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Sensitivity to cocaine in adult mice is due to interplay between genetic makeup, early environment and later experience

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1 **TITLE PAGE**

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3 **environment and later experience**

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19

Abstract

20 **Abstract**
21 Although early aversive postnatal events are known to increase the risk to develop psychiatric
22 disorders later in life, rarely they determine alone the nature and outcome of the psychopathology,
23 indicating that interaction with genetic factors is crucial for expression of psychopathologies in
24 adulthood. Moreover, it has been suggested that early life experiences could have negative
25 consequences or confer adaptive value in different individuals. Here we suggest that resilience or
26 vulnerability to adult cocaine sensitivity depends on a “triple interaction” between genetic makeup
27 x early environment x later experience. We have recently showed that Repeated Cross Fostering
28 (RCF; RCF pups were fostered by four adoptive mothers from postnatal day 1 to postnatal day 4.
29 Pups were left with the last adoptive mother until weaning) experienced by pups affected the
30 response to a negative experience in adulthood in opposite direction in two genotypes leading
31 DBA2/J, but not C57BL/6J mice, toward an “anhedonia-like” phenotype.

32 Here we investigate whether exposure to a rewarding stimulus, instead of a negative one, in
33 adulthood induces an opposite behavioral outcome. To test this hypothesis, we investigated the
34 long-lasting effects of RCF on cocaine sensitivity in C57 and DBA female mice by evaluating
35 conditioned place preference induced by different cocaine doses and catecholamine prefrontal-
36 accumbal response to cocaine using a “dual probe” *in vivo* microdialysis procedure. Moreover,
37 cocaine-induced c-Fos activity was assessed in different brain regions involved in processing of
38 rewarding stimuli. Finally, cocaine-induced spine changes were evaluated in the prefrontal-
39 accumbal system. RCF experience strongly affected the behavioral, neurochemical and
40 morphological responses to cocaine in adulthood in opposite direction in the two genotypes
41 increasing and reducing, respectively, the sensitivity to cocaine in C57 and DBA mice.

42
43 Keywords: unstable maternal environment, animal models, gene x environment interplay,
44 resilience, vulnerability, cocaine.

45

46 **Introduction**

47 Recent evidence suggests an association between exposure to aversive events during the early
48 postnatal life and increased vulnerability to a variety of neuropsychiatric and psychosocial disorders
49 later in life (Heim and Nemeroff, 2001; Matthews and Robbins 2003; McEwen 2000; McLaughlin
50 et al. 2015; Meaney, 2001; Nemeroff, 2004; Sánchez, et al., 2001; van der Veen, et al., 2008).

51 Although the effects of early experiences may be strong and pervasive, large individual differences
52 exist in their impact on health and behavior (Bakermans-Kranenburg and van Ijzendoorn, 2007;
53 Belsky and Pluess, 2009; Belsky et al., 2015), suggesting that genetic background can modulate the
54 capacity of an environmental risk factor to give rise to mental illness (Caspi and Moffitt, 2006).

55 Finally, even if early aversive experiences are generally believed to confer a great risk for
56 psychopathologies in adult life, different current points of view suggest that the same early events
57 might determine either susceptibility or resilience to mental illness (Belsky and Pluess, 2009;
58 Belsky et al., 2013; Der-Avakian and Markou, 2010; Maccari et al., 2014; Meaney, 2010; Parker
59 and Maestripieri 2001; Pryce and Feldon, 2003; Rana et al., 2015; van der Veen et al., 2008),
60 probably depending on the genetic makeup (Bakermans-Kranenburg and van Ijzendoorn 2007;
61 Belsky et al., 2013; Di Segni et al., 2016).

62 Based on the “three-hit concept” of resilience and vulnerability (Daskalakis et al., 2013) as well as
63 on the “plasticity genes” hypotheses (Belsky and Pluess, 2009), here we suggest that genetic
64 background-early environment interplay increases vulnerability to development/expression of
65 certain psychopathologies and resilience to others, also depending on the successive experiences in
66 adult life.

67 We have recently reported that Repeated Cross Fostering (RCF) experienced by pups, affected adult
68 behavioral and neurochemical responses to aversive events in opposite direction in two genotypes
69 leading DBA/2J (DBA) mice toward an “anhedonia-like” phenotype and C57BL/6J (C57) mice
70 toward an increased sensitivity for a natural reinforcing stimulus (Di Segni et al., 2016). In RCF
71 condition, pups from the same litter spend the first postnatal day (PND0) with their biological

72 mother. On PND1, experimental litter are fostered by replacing the mother with a novel lactating
73 female caring for pups of the same age; this procedure is repeated daily (4 times, from PND1 up to
74 PND4) until the fourth adoptive mother was reached. Pups are left with the last adoptive mother
75 until weaning (D'Amato et al., 2011; Ventura et al., 2013; Di Segni et al., 2016).

76 Instability of the early environment, modeled by the RCF procedure, is thus hypothesized also to
77 affect the response to a rewarding stimulus (cocaine) in opposite way, depending on the genetic
78 makeup. In particular we anticipate an increased and reduced, respectively, response to cocaine in
79 RCF C57 and RCF DBA female mice compared to their Control groups.

80 Females were chosen as subjects for this study because of the significant incidence of gender
81 differences in stress-related disorders and their greater stress sensitivity (Bale, 2006; Kendler et al.,
82 1992; Nestler et al., 2002) as well as on the basis of previous data reported by our laboratory (Di
83 Segni et al., 2016). Moreover, clinical research indicates that women show higher susceptibility to
84 cocaine abuse (Kosten et al., 1996; Scott et al., 2006, Cotto et al., 2010) and relapse (Becker and
85 Hu, 2008) in comparison to men. In fact, despite severity of childhood emotional abuse has been
86 correlated with increased risk for substance abuse in both sexes (Hyman et al. 2006), in woman
87 emotional abuse/neglect and maltreatment have been associated with a heightened severity of
88 substance abuse and increased risk of relapse (Hyman et al. 2006). Consistent with these reports,
89 preclinical studies suggest that female rodents are characterized by enhanced susceptibility to
90 develop addiction-related pathological phenotypes (Hu et al., 2004; Lynch, 2006; Van Swearingen
91 et al., 2013) and this outcome is increased by exposure to critical early life experiences (Kosten et
92 al., 2004; 2006).

93 To test our hypothesis, three experiments were carried out in RCF and Control (Cont) female mice
94 of the 2 strains. In the first experiment, we tested whether RCF experience affected the behavioral
95 response to cocaine in adult mice in a genotype-dependent manner using cocaine-induced
96 conditioned place preference (CPP), a drug seeking correlated behavioral test (Kiraly et al., 2010).
97 Clinical and pre-clinical studies have demonstrated the impact of early postnatal experiences on

98 prefrontal-accumbal catecholamine circuit (Barros et al., 2004; Huppertz-Kessler et al., 2012; Jahng
99 et al., 2010; Oswald et al., 2014; Peña et al., 2014; Rincón-Cortés and Sullivan, 2016; Ventura et
100 al., 2013; Yang et al., 2014), a brain network known to be involved in processing of rewarding
101 stimuli (Cabib and Puglisi-Allegra, 2012; Di Segni et al., 2016, Pascucci et al., 2007; Puglisi-
102 Allegra and Ventura 2012; Siciliano et al., 2015; Ventura et al., 2007; 2008; 2013). Based on this
103 evidence, in the second experiment, we evaluated the enduring effects of RCF on catecholamine
104 prefrontal-accumbal response to cocaine injection in adult mice using a “dual probes” *in vivo*
105 intracerebral microdialysis procedure (Di Segni et al., 2016). Moreover, c-Fos activity was assessed
106 in different brain areas involved in processing of rewarding stimuli and, based on literature pointing
107 to the role that dendritic spine alterations may play in drug-related behaviors (Kauer and Malenka,
108 2007; Kiraly et al., 2010; Pulipparacharuvil et al., 2008; Robinson and Kolb, 2004; Villalba and
109 Smith, 2013), in the last experiment we investigated the effects of repeated cocaine injection on
110 spine density on Medium-Sized Spiny Neurons (MSNs) in the NAc as well as on pyramidal neurons
111 in mpFC, two brain areas strongly involved in the effects of drugs of abuse in both humans and
112 rodents.

113

114 **Materials and Methods**

115 **Animals**

116 C57BL/6J and DBA2/J female mice (purchased when 6/7 weeks old from Charles River, Italy)
117 were mated when 12 weeks old.

118 Mating protocol was as previously described (Di Segni et al., 2016). Each experimental group was
119 composed by 6-8 animals. Adequate measures were taken to minimize pain or discomfort of mice.

120 All experiments were carried out in accordance with Italian national law (DL 116/92 and DL
121 26/2014) on the use of animals for research based on the European Communities Council Directives
122 (86/609/EEC and 2010/63/UE).

123

124 Repeated Cross-Fostering

125 As previously described (D'Amato et al., 2011; Di Segni et al., 2016; Ventura et al., 2013), pups
126 from the same litter spent the first postnatal day (PND0) with their biological mother. On PND1,
127 litters were randomly selected and assigned to experimental (RCF) or Control (Cont) treatment.
128 RCF pups were fostered by introducing the entire litter into the home cage of a different dam whose
129 pups had just been removed and moved to a different adoptive mother. This procedure was repeated
130 daily (4 times, from PND1 up to PND4) until the fourth adoptive mother was reached. Pups were
131 left with the last adoptive mother until weaning. Control litters were only picked up daily and
132 reintroduced in their home cage, for the same period, and had their mothers returned within 30 sec.
133 Animals were weaned at PND28, separated by sex and housed in groups of 4 littermates. All of the
134 experiments were performed on 3-month-old adult female mice. To prevent potential estrous cycle
135 group synchronization, experimental subjects for cross-fostered and control female groups were
136 sorted by collecting not >2 individuals per cage/litter (Ventura et al., 2013).

137

138 Conditioned Place Preference

139 Conditioned place preference experiments were performed using a place conditioning apparatus as
140 previously reported (Ventura et al., 2007; 2013). Briefly, on day 1 (pretest), mice were free to
141 explore the entire apparatus for 20 min. During the following 8 days (conditioning phase), mice
142 were confined daily for 40 min alternately in 1 of the 2 chambers. One of the patterns was
143 consistently paired with drug (P; cocaine-Paired chamber (C57: 2.5, 5, 7.5, mg/kg i.p.; DBA: 5, 7.5,
144 10 mg/kg i.p.)) and the other one with vehicle (U; Unpaired chamber (saline 0.9%)). Different doses
145 of cocaine were chosen on the basis of a previous work indicating that C57 and DBA mice differ for
146 the sensitivity to cocaine-induced CPP, and suggesting a different sensitivity to the positive
147 incentive properties of drugs of abuse (Kutlu et al. 2015; Orsini et al., 2005). Testing for the

148 expression of CPP was conducted on day 10 using the pretest procedure. Behavioral data were
149 collected and analyzed by the “EthoVision” (Noldus, The Netherlands) fully automated video
150 tracking system (Spink et al., 2001). The digital data were analyzed by means of the EthoVision
151 software to obtain “time spent” (seconds; sec.), which was used as raw data for preference scores in
152 each sector of the apparatus of each subject. Each experimental group was composed by 6-8
153 animals.

154

155 **In Vivo Microdialysis**

156 “Dual probe” microdialysis experiments in mpFC and NAc were carried out as previously described
157 (Di Segni et al., 2016). Animals were anesthetized with Zoletil and Rompun, mounted in a
158 stereotaxic frame (David Kopf Instruments, Tujunga, CA, USA) equipped with a mouse adapter.
159 Zoletil 100 (Virbac, Milano, Italy (tiletamine HCl 50 mg/ml+zolazepam HCl 50 mg/ml)) and
160 Rompun 20 (Bayer S.p.A Milano, Italy (xylazine 20 mg/ml)), purchased commercially, were
161 dissolved in a volume of 4.1mg/ml and 1.6mg/ml respectively, in saline and injected in a volume of
162 7.3 ml/kg.

163 Bilateral (mpFC and NAc) probe implantation was counterbalanced in either the right or left
164 hemisphere. Vertical concentric dialysis probes were prepared with AN69 fibers (Hospal Dasco,
165 Bologna, Italy), according to the method of Di Chiara et al. (Di Chiara et al., 1996). Different
166 coordinates for probe implantation were used for the 2 strains. The coordinates from bregma
167 (measured according to the atlas of Franklin and Paxinos (Franklin and Paxinos, 1997) and Mouse
168 Brain Atlases, The Mouse Brain Library, www.nervenet.org) were: mpFC: 2.5 Antero-Posterior
169 (AP), ± 0.6 Medio-Lateral (ML) (C57), and 2.0 AP, ± 0.6 L (DBA); NAc: +1.6 AP, ± 0.6 L (C57) and
170 +1.1 AP; ± 0.6 L (DBA). Different probe length was used for C57 and DBA NAc (mpFC: 3 mm
171 shaft length, dialysis membrane length 2 mm o.d. 0.24 mm; NAc: 4.5 mm (C57) or 4.0 mm (DBA)
172 shaft length, dialysis membrane length 1 mm o. d. 0.24 mm). The final depth of the probe was 3
173 mm for mpFC and 5.5 or 5.0 mm for C57 and DBA NAc, respectively, ventral to the skull surface.

174 The day before use, the membranes were tested to verify in vitro norepinephrine and/or dopamine
175 recovery. The microdialysis system was as previous reported (Di Segni et al., 2016). Experiments
176 were carried out 48 h after probe placement. The mean concentration of the 3 samples collected
177 immediately before treatment (<10% variation) was taken as baseline concentration. Following,
178 Cont and RCF C57 and DBA mice were injected (i.p.) with cocaine (C57: 2.5 mg/Kg; DBA: 5
179 mg/Kg). 2.5 and 5 mg/Kg cocaine doses were choose, respectively, in C57 and in DBA mice based
180 on CPP experiments showing that these doses only induced different effects (discriminative dose) in
181 RCF vs. Control groups of two strains. Dialysate was collected every 20 min for 120 min. Twenty
182 microliters of the dialysate sample was analyzed by performance liquid chromatography, as
183 previously reported (Di Segni et al., 2016; Latagliata et al., 2014; Ventura et al., 2013).
184 Concentrations (pg/20 μ L) were not corrected for probe recovery. Each experimental group was
185 composed by 6-8 animals.

186

187 **Immunohistochemistry**

188 At the end of the microdialysis experiments, mice were sacrificed and c-Fos expression was
189 evaluated in NAc core (NAcC) and shell (NAcS), caudate-putamen (CP), infralimbic cortex (ILC),
190 prelimbic cortex (PRLC), Hippocampus (CA3, CA1, dentate gyrus (DG)), Amygdala (Lateral
191 Amygdala (LA), Basolateral Amygdala (BLA), Central Amigdala (CEA)) in the contralateral areas
192 of probes implantation. Brains were fixed overnight in paraformaldehyde (4%), cryoprotected in
193 sucrose (30%), frozen with dry ice, and cut in transversal sections. For c-Fos immunohistochemistry,
194 brain sections (40 μ m) were collected approximately from 1.98 to -3.40 for C57 and from 2.40 to -
195 2.80 for DBA from bregma, according to the atlas of Franklin and Paxinos (Franklin and Paxinos,
196 1997) and Mouse Brain Atlases (The Mouse Brain Library; www.nervenet.org/mbl/).
197 Immunohistochemistry was performed as previously described (Conversi et al., 2004; Di Segni et
198 al., 2016). Each experimental group was composed by 6-8 animals.

199

200 Morphologic analysis

201 Based on a previous work (Kiraly et al., 2010), mice from the different groups were given 4 daily
202 injections of cocaine or saline (C57 Cont cocaine/saline, C57 RCF cocaine/saline, DBA Cont
203 cocaine/saline, DBA RCF cocaine/saline). Different doses of cocaine (C57 2.5mg/kg; DBA 5mg/kg)
204 were used based on the behavioral (CPP) and microdialysis experiments. Each experimental group
205 was composed by 4-6 animals.

206 Thirty minutes after the last injection, mice were perfused and collected brains were impregnated
207 with a standard Golgi-Cox solution as previously described (Andolina et al., 2011). Coronal
208 sections of 120 μ m were obtained using a vibratome, mounted on gelatinized slides, stained
209 according to the Gibb and Kolb method (Gibb and Kolb, 1998) and covered with Eukitt (Kindler
210 GmbH & Co., Germany). Measurements were performed on impregnated neurons identified under
211 low magnification (20X/0.4 numerical aperture). MpFC and NAc analysis was restricted,
212 respectively, to pyramidal-like (C57 from 1.98 to 1.58; DBA from 1.40 to 1.70) and medium-spiny
213 neurons (MSNs) (C57 from 1.94 to 0.74; DBA from 1.70 to 0.74 from bregma, measured according
214 to the atlas of Franklin and Paxinos (Franklin and Paxinos, 1997) and Mouse Brain Atlases (The
215 Mouse Brain Library; www.nervenet.org/mbl/)). Golgi-impregnated neurons were selected
216 according to criteria proposed by Vyas (Vyas et al., 2002) as reported in Figure 1. An average of
217 3/4 neurons for each mouse were analyzed and randomly selected from both hemispheres. An
218 experimenter blind to the experimental groups performed the morphological analyses. The analysis
219 of total spine density, mature and immature spine density from apical and basal dendrite of
220 pyramidal neurons from mpFC as well as total spine density, mature and immature spine density
221 from accumbal MSNs was performed by 3D reconstruction of the selected neurons using the
222 NeuroLucida image analysis system (mbf, Bioscience) connected to an Olympus BX53 microscope
223 (100X/1.25 numerical aperture). By using the same NeuroLucida system (100X, 1.25 numerical
224 aperture, Olympus BX53), all protrusions, respective of their morphological characteristics, were
225 counted on each-order branches as spines if they were in direct continuity with the dendritic shaft.

226 On each dendrite, all protrusions with a clearly recognizable neck were considered as spines and
227 were classified according to the categories proposed by Peters & Kaiserman-Abramof (1969) as
228 stubby, mushroom and thin types. Stubby spines protrude from spiny dendrites with no neck visible,
229 they have a length similar to the diameter of the neck and to the head width; mushroom spines have
230 a neck diameter much smaller than the diameter of the head, head width >2 neck width; thin spines
231 have a head width <2 neck width. Spine types were grouped as mature (stubby and mushroom) and
232 immature (thin) (Figure 1).

233

234 **Statistics**

235 Statistical analysis for place conditioning experiments was performed on preference scores assessed
236 by calculating the time spent in cocaine (Paired; P) and saline (Unpaired; U) compartments on the
237 test day minus the time spent in the same compartments on the pretest session (Ventura et al., 2007;
238 2013). Data were analyzed, separately for each strain (C57, DBA) and dose (C57: 2.5, 5, 7.5 mg/kg;
239 DBA: 5, 7.5, 10 mg/kg) by repeated-measures ANOVA with 1 between factor (early experience, 2
240 levels: Control, RCF) and 1 within factor (pairing, 2 levels: Paired, Unpaired). In order to evaluate
241 the difference in the time spent in paired and unpaired chambers, repeated-measures ANOVA
242 (pairing, 2 levels: Paired, Unpaired) was carried out separately for each experimental group.

243 C-Fos immunoreactivity in selected brain regions (NAcC, NAcS, CP, ILC, PRLC, Hip, Amy) was
244 analyzed by Student's t-test within each strain.

245 In vivo microdialysis data were analyzed by repeated-measures ANOVAs with one between factor
246 (early experience, 2 levels: Control, RCF) and one within factor (time, 7 levels: 0, 20, 40, 60, 80,
247 100, 120) within each strain for each brain area (mpFC, NAc). Statistical analyses were performed
248 on raw data (concentration of pg/20 μ L). Simple effects were assessed by one-way ANOVA for
249 each time point. Baseline extracellular levels in mpFC (NE, DA) and NAc (DA) of Control and
250 RCF animals were compared by Student's t-test within each strain.

251 Concerning morphological data, two-way ANOVA (early experience, 2 levels: Control, RCF;
252 treatment, 2 levels: Saline, Cocaine) was conducted separately for each strain. Three parameters
253 were analyzed for both mpFC and NAc: total spine density, immature spine density; mature spine
254 density. For all experiments, individual between-group comparisons were performed when
255 appropriate by post hoc test (Duncan's multiple range test).

256

257 **Results**

258 **Conditioned Place Preference**

259 The effects of RCF experience on CPP are shown in Figure 2 (A,B).

260 For C57 mice, repeated-measures ANOVA revealed that RCF animals showed a preference for the
261 cocaine-paired compartment at all the doses: 2.5 mg/kg ($F(1,14) = 15.76, P < 0.001$), 5 mg/kg
262 ($F(1,14) = 11.53, P < 0.005$) and 7.5 mg/kg ($F(1,14) = 9.37, P < 0.005$), while Control animals
263 showed a significant preference for the cocaine-paired compartment only at the high doses, 5 mg/kg
264 ($F(1,12) = 9.18, P < 0.005$) and 7.5 mg/kg ($F(1,14) = 25.26, P < 0.001$), but not at the lowest one
265 (2.5 mg/kg ($F(1,12) = 3.02, n.s.$)) (Figure 2A).

266 Concerning DBA mice, repeated-measures ANOVA revealed that RCF animals showed a
267 significant preference for the cocaine-paired compartment only at the high doses, 7.5 mg/kg ($F(1,14)$
268 $= 10.36, P < 0.001$) and 10 mg/kg ($F(1,14) = 16.44, P < 0.001$), but not at the lowest dose, 5 mg/kg
269 ($F(1,12) = .752, n.s.$), while Control animals showed a significant preference for the cocaine-paired
270 compartment at all the doses: 5 mg/kg ($F(1,12) = 11.34, P < 0.05$), 7.5 mg/kg ($F(1,14) = 17.35, P <$
271 0.001) and 10 mg/kg ($F(1,12) = 10.95, P < 0.05$) (Figure 2B).

272 These data strongly indicate that unstable maternal environment has opposite effects on adult C57
273 and DBA mice increasing and reducing, respectively, the sensitivity to cocaine.

274 **Microdialysis**

275 *Medial Prefrontal Cortex*

276 Data analysis for the effects of cocaine on prefrontal NE and DA outflow in C57 mice showed a
 277 significant early experience x time interaction for NE ($F(1,84) = 3.01, P < 0.05$) and a significant
 278 time effect for DA ($F(6,72) = 16.42, P < 0.005$).

279 Simple effect analyses revealed a significant time effect for DA and NE outflow in both (Cont, RCF)
 280 groups. Cont animals showed a time-dependent increase in NE and DA outflow reaching a maximal
 281 increase of 110 ± 10 and of 110 ± 10 at 20 min respectively. RCF animals showed a time-
 282 dependent increase in NE and DA outflow reaching a maximal increase of 150 ± 10 and of
 283 150 ± 10 at 20 min respectively. Moreover, cocaine produced a higher NE increase in the RCF
 284 group compared to the Control group (Figure 3A); no significant difference in DA outflow between
 285 Cont and RCF groups was evident (Figure 3B).

286 Concerning DBA mice, data analysis for the cocaine effects showed a significant early experience x
 287 time interaction for both NE ($F(1,66)=2.36, P<0.05$) and DA ($F(1,78)=2.26, P<0.05$). Simple effect
 288 analyses revealed a significant time effect for NE and DA outflow in Cont animals and a significant
 289 time effect for NE outflow in RCF animals (Figure 3 D,E). Cont animals showed a time-dependent
 290 increase in NE and DA outflow reaching a maximal increase of 75 ± 10 and of 75 ± 10 at 20 and
 291 at 40 min respectively. RCF animals showed a time-dependent increase in NE outflow reaching a
 292 maximal increase of 70 ± 10 at 20 min. No significant increase in DA outflow was evident. A
 293 significant difference in NE and DA outflow was evident between Cont and RCF animals injected
 294 with cocaine (Figure 3 D,E).

295 Finally, analyses of baseline levels of catecholamine did not reveal a significant difference between
 296 Cont and RCF animals within each strain (NE: C57 Cont mean= 0.754 ; SE= 0.112 ; C57 RCF
 297 mean= 0.561 ; SE= 0.046) ($t_{(14)}=-1.595, n.s.$); DBA Cont mean= 0.637 ; SE= 0.182 ; DBA RCF mean= 0.512 ;
 298 SE= 0.118) ($t_{(11)}=-.557, n.s.$) and DA: C57 Cont mean= 0.538 ; SE= 0.073 ; C57 RCF mean= 0.480 ;
 299 SE= 0.124) ($t_{(12)}=-.390, n.s.$); DBA Cont mean= 0.479 ; SE= 0.071 ; DBA RCF mean= 0.314 ; SE= 0.055)
 300 ($t_{(13)}=-1.797, n.s.$).

301 *Nucleus Accumbens*

302 Data analysis for the effects of cocaine on accumbal DA outflow showed a significant time effect
303 for both strains (C57: $F(1,6)=12.79$, $P<0.005$; DBA: $F(1,6)=2.86$, $P<0.05$).

304 Concerning C57 mice, simple effects analysis revealed a significant time effect for DA outflow in
305 both (Cont, RCF) groups. Cont animals showed a time-dependent increase in DA outflow reaching
306 a maximal increase of $\square 100\%$ at 20 min; RCF animals showed a time-dependent increase in DA
307 outflow reaching a maximal increase of $\square 180\%$ at 20 min. Moreover, a significant difference in DA
308 outflow between Cont and RCF mice injected with cocaine was observed (Figure 3C); cocaine
309 produced a longer and sustained increase in the RCF group compared to the Cont group.

310 Concerning DBA mice, simple effects analysis revealed a significant time effect for DA outflow in
311 both (Cont, RCF) groups. Both groups (Cont, RCF) showed a time-dependent increase in DA
312 outflow reaching a maximal increase of $\square 95\%$ at 20 min (Figure 3F). However, no significant
313 difference in DA outflow between Cont and RCF mice injected with cocaine was observed (Figure
314 3F).

315 Finally, analyses of baseline levels of dopamine did not reveal a significant difference between
316 Cont and RCF animals within each strain (C57 Cont mean=.323; SE=.026; C57 RCF mean=.314;
317 SE=.053) ($t_{(10)}=-.179$, n.s.); DBA Cont mean=.689; SE=.161; DBA RCF mean=.478; SE=.128)
318 ($t_{(13)}=-1.002$, n.s.).

319

320 **c-Fos Immunoreactivity**

321 The effects of the early experience on c-Fos positive nuclei are schematically illustrated in Table 1.
322 Concerning C57 mice, t-test analysis revealed a significant increase of c-Fos in PRLC ($t_{(45)}=-2.453$,
323 $P<0.05$), NAcC ($t_{(48)}=-3.246$, $P<0.01$), NAcS ($t_{(52)}=-3.051$, $P<0.05$), LA ($t_{(55)}=-2.303$, $P<0.05$) and
324 BLA ($t_{(58)}=-3.847$, $P<0.005$), a significant reduced activity in CA1 ($t_{(60)}=2.530$, $P<0.05$) and DG
325 ($t_{(56)}=3.779$, $P<0.001$) of RCF animals in comparison with Control animals. No significant

326 difference between RCF and Cont was evident in CP ($t_{(63)}=-0.546$, n.s.), CA3 ($t_{(51)}=1.707$, n.s.) and
327 CEA ($t_{(58)}=-0.537$, n.s.) and ILC ($t_{(43)}=0.270$, n.s.).

328 Concerning DBA mice, t-test analysis revealed a significant reduced activity of c-Fos in ILC
329 ($t_{(25)}=3.432$, $P<0.01$), PRLC ($t_{(23)}=3.317$, $P<0.01$), NAcC ($t_{(71)}=2.179$, $P<0.05$), NAcS ($t_{(21)}=2.919$,
330 $P<0.01$), CP ($t_{(69)}=2.152$, $P<0.05$) DG ($t_{(62)}=2.116$, $P<0.05$), LA ($t_{(55)}=3.522$, $P<0.01$), BLA
331 ($t_{(64)}=3.145$, $P<0.01$) and CEA ($t_{(62)}=3.387$, $P<0.01$) of RCF group in comparison with Control
332 animals. No significant difference was evident in CA1 ($t_{(66)}=-0.088$, n.s.) and in CA3 ($t_{(67)}=1.556$,
333 n.s.) between RCF and Cont groups.

334 **Morphological analysis**

335 *Medial Prefrontal Cortex*

336 The effects of cocaine on spine density and morphology of C57 and DBA mice are shown in Figure
337 4. Concerning C57 mice, two-way ANOVA for apical dendrites of prefrontal pyramidal neurons
338 reveals a significant treatment x early experiences interaction for mature spine density ($F(1,55) =$
339 4.21 , $P<0.05$) (Figure 4C) but not for total spine density ($F(1,55) = .32$, n.s.) (Figure 4A) and
340 immature spine density ($F(1,55) = .48$, n.s.) (Figure 4B). Analysis conducted within each group
341 (Cont, RCF) shows that cocaine increased mature spine density only in RCF mice ($P<0.05$) (Figure
342 4C).

343 Two-way ANOVA for basal dendrites of prefrontal pyramidal neurons reveals a significant
344 treatment x early experiences interaction for total spine density ($F(1,57) = 6.12$, $P<0.05$), immature
345 spine density ($F(1,57) = 6.05$, $P<0.05$) and mature spine density ($F(1,57) = 4.04$, $P<0.05$).

346 Comparison within Cont and RCF C57 groups showed that cocaine produced a significant increase,
347 compared to the respective saline-treated groups, in total spine density ($P<0.001$) (Figure 4D),
348 immature spine density ($P<0.001$) (Figure 4E), and mature spine density ($P<0.05$) (Figure 4F), in
349 RCF C57 mice but not in Cont C57.

350 Concerning DBA mice, two-way ANOVA for apical dendrites of prefrontal pyramidal neurons of

351 mpFC reveals a significant treatment x early experiences interaction for total spine density ($F(1,67)$
352 $=7.72$, $P < 0.05$), immature spines ($F(1,67) = 7.53$, $P < 0.05$) and mature spines ($F(1,67) = 4.76$, $P <$
353 0.05).

354 Analysis conducted within each group (Cont, RCF) showed that cocaine induced an increase in total
355 spine density ($P < 0.05$) (Figure 4G), immature spine density ($P < 0.05$) (Figure 4H) and mature
356 spine density ($P < 0.05$) (Figure 4I) in Cont DBA but not in RCF DBA mice, in comparison with the
357 respective saline-treated groups.

358 Two-way ANOVA for basal dendrites of prefrontal pyramidal neurons reveals a significant
359 treatment x early experiences interaction for total spine density (Figure 4L) and immature spine
360 (Figure 4M) (respectively: $F(1,67) = 4.09$, $P < 0.05$; $F(1,67) = 3.99$, $P < 0.05$) but not for mature
361 spines ($F(1,67) = 2.10$, n.s.) (Figure 4N). Analysis conducted within each group (Cont, RCF)
362 showed that cocaine produced in RCF DBA a reduction, in comparison with the saline-treated
363 group, in total spine density ($P < 0.05$) (Figure 4L) and no significant effect in Control DBA,
364 compared to the saline-treated group, in total spine density (Figure 4L) and immature spine density
365 (Figure 4M).

366

367 *Nucleus Accumbens*

368 The effects of cocaine on morphological changes in the NAc of C57 and DBA mice are shown in
369 Figure 5.

370 Concerning C57 mice, two-way ANOVA analysis conducted for MSNs in the NAc reveals a
371 significant treatment x early experiences interaction for total spine density ($F(1,85) = 15.28$, $P <$
372 0.001) and immature spine density ($F(1,85) = 8.45$, $P < 0.005$) but not for mature spine density
373 ($F(1,85) = 1.65$, n.s.) (Figure 5A, B, C).

374 Comparison conducted within each group (Cont, RCF) showed that cocaine produced in RCF mice
375 an increase, in comparison with the saline-treated group, in total spine density ($P < 0.001$) (Figure
376 5A) and immature spine density ($P < 0.05$) (Figure 5B), but not in mature spine density (Figure 5C).

377 No significant effect was evident for the same parameters in the Cont group (Figure 5 A,B,C).
378 Concerning DBA mice, two-way ANOVA showed a significant treatment x early experiences
379 interaction for total spine density ($F(1,65) = 9.53, P < 0.005$) and immature spine density ($F(1,65) =$
380 $9.28, P < 0.005$), but not for mature spine density ($F(1,65) = 3.4, n.s.$) (Figure 5 D,E,F).
381 Analysis conducted within each group (Cont, RCF) showed that cocaine produced a significant
382 reduction in total spine density ($P < 0.001$) (Figure 5 D) and immature spine density ($P < 0.001$)
383 (Figure 5 E) in Cont mice in comparison with the saline-treated group. No significant effect was
384 evident for the same parameters in RCF group (Figure 5 D,E,F).

385

386 Discussion

387 This paper provides experimental data supporting the current point of view that early life aversive
388 experiences affect response to drugs of abuse in adult life.
389 Contrasting data on long-term effects of postnatal events on responsiveness to psychostimulants in
390 adulthood have been reported showing increased (Der-Avakian and Markou, 2010; Kosten et al.,
391 2003; Meaney et al., 2002; Brake et al., 2004; Zhang et al., 2005) or reduced (Campbell et al., 1999;
392 Ewing Corcoran and Howell, 2010; O'Connor et al., 2015) sensitivity, probably affecting
393 catecholaminergic cortical-accumbal system (Brake et al., 2004; Moffett et al., 2007). Results
394 presented in this paper showing that an early experience (RCF) may either increase or reduce
395 response to cocaine in mice, permit to reconcile conflicting data supporting the point of view that
396 complex gene-environment interplay may determine either susceptibility or resilience to drugs
397 abuse (Der-Avakian and Markou, 2010; van der Veen et al., 2008).

398 Here we found that RCF C57 and DBA female mice show:

- 399 1) opposite behavioral response to cocaine as evaluated by cocaine-induced CPP;
- 400 2) different catecholamine outflow within the prefrontal-accumbal system induced by systemic
401 cocaine injection, as well as an opposite c-Fos activity in cortical and subcortical structures;

3) opposite morphological changes induced by systemic repeated cocaine in the prefrontal-
accumbens system.

CPP results indicate that unstable maternal environment induced by RCF produced opposite effects on behavioral response to cocaine in adult C57 and DBA mice. In agreement with previous reports, C57 strain shows a greater sensitivity to psychostimulants than DBA strain (Cabib et al., 2000; Orsini et al., 2005; Ventura et al., 2004) but, more importantly, our data indicate that early experience increased and reduced, respectively, the sensitivity to cocaine in adult C57 and DBA mice.

Neurochemical results confirmed behavioral data. Accordingly to previous reports on psychostimulants effects in C57 and DBA strains, Cont DBA mice showed lower sensitivity to cocaine effects on dopamine outflow in the NAc compared with Cont C57 mice (Ventura et al., 2004; Zocchi et al., 2001) as demonstrated by the similar mean percent increase of accumbal DA release shown by two strains using different doses (2.5 mg/kg for C57 and 5.0 mg/kg for DBA). Most importantly, here we found that RCF induced a significantly different prefrontal-accumbal response to cocaine in adult C57 and DBA mice paralleling behavioral results. Cocaine injection induced a lower NE and DA release in the mpFC of RCF DBA mice in comparison with their control and no significant difference in accumbal DA outflow between Cont and RCF groups. A different scenario was evident in RCF C57 mice showing higher prefrontal NE as well as accumbal DA release and no significant difference between Cont and RCF groups in prefrontal DA release. The different prefrontal-accumbal response observed in two strains is consistent with previous reports suggesting a facilitating and inhibitor role of prefrontal NE and DA release, respectively, on accumbal DA outflow (Darracq et al., 1998; Di Segni et al., 2016; Pascucci et al., 2007; Ventura et al., 2003; 2005; 2007). In fact, RCF C57 mice showed a parallel prefrontal NE and accumbal DA cocaine-induced increase. Concerning DBA groups, the reduced increase in both prefrontal NE and DA shown by RCF DBA in comparison with Cont DBA mice is associated with no significant difference in accumbal DA release between RCF and Cont groups. This apparently odd effect can

428 be easily explained by opposite role of prefrontal NE and DA on accumbal DA outflow (Puglisi-
429 Allegra and Ventura 2012; Ventura et al., 2003; 2007; 2015). In fact, the facilitating effect of
430 prefrontal NE release on accumbal DA outflow is probably counteracts by inhibitor effect of
431 prefrontal DA increase, so producing the same “net” effect on accumbal DA release shown by RCF
432 group compared to Cont group.

433 According to behavioral and neurochemical results, c-Fos results indicated an opposite cocaine-
434 induced activity pattern in cortical and subcortical brain areas depending on the genotype. In fact,
435 while RCF C57 mice showed an enhancement of c-Fos positive nuclei in most of the areas
436 investigated in comparison with Cont C57, a reduction was evident in almost all brain areas (except
437 for CA1 and CA3) of RCF DBA mice compared with their controls (Table 1).

438 Exposure of the developing nervous system to early adversity in life can shape synaptic plasticity
439 and neurogenesis (Korosi et al., 2011; Lupien et al., 2009) permanently changing brain functioning
440 and increasing the probability of dysfunctional behavior in adulthood (Cabib and Puglisi-Allegra,
441 2012; Coccorello et al., 2009; McClelland et al., 2001). Mesocorticolimbic system is critically
442 involved in emotion and motivation related processes and dendritic features in mesocorticolimbic
443 brain areas are very sensitive to many psychological conditions. Different levels of maternal care
444 and stress exposure during early postnatal development in rodents, as well as early life trauma in
445 humans, have been shown to induce long-lasting consequences on morphology, especially in
446 corticolimbic areas (Maccari et al., 2014; Musazzi and Marrocco, 2016; Romano-López et al., 2015;
447 Wang et al., 2012).

448 Moreover, neuroplastic changes induced by drug exposure have been suggested to mediate cocaine-
449 related behaviors (Kauer and Malenka; 2007; Kiraly et al., 2010; Pulipparacharuvil et al., 2008;
450 Robinson and Kolb, 2004).

451 Based on these evidences, in the last experiment we evaluated whether RCF experience affected, in
452 a genotype-dependent manner, dendritic spine density induced by repeated cocaine injection. Also

453 for this experiment, we used the two discriminative cocaine doses able to induce conditioned place
454 preference in C57 (2.5 mg/kg i.p.) and DBA (5 mg/kg i.p.) mice.

455 Repeated cocaine injection produced increased spine density in apical portion of prefrontal
456 pyramidal neurons of animals belonging to the groups showing CPP (RCF C57: mature spine
457 density; Cont DBA: total, immature and mature spine density) and increased spine density in basal
458 portion of prefrontal pyramidal neurons in RCF C57 mice (total, immature and mature spine
459 density). Increased spine density in mpFC induced by cocaine injection is in agreement with
460 previous reports (Robinson and Kolb, 1999; 2004; Robinson et al., 2001). Concerning NAc neurons,
461 cocaine injection induced increased spine density in RCF C57 (total and immature spine density)
462 and reduced spine density (total and immature spine density) in Cont DBA mice. While results from
463 C57 are in line with literature (Dumitriu et al., 2012; Martin et al., 2011), data from Cont DBA are
464 more puzzling and unexpected. However, different interpretations could be proposed. First,
465 cocaine-induced behavior has been suggested to be not necessarily related with increased spine
466 density or also to be correlated with decreased spine density (Kiraly et al., 2010; Norrholm et al.,
467 2003; Pulipparacharuvil et al., 2008; Taylor et al., 2007). Moreover, in agreement with our results,
468 it has been recently reported that cocaine can induce opposite morphological modifications in rats
469 depending on the genotype (Miguéns et al., 2015). Finally, note that this is the first study
470 investigating the cocaine-induced morphological alterations in DBA mice and a comparison with
471 other studies is to date not possible.

472 Adverse early experiences (i.e. maternal separation) have been shown to affect drug-related
473 behaviors in adult animals inducing morphological changes in some areas of the corticolimbic
474 system (Higley et al., 1991; Huot et al., 2001; Muhammad and Kolb, 2011).

475 According to these reports, our morphological results strongly confirm a critical role of early
476 postnatal environment, in interaction with the genetic background, on cocaine response in adult
477 animals.

478

479 **Conclusions**

480 It has been recently suggested that early life experiences could have negative consequences or
481 confer adaptive value in different individuals (Rana et al., 2015), and we have previously suggested
482 that RCF experience makes DBA a “model” of anhedonia-like state while increases sensitivity
483 toward rewarding natural stimuli in C57 mice that appear instead resilient to develop an anhedonia-
484 like state (Di Segni et al., 2016), thus supporting the hypothesis of differences in the direction rather
485 than in the intensity of stress-induced effects (Alcaro et al., 2002; Cabib et al., 1997; 2000; Ventura
486 et al., 2001) shown by these inbred strains. Present data confirm the opposite effects that the same
487 early experience has in different individuals depending on the genetic makeup also shedding new
488 light on the important role that later experiences in adulthood have on complex behavioral outcomes.
489 Accordingly to the “three-hit concept” of resilience and vulnerability hypothesis (Daskalakis et al.,
490 2013) as well as to the “plasticity genes” concept (Belsky and Pluess, 2009; Belsky et al., 2013), we
491 suggest that the interaction between genetic factors (inbred strains) and early-life environmental
492 condition (RCF experience) induces a sort of “programmed phenotype” that when exposed to
493 following challenges may promote resilience to expression of a pathological phenotype but when
494 exposed to a different challenge may push the same individual toward a higher vulnerability to
495 develop a different kind of pathological phenotype. In particular, here we present data supporting
496 the hypothesis that C57 mice subjected to RCF, that appear resilience to anhedonia-like phenotype
497 when exposed to a stressful experience in adult life (Di Segni et al., 2016), show instead an
498 increased sensitivity to cocaine whether they get in touch with its effects in adulthood.

499 The implicit concept that underlies our hypothesis is that when subjected to early aversive
500 experiences, it is difficult to anticipate the subsequent development of resilience or vulnerability to
501 a specific psychopathology *tout court* in adulthood, because it is challenging to forecast what type
502 of subsequent experience with which an individual must cope, with the final outcome depending on
503 the interaction between early and later events and his genetic makeup. Detailed molecular
504 investigations will shed light on causal mechanisms responsible for the opposite effects observed in

505 the two strains.

506

507

508

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513

514 **Conflict of Interest:**

515 The authors declare no competing financial interests.

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868 **Figure Legends**

869 **Figure 1.** Schematic representation of medial pre-Frontal Cortex (mpFC; A) and Nucleus
 870 Accumbens (NAc; C) and relative photomicrographs of representative Golgi–Cox impregnated
 871 pyramidal (mpFC) and medium spiny neurons (NAc). Scale bar, 100 μ m.

872 High-power photomicrographs (B) of the representivel dendritic segment and of categories of
 873 spines considered (mature: M=mushroom; S=stubby; and immature: T=thin). Scale bar, 10 μ m.

874

875 **Figure 2. Effect of RCF on cocaine-induced Conditioned Place Preference in C57 and DBA**
 876 **mice.**

877 Results from experiments performed with different doses of cocaine (A; C57: 2.5; 5; 7.5 mg/kg i.p.;
 878 B; DBA: 5; 7.5; 10 mg/kg i.p.; lower line). All data are expressed as mean (X) of time spent (\pm
 879 SEM) in compartments Paired (P) / Unpaired (U) with cocaine on Test day minus the time spent in
 880 the same compartments during the Pre-Test session. * $P < .05$ in comparison with Unpaired
 881 compartment. [A; C57: 2.5 mg/kg; Cont: P, X 45,634 \pm 30,853; U, X -41,794 \pm 39,715; RCF: P, X
 882 89,95 \pm 33,979; U, X -98,65 \pm 33,183; 5 mg/kg; Cont: P, X95,371 \pm 34,541; U, X -56,834 \pm 36,46;
 883 RCF: P, X 69,13 \pm 23,512; U, X -59,44 \pm 30,484; 7.5 mg/kg; Cont: P, X 73,58 \pm 22,75; U, X -89,66
 884 \pm 23,175; RCF: P, X 82,18 \pm 32,732; U, X -52,75 \pm 29,521; B; DBA: 5 mg/kg; Cont: P, X 130,469
 885 \pm 40,51; U, X -97,634 \pm 54,272; RCF: P, X 68,42 \pm 38,477; U, X -32,332 \pm 29,463; 7.5 mg/kg; Cont:
 886 P, X 110,84 \pm 32,046; U, X -90,13 \pm 36,043; RCF: P, X 107,6 \pm 41,156; U, X -89,73 \pm 45,418; 10
 887 mg/kg; Cont: P, X 138,789 \pm 48,463; U, X -110,469 \pm 42,046; RCF: P, X 99,69 \pm 42,807; U, X-99,28
 888 \pm 23,981].

889

890 **Figure 3. Effect of RCF on cortico-accumbal cocaine-induced catecholamine release in C57**
 891 **mice.**

892 Results of cocaine administration in C57 (upper line; 2.5 mg/kg i.p.) and DBA (lower line; 5 mg/kg
 893 i.p.) mice on NE (A,D) and DA (B,E) release in mpFC and DA in NAc (C,F) in Control and RCF

894 mice and relative photomicrographs of representative crazy-violet impregnated brain coronal
 895 section; the arrow indicates the tip of the probe, scale bar = 500 μ m. Cocaine was administrated at
 896 time 0. All data are expressed as percentage changes from baseline level of each group as mean \pm
 897 SEM. * $P < .05$ in comparison with Control (Cont) group of same strain.

898

899 **Figure 4. Cocaine effects on morphological changes in cortical Pyramidal neurons of RCF**

900 **C57 and DBA mice**

901 Results of cocaine (C) administration (A-F, C57; 2.5 mg/kg i.p.; G-N, DBA, lower portion; 5 mg/kg
 902 i.p.) on apical and basal Total Spine Density, Immature and Mature Spine Density. All data are
 903 expressed as mean (X) \pm SEM. * $P < .05$ in comparison with saline (S)-treated mice of same group.

904 [A; Cont: S, X 2,272 \pm 0,2; C, X 3,659 \pm 0,323; RCF: S, X 2,296 \pm 0,2; C, X 3,177 \pm 0,314; B; Cont: S,
 905 X 2,21 \pm 0,196; C, X 3,588 \pm 0,319; RCF: S, X 2,242 \pm 0,198; C, X 3,064 \pm 0,303; C; Cont: S, X
 906 0,062 \pm 0,008; C, X 0,071 \pm 0,011; RCF: S, X 0,054 \pm 0,005; C, X 0,113 \pm 0,017; D; Cont: S, X
 907 0,876 \pm 0,095; C, X 0,803 \pm 0,07; RCF: S, X 0,729 \pm 0,074; C, X 1,218 \pm 0,114; E; Cont: S, X
 908 0,858 \pm 0,092; C, X 0,79 \pm 0,068; RCF: S, X 0,719 \pm 0,073; C, X 1,194 \pm 0,113; F; Cont: S, X
 909 0,018 \pm 0,004; C, X 0,013 \pm 0,003; RCF: S, X 0,01 \pm 0,002; C, X 0,023 \pm 0,005; G; Cont: S, X
 910 2,228 \pm 0,28 C, X 3,246 \pm 0,242; RCF: S, X 2,263 \pm 0,231; C, X 2,063 \pm 0,254; H; Cont: S, X
 911 2,145 \pm 0,231; C, X 3,103 \pm 0,266; RCF: S, X 2,193 \pm 0,225; C, X 1,997 \pm 0,248; I; Cont: S, X
 912 0,084 \pm 0,015; C, X 0,143 \pm 0,02; RCF: S, X 0,07 \pm 0,011; C, X 0,065 \pm 0,012; L; Cont: S, X
 913 0,776 \pm 0,04; C, X 0,83 \pm 0,098; RCF: S, X 0,795 \pm 0,063; C, X 0,617 \pm 0,049; M; Cont: S, X
 914 0,759 \pm 0,039; C, X 0,81 \pm 0,095; RCF: S, X 0,772 \pm 0,06; C, X 0,603 \pm 0,047; N; Cont: S, X
 915 0,017 \pm 0,004; C, X 0,02 \pm 0,004; RCF: S, X 0,023 \pm 0,005; C, X 0,014 \pm 0,003]

916

917 **Figure 5. Cocaine effects on morphological changes in accumbal Medium Spiny Neurons of**

918 **RCF C57 and DBA mice**

919 Results of cocaine (C) administration (A, B, C, C57; 2.5 mg/kg i.p.; D, E, F, DBA; 5 mg/kg i.p.) on

920 Total Spine Density, Immature and Mature Spine Density. All data are expressed as mean (X) ±
 921 SEM. * P< .05 in comparison with saline (S)-treated mice of same group. [A; Cont: S, X
 922 0,896±0,141; C, X 0,828±0,087; RCF: S, X 0,738±0,077; C, X 1,82±0,143; B; Cont: S, X
 923 0,876±0,141; C, X 0,804±0,084; RCF: S, X 0,845±0,148; C, X 1,416±0,15; C; Cont: S, X
 924 0,019±0,004; C, X 0,04±0,007; RCF: S, X 0,024±0,005; C, X 0,054±0,01_D; Cont: S, X
 925 1,156±0,114; C, X 0,645±0,046; RCF: S, X 0,835±0,113; C, X 1,041±0,13; E; Cont: S, X
 926 1,101±0,109; C, X 0,634±0,038; RCF: S, X 0,783±0,104; C, X 0,979±0,12; F; Cont: S, X
 927 0,061±0,016; C, X 0,036±0,01; RCF: S, X 0,041±0,009; C, X 0,062±0,014].

928

929 **Tab 1. C-Fos expression in different brain structures**

930 Cocaine-induced c-Fos expression in RCF C57 and DBA mice in comparison with the respective
 931 Control groups.

932 Results of cocaine administration (C57; 2.5 mg/kg i.p.; DBA; 5 mg/kg i.p.) on c-Fos
 933 immunoreactive cell nuclei in different brain areas (infralimbic Cortex (ILC), prelimbic Cortex
 934 (PRLC), Nucleus Accumbens Core (NAcC), Nucleus Accumbens Shell (NAcS), Caudate-Putamen
 935 (CP), Hippocampus (CA3, CA1, Dentate Gyrus (DG)), Amygdala (Lateral Amygdala (LA),
 936 Basolateral Amygdala (BLA), Central Amygdala (CEA)).

937 All data are expressed as trend arrows. *P<0.05 in comparison with Control (Cont) group of the
 938 same strain. [NAcC: DBA: Cont X 0.174±0.036, RCF X 0.505 ±0.082; C57: Cont X 0.695±0.0772,
 939 RCF X 0.121±0.013; NAcS: DBA Cont X 0.190±0.031, RCF X 0.489±0.0129; C57: Cont X
 940 0.654±0.070, RCF X 0.114±0.141; LA: DBA Cont X 0.651±0.013 RCF X 0.579±0.010; C57: Cont
 941 X 0.107±0.020, RCF X 0.258±0.063; CEA: DBA Cont X 0.740±0.015, RCF X 0.506±0.011; C57:
 942 Cont X 0.132±0.314, RCF X 0.276±0.057; BLA: DBA Cont X 0.966±0.020, RCF X 0.110±0.033;
 943 C57: Cont X 0.131±0.02, RCF X 0.286±0.0401; CA1: DBA Cont X 0.652±0.018, RCF X
 944 0.672±0.099; C57: Cont X 0.269±0.084, RCF X 0.4870±0.0238; CA3: DBA Cont X 0.131±0.0218,

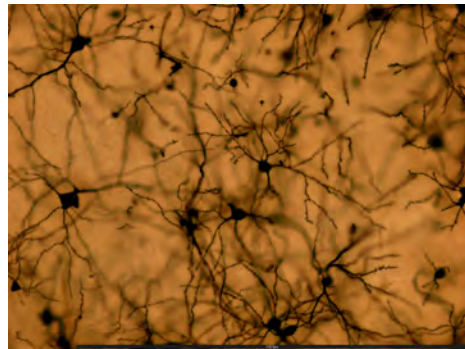
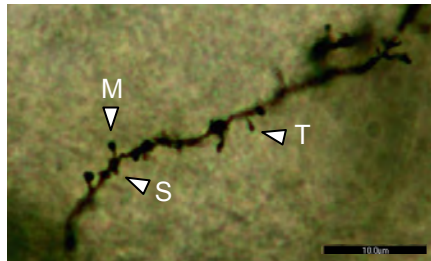
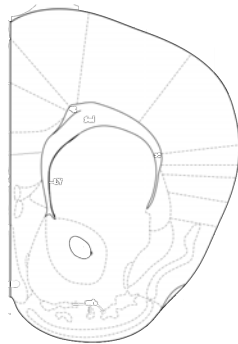
945 RCF X 0.126 ± 0.399 ; C57: Cont X 0.552 ± 0.086 , RCF X 0.329 ± 0.098 ; DG: DBA Cont X
946 0.547 ± 0.091 , RCF X 0.358 ± 0.045 ; C57: Cont X 0.926 ± 0.022 , RCF X 0.393 ± 0.088 ; CP: DBA Cont
947 X 0.113 ± 0.0172 , RCF X 0.143 ± 0.054 ; C57: Cont X 0.238 ± 0.057 , RCF X 0.457 ± 0.010 ; IL: DBA
948 Cont X 0.762 ± 0.092 , RCF X 0.245 ± 0.017 ; C57: Cont X 0.322 ± 0.059 , RCF X 0.270 ± 0.044 ; PRLC:
949 DBA Cont X 0.964 ± 0.012 , RCF X 0.209 ± 0.042 ; C57: Cont X 0.214 ± 0.028 , RCF X 0.409 ± 0.059].

950 **Tab 1.** Effects of RCF on c-fos expression in different brain structures of C57 and DBA mice.

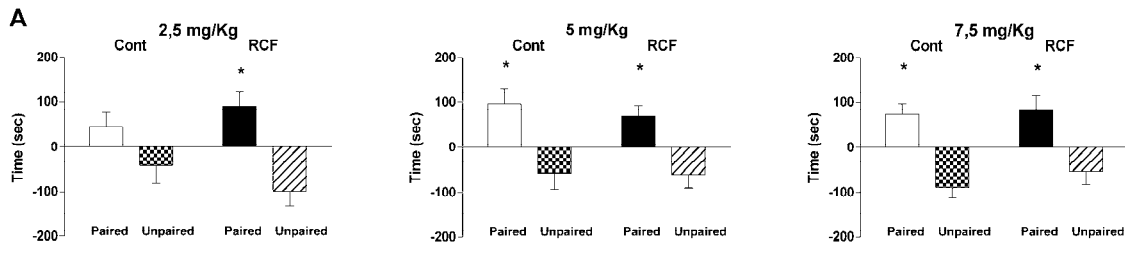
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Brain area	C57 RCF	DBA RCF
ILC	=	↓
PRLC	↑	↓
NAcC	↑	↓
NAcS	↑	↓
CP	=	↓
CA3	=	=
CA1	↓	=
DG	↓	↓
LA	↑	↓
BLA	↑	↓
CEA	=	↓

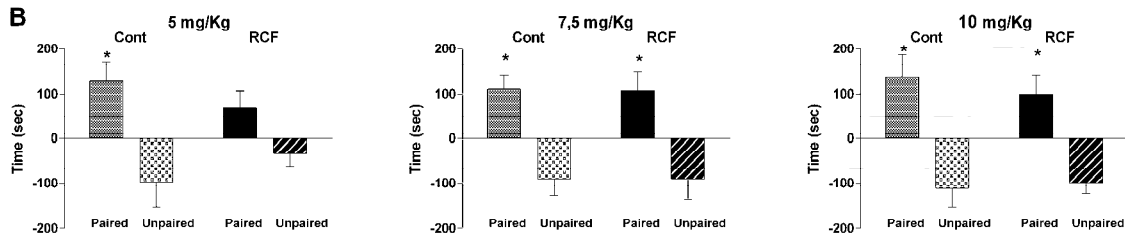
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A**B****C**

C57

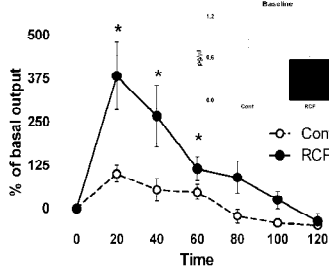
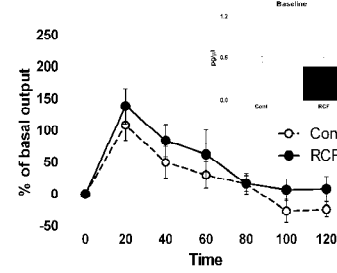


DBA

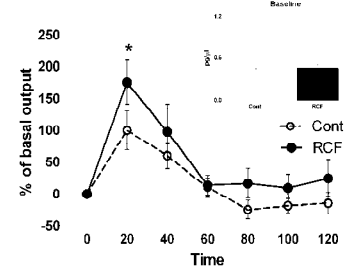


C57 - 2.5 mg/kg

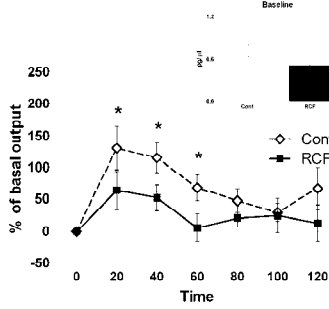
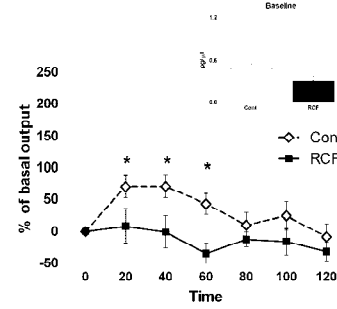
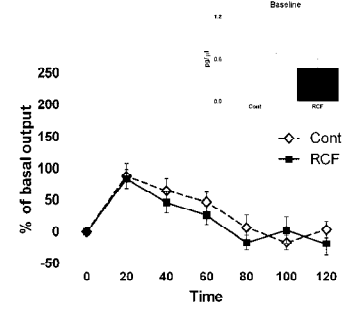
mpFC

A Norepinephrine**B** Dopamine

NAC

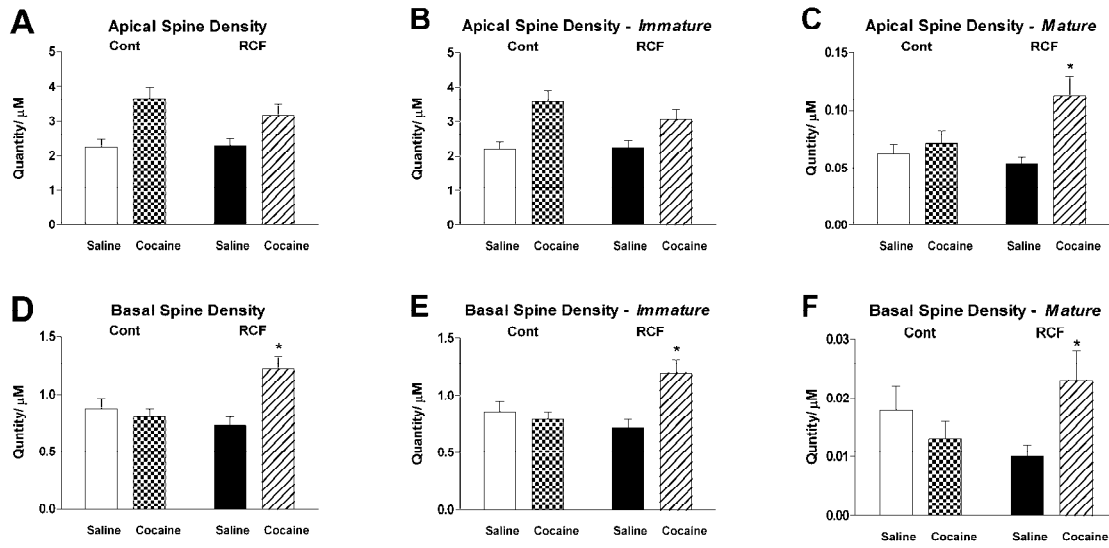
C Dopamine

DBA - 5 mg/kg

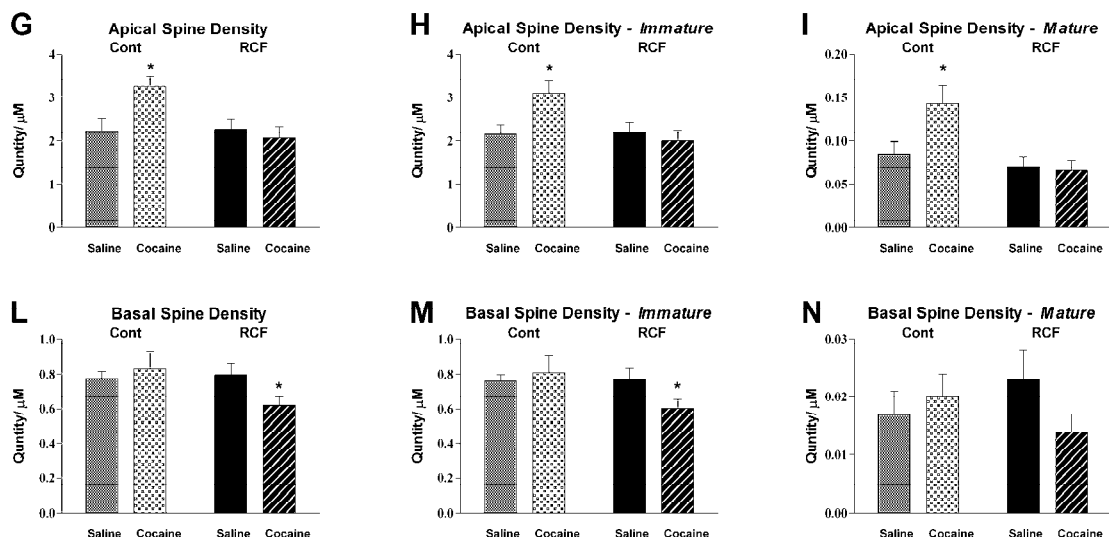
D Norepinephrine**E** Dopamine**F** Dopamine

ACCEPTED MANUSCRIPT

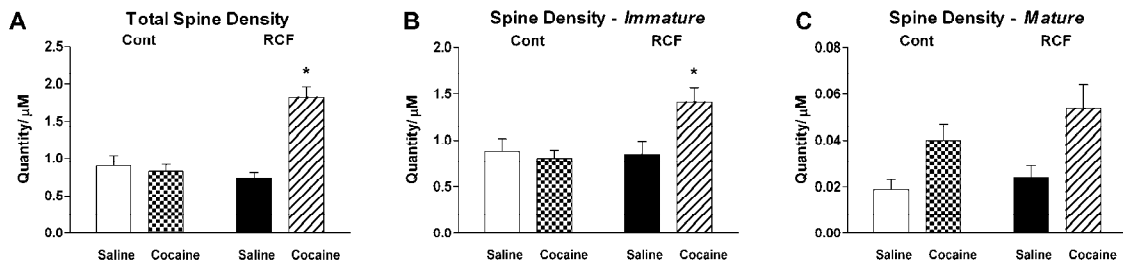
C57



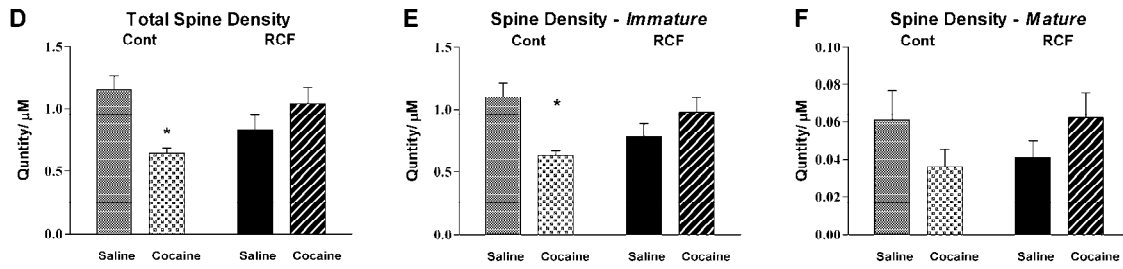
DBA



C57



DBA



Highlights

Genetic makeup interacts with early experience and later experiences in adulthood

Early environment affects cocaine response in adulthood depending on the genotype

Early environment modifies behavioral, neurochemical and morphological response to cocaine

Early environment increases cocaine susceptibility in C57BL6/J mice