefore, the cantilever torsional signal can be analyzed using inverse Fourier transform analysis in order to extract a complete loading/unloading force-distance curve during each tapping cycle. Unloading force-distance curves are analyzed in real time to evaluate  $M_s$  through the DMT model (Derjaguin et al., 1975) which is currently implemented in HarmoniX<sup>TM</sup> software, although the Johnson-Kendall-Roberts (JKR) model (Johnson et al., 1971) has been demonstrated to be more accurate in the range of materials analyzed in this work (Dokukin and Sokolov, 2012). To overcome the need for accurate calibration of the cantilever geometrical and elastic parameters, a phenomenological calibration is performed using a reference sample with well-known indentation modulus  $M_{ref}$ .

### 3.2. Contact Resonance Atomic Force Microscopy

In contact resonance AFM (CR-AFM) the tip is in contact with the sample surface and the system constituted by the cantilever, the tip, and a volume of the sample under the tip is set into oscillation using an ultrasonic transducer coupled to the sample (Rabe and Arnold, 1994) or to the cantilever (Yamanaka et al.,

2008), or by the direct photothermal excitation of the cantilever (Wagner and Killgore, 2015). In these conditions, the resonance frequencies of the cantilever, namely the contact resonance frequencies (CRFs)  $f_n$ , can be detected. When the investigated material is considered elastic, the system can be described as sketched in **Figure 1A**. In this model: the cantilever is considered inclined by an angle  $\alpha$  respect to the sample surface; the tip is located at distance  $L_1$  from the cantilever clamped end, so that the parameter  $r = L_1/L$  can be defined being  $L$  the cantilever length; the height of the tip is  $h_t$ ; the tip interacts with the sample surface via both normal and lateral forces, and the contact is thus described by the normal and lateral contact stiffness, modeled through the linear springs  $k^*$  and  $k_{lat}^*$ , respectively. The values of the CRFs are determined by  $k^*$  and  $k_{lat}^*$ , which depends on the elastic parameters of the sample, i.e., on the indentation and shear moduli of the sample ( $M_s$  and  $G_s$ , respectively). Therefore, the measured values of  $f_n$  can be used to evaluate the elastic properties of the sample. Nevertheless, due to the complexity of the model in **Figure 1A**, the analysis of CRFs data are generally carried out considering the simplified model depicted in **Figure 1B**, in which the cantilever inclination, the tip height, and the lateral tip-sample coupling are neglected. In the standard procedure, the values of two different CRFs (say  $f_n$  and  $f_m$  with  $n \neq m$ ) are measured and used to determine  $r$  and  $k^*$  by numerically solving  $k^*(f_n, r) = k^*(f_m, r)$  (Kester et al., 2000). The value of  $k^*$  is related to that of  $M_s$  through the relation

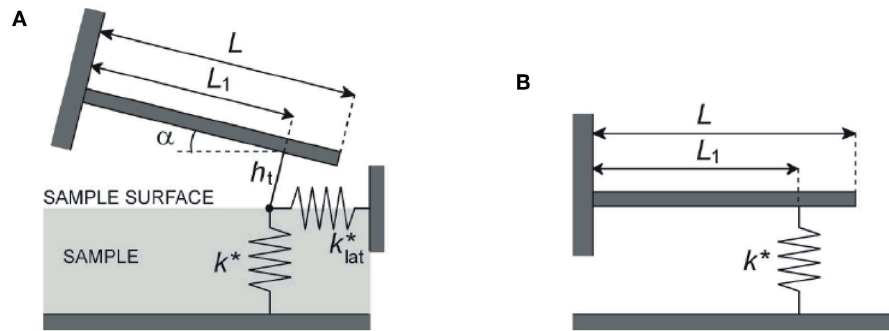
$$k^* = 2a_c M_s \quad (2)$$

where  $a_c$  is the tip-sample contact radius and assuming  $M_s \ll M_t$  and, thus,  $E^* \approx M_s$ . Using the Hertzian model to describe the tip-sample contact, Equation (2) can be rewritten as

$$k^* = \sqrt[3]{6M_s^2 R_t F_N} \quad (3)$$

where  $F_N$  is the static normal load applied by the tip on the sample as a result of the selected deflection set point. To evaluate  $M_s$  from  $k^*$ , the knowledge of the  $R_t$  is required. As  $R_t$  may change during a measurement session due to abrasion, especially when relatively stiff materials are investigated (Amelio et al., 2001), and thus the use of predetermined values of  $R_t$  in Equation (3) may result in a not accurate estimation of  $M_s$ . The uncertainty in the value of  $R_t$  is one of the major factors affecting the accuracy of CR-AFM measurements and, therefore, the actual tip radius (and also the tip shape) should be determined during each experimental session by calibrating the tip radius using one sample with well-known indentation modulus as reference material (Yamanaka et al., 2000; Kopycinska-Müller et al., 2006; Marinello et al., 2010, 2011). Really, to avoid the explicit calculation of  $R_t$ , after the contact stiffness  $k_{ref}^*$  is determined on the reference material with reduced modulus  $E_{ref}^*$ , the reduced modulus of the investigated sample  $E_s^*$  is calculated from the measured contact stiffness  $k_s^*$  as

$$E_s^* = E_{ref}^* \left( \frac{k_s^*}{k_{ref}^*} \right)^{3/2} \quad (4)$$



**FIGURE 1 | (A)** Beam model of the cantilever with length  $L$ , inclined of an angle  $\alpha$  respect to the sample surface; the tip is located at distance  $L_1$  from the cantilever clamped end, its height being  $h_t$ ; the tip interacts with the sample surface via both normal and lateral forces, and the contact is thus described by the normal and lateral contact stiffness modeled through the linear springs  $k^*$  and  $k_{lat}^*$ , respectively. **(B)** Simplified beam model of the cantilever, supposed parallel to the sample surface; the tip, the height of which is neglected, interacts with the sample only through normal forces modeled by the contact stiffness  $k^*$ . Reprinted and adapted from Passeri et al. (2013b).

if measurements on the investigated sample and on the reference material are performed at the same value of  $F_N$  (Rabe et al., 1996; Hurley et al., 2005; Passeri et al., 2005). For the sake of clarity, this procedure—which is admittedly the most widespread one—will be referred to as “single reference” approach in the rest of the paper. In more accurate calibration procedures not only  $R_t$  but also the indentation modulus of the tip  $M_t$  can be calibrated using two (or more) reference materials (Stan and Price, 2006). In particular, using two reference materials,  $M_s$  and  $M_t$

$$M_s = \frac{(k_{ref1}^*/k_{ref2}^* - 1)^{3/2}}{(k_{ref1}^*/k_s^*)^{3/2} (1/M_{ref2} - 1/M_{ref1}) + (k_{ref1}^*/k_{ref2}^*)^{3/2} 1/M_{ref1} - 1/M_{ref2}} \quad (5)$$

and

$$M_t = \frac{M_{ref1} M_{ref2} (k_{ref1}^{*3/2} - k_{ref2}^{*3/2})}{k_{ref2}^{*3/2} M_{ref1} - k_{ref1}^{*3/2} M_{ref2}} \quad (6)$$

where  $k_{ref1}^*$  and  $k_{ref2}^*$  are the contact stiffness measured on the first and second reference sample, respectively, and  $M_{ref1}$  and  $M_{ref2}$  are the indentation moduli of the two reference materials (Stan and Price, 2006). In particular, despite in a relatively limited range of elastic moduli, the use of multiple reference materials allows one to neglect lateral forces, thus using the model in **Figure 1B** instead of that in **Figure 1A** although apparent values of  $R_t$  and  $M_t$  (different from the real ones) are determined (Passeri et al., 2013b).

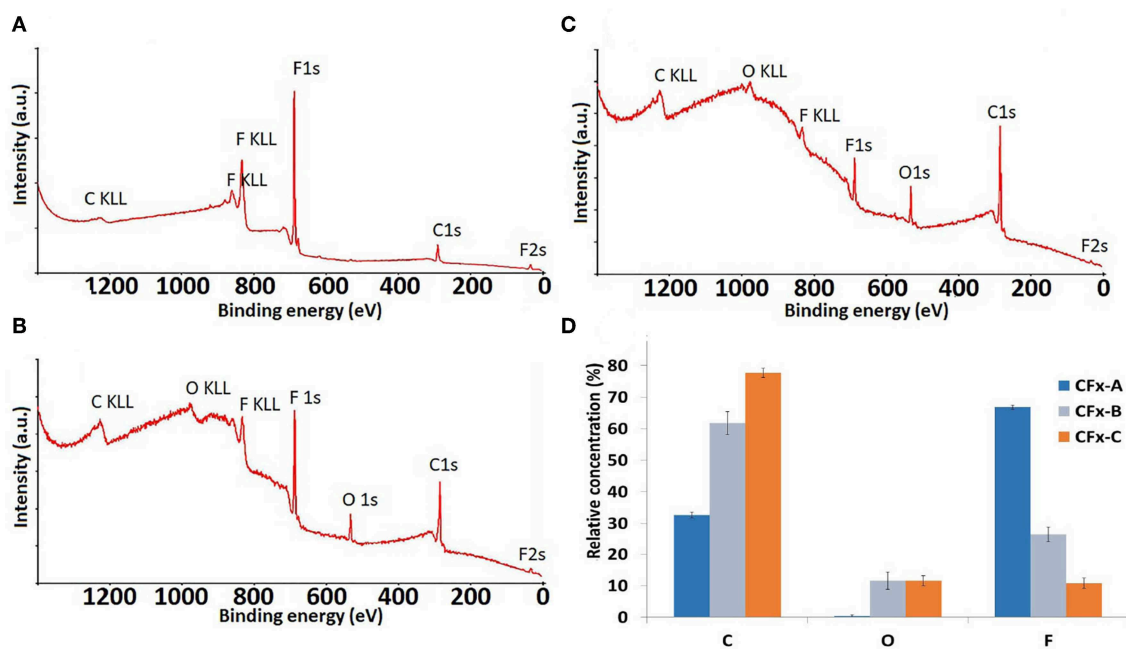
In this work, results of CR-AFM data analysis through the “single reference” approach were compared to those obtained using an original method, which we refer to as the “apparent stiffness” method. In the “apparent stiffness” method, the output of numerical simulations of CR-AFM experiments performed using the model in **Figure 1A** is analyzed using the simplified model reported in **Figure 1B** in order to obtain calibration curves

in which the apparent value of  $k^*$  is related to that of  $M_s$ . Therefore, these calibration curves are used to evaluate  $M_s$  from the values of  $k^*$  determined in the first steps of the “single reference” procedure.

Finally, not only can CR-AFM be used to study elastic but also viscoelastic materials (Killgore and DelRio, 2018). In order to account for the viscoelastic behavior of samples, a dashpot of dumping  $\sigma$  is included in parallel to  $k^*$  in the model in **Figure 1B** (Yuya et al., 2008). The tip-sample contact is described by the normalized tip-sample contact stiffness  $\alpha$  and the normalized damping coefficient  $\beta$ . These parameters can be evaluated measuring the frequency  $f_n$  of a given contact mode of the cantilever and the corresponding quality factor  $Q_n$  (Yuya et al., 2011). Finally, the values of  $\alpha$  and  $\beta$  can be used to evaluate the storage and the loss modulus of the sample ( $E'_s$  and  $E''_s$ , respectively) and the loss tangent defined as  $\tan \delta = E''_s/E'_s$ . In particular, a “single reference” approach can be followed, using a material with well-known storage and loss modulus ( $E'_{ref}$  and  $E''_{ref}$ , respectively). In this case, considering the tip much stiffer than the sample,  $E'_s$  and  $E''_s$  can be calculated as  $E'_s = E'_{ref} (\alpha/\alpha_{ref})^{3/2}$  and  $E''_s = E''_{ref} (f_n \beta / f_{n,ref} \beta_{ref})^{3/2}$ , where  $f_{n,ref}$ ,  $\alpha_{ref}$ , and  $\beta_{ref}$  are measured and calculated on the reference sample (Killgore et al., 2011). Conversely,  $\tan \delta$  can be evaluated without the need for calibration using a reference material through the relation

$$\tan \delta = (k_n L)^2 r^2 \frac{\beta}{\alpha} \frac{f_n}{f_{0,n}} \quad (7)$$

where  $k_n L = 7.855$  if the third mode of the cantilever is analyzed ( $n = 3$ ) (Hurley et al., 2013), which we used in this work as it was demonstrated to be more sensitive than, e.g., the first one (Killgore and Hurley, 2012). In this work, we evaluated  $E'_s$  using the “single reference” approach assuming the PS as the reference material,  $\tan \delta$  was evaluated using Equation (7), and finally  $E''_s$  was calculated as  $E''_s = E'_s \tan \delta$ .



**FIGURE 2 |** XPS survey spectra of the untreated CF<sub>x</sub> film (A) and CF<sub>x</sub> films treated with 30 s (B) and 90 s (C) methanol plasma treatment and histogram of the relative concentration of C, O, and F (D).

## 4. RESULTS

### 4.1. Chemical Analysis

Survey XPS spectra of CF<sub>x</sub> coatings before and after 30 and 90 s methanol plasma treatments are reported in **Figure 2** and show that methanol plasma treatment produced an increase of the oxygen amount of the coating (from 0% in CF<sub>x</sub>-A to 12% in both CF<sub>x</sub>-B and CF<sub>x</sub>-C); the increase of the time of methanol plasma treatment produced the increase of carbon amount (from 32.75 in CF<sub>x</sub>-A to 61.8% in CF<sub>x</sub>-B and 77.75 in CF<sub>x</sub>-C) and the corresponding decrease of fluorine content (from 66.9 in CF<sub>x</sub>-A to 26.5% and 10.7% in CF<sub>x</sub>-B and CF<sub>x</sub>-C, respectively). No metal species were detected, demonstrating that the coating was not damaged by methanol plasma treatment.

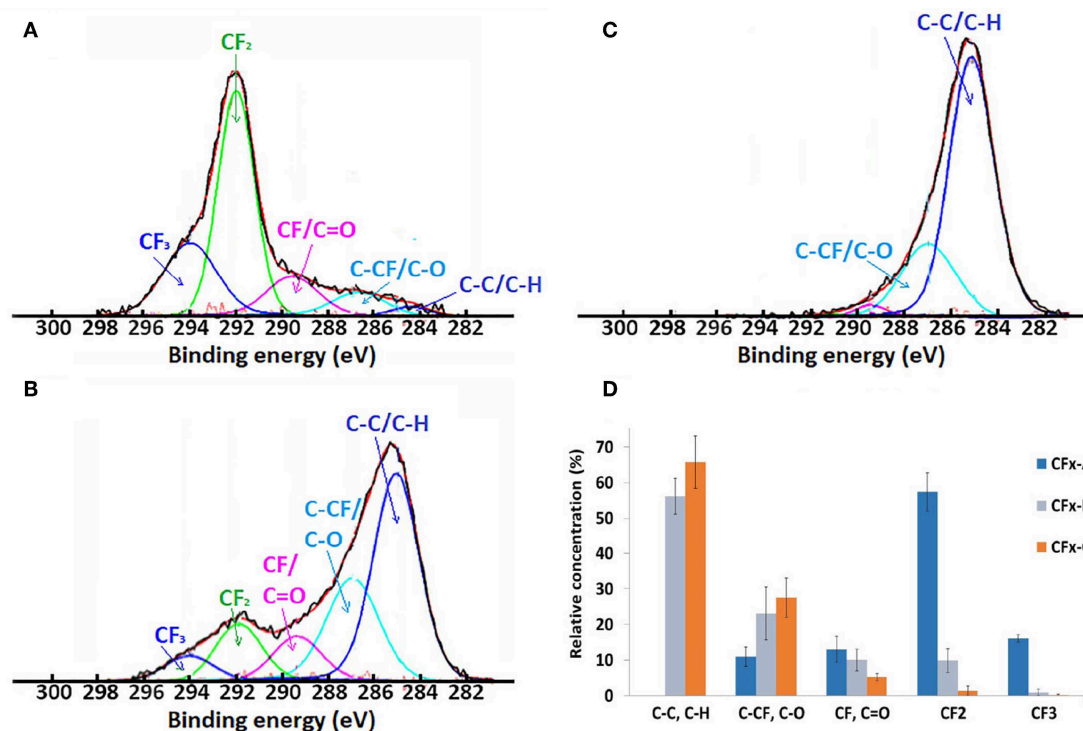
**Figure 3** shows high resolution XPS spectra for C(1s) of the unmodified CF<sub>x</sub> coating (a) and CF<sub>x</sub> coatings after 30 and 90 s methanol plasma treatments. The C(1s) spectra of the CF<sub>x</sub> films are fitted with five spectral components, as assigned to C-H/C-C- (BE = 285 eV), C-CF/C-O (BE = 287), CF/C=O (BE = 289.5 eV), -CF<sub>2</sub> (BE = 292), and CF<sub>3</sub> (BE = 294 eV) groups, according to literature (Horie, 1995; Mackie et al., 1997; Bourgoignie et al., 1999; Boehm, 2002). As <1 at.% of oxygen was detected at the surface of the untreated CF<sub>x</sub> coating, no attribution due to C-O (286.9) or C=O (288.1) is proposed, while for methanol treated samples, peaks at BE = 287 and 289.5 can be attributed to oxygen containing groups. **Figure 3A** shows the typical C(1s) spectrum of CF<sub>x</sub> coating without any methanol plasma treatment, which consists mainly of C-CF groups, CF<sub>2</sub> and CF<sub>3</sub> groups that are characteristic of fluorocarbon plasma-polymer films. High resolution spectra of CF<sub>x</sub> coating after 30 and 90 s methanol

plasma treatments (**Figures 3B,C**) show the increasing hindrance of the CF<sub>2</sub> and CF<sub>3</sub> bands—characteristic of CF<sub>x</sub> coatings—with the increase of the time of oxidative treatment, from 55 to 15% and <2%, respectively, and the corresponding increase of C-C and C-O containing species. This demonstrates that methanol plasma treatments allows the deposition of oxygen species not containing fluoride on the surface of fluorocarbon coating, without damaging it. The increase of the time of methanol plasma treatment produces the increasing covering of the fluorocarbon coating by oxygen species, which is also consistent with the previously reported time-of-flight secondary ion mass spectrometry (ToF-SIMS) analysis and the previously observed increased thickness of the coating (from 30 nm for untreated CF<sub>x</sub> coating to 41 nm for 30 s and 64 nm for 90 s methanol treated samples) (Montaño-Machado et al., 2019).

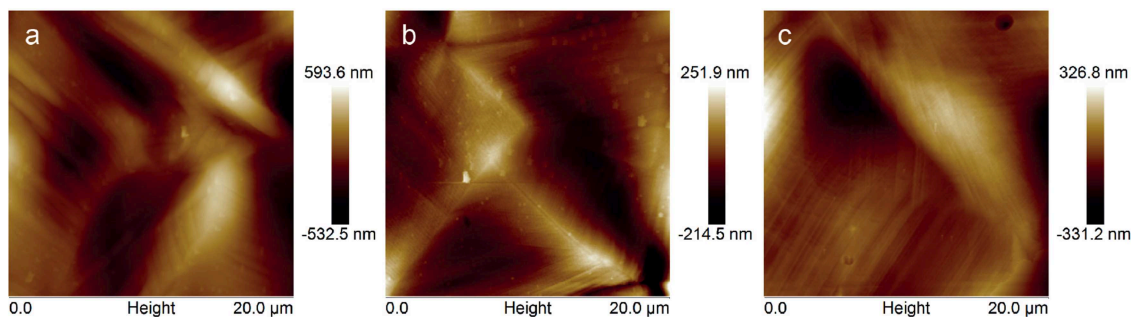
### 4.2. Cohesion and Adhesion After Deformation (Small Punch Test)

In **Figure 4** typical AFM topographies of the areas corresponding to the topmost part of the deformed samples are reported. AFM analysis of all the deformed samples (unmodified CF<sub>x</sub> and CF<sub>x</sub> after methanol treatment of 30 and 90 s) shows the occurrence of slip bands due to the plastic deformation of the underlying stainless steel substrate, but does not show any cracks, delamination or failure. This demonstrates that the methanol treatment did not significantly affect cohesion and adhesion properties of the CF<sub>x</sub> films, which resulted in having sufficient interfacial adhesion and cohesion to be employed as stent coatings.





**FIGURE 3 |** XPS high resolution C(1s) spectra of CF<sub>x</sub> film (A) and CF<sub>x</sub> films treated with 30 s (B) and 90 s (C) methanol plasma treatment and histogram of the relative concentration of the characteristic bands (D).



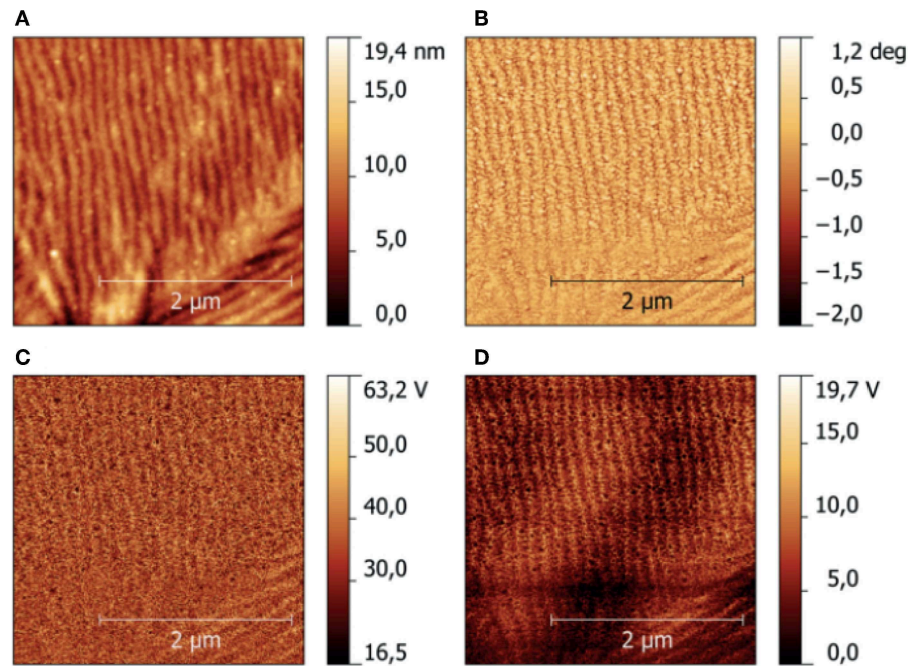
**FIGURE 4 |** AFM topography images of the topmost part of CF<sub>x</sub>-A (a), CF<sub>x</sub>-B (b), and CF<sub>x</sub>-C (c) samples after application of 25% deformation.

## 4.3. Nanomechanical Characterizations

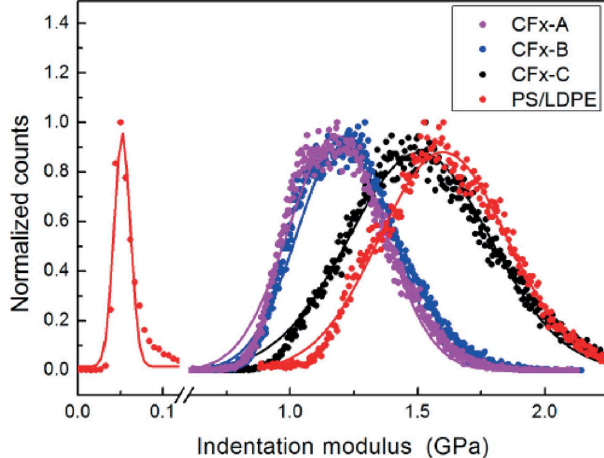
### 4.3.1. Elastic Modulus

A typical result of the characterization of CF<sub>x</sub> samples using HarmoniX<sup>TM</sup> is shown in Figure 5, where topography (Figure 5A), phase image (Figure 5B), map of the (not calibrated) indentation modulus (Figure 5C), and map of the (not calibrated) tip-sample adhesion force (Figure 5D) obtained on the sample CF<sub>x</sub>-C are shown. Morphological reconstruction shows the presence of ripples on the surface, indicating that the film reproduces the features typical of the stainless-steel substrate. These features are observed also in phase, indentation modulus, and adhesion maps, and can be ascribed to the

modulation of the local value of the tip-sample contact area (Stan and Cook, 2008) and to the variation of the local inclination of the surface with respect to the tip axis (Passeri et al., 2013b). Calibration of the indentation modulus maps was performed on the PS/LDPE reference sample (Passeri et al., 2013a). More specifically, the PS regions were used for calibration purposes, while LDPE regions were used to check the calibration range by comparing the obtained value to the one indicated by the vendor (100 MPa). In Figure 6, the histograms of the indentation modulus of the investigated CF<sub>x</sub> samples and those of the reference sample are reported. The obtained value of the indentation modulus of the LPDE is compatible with that



**FIGURE 5** | Example of nanomechanical mapping of the CF<sub>x</sub>-C sample using HarmoniX™: (A) height, (B) phase, (C) not calibrated indentation modulus, (D) not calibrated adhesion force.



**FIGURE 6** | Distribution of the values of indentation modulus of the three CF<sub>x</sub> samples and of the PS/LDPE reference, measured using HarmoniX™.

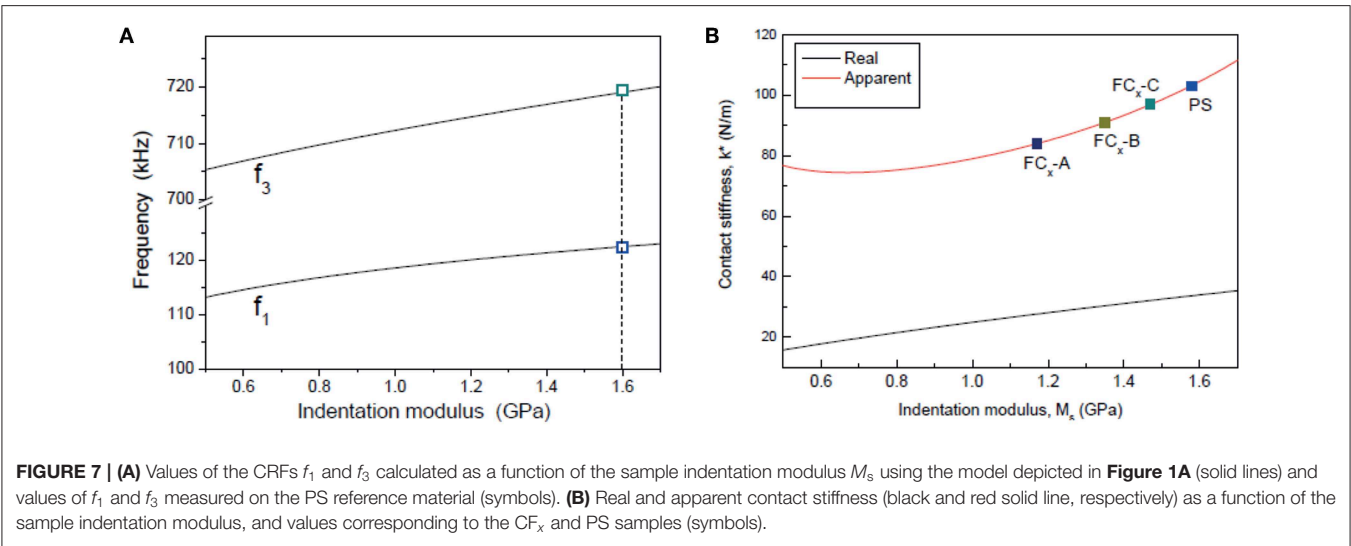
expected but lower ( $51 \pm 1$  MPa), indicating a not perfect calibration in low modulus range. Nevertheless, the moduli of CF<sub>x</sub> samples are comparable to those of PS, and thus the calibration can be considered accurate enough in the range of interest. Indentation modulus values with the corresponding uncertainty were obtained by Gaussian fitting of data in **Figure 6** and are reported in **Table 1**.

CR-AFM was first used to evaluate mechanical properties of the CF<sub>x</sub> thin films in elastic approximation. After

characterization of the free cantilever resonance in air, the tip was brought in contact with the sample which was made oscillate by the piezoelectric transducer coupled with its back side. Values of  $f_1$  and  $f_3$  were obtained from statistics on the corresponding CRFs maps acquired simultaneously to the topographical images. For calibration purposes, CRFs maps on the reference sample were acquired before and after each measurement session. To limit the effect of tip wear, CF<sub>x</sub> samples were purposely analyzed following the decreasing order of their expected stiffness, i.e., starting from the sample expected to be the stiffer one. **Table 1** reports the values of  $f_1$  and  $f_3$  measured on the CF<sub>x</sub> samples and the corresponding value of  $k^*$  calculated using the model in **Figure 1B**. The corresponding values of indentation modulus calculated through the “single reference” approach ( $M_s^{s+r}$ ) are then reported. The measured values of CRFs confirm that the mechanical properties of the CF<sub>x</sub> samples are close to those of the reference PS film ( $f_1 = 121–123$  kHz and  $f_3 = 704–715$  kHz), encouraging the use of the “single reference” approach. Conversely, CRFs measured on LDPE seemed too low use LDPE as a second reference sample ( $f_1 = 111–112$  kHz and  $f_3 = 660–685$  kHz). To verify the accuracy of the method, experimental data were analyzed through a numerical simulation (the “apparent stiffness” method). CR-AFM experiment was first simulated using the model in **Figure 1A** to obtain as output the pairs of CRFs  $f_1$  and  $f_3$  with a code which received as input mechanical properties of the cantilever and the tip, their geometrical parameters, and the mechanical properties of the sample (Passeri et al., 2013b). The characteristic parameters of the system were optimized in order to simultaneously match  $f_1$  and  $f_3$  experimentally

**TABLE 1** | Result of the nanomechanical characterizations using **HarmoniX™** and CR-AFM: indentation modulus obtained with **HarmoniX™** ( $M_s$ ); first and third flexural CRFs ( $f_1$  and  $f_3$ , respectively); calculated contact stiffness  $k^*$ ; indentation modulus calculated using the “single reference” method and the “apparent stiffness” method ( $M_s^{s-r}$  and  $M_s^{a-s}$ , respectively).

Sample	HarmoniX™			CR-AFM		
	$M_s$ (GPa)	$f_1$ (kHz)	$f_3$ (kHz)	$k^*$ (N/m)	$M_s^{s-r}$ (GPa)	$M_s^{a-s}$ (GPa)
CFx-A	$1.18 \pm 0.19$	$120.2 \pm 0.4$	$711.5 \pm 1.1$	84	$1.26 \pm 0.13$	$1.27 \pm 0.14$
CFx-B	$1.22 \pm 0.20$	$121.2 \pm 0.1$	$713.2 \pm 1.5$	91	$1.43 \pm 0.15$	$1.46 \pm 0.16$
CFx-C	$1.51 \pm 0.28$	$121.8 \pm 0.3$	$715.2 \pm 0.8$	97	$1.57 \pm 0.16$	$1.59 \pm 0.18$



**TABLE 2** | Measured values of the contact resonance frequency and quality factor of the third mode of the cantilever ( $f_3$  and  $Q_3$ , respectively), calculated values of the normalized contact stiffness ( $\alpha$ ) and the normalized damping ( $\beta$ ) and calculated values of the loss tangent  $\tan \delta$ , storage modulus ( $E'$ ) and loss modulus ( $E''$ ).

Sample	$f_3$ (kHz)	$Q_3$	$\alpha$	$\beta$	$\tan \delta$	$E'$ (GPa)	$E''$ (MPa)
CFx-A	$711.5 \pm 1.1$	$111 \pm 7$	724	0.057	$7.37 \times 10^{-3}$	1.26	9.31
CFx-B	$713.2 \pm 1.5$	$137 \pm 6$	792	0.051	$5.75 \times 10^{-3}$	1.45	8.31
CFx-C	$715.2 \pm 0.8$	$88 \pm 5$	833	0.011	$1.21 \times 10^{-3}$	1.56	1.88

obtained in contact with the PS film. Pair of  $f_1$  and  $f_3$  were than calculated as a function of  $M_s$  by varying  $M_s$  in the range 0.5–1.7 GPa, obtaining the curves reported in **Figure 7A**, where the values of CRFs corresponding to the PS reference sample are indicated (symbols). Then, assuming the model in **Figure 1B**, for each  $M_s$  the values of  $f_1$  and  $f_3$  were used to calculate  $k^*$ , which represents the “apparent” value of the tip-sample contact stiffness. Values of the apparent contact stiffness as a function of  $M_s$  are shown in **Figure 7B** (red solid line). For comparison, the values of real contact stiffness calculated as  $k^* \approx \sqrt[3]{6R_t F_N M_s^2}$  (black solid line), where  $F_N$  is the static normal load applied by the cantilever during the contact and  $M_s$  is used instead of the reduced modulus as  $M_s \ll M_t$ . As expected, the apparent value of  $k^*$  is bigger than the real one as a result of neglecting lateral forces (Passeri et al., 2013b). Finally, the values of  $k^*$  were calculated for the CF<sub>x</sub> samples using the model in **Figure 1B** and the curve of the apparent stiffness was used to determine the corresponding values of  $M_s$  (symbols in **Figure 7B**). The

procedure was repeated using the CRFs measured on the PS reference before and after the analysis of the CF<sub>x</sub> samples and the corresponding values of indentation modulus calculated using the “apparent stiffness” method ( $M_s^{a-s}$ ) are reported in **Table 1**.

#### 4.3.2. Viscoelastic Modulus

To characterize the viscoelastic response of the CF<sub>x</sub> films using CR-AFM, maps of the quality factor of the third mode ( $Q_3$ ) have been acquired in addition to those of  $f_3$  which are reported in **Table 2**. First, by approximating the samples as elastic,  $f_1$  and  $f_3$  were used as described in section 4.3.1 to determine the value of  $r$ , which was eventually used to calculate  $\alpha$  and  $\beta$  using a in-house written Matlab code (Passeri et al., 2013c) which are reported in **Table 2** together with the values of  $\tan \delta$  calculated using Equation (7). The values of  $\beta$  were used to calculate  $E'$  through the “single reference” approach after determining  $\beta$  on the PS reference sample and the values of  $E''$  were determined as



$E' \times \tan \delta$ .  $E'$  and  $E''$  calculated for the CF<sub>x</sub> samples are reported in Table 2.

## 5. DISCUSSION

CF<sub>x</sub> materials are attractive for the use as stent coatings, in particular after oxidative methanol plasma treatment which has been demonstrated to modulate the hydrophobicity of their surface in order to improve their performances, e.g., as far as blood contact behavior is regarded, without altering integrity, uniformity, and morphology (Montaño-Machado et al., 2019). Mechanical properties of CF<sub>x</sub> coatings can undergo significant changes when changes in the chemical composition and in the thickness occur. Indeed, it has been shown that fluorocarbon coatings thicker than 100 nm do not exhibit the required cohesion and adhesion properties to resist during stent expansion (Lewis et al., 2008). Furthermore, the elastic modulus of CF<sub>x</sub> coatings has been demonstrated to be strongly dependent on fluorine and crosslinking C–C units content (Tang et al., 2005). We showed that our methanol plasma treatment produces a significant increase of coating thickness and the decrease of fluorine content at the surface, which could significantly affect adhesion, cohesion and elastic modulus of the coatings. A comprehensive mechanical characterization was therefore necessary in order to evaluate the effect of methanol plasma treatment on the mechanical properties of the modified coatings.

Small punch tests have demonstrated that the innovative methanol plasma treatment does not undermine interfacial adhesion and cohesion to be employed as stent coatings as no delamination or cracks were observed even at plastic deformations as high as 25%, corresponding to the estimated maximum deformation which a stent may undergo during deployment (Migliavacca et al., 2005).

The effect of plasma treatment on mechanical properties of CF<sub>x</sub> films is another key issue that must be assessed to evaluate the suitability of CF<sub>x</sub> thin films as stent coatings. Indeed, depending on the polymerization and the post-deposition treatments, elastic modulus of CF<sub>x</sub> film is reported to vary from hundreds of megapascals like that of polytetrafluoroethylene (PTFE) (Ianev and Schwesinger, 2001), to a few gigapascals (Sirghi et al., 2009) and up to tens of gigapascals (Tang et al., 2005; Li et al., 2008; Koumoulos et al., 2012). Results reported in the present work show a good agreement between HarmoniX<sup>TM</sup> and CR-AFM. In particular, no discrepancies were observed between the two different methods for CR-AFM data analysis, i.e., the “single reference” approach and the simulation of the actual CRFs using the more comprehensive model depicted in Figure 1A and calculating the “apparent stiffness” using the simpler model in Figure 1B. The agreement between the two different approaches can be rationalized observing that all the investigated materials (CF<sub>x</sub> and PS) correspond to a region of the curve in Figure 7B in which  $k^*$  is proportional to  $M_s$ , i.e.,  $k^* = cM_s$  being  $c = 66$  nm. It must be observed that if softer materials were included among the investigated samples, the use of the single reference method using PS would imply an overestimation of their indentation modulus. Notably, considering the relatively

large thickness of the coatings (i.e., in the range 30–60 nm), the similar nanometer tip-sample contact radius values typical of CR-AFM and HarmoniX<sup>TM</sup> make the results of both the techniques representative of the sole CF<sub>x</sub> thin films indentation modulus, without the effect of the stainless steel substrate (Reggente et al., 2017). This undoubtedly represents an advantage of these (and similar) techniques over AFM-based nanoindentation, which more extensively suffers from substrate effect in case of thin compliant films on stiff substrates (Kovalev et al., 2004; Shulha et al., 2004; Clifford and Seah, 2006; Passeri et al., 2011). Nevertheless, in case of thinner films are investigated and/or tip with larger  $R_t$  are used, substrate effects may affect also CR-AFM and HarmoniX<sup>TM</sup> measurements and must be subtracted to obtain the elastic modulus of the sole film (Passeri et al., 2008, 2015). The obtained indentation modulus values, in the range 1.0–1.6 GPa, are compatible with those obtained on analogous materials using AFM based nanoindentation or conventional nanoindentation. Indeed, Sirghi et al. (2009) measured by AFM nanoindentation the elastic modulus of CF<sub>x</sub> films deposited by PECVD using C<sub>2</sub>F<sub>8</sub> as precursor and found values between 1.75 GPa and 3.2 GPa depending on the deposition parameters (power of the RF discharge and dc bias potential). Tang et al. (2005) studied, by nanoindentation, the hardness and the elastic modulus of fluorocarbon coatings deposited by RF magnetron sputtering using a PTFE target using different process parameters (RF power, Ar and H<sub>2</sub> flux) and found a dependence of the studied properties with the fluorine and carbon content and, more specifically, on the content of CF<sub>x</sub> or C–C crosslinking units. The measured elastic modulus decreased with the increase of fluorine content (i.e., CF<sub>x</sub> units) and the decrease of carbon (i.e., crosslinking units) content from 18 GPa (for films having 33% fluorine and 67% carbon content) to 12 GPa (for films having 43% fluorine and 57% carbon content) (Tang et al., 2005). Considering the significantly higher fluorine content of our untreated CF<sub>x</sub> coating (66.9%), the value of the elastic modulus we measured by HarmoniX<sup>TM</sup> (1.18 GPa) and CR-AFM (1.26 GPa) appears to be coherent with the results of Tang et al. (2005). As shown in Table 1, indentation modulus of CF<sub>x</sub> films has been found dependent on the methanol plasma treatment. Indeed, a slight increase of the elastic modulus of the coating with the increase of the time of methanol plasma treatment was observed. An increase of about 30% is observed between the as prepared sample and that treated with methanol plasma for 90 s, which is coherent with the covering of the fluorocarbon coating by a thin layer of carbon and oxygen species not containing fluoride.

As expected, an analogous dependence with plasma exposure time is observed in the measured viscoelastic parameters, i.e., in  $E'$ ,  $E''$ , and  $\tan \delta$ . In particular, although the comparison with PTFE properties is not always straightforward, we observe that the values of  $E'$  obtained on both untreated and plasma treated CF<sub>x</sub> films are coherent with those reported for PTFE (Faughnan et al., 1998; Fu and Chung, 2001; Blumm et al., 2010). Conversely, the values of  $\tan \delta$  and thus of  $E''$  are definitely lower than those observed on PTFE, indicating that the mechanical behavior of the films is characterized by a very low viscous component. However, it should be observed that viscoelastic parameters generally depend on the frequency at which the mechanical response of

the material is investigated, e.g., the viscous component may either increase or decrease with the frequency and depends on the specific polymer (Le Rouzic et al., 2009). Results by standard dynamical mechanical analysis (DMA) usually reported in literature are generally obtained for low frequency values, e.g., not exceeding 100 Hz, while CR-AFM investigates the sample at much higher frequencies, e.g., from tens to hundreds kilohertz or even a few megahertz (Hurley et al., 2013). In any case, plasma treatment is observed to be responsible for the reduction of the viscous component of the mechanical response of the films with respect to the untreated sample.

## 6. CONCLUSION

In conclusion, mechanical characterization of methanol plasma treated CF<sub>x</sub> ultrathin films on stainless steel demonstrated that methanol plasma treatment does not affect cohesion of the films and their adhesion to the substrate after deformation. Also, nanoscale analysis of elastic and viscoelastic response of films indicated that although these are slightly affected by methanol plasma, which is responsible for the stiffening and reduction of viscosity of the films. Nevertheless, such an effect is admittedly marginal and, especially from the point of view of the biological response, methanol plasma treatment does not significantly modify mechanical properties of the films. Thus, methanol plasma is a promising route to treat CF<sub>x</sub> ultrathin stent coating, which allows the modulation of the wettability

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- ## DATA AVAILABILITY STATEMENT
- The datasets generated for this study are available on request to the corresponding author.
- ## AUTHOR CONTRIBUTIONS
- LA realized the CF<sub>x</sub> thin films and carried out chemical and ancillary characterizations, small punch test, and AFM morphological analysis under the supervision of PC, ST, and DM. MRe performed the AFM based mechanical nanocharacterizations with the supervision of DP. MRe and DP analyzed the results of mechanical tests, with the supervision of MRo. MRe, LA, and DP wrote the first draft of the manuscript which was then revised by all the authors.
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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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