Absorption, metabolism and protective role of fruits and vegetables polyphenols against gastric cancer

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Abstract. – Growing evidence links free radicals to the aging processes, degenerative diseases and cancer, underlying the important role played by some antioxidants, as polyphenols, present in fruits and vegetables, which seem able to counteract the toxic effects induced by oxidative stress. The gastrointestinal tract is continuously exposed to oxidant and antioxidant substances and, in particular in this district, the food rich in antioxidants could exert a protective effect against the risk of cancer. Polyphenols have a direct protective effect on the gastrointestinal tract, detoxifying the Reactive Oxygen Species (ROS) and Reactive Nitrogen Species (RNS), preserving antioxidant proteins and complexing metals. Although polyphenols are a class of antioxidant largely represented in vegetables and fruits, we are still uncertain whether the beneficial effects of a diet rich in plant products, are mainly due to these compounds. Our knowledge does not allow to be sure about which antioxidants are capable of having therapeutic effects, through which mechanism, the exact therapeutic dose or how long they have to be taken to have a significant protective effect. In this review we take into account the most common antioxidants, usually found in the diet and the processes regulating their absorption, metabolism and excretion, in order to elucidate the mechanism that could be responsible for the protection against cancer.

Key Words:

Oxidative stress, Gastric cancer, Polyphenols, Antioxidants metabolism, Polyphenols in fruit and vegetables.

Introduction

Free radicals are mainly constituted by unstable substances able to react with a large number of molecules, including those present in the cells, altering their structure and function¹. Consequently, these interactions can lead to toxic reactions causing changes of the cellular biological system,

that often result in close association with some pathologies. There is many evidence that links radicals to the aging processes, tumor and cardiovascular, neurodegenerative, infectious diseases²⁻¹². On the other hand, another interesting aspect is the role played by antioxidants, mainly found in vegetable foods, which can counteract the toxic radical action. A large number of studies shows that the consumption of foods and drinks rich in antioxidants is often associated with a decreased risk of developing these diseases¹³⁻¹⁹. Taking into account these observations, it would seem appropriate to prescribe antioxidant drugs to prevent degenerative diseases or slow down the natural aging process. Moreover, diet could be enriched with products capable of defending the organism from the radicals. With our knowledge we cannot determine which substances are capable of having preventative or therapeutic effects, how long they have to be taken, nor the therapeutic dose. A class of antioxidant compounds (flavonoids and biophenols) are largely represented in vegetable and fruits, but there is still uncertainty on the beneficial effects of a diet rich in plant products.

Physiological and Pathological Role of Free Radicals

The term "free radicals" is usually associated to something dangerous and harmful to health or otherwise to avoid. This is supported by the fact that radicals are produced by radiation or environmental pollutants becoming radical, during their metabolism in our body. In fact, the free radicals are typically substances characterized by a high reactivity, that is responsible of their cytotoxic and genotoxic effects. However, less is known about the concept that some radicals are physiologically produced by the body (bio-radicals) and are essential to life. These are involved in enzymatic catalysis and other critical processes as, for example, the regulation of embryogenesis and the

functioning of neuronal cells. The production and metabolism of free radicals, ROS and RNS in the human body are finely regulated processes, and an alteration of these mechanisms can lead to cellular injury as in chronical inflammatory state. In fact, the persistence of the inflammatory status is one of the major causes for the development of some types of cancer and neurodegenerative or cardiovascular diseases²⁰⁻²². Bio-radicals play a major role in signaling pathways, modulating the cell's reduction-oxidation reactions (redox status) and, paradoxically, also playing an antioxidant action as "scavenger" of other radicals (radical-radical reactions). The importance of the cell redox status is due to its capability to modulate reversibly the transport, the metabolism, the fat accumulation, the activity of enzymes and transcription factors. In fact, many of these processes are regulated by redox centers (cysteines and metal-containing centers) whose oxidation or reduction, is responsible for biological events such as cell cycle, metabolism, neurotransmission, differentiation, cell-cell communication and apoptosis²³. The excess of free radicals (oxidative stress), plays a pathogenic role: hyper-activation of some cellular signals (phosphorylation, transport), and the oxidation of specific protein sites (metal centers, SH, methionine, tyrosine, tryptophan, histidine) or DNA (guanine, xanthine, adenine). All of this leads to chronic inflammation, uncontrolled cellular proliferation, apoptosis or necrosis, resulting in cellular degeneration and a higher risk of developing cancer²⁴. The complexity of cellular processes involving the oxidative stress suggests that the degenerative diseases and cancer are related to the excessive production of free radicals. A critical step is the understanding of the mechanisms that modulate the effects leading to the formation of chronic inflammatory process. Recently, several authors underlined the importance of two free radicals, anion superoxide ($\bullet O_2$) and nitric oxide (•NO), carrying out essential functions as second messenger in the cells. These two radicals can react with each other to form a dangerous oxidizing species, peroxynitrite (\bullet NO + \bullet O, \rightarrow ONOO)25, that if produced in excessive quantities or in a compartment where the detoxification systems are unable to counteract their effects, can lead to a significant cellular injuries. The reaction between •O₂ and •NO is a radical-radical reaction and its velocity is controlled by the diffusion and consequently, every time the two molecules meet produce peroxynitrite. Moreover, this reaction is faster than the superoxide reduction reaction carried out by the superoxide dismutase, the antioxidant enzyme able to counteract the excess of $\bullet O_a$. This implies that small amounts of peroxynitrite may form physiologically and probably be useful for the cell (for example in the redox signals)²⁶ and be metabolized by the tissue without causing cytotoxic reactions²⁷. On the other hand, micromolar peroxynitrite levels produce sometimes irreversible protein and DNA oxidation, confusing the border between physiological and pathological activity not entirely clear²⁸. In inflammatory neurodegenerative pathologies, for example, peroxynitrite probably plays an important cytotoxic role, as shown by the presence of 3-nitrothyrosine found in inflammatory or degenerative tissues^{29,30}. In table I are shown the most common chemical modifications induced by an excess of free radicals that involve the macromolecules present in cells.

Assessment of Oxidative Stress in Man

As previously described, oxidative stress has been associated with a large number of diseases in humans, even if the relationship between the free radicals and some specific pathological processes has not definitively been established. In many pathologies, oxidative stress is not the primary cause of the disease, but a consequence of the cellular alteration caused by the disease. In order to protect themselves from oxidative damage, aerobic organisms, including humans, use a variety of antioxidant defense systems, both en-

Table I. Chemical modifications induced by oxidative stress.

Protein	Lipids	DNA
Carbonyl group	Hydroperoxides	8-hydroxyldeoxyguanosine
Hydroperoxides	Conjugated dienes	8-oxy-2-deoxyguanosine
Val and Leu Hydroperoxides	Isoprostane	8-hydroxylguanosine
3-nitrotyrosine	Malondialdehyde	5 hydroxylmethyluracil
2-oxo-histidine	4-hydroxylnonenal	8-oxyguanosine
Dityrosine	Total aldehydes	J.C

Table II. Reduction potential of different antioxidants (pH 7.0, 20°C).

Antioxidant	Reduction potential
Ascorbate (Vitamin C)	282
Epigallocatechin	430
A-Tocopherol (Vitamin E)	500
Theaflavin	510
Caffeic acid	540
Epicatechin	570
Uric acid	590
Glutathione	920

dogenous and exogenous, strategically located in the various cellular districts. Inside the cells is possible to find specific enzymes that interact with ROS: superoxide dismutase, catalase and glutathione peroxidase, while most of membrane protection is due to the α -tocopherol, β -carotene and coenzyme Q. The antioxidant protection of extra- and intra-cellular fluids, is based on the action of metal binding proteins as ferritin, transferrin, ceruloplasmin, which retain metal ions. In particular, iron and copper in non-reactive forms, are able to prevent and to limit radical reactions. Moreover, the biological fluids are also protected by some important antioxidants such as vitamin E, uric acid, bilirubin, ascorbate and thiol groups³¹. It is, therefore, clear that the determination of the balance between pro-oxidant and antioxidant species can play an important role in the diagnosis and therapeutic treatment of certain pathologies related to oxidative stress³². A large number of methods has been proposed in the literature to measure oxidative stress or oxidative damage reaction products³³⁻³⁵, notwithstanding free radicals are short life species, thus difficult to determine directly³⁶. To overtake this, a complementary approach could be the measurement

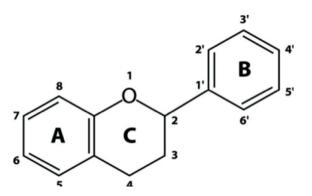


Figure 1. The basic structure of flavonoids.

of the endogenous antioxidant defense systems, as a biological marker of oxidative damage. However, this approach is relatively simple to use in routine clinical biochemical analysis, but it presents some obstacles that prevent its current use. In fact, the scientific literature lacks in studies reporting the antioxidants plasma profile of patients affected by pathologies associated to oxidative stress, with respect to healthy people. Moreover, the depletion of endogenous antioxidants due to a particular diet, is not distinguishable from the one caused by the disease³⁷. Consequently, the lack of universally accepted and standardized reference values for such measurements makes improbable, nowadays, the use of these biomarkers for clinical diagnosis. Another aspect to take into account is the "basal production" of oxidation products, that should be evaluated with care in order to differentiate it from the "pathological" ones. Likewise, we cannot rule out circadian variations of free radicals production.

Antioxidant Hypothesis

There are more than 5000 polyphenols in plant foods, most of which are flavonoids, including flavanols, flavonols, flavones, isoflavones, hydroxycinnamates and anthocyanins. These compounds are distributed among plants, fruits and vegetables in different amounts, all with different antioxidant activity. They represent the most abundant antioxidants in our diet; for example, in 200 g of fruits there are 500 mg of total polyphenols³⁸. The antioxidant activity of polyphenols is related to their structure. In fact, it depends on the hydroxylation pattern, the position and the substitution of specific hydroxyl groups that differentiate the individual compounds³⁹. Figure 1 shows the basic structure of flavonoids.

The following structures are important for antioxidant activity and detoxification of radicals: (1) the presence of two or three hydroxyl groups adjacent to ring B (catechol function at position 3' and 4' and pyrogallol function at position 3, 4' and 5', respectively); (2) the presence of other hydroxyl groups in the other rings (positions 5 and 7 of ring A, position 3 of ring C); (3) a double bond C2-C3 in ring C. Other hydroxyl groups may be present, for example, in position 5' of ring B and three more in positions 3', 4' and 5'of an esterified gallate group in position 3 of ring C⁴⁰. The flavonoid structures can be very simple, such as caffeic acid, or complex, such as gallate epigallocatechin (Figure 2).

The reduction potential is a measure of the reactivity of an antioxidant, as a hydrogen or an

Figure 2. (A) Caffeic acid. (B) Gallate epigallocatechin.

electron donor, under standard conditions (Table II): a low reduction potential indicates greater capacity in hydrogen or electron donation, therefore reducing reactive species with unpaired electrons. Polyphenols generally have an average reduction potential in the natural antioxidant scale comparable to that of vitamin E⁴¹, while ascorbic acid is considered the best natural antioxidant⁴². Polyphenols are able to detoxify most of the ROS and RNS, such as superoxide and hydroxyl radicals, hypochlorous acid, singlet oxygen, peroxyl radicals and peroxynitrite⁴³. In addition, a major property tied mainly to the catechin structure is the ability to complex transition metals (copper and iron principally), preventing their catalytic activity in redox reactions^{44,45}. Moreover, polyphenols also exhibit anti-inflammatory activity attributed to the inhibition of cyclooxygenase, lipoxygenase, myeloperoxidase, nitric oxide synthase and xanthine oxidase46-49.

Factors that Contribute to the Different Biological Activities of polyphenols in vitro and in vivo

Although there is a huge amount of data on the antioxidant capacity *in vitro* of individual polyphenolic compounds in scientific literature, the efficacy of their consumption has not always been confirmed in clinical trials conducted on humans. In some cases, evidence of a protective effect was obtained^{46,50-53}, while in others it was not possible to correlate the efficacy of polyphenol consumption, with a decrease in oxidative stress markers (F2-isoprostanes, oxidized LDL, oxidation products of plasmatic proteins, damaged DNA)⁵⁴⁻⁵⁷. Moreover, some researches showed an increase in plasma protein oxidation after the consumption of fruit juice, supporting the fact that the mechanism of action of polyphenols is not completely

clear^{56,58}. It has also been observed that frequent consumption of drinks and foods (fruit juice and apples) rich in polyphenols, leads to a modest and transient increase in the total plasma antioxidant capacity. This could be explained by the increased levels of urate caused by fructose, contained in drinks and foods, rather than polyphenols^{59,60}. The different biological activities shown *in vitro* and *in vivo* by polyphenols, are not only related to the heterogeneity of these molecules or to their structural and functional characteristics, but also reflect how these compounds are absorbed and metabolized in the gastrointestinal tract⁶¹.

Transport and Absorption

The antioxidants' concentration in the stomach is comparable to that present in foods, while is not always predictable the amount of antioxidants actually available at the intestinal and hepatic level, because of the digestive processes happening there⁶²⁻⁶⁴. The absorption and the effects of polyphenols already begin at the level of the oral cavity^{65,66}, although greater absorption occurs in the gastric tract and, to a lesser extent, in the intestinal tract. After the consumption of 10 to 500 mg of polyphenols, the maximum plasma concentration generally does not exceed 1 µM, mainly due to poor absorption and metabolism by tissues and gastrointestinal microflora⁶⁷⁻⁶⁹. Consequently, polyphenols are less available than ascorbate and tocopherols, which have a specific absorption system. It was demonstrated that the antioxidant activity due to micromolar concentrations of polyphenols is significantly lower than that detected in plasma (> $10^3 \mu M$); in fact, at least 20-50 μM of polyphenols would be needed to effectively compete with endogenous antioxidants. In vitro, polyphenols and their metabolites, can decrease or inhibit the activity of some cell's membrane transport systems, as demonstrated for glucose (GLUT2) and ascorbic acid (SVCT1), whose transport activity is inhibited by the presence of flavonoids⁷⁰. The flavonoid glucosides, the most abundant polyphenol in the diet, can be transported to the enterocytes, via the sodium-dependent glucose transporter 1 (SGLT1) and metabolized to intracellular aglycones, by a β -glucosidase⁷¹. However, this pathway is less important because of the presence of an effluent intracellular glucose system directed towards the digestive tract lumen, through the Multidrug Resistance Associated Protein 2 (MRP2) transporter. In the lumen of the digestive tract, with the contribution of the intestinal bacterial flora, the flavonoids glucosides are definitively transformed into aglycones and absorbed from the intestinal and the blood cells through the MRP3⁷².

Polyphenols Biotrasformation

The polyphenols undergo some significant biochemical modification in the oral cavity and gastrointestinal tract, before entering the circulatory system and reach the liver to be metabolized to several active metabolites. In fact, in saliva, the flavonoids glucosides are hydrolyzed to aglycones and then converted into compounds that can be absorbed by the epithelium of the oral cavity⁷³. The reduction of polyphenols to monomeric units occurs mainly the stomach (Figure 3). In the small bowel and later in the liver, the two step transformation occurr: phase I, deglycosylation and formation of aglycones; phase II, transformation through the oxidative metabolism by the enzymes belonging to the P450 cytochrome family⁷⁴. This last step leads to the formation of (a) methylated, sulphate and glucuronidated products; (b) protein or thiols adduct (glutathione); (c) RNA or DNA adduct⁷⁵. A further transformation takes place at large bowel level, where the intestinal microflora degrades flavonoids to simple phenolic acids, subsequently absorbed and metabolized in the liver, through the enterohepatic circulation. The methylated, sulphate and glucuronidated products continue to have biological activity⁷⁶, although lower with respect to the original molecule, because of the chemical modifications of the same (hydroxyl) groups responsible for the antioxidant activity.

Interactions and Synergies Between food Antioxidants and Endogenous Antioxidants

The interactions and synergies between food antioxidants and endogenous antioxidants are due to the redox potential of antioxidants (Table II). Antioxidants with lower redox potential are able to regenerate (reduce) other antioxidants, which have been previously oxidized into radicals. Regeneration reactions occur, for example, between polyphenols, ascorbic acid and vitamin E⁷⁷.

Gastric Cancer, Antioxidant Polyphenols, ROS and RNS

The highest concentration of polyphenols introduced with the diet can be found in the oropharyngeal tract, in the stomach, and partly in the intestine, before the absorption, metabolism or excretion process begins. In fact, after a diet rich in these antioxidant compounds, in the gastrointestinal tract can be detected µM concentrations of polyphenols, that could exert the protective effect against gastric and colorectal cancer.

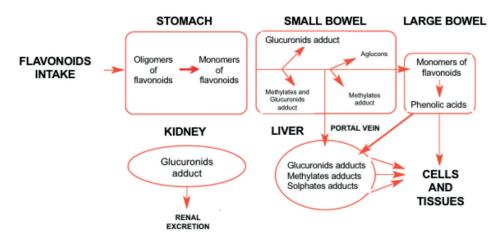


Figure 3. Metabolism of flavonoids.

Furthermore, the gastrointestinal tract is particularly exposed to the oxidative stress due to the ROS and RNS species from the diet and from the activation of the intestinal inflammatory cells⁷⁸. In particular, RNS play an important role in the onset of stomach cancer, because in the presence of acidic pH they are able to form nitrating/nitrosating species that support the formation of carcinogenic nitrosamines⁷⁹. Polyphenols have a direct protective effect on the gastrointestinal tract, detoxifying the ROS and RNS, preserving antioxidant proteins and complexing metals⁸⁰. In the stomach the acid amplifies the peroxidation reactions catalyzed by the pro-oxidation compounds present in food, for example meat (i.e. Fe³⁺-myoglobin, non-conjugated iron, fatty acids and hydroperoxides). These reactions are inhibited by catechin and red wine polyphenols, that in the gastric model were able to shift the reactions from pro-oxidants to antioxidant⁸¹. Similar results were obtained in other studies in conditions closer to physiological ones, where the polyphenols concentrations were comparable to those found in the stomach after a meal. All these results show that the polyphenols prevented the formation of nitrosamines involved in the pathogenesis of gastric cancer82,83.

Conclusions

The available epidemiological data has shown a strong reverse correlation between the intake of fruits and vegetables and the occurrence of degenerative diseases and cancer. The observed effect, seems to be due to the synergistic action between the compounds of the endogenous antioxidant system (superoxide dismutase, glutathione peroxidase, glutathione-S-transferase and catalase) and the antioxidants from the diet. However, it is still unclear whether some foods are able to exert an optimal protective effect against cancer and much less we know about the hypothetical recommended levels of the antioxidants intake. Despite the interest in antioxidants as protective agents against cardiovascular, degenerative pathologies and cancer, the actual contribution of these compounds to the maintenance of health and their in vivo mechanism of action are not yet known. In fact, there is no substantial evidence that the in vitro antioxidant effect is the same that actually occurs in vivo. Flavonoids are poorly absorbed but are extensively metabolized in the gastrointestinal tract, mainly forming glucuronidated, sulphate and methylated conjugates, whose adducts are subsequently absorbed. Since the flavonoids and their metabolites are potent bioactive molecules able to interact with the intracellular signal pathways, it is mandatory to clarify their mechanisms of action such as antioxidants or signaling molecules, in order to assess their potential anti-tumoral role. More attention should be given to the effects of polyphenols in the gastrointestinal tract, where these compounds probably exhibit the highest antioxidant capacity, before they are metabolized and absorbed in the circulatory stream.

Acknowledgements

This work represents a Ph.D research project of the Doctorate Biotechnology in Clinical Medicine of the Sapienza University of Rome. We are grateful to prof. S. Filetti, and prof. M. Arca for the helpful advice.

Conflict of Interest

The Authors declare that they have no conflict of interest.

References

- Lushchak VI. Free radicals, reactive oxygen species, oxidative stresses and their classifications. Ukr Biochem J 2015; 87: 11-18.
- CAGLAYAN A, KATLAN DC, SELÇUK TUNCER Z, YÜCE K, SAYAL HB, COÐKUN SALMAN M, KOCER-GUMUSEL B. Impaired antioxidant enzyme functions with increased lipid peroxidation in epithelial ovarian cancer. IUBMB Life 2017; 69: 802-813.
- CARINI, F, MAZZOLA M, RAPPA F, JURJUS A, GEAGEA AG, AL KATTAR S, BOU-ASSI T, JURJUS R, DAMIANI P, LEONE A, TOMASELLO G. Colorectal carcinogenesis: role of oxidative stress and antioxidants. Anticancer Res 2017; 37: 4759-4766.
- WILKES JG, ALEXANDER MS, CULLEN JJ. Superoxide dismutases in pancreatic cancer. Antioxidants (Basel) 2017; 6: pii: E66.
- WHONGSIRI P, PHOYEN S, BOONLA C. Oxidative stress in urothelial carcinogenesis: measurements of protein carbonylation and intracellular production of reactive oxygen species. Methods Mol Biol 2018; 1655: 109-117.
- 6) Moris D, Spartalis M, Spartalis E, Karachaliou GS, Karachanis GI, Tsourouflis G, Tsilimigras DI, Tzatzaki E, Theocharis S. The role of reactive oxygen species in the pathophysiology of cardiovascular diseases and the clinical significance of myocardial redox. Ann Transl Med 2017; 5: 326.
- MANOHARAN S, GUILLEMIN GJ, ABIRAMASUNDARI RS, ESSA MM, AKBAR M, AKBAR MD. The role of reactive oxygen species in the pathogenesis of Alzheimer's disease, Parkinson's disease, and Huntington's

- disease: a mini review. Oxid Med Cell Longev 2016; 2016: 8590578.
- METERE A, CHIESA C, DI COSIMO C, FIERRO G, GIACOMELLI L, PIETRAFORTE D. A novel approach to study oxidative stress in thyroid diseases: a preliminary study. Eur Rev Med Pharmacol Sci 2012; 16: 646-652.
- MALLOZZI C, MARTIRE A, DOMENICI MR, METERE A, POPOLI P, DI STASI AM. L-NAME reverses quinolinic acid-induced toxicity in rat corticostriatal slices: involvement of SRC family kinases. J Neurosci Res 2007; 85: 2770-2777.
- 10) Straface E, Marchesi A, Gambardella L, Metere A, Tarissi de Jacobis I, Viora M, Giordani L, Villani A, Del Principe D, Malorni W, Pietraforte D. Does oxidative stress play a critical role in cardiovascular complications of Kawasaki disease? Antioxid Redox Signal 2012; 17: 1441-1446.
- 11) ZHUAN B, YU Y, YANG Z, ZHAO X, LI P. Mechanisms of oxidative stress effects of the NADPH oxidase-ROS-NF-κB transduction pathway and VPO1 on patients with chronic obstructive pulmonary disease combined with pulmonary hypertension. Eur Rev Med Pharmacol Sci 2017; 21: 3459-3464.
- 12) DURANTE C, TALLINI G, PUXEDDU E, SPONZIELLO M, MORETTI S, LIGORIO C, CAVALIERE A, RHODEN KJ, VERRIENTI A, MARANGHI M, GIACOMELLI L, RUSSO D, FILETTI S. BRAF(V600E) mutation and expression of proangiogenic molecular markers in papillary thyroid carcinomas. Eur J Endocrinol 2011; 165: 455-463.
- 13) SOUILLARO T, SCHETTINO C, SAMPAOLO S, GALDERISI U, DI IORIO G, GIORDANO A, MELONE M. Adult-onset brain tumors and neurodegeneration: are polyphenols protective? J Cell Physiol 2017 Sep 8. doi: 10.1002/jcp.26170. [Epub ahead of print]
- 14) ABD EL-RAHMAN SS, SHEHAB G, NASHAAT H. Epigallocatechin-3-Gallate: the prospective targeting of cancer stem cells and preventing metastasis of chemically-induced mammary cancer in rats. Am J Med Sci 2017; 354: 54-63.
- BADIMON L, CHAGAS P, CHIVA-BLANCH G. Diet and cardiovascular disease: effects of foods and nutrients in classical and emerging cardiovascular risk factors. Curr Med Chem 2017 Apr 27. doi: 10.2174/09 29867324666170428103206. [Epub ahead of print]
- 16) ZIELIŃSKA MA, BIAŁECKA A, PIETRUSZKA B, HAMUŁKA J. Vegetables and fruit, as a source of bioactive substances, and impact on memory and cognitive function of elderly. Postepy Hig Med Dosw 2017; 71: 267-280.
- OMAR SH, SCOTT CJ, HAMLIN AS, OBIED HK. The protective role of plant biophenols in mechanisms of Alzheimer's disease. J Nutr Biochem 2017; 47: 1-20.
- Wu X, Cheng J, Wang X. Dietary antioxidants: potential anticancer agents. Nutr Cancer 2017; 69: 521-533.
- 19) ALAMOLHODAEI NS, TSATSAKIS AM, RAMEZANI M, HAYES AW, KARIMI G. Resveratrol as MDR reversion molecule in breast cancer: an overview. Food Chem Toxicol 2017; 103: 223-232.
- MACDONALD N. Chronic inflammatory states: their relationship to cancer prognosis and symptoms. J R Coll Physicians Edinb 2011; 41: 246-253.

- AGGARWAL BB, GEHLOT P. Inflammation and cancer: how friendly is the relationship for cancer patients? Curr Opin Pharmacol 2009; 9: 351-369.
- 22) Khansari N, Shakiba Y, Mahmoudi M. Chronic inflammation and oxidative stress as a major cause of age-related diseases and cancer. Recent Pat Inflamm Allergy Drug Discov 2009; 3: 73-80.
- 23) Вокком I. Recognition of functional roles of free radicals. Curr Neuropharmacol 2012; 10: 287-288.
- 24) LIOU GY, STORZ P. Reactive oxygen species in cancer. Free Radic Res 2010; 44: 479-496.
- Beckman JS. Understanding peroxynitrite biochemistry and its potential for treating human diseases. Arch Biochem Biophys 2009; 484: 114-116.
- 26) METERE A, IORIO E, PIETRAFORTE D, PODO F, MINETTI M. Peroxynitrite signaling in human erythrocytes: synergistic role of hemoglobin oxidation and band 3 tyrosine phosphorylation. Arch Biochem Biophys 2009; 484: 173-182.
- 27) MINETTI M, PIETRAFORTE D, STRAFACE E, METERE A, MATARRESE P, MALORNI W. Red blood cells as a model to differentiate between direct and indirect oxidation pathways of peroxynitrite. Methods Enzymol 2008; 440: 253-272.
- 28) RADI, R. Peroxynitrite, a stealthy biological oxidant. J Biol Chem 2013; 288: 26464-26472.
- 29) NAKAMURA T, LIPTON SA. Protein S-Nitrosylation as a therapeutic target for neurodegenerative diseases. Trends Pharmacol Sci 2016; 37: 73-84.
- 30) NAKAMURA T, PRIKHODKO OA, PIRIE E, NAGAR S, AKHTAR MW, OH CK, McKercher SR, Ambasudhan R, OKAMOTO S, LIPTON SA. Aberrant protein S-nitrosylation contributes to the pathophysiology of neurodegenerative diseases. Neurobiol Dis 2015; 84: 99-108.
- 31) FORMAN HJ, DAVIES KJ, URSINI F. How do nutritional antioxidants really work: nucleophilic tone and para-hormesis versus free radical scavenging in vivo. Free Radic Biol Med 2014; 66: 24-35.
- 32) FRIJHOFF J, WINYARD PG, ZARKOVIC N, DAVIES SS, STOCKER R, CHENG D, KNIGHT AR, TAYLOR EL, OETTRICH J, RUSKOVSKA T, GASPAROVIC AC, CUADRADO A, WEBER D, POULSEN HE, GRUNE T, SCHMIDT HH, GHEZZI P. Clinical relevance of biomarkers of oxidative stress. Antioxid Redox Signal 2015; 23: 1144-1170.
- 33) BALASUBRAMANYAM M, ADAIKALAKOTESWARI A, SAMEER-MAHMOOD Z, MOHAN V. Biomarkers of oxidative stress: methods and measures of oxidative DNA damage (COMET assay) and telomere shortening. Methods Mol Biol 2010; 610: 245-261.
- 34) ROBITAILLE L, HOFFER LJ. A simple method for plasma total vitamin C analysis suitable for routine clinical laboratory use. Nutr J 2016; 15: 40.
- HARMA M, EREL O. Measurement of the total antioxidant response in preeclampsia with a novel automated method. Eur J Obstet Gynecol Reprod Biol 2005; 118: 47-51.
- HARRIS C, HANSEN JM. Oxidative stress, thiols, and redox profiles. Methods Mol Biol 2012; 889: 325-346.
- 37) SPENCER JP, ABD EL MOHSEN MM, MINIHANE AM, MATHERS JC. Biomarkers of the intake of dietary polyphenols:

- strengths, limitations and application in nutrition research. Br J Nutr 2008; 99: 12-22.
- 38) PANDEY KB, RIZVI SI. Plant polyphenols as dietary antioxidants in human health and disease. Oxid Med Cell Longev 2009; 2: 270-278.
- 39) TsAO R. Chemistry and biochemistry of dietary polyphenols. Nutrients 2010; 2: 1231-1246.
- Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: an overview. Scientific World Journal 2013; 2013: 162750.
- 41) Duncan KR, Suzuki YJ. Vitamin E nicotinate. Antioxidants (Basel) 2017; 6. pii: E20.
- SARDI W. Narrow scope of vitamin C review. PLoS Med 2005; 2: e308.
- 43) KASOTE DM, KATYARE SS, HEGDE MV, BAE H. Significance of antioxidant potential of plants and its relevance to therapeutic applications. Int J Biol Sci 2015; 11: 982-991.
- 44) Chobot V, Hadacek F. Iron and its complexation by phenolic cellular metabolites: from oxidative stress to chemical weapons. Plant Signal Behav 2010; 5: 4-8.
- 45) BROWN JE, KHODR H, HIDER RC, RICE-EVANS CA. Structural dependence of flavonoid interactions with Cu2+ions: implications for their antioxidant properties. Biochem J 1998; 330 (Pt 3): 1173-1178.
- 46) LITTLE CH, COMBET E, McMILLAN DC, HORGAN PG, ROXBURGH CS. The role of dietary polyphenols in the moderation of the inflammatory response in early stage colorectal cancer. Crit Rev Food Sci Nutr 2017; 57: 2310-2320.
- UPADHYAY S, DIXIT M. Role of polyphenols and other phytochemicals on molecular signaling. Oxid Med Cell Longev 2015; 2015: 504253.
- 48) Zeinali M, Rezaee SA, Hosseinzadeh H. An overview on immunoregulatory and anti-inflammatory properties of chrysin and flavonoids substances. Biomed Pharmacother 2017; 92: 998-1009.
- 49) SARKAR S, MAZUMDER S, SAHA SJ, BANDYOPADHYAY U. Management of inflammation by natural polyphenols: a comprehensive mechanistic update. Curr Med Chem 2016; 23: 1657-1695.
- CHIVA-BLANCH G, BADIMON L. Effects of polyphenol intake on metabolic syndrome: current evidences from human trials. Oxid Med Cell Longev 2017; 2017: 5812401.
- 51) LANDEKA I, JURČEVIĆ DORA M, GUBEROVIĆ I, PETRAS M, RIMAC S, BRNČIĆ, ĐIKIĆ D. Polyphenols from wine lees as a novel functional bioactive compound in the protection against oxidative stress and hyperlipidaemia. Food Technol Biotechnol 2017; 55: 109-116.
- 52) VISIOLI F, DAVALOS A. Polyphenols and cardiovascular disease: a critical summary of the evidence. Mini Rev Med Chem 2011; 11: 1186-1190.
- 53) GORECKI P, BURKE DL, CHAPPLE ILC, HEMMING K, SAUND D, PEARSON D, STAHL W, LELLO R, DIETRICH T. Perioperative supplementation with a fruit and vegetable juice powder concentrate and postsurgical morbidity: a double-blind, randomised, placebo-control-

- led clinical trial. Clin Nutr 2017 Aug 10. pii: S0261-5614(17)30272-8. doi: 10.1016/j.clnu.2017.08.004. [Epub ahead of print]
- 54) SKIBOLA CF, SMITH MT. Potential health impacts of excessive flavonoid intake. Free Radic Biol Med 2000; 29: 375-383.
- 55) O'REILLY JD, MALLET AI, McANLIS GT, YOUNG IS, HALLIWELL B, SANDERS TA, WISEMAN H. Consumption of flavonoids in onions and black tea: lack of effect on F2-isoprostanes and autoantibodies to oxidized LDL in healthy humans. Am J Clin Nutr 2001; 73: 1040-1044.
- 56) SÁNCHEZ-MORENO C, CANO MP, DE ANCOS B, PLAZA L, OLMEDILLA B, GRANADO F, MARTÍN A. Effect of orange juice intake on vitamin C concentrations and biomarkers of antioxidant status in humans. Am J Clin Nutr 2003; 78: 454-460.
- 57) MØLLER P, VOGEL U, PEDERSEN A, DRAGSTED LO, SAND-STRÖM B, LOFT S. No effect of 600 grams fruit and vegetables per day on oxidative DNA damage and repair in healthy nonsmokers. Cancer Epidemiol Biomarkers Prev 2003; 12: 1016-1022.
- 58) YOUNG JF, NIELSEN SE, HARALDSDÓTTIR J, DANESHVAR B, LAURIDSEN ST, KNUTHSEN P, CROZIER A, SANDSTRÖM B, DRAGSTED LO. Effect of fruit juice intake on urinary quercetin excretion and biomarkers of antioxidative status. Am J Clin Nutr 1999; 69: 87-94.
- 59) LOTITO SB, FREI B. The increase in human plasma antioxidant capacity after apple consumption is due to the metabolic effect of fructose on urate, not apple-derived antioxidant flavonoids. Free Radic Biol Med 2004; 37: 251-258.
- 60) Godycki-Cwirko M, Krol M, Krol B, Zwolinska A, Kolodziejczyk K, Kasielski M, Padula G, Grebowski J, Græbocki J, Kazmierska P, Kazimierska P, Miatkowski M, Markowski J, Nowak D. Uric acid but not apple polyphenols is responsible for the rise of plasma antioxidant activity after apple juice consumption in healthy subjects. J Am Coll Nutr 2010; 29: 397-406.
- 61) KAHLE K, KEMPF M, SCHREIER P, SCHEPPACH W, SCHRENK D, KAUTENBURGER T, HECKER D, HUEMMER W, ACKERMANN M, RICHLING E. Intestinal transit and systemic metabolism of apple polyphenols. Eur J Nutr 2011; 50: 507-522.
- 62) Hu M, Wu B, Liu Z. bioavailability of polyphenols and flavonoids in the era of precision medicine. Mol Pharm 2017; 14: 2861-2863.
- 63) Hu Y, Guo DH, Liu P, Cao JJ, Wang YP, Yin J, Zhu Y, Rahman K. Bioactive components from the tea polyphenols influence on endogenous antioxidant defense system and modulate inflammatory cytokines after total-body irradiation in mice. Phytomedicine 2011; 18: 970-975.
- 64) TOMMONARO G, SPERANZA G, DE PRISCO R, IODICE C, CRUDELE E, ABBAMONDI GR, NICOLAUS B. Antioxidant activity and bioactive compound contents before and after in vitro digestion of new tomato hybrids. J Sci Food Agric 2017; 97: 5241-5246.
- 65) SILVA MS, GARCÍA-ESTÉVEZ I, BRANDÃO E, MATEUS N, DE FREITAS V, SOARES S. Molecular interaction between salivary proteins and food tannins. J Agric Food Chem 2017; 65: 6415-6424.

- 66) GAUR S, AGNIHOTRI R. Green tea: a novel functional food for the oral health of older adults. Geriatr Gerontol Int 2014; 14: 238-250.
- 67) Mosele JI, Macià A, Motilva MJ. Metabolic and microbial modulation of the large intestine ecosystem by non-absorbed diet phenolic compounds: a review. Molecules 2015; 20: 17429-17468.
- SCALBERT A, WILLIAMSON G. Dietary intake and bioavailability of polyphenols. J Nutr 2000; 130: 2073S-2085S.
- 69) BLAUT M, SCHOEFER L, BRAUNE A. Transformation of flavonoids by intestinal microorganisms. Int J Vitam Nutr Res 2003; 73: 79-87.
- 70) Song J, Kwon O, Chen S, Daruwala R, Eck P, Park JB, Levine M. Flavonoid inhibition of sodium-dependent vitamin C transporter 1 (SVCT1) and glucose transporter isoform 2 (GLUT2), intestinal transporters for vitamin C and Glucose. J Biol Chem 2002; 277: 15252-15260.
- 71) HOSTETLER GL, RALSTON RA, SCHWARTZ SJ. Flavones: food sources, bioavailability, metabolism, and bioactivity. Adv Nutr 2017; 8: 423-435.
- 72) WILLIAMSON G, CLIFFORD MN. Role of the small intestine, colon and microbiota in determining the metabolic fate of polyphenols. Biochem Pharmacol 2017; 139: 24-39.
- 73) GINSBURG I, KOREN E, SHALISH M, KANNER J, KOHEN R. Saliva increases the availability of lipophilic polyphenols as antioxidants and enhances their retention in the oral cavity. Arch Oral Biol 2012; 57: 1327-1334.
- 74) BASHEER L, KEREM Z. Interactions between CYP3A4 and dietary polyphenols. Oxid Med Cell Longev 2015; 2015: 854015.
- 75) RECHNER AR, KUHNLE G, HU H, ROEDIG-PENMAN A, VAN DEN BRAAK MH, MOORE KP, RICE-EVANS CA. The me-

- tabolism of dietary polyphenols and the relevance to circulating levels of conjugated metabolites. Free Radic Res 2002; 36: 1229-1241.
- 76) DELGADO L, FERNANDES I, GONZÁLEZ-MANZANO S, DE FREITAS V, MATEUS N, SANTOS-BUELGA C. Anti-proliferative effects of quercetin and catechin metabolites. Food Funct 2014; 5: 797-803.
- 77) KADOMA Y, ISHIHARA M, OKADA N, FUJISAWA S. Free radical interaction between vitamin E (alpha-, beta-, gamma- and delta-tocopherol), ascorbate and flavonoids. In Vivo 2006; 20: 823-827.
- 78) BHATTACHARYYA A, CHATTOPADHYAY R, MITRA S, CROWE SE. Oxidative stress: an essential factor in the pathogenesis of gastrointestinal mucosal diseases. Physiol Rev 2014; 94: 329-354.
- 79) Cui J, Guo XM, Bao HL, Tan JB. Relationship between N-nitrosodimethylamine and risk of digestive tract cancers: a meta analysis based on cohort studies. Zhonghua Liu Xing Bing Xue Za Zhi 2016; 37: 725-729.
- DRYDEN GW, SONG M, McCLAIN C. Polyphenols and gastrointestinal diseases. Curr Opin Gastroenterol 2006; 22: 165-170.
- 81) GORELIK S, LIGUMSKY M, KOHEN R, KANNER J. The stomach as a "bioreactor": when red meat meets red wine. J Agric Food Chem 2008; 56: 5002-5007.
- 82) HERNÁNDEZ-RAMÍREZ RU, GALVÁN-PORTILLO MV, WARD MH, AGUDO A, GONZÁLEZ CA, OÑATE-OCAÑA LF, HERRERA-GOEPFERT R, PALMA-COCA O, LÓPEZ-CARRILLO L. DIETARY intake of polyphenols, nitrate and nitrite and gastric cancer risk in Mexico City. Int J Cancer 2009; 125: 1424-1430.
- 83) ZULUETA A, CARETTI A, SIGNORELLI P, GHIDONI R. Resveratrol: a potential challenger against gastric cancer. World J Gastroenterol 2015; 21: 10636-10643.