





Figure 1 – Geographical position of Coste San Giacomo site.

Sulimsky (1964) for *Sciurus*, to Van Weers (1994) for *Hystrix*, to Angelone (2008) for *Prolagus*, to Fanfani and Masini (1997) and Rümke (1985) for Eulipotyphla.

The small mammal collection is now housed at the IsIPU laboratory in Anagni.

### Systematic paleontology

Order Rodentia

Family Cricetidae FISHER, 1817

Subfamily Arvicolinae GRAY, 1821

Genus *Mimomys* FORSYTH-MAJOR, 1902

*Mimomys pliocaenicus* FORSYTH-MAJOR, 1902

Plate 1, Figs. 1, 2, 3, 4

Material: 39 M/1 (CSG 1-4, 32-33, 36, 38-40, 62-72, 74-77, 79, 81-85, 89-94, 96, 208); 29 M/2 (CSG 13-15, 41-43, 56-57, 60, 124-129, 138-139, 154, 169-1772, 182, 200-203, 215, 237); 18 M/3 (CSG 16-18, 130, 142-143, 173, 184-185, 205-207, 212-213, 231-232, 239-240); 41 M1/ (CSG 5-9, 44-46, 61, 98-104, 112, 133-135, 146-149, 164-165, 178-181, 187-193, 214, 217, 222-223); 38 M2 (CSG 10-12, 47-50, 54-55, 113-123, 136-137, 152-153, 166-167, 176, 183, 194-196,

Table 1 – List of small mammals remains from Coste San Giacomo (NR = numbers of remains; %NR = percentage of NR; MNI = minimum numbers of individual estimated; %MNI = percentage of MNI).

Taxon	NR	%NR	MNI	%MNI
<b>Rodentia</b>				
<i>Apodemus</i> sp.	1	0.36	1	1.89
<i>Mimomys pliocaenicus</i>	228	82.61	20	37.74
<i>Mimomys</i> gr. <i>tigliensis/tornensis</i>	6	2.17	3	5.66
<i>Sciurus</i> cf. <i>wartae</i>	1	0.36	1	1.89
<i>Castor fiber</i>	2	0.72	2	3.77
<i>Hystrix refossa</i>	7	2.54	4	7.55
<b>Lagomorpha</b>				
<i>Prolagus italicus</i>	10	3.62	4	7.55
<b>Eulipotyphla</b>				
<i>Beremendia fissidens</i>	6	2.17	6	11.32
Soricinae indet.	1	0.36	1	1.89
<i>Sorex</i> cf. <i>minutus</i>	1	0.36	1	1.89
<i>Galemys kormosi</i>	3	1.09	3	5.66
<i>Talpa minor</i>	8	2.90	5	9.43
<i>Talpa</i> sp.	3	1.09	3	5.66
<b>Total</b>	<b>276</b>	<b>100</b>	<b>53</b>	<b>100</b>

210, 218-220, 224, 235-236); 24 M3 (CSG 30-31, 59, 105-111, 140, 156-159, 163, 197, 211, 225-230); 7 hemi-mandibles (two specimens without the teeth and four with at least the M/1) (CSG 34, 73, 78, 80, 87-88, 258).

The dentine fields show a moderate confluence and the differentiation of the enamel thickness presents the “*Mimomys*” or negative type. The crown cement occurs, abundant in older and moderate in younger specimens. The anteroconid complex is fairly simple and the “*Mimomys* ridge” is always present. In the younger specimens a well-developed islet occurs, absent in the older specimens.

The hyposinuid and hyposinulid reach the occlusal surface in the adult to old individuals whereas in few young to very young specimens the hyposinulid does not reach the occlusal surface. This has been observed in all the complete teeth. The length of the m1 spans from 2.98 mm to 3.7 mm with an average of 3.31 mm (Tab. 2).

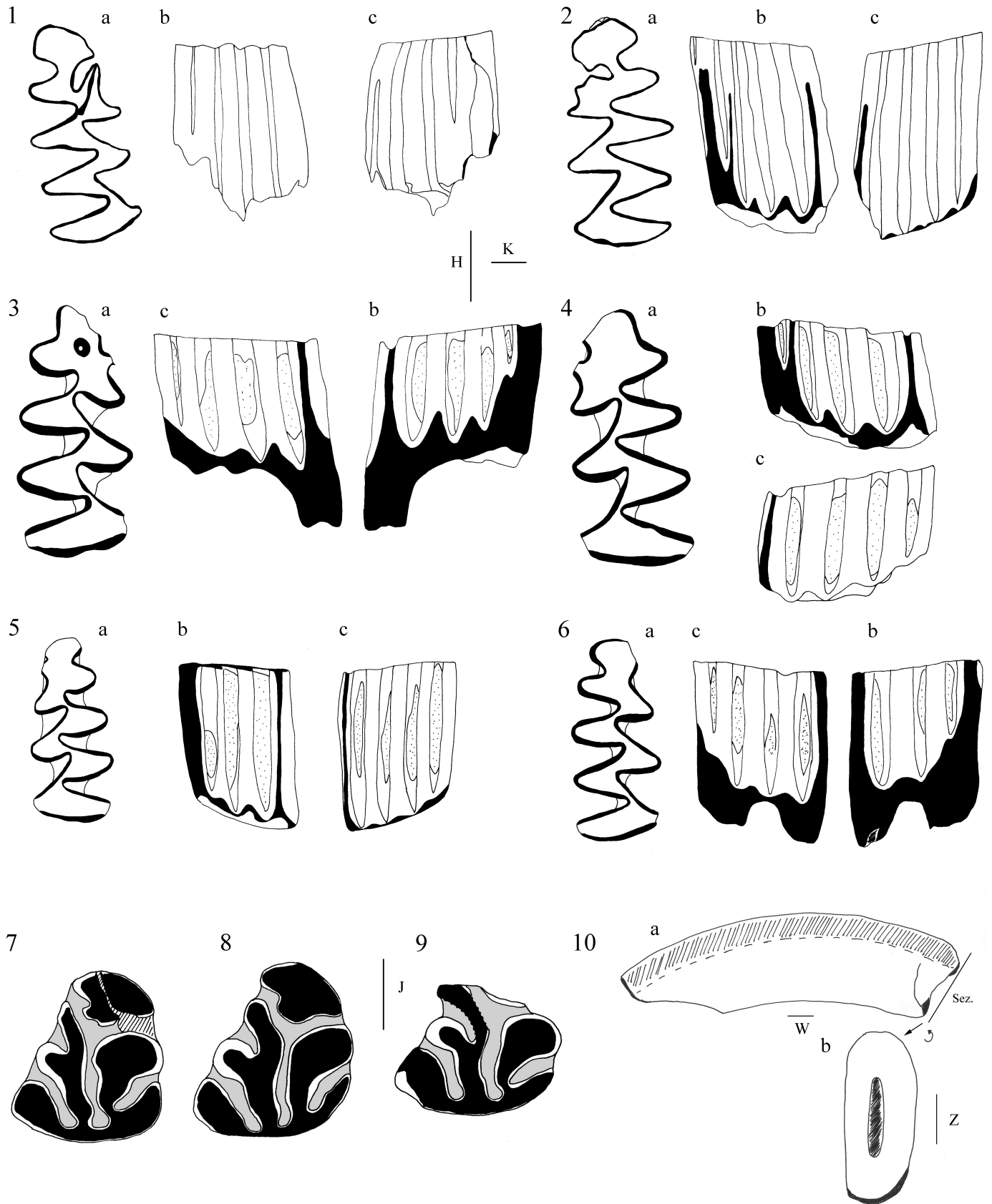
Table 2 – Measurements of lower first molar of *Mimomys pliocaenicus* from CSG (L: maximum occlusal length; W: maximum occlusal width; AL: anteroconid length; LHC: lingual height of the crown; A/L: (A/L)/L. Measures in mm).

M/1	n	max	min	sd	mean
L	25	3.70	2.86	0.23	3.31
W	37	1.77	0.98	0.15	1.42
AL	32	1.75	1.23	0.14	1.47
LHC	19	5.69	1.84	1.17	4.02
A/L	21	47.60	37.89	2.86	44.00

The great hypsodonty of the m1 of CSG permits to discard its attribution to *Mimomys polonicus*; the dimensions fall within the range of *Mimomys pliocaenicus* (Tesakov, 1998; Sala et al., 1994). The M/1 seems to be smaller and it shows a less evolved sinuous line than *Mimomys ostramosensis* (Carls and Rabeder, 1988). The first lower molars of CSG show more archaic features than the holotype if compared with *Mimomys pliocaenicus* from Castelfranco di Sopra (see Fig. 3, drawing 2 in Masini and Torre, 1987). The Castelfranco di Sopra specimen is in fact more similar to *Mimomys ostramosensis* than to *Mimomys pliocaenicus* of Tegelen considering the development of the sinuous line. It is worth noting that the CSG large *Mimomys* specimens represent the largest collection of this arvicolid species and therefore can be considered as a reference. A detailed work on the morphological and biometrical data of *Mimomys pliocaenicus* from CSG is in progress.

*Mimomys* gr. *tigliensis-tornensis*

Plate 1, Figs. 5, 6



**Plate 1** – *Mimomys pliocaenicus*: 1-4, four different age development; 1 – M/I right, very juvenile specimen (CSG 81); 2 – M/I left, juvenile specimen (CSG 94); 3 – M/I right, adult specimen (CSG 84); 4 – M/I left, senile specimen (CSG 91). *Mimomys gr. tigliensis-tornensis*: 5 – M/I left (CSG 95); 6 – M/I right (CSG 97). a: occlusal view; b: lingual view; c: buccal view. H-K: scale bar = 1 mm, H occlusal view and K lateral view. *Prolagus italicus*: 7 – P/3 left (CSG 265); 8 – P/3 left (CSG 266); 9 – P/3 left (CSG 269). J: scale bar = 1 mm. *Sciurus cf. warthea*: 10 lower right incisor. a: lateral view; b: section. W-Z: scale bar = 1mm, W lateral view and Z section.

Material: 3 M/1 (CSG 86, 95, 97); 2 M/2 (CSG 145, 204); 1 M3/ (CSG 141).

The small representatives of the genus *Mimomys* are very scarce in the Coste San Giacomo collection, being the ratio of small *Mimomys* M/1 to *M. pliocaenicus* M/1 is 1 to 13. This fact is quite unusual, since in other coeval sites in Europe (Tegelen, Rivoli Veronese, etc.) the small *Mimomys* teeth are more numerous than those of the large *Mimomys pliocaenicus*. At Rivoli Veronese the ratio is 1 to 7 (Sala et al., 1994) and at Tegelen is 1 to 4 (Tesakov, 1998). Large arvicolid are usually less represented than the small ones also in the Middle and Late Pleistocene sites where the ratio *Arvicola/Microtus* is always much less than one (see for example: Fontana Marella cave – Bona et al., 2008; Caverna Generosa – Bona et al., 2009; Salnova quarry – Bona, 2011). This peculiar ratio in the CSG collection, considering the good accuracy of the sieving, may be due to the environmental condition surely more favourable to the large *Mimomys*.

The two complete first lower molars of the small *Mimomys* show a mix of features between two very similar species: *M. tigliensis* and *M. tornensis*. In particular, the small specimen CSG-95 shows all the typical dimensions and features of *M. tigliensis*: abundant cement in the re-entrant angles lacking the enamel islet, prominent *Mimomys*-ridge in younger stages of wear, rounded anterior cap of the anteroconid (Tesakov, 1998, Fig. 24 numbers 5 and 6). On the other hand, the specimen CSG 97 shows the *M. tornensis* size and features (Tab.3). According to Tesakov (1998) *M. tornensis* differs from *M. tigliensis* in the following features: larger size, more advanced hypsodonty, lower percentage of *Mimomys*-ridge occurrence and less developed roots.

At the present day it is neither possible to ascribe our specimens to *M. tigliensis* or *M. tornensis* nor to establish the contemporary occurrence of these two taxa.

**Table 3** – Measurements of lower first molar of *Mimomys* gr. *tornensis-tigliensis* from CSG (L: maximum occlusal length; W: maximum occlusal width; AL: anteroconid length; LHC: lingual height of the crown; A/L: (A/L)/L. Measures in mm).

M/1	n	max	min	sd	mean
L	2	2.86	2.58	0.20	2.72
W	2	1.14	1.05	0.06	1.10
AL	2	1.30	1.16	0.10	1.23
LHC	2	3.90	3.03	0.54	3.52
A/L	2	45.45	44.96	0.35	45.21

Family Muridae GRAY, 1821

Genus *Apodemus* KAUP, 1829

Material: 1 damaged left hemi-mandible without teeth (CSG 23)

The specimen CSG 23 is a horizontal ramus of left hemi-mandible without teeth. The overall morphology and distribution of the alveoli, six in line, allow to ascribe this fossil remain to *Apodemus* sp.

Family Gliridae THOMAS, 1897

Genus *Sciurus* LINNAEUS, 1758

*Sciurus* cf. *S. warthae* SULIMSKY, 1964

Plate 1, Fig. 10

Material: 1 right lower incisor (CSG 22)

The dimensions (antero-posterior diameter: 3.52 mm; transverse diameter: 1.60 mm) and the overall morphology permit a positive comparison of CSG 22 with the remains of *Sciurus warthae* from Weze I (Sulimsky, 1964). The measurements of CSG 22 clearly fall within the range of *Sciurus warthae* and the broad transversal diameter strengthens this determination.

Family Castoridae HEMPRICH, 1820

Genus *Castor* LINNAEUS, 1758

*Castor fiber* LINNAEUS, 1758

Material: 2 right M/1-2 (CSG 25, CSG 242)

The hypoflexid does not reach the half of the tooth width and it is slightly curved backwards. The paraflexid and the mesoflexid are coalescent forming an oval island between the flexids. The absence of

secondary enamel folds permits to rule out its attribution to the sub-species *Castor fiber plicidens* Barisone et al., 2006). The measurements of the specimen are CSG 25: L= 9.7 mm, W= 9.8 mm; CSG 242: L= 8.7 mm, W= 9.1 mm.

Family Hystricidae Burnett, 1830

Genus *Hystrix* LINNAEUS, 1758

*Hystrix refossa* GERVAIS, 1852

Material: 2 left M1/ or M2/ (CSG 243, 244), 1 left DP/4 (CSG 28), 1 left P/4 (CSG 29), 1 P/4? (not well preserved -CSG 245-), 1 right M/1 or M/2 (CSG 247), 1 right M/3 (unworn and rootless -CSG 246-)

The permanent teeth are strongly hypsodont (Tab. 4) and fit perfectly fit within the *Hystrix refossa* range (Van Weers, 1994; Rook and Sardella, 2005); the teeth are larger than *H. vinogradovi* and they are similar in size to those of *H. primigenia* but they are more hypsodont.

The analysis of the wear stage (reported according to Van Weers and Rook, 2003) allows to hypothesize the occurrence of 4–5 individuals at least (Tab. 4, last line).

The deciduous DP/4 (CSG 28) shows a high wear stage on the occlusal surface with seven enamel islands and a very low crown, testified by E/L index (Tab. 4). The two M1/ or M2/ are teeth with three roots folded in the bucco-lingual direction of different wear stage and with quite different dimensions (Tab. 4); the occlusal surface shape ranges from sub-quadrangular to sub-rectangular. The P/4 (CSG 29) shows a sub-rectangular occlusal surface and an intermediate wear stage with three folds and three islands; the crown is straight and well developed. The M/1 or M/2 is large and slightly bent in bucco-lingual direction; the occlusal surface has a sub-rectangular shape.

**Table 4** – Measurements of *Hystrix refossa* teeth (L: maximum occlusal length; W: maximum occlusal width; E: enamel buccally maximum length; El: enamel lingually maximum length; E/L: (E/L)×100. Measures in mm).

	28	29	243	244	245	246	247
	DP/4	P/4	M1/ or M2/	M1/ or M2/	P4?	M/3	M/1 or M/2
L	8.9	10.2	9.0	10.3	8.8	10.1	10.4
W	7.1	8.0	7.3	10.2	6.3	7.5	8.5
E	8.0	14.6	11.8	10.9		14.0	14.3
E1	6.6	15.9	19.0	17.8		15.1	
E/L	90	143	131	106		139	137
Wear class	T	Q-R	D	G		O	R-S

Order Lagomorpha

Family Ochotonidae THOMAS, 1897

Genus *Prolagus* POMEL, 1853

*Prolagus italicus* ANGELONE, 2008

Plate 1, Figs. 7, 8, 9

Material: 4 P/3 (CSG 265, 266, 267, 269), 4 fragments of P/4 or M/1 or M/2 (CSG 268, 272, 273, 274), 1 P2/ (CSG 271)

The morphological features of the P/3 permit to attribute the CSG Lagomorpha to the genus *Prolagus* and, in particular, to *Prolagus italicus* ANGELONE, 2008. The characteristics of the P/3 are: i) all P/3 lack the crochet, in holotype population described by Angelone (2008) the crochet occurs in more than 50% of samples; ii) the hypoconid and the entoconid are well developed; iii) the mesoflexid is developed and has a “S” shape; iv) the Anteroconid is sub-triangular and well developed; v) the Metacone and Anterocone have approximately the same dimensions; vi) the Protoconid is undeveloped vii) the Protoconulid is developed; viii) the P/3 fall in the dimensional range (Tab. 5) of *P. italicus* whereas they are slightly larger than of *P. savagei* and *P. sorbini*.

The P2/ is not well preserved but the overall morphology shows characters similar to those highlighted by Angelone (2008) for *P. italicus*. The fragments of molars are poorly preserved and not allow further analyses.

Order Eulipotyphla

Family Soricidae GRAY, 1821

**Table 5** – Measurements of *Prolagus italicus* teeth (L: maximum occlusal length; W: maximum occlusal width. Measures in mm).

P/3	CSG 265	CSG 266	CSG 267	CSG 269	TP*	MS*
L	2.41	2.28		2.42	2.03	2.17
W	1.98	2.03	1.86	2.13	2.10	2.63
P2/	CSG 271				TP*	MS*
L	1.40				1.20	1.51
W	2.00				1.65	2.30

\* average data from Angelone (2008), TP means “Cava Toppetti” and MS means “Montagnola Senese”.

Subfamily Soricinae FISHER, 1817

Soricinae indet.

Plate 2, Fig. 9

Material: 1 damaged right hemi-mandible with alveoli of M/2-3 (CSG 53)

The specimen CSG 53 is not well preserved; it lacks all the teeth and about the mesial half of the horizontal ramus. The morphology of CSG 53 is similar to that of CSG 20 but CSG 53 is significantly smaller (Tab. 6).

**Table 6** – Measurements of Soricidae remains (I/I. L: length of crown. Hemi-mandible. H: height of ascending ramus; L: length from point P to mental foramen (see ?); LI: length from point P to end of I/I alveolus; S1, S2, S3: height of the horizontal ramus in correspondence of the distal end of M/1, M/2 and M/3 respectively; M1, M2, M3: length of M/1, M/2 and M/3; SA1, SA2, SA3: width of the trigonid of M/1, M/2 and M/3; SP1, SP2, SP3: width of the talonid of M/1, M/2 and M/3; Condyle. LUF: length of upper condilar facet; LLF: length of lower facet; HC: maximum height of condyle. Measures in mm.

I/I	CSG 262	CSG 263		
L	5.57	6.11		
Hemimandible	CSG 20	CSG 261	CSG 53	CSG 275
H				3.22
L				5.44
L1				4.74
S1		2.05		0.74
S2		2.42		0.85
S3		2.71	2.09	0.97
M1		2.95		
SA1	1.50	1.51		
SP1		1.64		
M2	2.58*	2.49	1.74*	1.04
SA2		1.55		0.69
SP2		1.46		0.74
M3	1.51*	1.60	1.37*	
SA3		1.12		
SP3		0.87		
M1-M3	5.67*			
TI				0.82
TM3				2.11
Condyle				
LUF				0.53
HC				0.97
LLF				0.77

\* alveolar length.

Tribe Beremendiini REUMER, 1984

Genus *Beremendia* KORMOS, 1934

*Beremendia fissidens* PETÉNYI, 1864

Plate 2, Figs. 8, 10, 11

Material: 3 damaged right hemi-mandibles: 1 horizontal ramus with fragmented M/1 (CSG 20), 1 horizontal ramus with M/1-3 (CSG 261),

1 fragmented distal portion with alveoli of M/2-3 (CSG 264); 3 right I/1 (CSG 21, 262, 263)

Measurements: see Tab. 6

The incomplete right hemi-mandible CSG 20 is very large, showing a robust morphology and the fragmented M/1 has an intense dark red colouration. Specimen CSG 261 is similar to CSG 20, but it is better preserved, conserving the M/1, M/2 and M/3. According to the dimensions and the overall morphology described by many authors (e.g., Sala et al., 1994; Rzebik-Kowalska, 1976) it is possible to ascribe the specimens CSG 20, 261, 264 to *Beremendia fissidens*. CSG 262 and CSG 263 are well preserved I/1. They are acuspidate large teeth, long, knife-shaped with apex curved up and strongly coloured. There is a characteristic thin cingulum on the posterior dorsal margin of the buccal side. CSG 21 is a very small fragment of I/1 but according to its relative dimension and to the very dark red colour, it can be assigned tentatively to *Beremendia cf. fissidens*.

Tribe Soricini FISHER, 1817

Genus *Sorex* LINNAEUS, 1758

*Sorex cf. S. minutus*

Plate 2, Fig. 12

Material: 1 right hemi-mandible with half M/1 and a complete M/2 (CSG 275)

Measurements: see Tab. 6

The hemi-mandible, despite the absence of redness on the teeth cusps, has a slender horizontal ramus with a very thin symphysis portion, which contrasts with others crocidurinae shrews of similar size, like *Suncus*. According to these features, CSG 275 is attributable to the genus *Sorex*. The mandibular ramus is low and the central part of its lower border is concave. The condyle presents a small upper facet that is large, if compared to others soricinae, and it bends down on the lingual side and a broad interarticular area. The mental foramen is located below the first root of M/1. According to the hemi-mandible and teeth sizes, it is possible to attribute the specimen, with some doubts, to *Sorex cf. minutus*.

*Sorex minutus* is one of the smallest representative of the genus *Sorex*. This shrew species was widespread in Europe since the earliest Pliocene (Rzebik-Kowalska, 1998). The *S. minutus* of CSG therefore represents the oldest finding in the Italian peninsula. Until now, in Italy, *S. minutus* had been only reported in the Early Biharian sites of Monte La Mesa (Marchetti et al., 2000).

Family Talpidae FISCHER, 1817

Subfamily Talpinae FISCHER, 1817

Tribe Talpini FISCHER, 1817

Genus *Talpa* LINNAEUS, 1758

*Talpa minor* PETÉNYI, 1864

Plate 2, Figs. 6, 7

Material: 3 hemi-mandibles (CSG 37 is a left mesial portion of horizontal ramus with P/2, P/4 and M/1, CSG 251 -left- and CSG 253 -right- are fragments of horizontal ramus without teeth), 3 ulna fragments (2 right and 1 left), 1 left radius distal fragment, 1 left M/1.

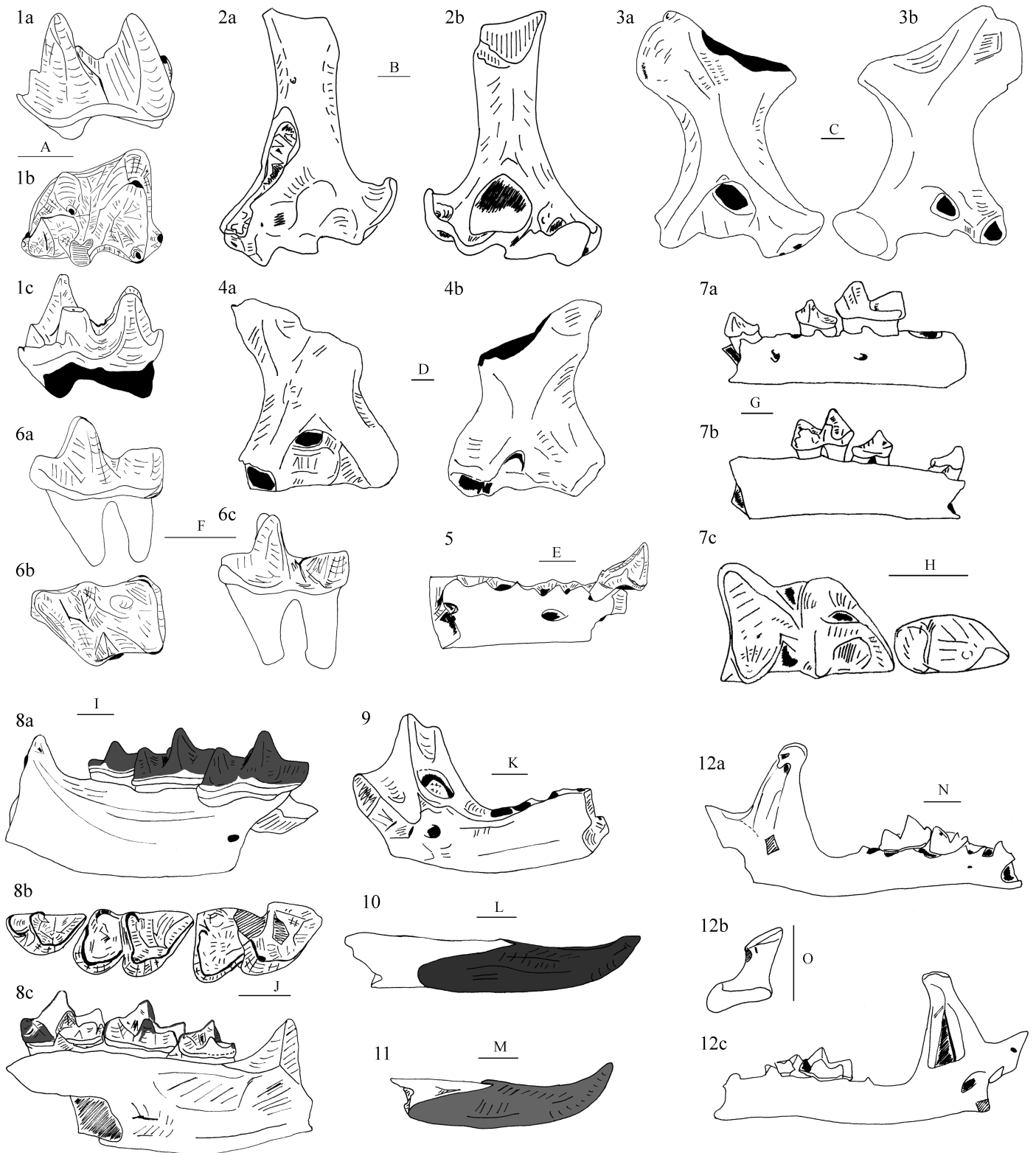
All morphological characters shown by specimen are typical of *Talpa* genus, the size of the hemi-mandibular fragments and post-cranial elements are smaller than those of *Talpa europaea* LINNAEUS, 1758 and they fall within the range of the *minor-caeca* group (Tab. 7).

*Talpa* sp.

Plate 2, Figs. 3, 4, 5

Material: 2 damaged humeri, 1 right (CSG 26) and 1 left (CSG 27), 1 mesial fragment of right hemi-mandible with p1 (CSG 260).

The two humeri are heavily damaged and it has been possible take only a few measurements. These measurements (Tab. 7) fall within the range of *Talpa cf. T. fossilis* from Monte Peglia (Van der Meulen, 1973), but also with that of *Talpa cf. T. minor* from Monte La Mesa (Marchetti et al., 2000). New finds are necessary to give more precise information about the taxonomic status of the Coste San Giacomo moles.



**Plate 2** – *Galemys kormosi*: 1 – right M/2 (CSG 259) (A); 2 – left humerus (CSG 24) (B). *Talpa* sp.: 3 – right humerus (CSG 26) (C); 4 – left humerus (CSG 27) (D); 5 – right hemi-mandible fragment with P/1(CSG 260) (E). *Talpa minor*: 6 – left M/1 (CSG 254) (F); 7 – left hemi-mandible fragment with with P/2, P/4 and M/1 (CSG 37) (G). Soricinae indet.: 9 – left hemi-mandible fragment without teeth (CSG 53) (K). *Beremendia fissidens*: 8 – right hemi-mandible with M1-M3 (CSG 261) (I: lateral view, J: occlusal view); 10 – right lower Incisive (CSG 283) (L); 11 – right lower Incisive (CSG 262) (O). *Sorex* cf. *minutus*: 12 – right hemi-mandible with M1-2 (CSG 275) (N: lateral view, O: condyle view). All scale bar are 1 mm (in capital letter in the brackets at the end of the description of specimen).

**Table 7** – Measurements of Talpidae remains (Hemi-mandible and teeth. Sl: height of the horizontal ramus in correspondence of the distal end of M/1; M1, M2: length of M/1 and M/2; SA1, SA2: width of the trigonid of M/1 and M/2; SP1, SP2: width of the talonid of M/1 and M/2; P4: length of P/4; SP4: width of P/4; P1: length of P1; PIW: width of P/1. Measures in mm).

Humerus	CSG 26	CSG 27	CSG 24	
Minimum width	3.23	3.88		
Distal width	6.96		5.67	
Hemimandible and M/1	CSG 37	CSG 254	CSG 52	CSG 259
S1	2.06			
M1	2.13	1.84		
SA1	1.24	1.19		
SP1	1.41	1.10		
M2			2.35	2.40
SA2			1.42	1.45
SP2			1.67	1.75
P4	1.39			
SP4	0.69			
P1	1.11			
P1 W	0.53			

Subfamily Desmaninae RÜMKE, 1985

Genus *Galemys* KAUP, 1829

*Galemys kormosi* (SCHREUDER, 1940)

Plate 2, Figs. 1, 2

Material: 1 distal half of left humerus (CSG 24); 2 M/2, 1 right (CSG 259) and 1 left (CSG 52).

The M/2 show the typical talpids morphology. The cusps are sturdy than in the genus *Talpa*. The oblique crista is long and it reaches the metaconid tip. A strong anterior cingulum is present, widening just before reaching the lingual border forming a kind of parastylid. A strong entostylid is present on the posterior side.

The humerus shows a shape not typical of fossorial talpids but typical of all representatives of this subfamily (Rümke, 1985). The bone preserves a few more that the distal half with a narrow diaphysis and an enlarged distal epiphysis.

The morphology and the dimensions (Tab. 7) fall in the range of the *Galemys kormosi*, in particular with the *G. kormosi* from Tegelen (Rümke, 1985).

Therefore, the CSG water mole findings represent the first report of the Desmaninae subfamily in Italy.

## Discussion and final remarks

Rodents are very important elements in fossil mammal faunal assemblages and, among them, arvicoline have an extraordinary biochronological value. The evolutionary lineage including the Pliocene-Early Pleistocene *Mimomys* (with rooted molars) and the Middle to Late Pleistocene *Arvicola* (with not rooted molars) is of crucial importance for defining a biochronological scheme at an European scale (Maul et al., 1998). Since *Mimomys pliocaenicus* characterizes the earliest Pleistocene micromammal assemblages the occurrence of the rich sample at CSG is of particular interest and provides a valuable amount of biochronological and palaeoecological information.

The morphology of the lower first molar of the large vole *Mimomys pliocaenicus* permits to compare the CSG teeth with those from other Italian and European sites such as Osztramos 3 (Hungary) and, Tegelen (The Netherlands). The large *Mimomys* of CSG shows a greater hypsodonty than the archaic species *Mimomys hajnackensis* and *Mimomys polonicus* (Sabot et al., 2006; Mayhew et al., 2008). At the same time, it seems to be more brachyodont than the type specimen of *M. pliocaenicus* from Castelfranco di Sopra (Upper Valdarno, Tuscany) correlated with the Olivola FU (Gliozzi et al., 1997), and therefore it is considered less advanced.

According to Masini and Torre (1987), the type of *M. pliocaenicus* is comparable in size and hypsodonty degree to *Mimomys ostramosensis* from Osztramos 3 (for discussion see Sala et al., 1994). The *Mimomys*

*pliocaenicus* of CSG can also be compared with *Mimomys* cf. *pliocaenicus* of Rivoli Veronese (Sala et al., 1994) and the specimens of Tegelen (Tesakov, 1998), attributed to the Middle Villafranchian or Late Villafranchian Small Mammal Age (Fejfar and Heinrich, 1990). Compared with the minor degree of hypsodonty seen in *Mimomys* of Castelfranco di Sopra, the *M. pliocaenicus* of CSG can be considered therefore older than the Olivola FU (Tab. 8). This is in accordance with the age inferred from the study of the large mammals of the site, FU Coste San Giacomo (Gliozzi et al., 1997).

The CSG small mammal faunal assemblage provides important and new data on the biochronology and palaeoenvironment of the Mediterranean fauna during the Gelasian (Early Pleistocene).

In Tab. 8 is presented a detailed comparison between CSG and some selected European Late Villafranchian/Early Biharian sites. It is worth noting that the Eastern European small mammal taxa are absent in the CSG mammal assemblage. Thereafter the CSG small mammals fauna represents, during the Early Pleistocene in central Italy, a phase prior to the expansion of eastern taxa into South-western Europe as demonstrated by the lack of the genera *Villanyia*, *Ungaromys*, *Dinaromys* and *Ellobius* that are present among others at Rivoli Veronese (Sala et al., 1994) in the North-eastern Italy. The occurrence of the endemic *Prolagus* also strengthens this hypothesis. Moreover, the CSG fauna confirms the persistence in central Italy of environmental conditions, maybe characterized by a less continental climate than Rivoli Veronese, that have retarded the distribution of Eastern taxa already present in North Italy.

The association of both small and large mammalian assemblages is of great and unique biochronological importance and confirms the correlation of the CSG mammal assemblage at the end of the Middle Villafranchian and at the end of Villafranchian (Masini and Sala, 2007, 2011), in a crucial moment influenced by the diffusion of open environments, including steppe taxa, in the Mediterranean Europe (Bertini, 2010).

During the Gelasian the area of CSG was characterized by the presence of a braided streaming (data from geological and stratigraphical analyses, see Bellucci et al., 2014). The small and large mammal assemblage allowed to hypothesize that the surroundings of this watercourse was characterized by the presence of mainly grasslands, testified by the presence of abundant rests of *Equus stenorhinus* and abundant arvicolids. The wooded areas are, probably, in regressive phase; the last *Anancus* and the very poorly represented Eulipotyphla and Muridae testified the presence of reduced and unstructured covered areas. The humid environment is limited to the surroundings of the watercourse and it is characterized by the presence of the beavers, the hippopotamus, the large arvicolid *Mimomys pliocaenicus* and the water mole *Galemys kormosi*.

In conclusion, the new palaeontological data from the small mammals from CSG, in particular of large vole *Mimomys pliocaenicus*, have confirmed an age of the mammal assemblage older than Olivola FU (Gliozzi et al., 1997). Furthermore the small mammal assemblage provides evidence of the persistence in central Italy of environmental conditions that have retarded the distribution of Eastern taxa already present in Northern Italy. Finally, in this work, it has been reported for the first time, in Italian peninsula, the occurrence of the water mole *Galemys kormosi* belonging to the Desmaninae subfamily. ☞

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Table 8 – Late Villanyan/Early Biharian selected European sites.

Site	Galera 2 <sup>1</sup>	Barranco conejos <sup>2</sup>	Saint-Vallier <sup>3</sup>	Tegelien <sup>4</sup>	Schernfeld <sup>5</sup>	Kadzielinia <sup>1</sup> <sub>6</sub>	Oztramos-3 <sup>7</sup>	Coste San Giacomo <sup>8</sup>	Montagnola senese <sup>9</sup>	Rivoli Veronese <sup>10</sup>	Torre Picchio <sup>11</sup>	Casa Sgherri <sup>12</sup>	Monte la Mesa <sup>13</sup>
Age	Late Villanyan	Early Biharian	Late Villanyan	Late Villanyan	Early Biharian	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Early Biharian
MN unit	MN17	MN17	MN17	MN17	MNQ1	MN17	MN17	MN17	MN17	MN17	MN17	MN17	MNQ1
Magnetostatigraphy	1.8–1.7 M.a.	2.58–1.95 M.a.						2.1 M.a.					
Country	Spain	Spain	France	The Netherlands	Germany	Poland	Hungary	Italy	Italy	Italy	Italy	Italy	Italy
<b>Eulipotyphla</b>													
<i>Erinaceus</i> sp.						•	•						•
<i>Talpa fossilis</i>			•		•	•	•	•	•				•
<i>Talpa minor</i>				•									
<i>Talpa</i> sp.													
<i>Desmana thermalis</i>	•												
<i>Desmana</i> sp.							•						
<i>Galemys kormosi</i>				•									
<i>Galemys</i> cf. <i>kormosi</i>		•		•									
<i>Asoriculus gibberodon</i>			•										
<i>Beremendia fissidens</i>			•										
<i>Blarinoidea mariae</i>													
<i>Crociodura</i> sp.													
<i>Crociodura</i> cf. <i>kornfeldi</i>						•							
<i>Deinsdorfia hibbardii</i>						•							
<i>Petenyia hungarica</i>						•							
<i>Myosorex meini</i>						•							
<i>Urotrichus</i> sp.						•							
<i>Sulimskita kretzoi</i>						•							
<i>Sorex bor</i>						•							
<i>Sorex minutus</i>	•	•	•	•	•	•	•	•	•	•	•	•	•
<i>Sorex</i> sp.		•											
<i>Sorex subaraneus</i>		•											
<i>Sorex prealpinus</i>													
<i>Sorex (D.) praearaneus</i>										•			
<b>Rodentia</b>													
<i>Allocrietus bursae</i>													•
<i>Allocrietus ehiki</i>													•
<i>Castor fiber</i>			•										
<i>Castor</i> sp.													
<i>Trogontherium caveri</i>				•									•
<i>Histryx refossa</i>			•	•				•					•

References 1: Agustí et al., 2010; 2: Agustí et al., 2010; Agustí et al., 2013; 3: Guerin et al., 2004; Sen, 2004; Suarez and Mein, 2004; 4: van den Hoek Ostende and de Vos, 2006; 5: Carls and Rabeder, 1988; 6: Kowalsky, 1958; Rzebiak-Kowalska, 2009; 7: Janossy and van der Meulen, 1975 Janossy, 1986; 8: in this paper; Bellucci et al., 2014; 9: Fondi, 1972; Maul et al., 1998; 10: Sala et al., 1994; 11: Girotti et al., 2003; 12: Marcolini, 2003; 13: Marchetti et al., 2000.



Table 8 – Late Villanyan/Early Biharian selected European sites (continued).

Site	Galera 2 <sup>1</sup>	Barranco conejos <sup>2</sup>	Saint-Vallier <sup>3</sup>	Tegelen <sup>4</sup>	Schernfeld <sup>5</sup>	Kadzielinia 1 <sup>6</sup>	Osztramos-3 <sup>7</sup>	Coste San Giacomo <sup>8</sup>	Montagnola senese <sup>9</sup>	Rivoli Veronese <sup>10</sup>	Torre Picchio <sup>11</sup>	Casa Sgherri <sup>12</sup>	Monte la Mesa <sup>13</sup>
Age	Late Villanyan	Early Biharian	Late Villanyan	Late Villanyan	Early Biharian	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Early Biharian
MN unit	MN17	MNQ1	MN17	MN17	MNQ1	MN17	MN17	MN17	MN17	MN17	MN17	MN17	MNQ1
Magnetostratigraphy		1.8–1.7 M.a.	2.58–1.95 M.a.					2.1 M.a.					
Country	Spain	Spain	France	The Netherlands	Germany	Poland	Hungary	Italy	Italy	Italy	Italy	Italy	Italy
<b>Rodentia</b> (continued)													
<i>Apodemus atavus</i>	•		•										•
<i>Apodemus dominians</i>				•				•				•	
<i>Apodemus</i> sp.		•					•			•			
<i>Castillomys crusafonti</i>	•												
<i>Castillomys rivus</i>		•											
<i>Micromys</i> cf. <i>praeminutus</i>							•						
<i>Micromys</i> sp.				•									
<i>Prospalax priscus</i>													
<i>Sciurus warthae</i>								•					•
<i>Sciurus</i> sp.													
<i>Pliopetaurisia</i>													
<i>Hylopletes debnuijii</i>				•									
<i>Spermophilus primigenius</i>				•									
<i>Glirulus pusillus</i>				•									
<i>Glis sackdiligensis</i>													
<i>Glis</i> sp.													
<i>Eltonys</i> sp.		•											
<i>Muscardinus</i> sp.													
<i>Muscardinus pliocenicus</i>													
<i>Muscardinus</i> cf. <i>dacteus</i>													
<i>Borsodia newtoni</i>					•								•
<i>Clethrionomys</i> sp.													
<i>Clethrionomys kretzoi</i>													
<i>Dinaromys allegranzi</i>													
<i>Dinaromys dalmatinum</i>													
Cf. <i>Ellobius</i>													
<i>Kislangia gusti</i>													
<i>Lemmus kovalskii</i>					•								
<i>Lemmus</i> cf. <i>lemmus</i>													
<i>Microtus (A.) pliocenicus</i>													
<i>Mimomys</i> sp.		•											•

**References** 1: Agusti et al., 2010; 2: Agusti et al., 2010; Agusti et al., 2013; 3: Guerin et al., 2004; Sen, 2004; Suarez and Mein, 2004; 4: van den Hoek Ostende and de Vos, 2006; 5: Carls and Rabeder, 1988; 6: Kowalsky, 1958; Rzebik-Kowalska, 2009; 7: Janossy and van der Meulen, 1975 Janossy, 1986; 8: in this paper; Bellucci et al., 2014; 9: Fondi, 1972; Maut et al., 1998; 10: Sala et al., 1994; 11: Girotti et al., 2003; 12: Marcolini, 2003; 13: Marchetti et al., 2000.

Table 8 – Late Villanyan/Early Biharian selected European sites (continued).

Site	Galera 2 <sup>1</sup>	Barranco conejos <sup>2</sup>	Saint-Vaillier <sup>3</sup>	Tegelien <sup>4</sup>	Schernfeld <sup>5</sup>	Kadzielinia <sup>16</sup>	Oztramos-3 <sup>7</sup>	Coste San Giacomo <sup>8</sup>	Montagnola senese <sup>9</sup>	Rivoli Veronese <sup>10</sup>	Torre Picchio <sup>11</sup>	Casa Sgherri <sup>12</sup>	Monte la Mesa <sup>13</sup>
Age	Late Villanyan	Early Biharian	Late Villanyan	Late Villanyan	Early Biharian	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Late Villanyan	Early Biharian
MN unit	MN17	MNQ1	MN17	MN17	MNQ1	MN17	MN17	MN17	MN17	MN17	MN17	MN17	MNQ1
Magnetostratigraphy	1.8–1.7 M.a.	1.8–1.7 M.a.	2.58–1.95 M.a.				2.1 M.a.						
Country	Spain	Spain	France	The Netherlands	Germany	Poland	Hungary	Italy	Italy	Italy	Italy	Italy	Italy
<b>Rodentia</b> (continued)													
<i>Mimomys ostramosensis</i>					•							•	
<i>Mimomys</i> cf. <i>ostramosensis</i>				•			•						•
<i>Mimomys pliocenicus</i>			•					•					
<i>Mimomys</i> cf. <i>pliocenicus</i>									•				
<i>Mimomys pusillus</i>			•										•
<i>Mimomys</i> cf. <i>pusillus</i>													•
<i>Mimomys</i> cf. <i>malezi</i>													•
<i>Mimomys medasensis</i>	•										•		
<i>Mimomys coelodus</i>												•	
<i>Mimomys redai</i>				•									
<i>Mimomys</i> gr. <i>tigliensis/tornensis</i>								•					
<i>Mimomys tornensis</i>					•								•
<i>Mimomys tigliensis</i>				•									
<i>Mimomys pitmyoides</i>				•									
<i>Pliomys schemfeldensis</i>					•								
<i>Pliomys episcopalii</i>					•								
<i>Tcharinomyos oswaldoreigi</i>		•											
<i>Tibericola vandermeulenti</i>		•											
<i>Ungaromys</i> sp.													
<i>Ungaromys dehmi</i>					•								
<i>Ungaromys nanus</i>										•			
<i>Villanya</i> cf. <i>exilis</i>				•									
<b>Lagomorpha</b>													
<i>Hypolagus beremendensis</i>													•
<i>Hypolagus brachygnathus</i>				•									
<i>Ochotona</i> sp.													
<i>Oryctolagus lacosti</i>													
<i>Oryctolagus</i> cf. <i>lacosti</i>													•
<i>Oryctolagus</i> sp.													•
<i>Prolagus</i> sp.													•
<i>Prolagus calpensis</i>													•
<i>Prolagus italicus</i>								•					

References 1: Agustí et al., 2010; 2: Agustí et al., 2010; Agustí et al., 2013; 3: Guerin et al., 2004; Sen, 2004; Suarez and Mein, 2004; 4: van den Hoek Ostende and de Vos, 2006; 5: Carls and Rabeder, 1988; 6: Kowalsky, 1958; Rzebiak-Kowalska, 2009; 7: Janossy and van der Meulen, 1975; Janossy, 1986; 8: in this paper; Bellucci et al., 2014; 9: Fondi, 1972; Maul et al., 1998; 10: Sala et al., 1994; 11: Girotti et al., 2003; 12: Marcolini, 2003; 13: Marchetti et al., 2000.

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