

Nasal histamine responses in nonallergic rhinitis with eosinophilic syndrome

Giampietro Zambetti, M.D., Andrea Ciofalo, M.D., Raffaello Romeo, M.D., Pietro Soldo, M.D., Massimo Fusconi, M.D., Antonio Greco, M.D., Giuseppe Magliulo, M.D., and Marco de Vincentiis, M.D.

ABSTRACT

Background: Nonallergic rhinitis with eosinophilic syndrome (NARES) is persistent, without atopy, but with $\geq 25\%$ nasal eosinophilia. Hypereosinophilia seems to contribute to nasal mucosa dysfunction.

Objectives: This analytical case-control study aimed at assessing the presence and severity of nonspecific nasal hyperactivity and at finding out whether eosinophilia may be correlated with the respiratory and mucociliary clearance functions.

Materials: The symptom score was assessed in 38 patients and 15 controls whose nasal smear was also tested for eosinophils and mucociliary transport (MCT). Nonspecific nasal provocation tests (NSNPT) with histamine were also carried out, and total nasal resistance (TNR) was determined.

Results: The symptom score of NARES after NSNPT were not significantly different from the control group, and there was poor or no correlation among the single symptoms and the differences studied for every nasal reactivity class. This correlation improved when using the composite symptom score. The most severe eosinophilia was observed in high reactivity groups, and it was correlated with an increase in TNR. MCT worsened as eosinophilia and nasal reactivity increased. Unlike controls, a significant correlation was observed between the increase in MCT and TNR.

Conclusions: In NARES, nonspecific nasal hyperreactivity is the result of epithelial damage produced by eosinophilic inflammation, which causes MCT slow down, an increase in TNR, and nasal reactivity classes, with possible impact on classification, prognosis, and treatment control.

(Allergy Rhinol 6:e94–e100, 2015; doi: 10.2500/ar.2015.6.0125)

NARES, nonallergic rhinitis with eosinophilic syndrome, is a special form of chronic and persistent inflammatory rhinitis, which occurs in patients who are nonatopic. It is characterized by sneezing attacks, profuse aqueous rhinorrhea, nose-palate pruritus, nasal obstruction, and, occasionally, loss of the sense of smell.¹ Its acute stages are triggered by various factors, including, for instance, temperature changes, intense perfume, and particular climatic conditions. Nasal cytology shows a predominance of eosinophils.² Nonatopy is characterized by negative diagnostic allergy tests.³ Patients with NARES show more severe nasal symptoms compared with other types of allergic and nonallergic rhinitis, and they risk developing obstructive sleep apnea.⁴ Patients with chronic rhinitis show a 2–14% occurrence. Usually, it occurs as an isolated form, but it can be associated with bronchial hyperreactivity or as intrinsic asthma in 50% of patients, with no correlation between the increase in eosinophils count in induced sputum, and in nasal secre-

tion⁵, with aspirin intolerance and nasal polyps. NARES may be an early sign of aspirin-exacerbated respiratory disease, characterized by asthma, nasal polyps, and nonsteroidal anti-inflammatory drug intolerance.⁶ Pathogenetic factors could include chronic, non-specific release of histamine, and self-perpetuating eosinophilic infiltration, with nasal smear that shows more than 25% eosinophils as a diagnostic criterion.² Nonspecific nasal hyperactivity (NSNH), which is the correlation between nasal responsiveness to non-specific nasal provocation tests (NSNPT) and the response to everyday life stimuli, is typical of allergic and nonallergic rhinitis, and can entail vascular and glandular activation as well as the activation of different neuroregulating systems, which thus determines the type of response to nasal stimulation.⁷ In particular, in allergic rhinitis, NSNH requires a certain degree of inflammation, which is ensured by the eosinophil cationic protein and by eosinophils at a later stage of the allergic reaction.⁸ Hence, NSNH and the inflammation of the upper airways could be correlated with each other. This study aimed at determining whether NSNH is present in NARES and to what extent, whether eosinophilia diagnosed through nasal cytology is correlated with the respiratory and mucociliary clearance functions, and whether it is possible to grade NSNH based on the nasal reactivity classes.⁹

From the Rhinology and Immuno-Allergy Unit, Sense Organs Department, Otolaryngology Section, Rome "Umberto I" General Hospital, "La Sapienza" University, Rome, Italy

The authors have no conflicts of interest to declare pertaining to this article

Address correspondence to Giampietro Zambetti, M.D., Rhinology and Immuno-Allergy Unit, E.N.T. Department, "Umberto I" General Hospital, Viale del Policlinico 155, 00161 Rome, Italy

E-mail address: giampietro.zambetti@uniroma1.it

Copyright © 2015, OceanSide Publications, Inc., U.S.A.

MATERIALS AND METHODS

Study Design

Patients with NARES and healthy adult controls were screened at the Rhinology and Immuno-allergy Unit of the Sense Organs Department, Otolaryngology Section of "Umberto I" General Hospital between March 2010 and June 2013. Six weeks after screening, at the first checkup, a prick test was performed, the symptom score was assessed as well as nasal cytology and mucociliary transport (MCT). A week later, at the second checkup, an NSNPT with histamine and rhinomanometric evaluation of the response was carried out, and the evoked symptom score was recorded. The study's protocol was approved by the ethics committee, and all the people enrolled were asked to sign an informed consent form. All aspects of the study complied with the provisions of the Declaration of Helsinki.

Subjects Enrolled

Forty patients with NARES (18 men, 22 women; mean age, 50.7 years) and 15 healthy volunteers (7 men, 8 women; mean age, 39.4 years) were selected and enrolled in this study. The clinical history of the patients with NARES showed persistent symptoms, including sneezing, aqueous rhinorrhea, nasal and palatal pruritus, and nasal congestion that dated back to 2–5 years earlier, with an on-again-off-again pattern; the controls, instead, had a negative clinical history. The two groups underwent a skin-prick test, a radioallergosorbent test (RAST) to detect specific Immunoglobulin E antibodies to the most common allergens in the serum, and, if necessary, a specific nasal provocation test (SNPT) to ascertain the absence of any atopy. None of them were on any medical treatment with drugs that may cause nasal hypo- or hyperreactivity (which included tricyclic antidepressants, antihypertensive drugs such as clonidine and reserpine, decongestants, etc.) or with nonsteroidal anti-inflammatory drugs. Anti-allergenic drugs were discontinued for a 7-week period before the second checkup. Pregnancy and breast-feeding were exclusion criteria, as were concomitant diseases. They all showed an unmistakably negative clinical history of asthma. Rigid fiberoptic nasal endoscopy (LUT, Denzlingen, Germany) was carried out on all the patients and did not show any nasal polyps, any nasal septum deviation, or any other significant alteration. None of the patients enrolled had infectious rhinitis in the 2 weeks before the study. Exclusion criteria also included smoking and exposure to toxic powders or substances. Sinus radiography, which was performed as an ordinary diagnostic procedure, excluded any radiologic and clinical involvement of the paranasal sinuses. Hence, the patients with NARES were still sthenic, that is, in a phase with full nasal symptoms, with frequent "flares," but showing no anosmia or sinusitis.

Skin-Prick Test

The allergy status was assessed in the two groups based on their sensitization to the most common categories of allergens by means of a skin-prick test. The test was considered as positive for sensitization when it revealed a ≥ 3 mm-diameter wheal. The allergens panel (Stallergenes, Milan, Italy) included *Dermatophagoides pteronyssinus*, *Parietaria officinalis*, *Alternaria tenuis*, *Aspergillus fumigatus*, graminaceae mix, dog, cat, composite mix, olea, cypress, and hazel.

Nasal Cytology Test and Staining

For all the patients, nasal smear was collected by scraping the medial surface of the inferior turbinate. The material collected was spread on a glass slide and fixed with cytologic fixative. The material was then stained with May-Grünwald-Giemsa stain (C. Erba, Milan, Italy); then it was mounted, cover slipped, and observed under an optical microscope (BM100FL, Seben, Berlin, Germany). The eosinophil count was performed on 10 immersion microscope fields ($\times 1000$) and was expressed as a percentage over the cell count total. NARES was diagnosed when there were $\geq 25\%$ eosinophils.

Mucociliary Transport

A small amount of Edicol Orange (Edex, Newcastle, Australia) powder was spread on the anterior section of the inferior turbinate by means of a swab. Said powder was anteroposteriorly transported by the cilia of the nasal epithelium toward the nasopharynx, and the time required to cover such distance was the mucociliary clearance time. Edicol Orange left reddish traces along the way, therefore, it was possible to highlight with the staining agent collected by means of a swab positioned behind the soft palate. With this technique, the mucociliary clearance time for healthy patients was 15–18 minutes.

NSNPT With Histamine

Endonasal administration of a solution that contained 2.5 mg of histamine phosphate per milliliter (Sigma Co, St. Louis, MO) was performed in both nasal cavities of each subject, with a predosed spray in a volume of 0.1 mL to give a total dose of 0.5 mg.⁹ This dose was the minimum provocation histamine quantity that allowed a 100% increase in total nasal resistance (TNR) (expressed as $\text{Pa} \times \text{sec} \times \text{cm}^{-3}$) and determined, based on regression coefficients, the NSNPT response magnitude according to four nasal reactivity classes, including low (TNR between 0.51 and 1.1 $\text{Pa} \times \text{sec} \times \text{cm}^{-3}$), medium (TNR between 1.11 and 2.5 $\text{Pa} \times \text{sec} \times \text{cm}^{-3}$), high (TNR between 2.51 and 4.7 $\text{Pa} \times \text{sec} \times \text{cm}^{-3}$), and very high ($> \text{TNR}$ 4.7 $\text{Pa} \times \text{sec} \times \text{cm}^{-3}$). TNR values were matched with histamine doses by using the following exponential equation: $\ln(\text{TNR}) = a + bH$, in which "a" and

"b" were generic regression coefficients, TNR was the nasal resistance response, and "H" was the independent variable, that is, the quantity of histamine inhaled (0.2–0.8 mg). Regression coefficient "b" designed curves that represented the borders of the areas that encompassed the nasal reactivity classes.⁹ Active anterior rhinomanometry using Version 5 of the Jaeger Master-Scope (Carefusion, Yorba Linda, California) was performed 5 minutes before and 5 minutes after the NSNPT with histamine, and TNR was determined at a 150 Pa based on the following calculation: $TNR = (\text{right NR} \times \text{left NR}) / (\text{right NR} + \text{left NR})$. A hundred TNR increase was chosen as the basic value because such an increase cannot be due to the variations of a normal nasal cycle because other studies showed important increases already starting from 75–100%, although without a proper standardization^{9,10} and because excellent correlation has been observed between a minimum 100% TNR increase and the SNPT. Such an increase was the clinical expression of NSNH.

Symptom Score

The following symptom score¹¹ was adopted: 1 point, 3–4 sneezes; 3 points, ≥ 5 sneezes; 1 point, anterior rhinorrhea; 1 point, posterior rhinorrhea; 1 point, nasal breathing problems; 2 points, one blocked nostril; 3 points, both blocked nostrils; 1 point, nose pruritus; 1 point, palatal or ear pruritus; 1 point, conjunctivitis. The overall symptom score was deemed positive if it was ≥ 5 .

Statistics

The VassarStats software (Vassar College, New York, NY) was used to carry out the statistical analysis. Depending on the type of data distribution and on the differences between the two groups, the nonparametric Mann-Whitney *U*-test or the parametric Student *t*-test was performed. Linear regression analyses were calculated with a view to determining the existing correlations of all parameters examined, and $p < 0.05$ values were deemed statistically significant.

RESULTS

Two patients in the NARES group (5% of cases) were prick test results positive for *D. pteronyssinus* (+++) and *P. officinalis* (++), which also was confirmed by RAST. Another patient, who showed an uncertain skin response for the first allergen and was RAST results negative, underwent the SNPT and had a negative result after SNPT. The first two patients were excluded from the study.

The nasal response to NSNPT with histamine in the two groups, which was measured based on the symptom score, is shown in Fig. 1. The data are expressed as median values with 75% confidence intervals. The comparison of data referred to the symptom score

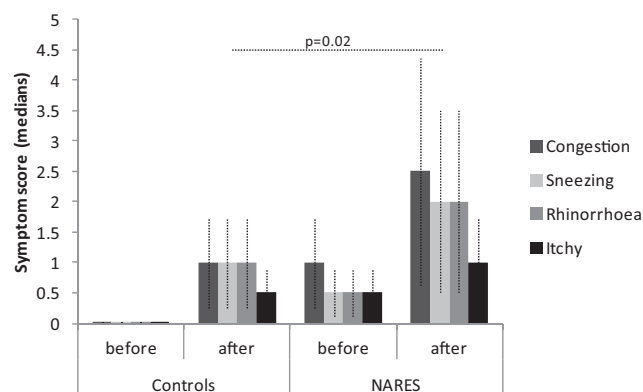


Figure 1. Congestion, sneezing, rhinorrhoea, and pruritus symptom scores for controls and patients with NARES, before and after NSNPT with histamine. Data are presented as medians with 75% confidence intervals.

obtained after and before the NSNPT with histamine has not allowed differentiation of the NARES from the control groups. Only the symptom of sneezing reached a reduced statistical significance ($p = 0.02$). In the NARES, the index of correlation between the score of single symptoms and the TNR, MCT, and the eosinophilia for each reactivity nasal class did not reach, or reached only for some symptoms, low levels of correlation, which, however, improved when using the composite symptom score (r , from 0.75 to 0.87) (Table 1). The median values with 75% confidence intervals for the nasal obstruction symptom score after NSNPT with histamine in the NARES group are shown in Fig. 2. Such values were compared with those of TNRs and then were matched with the nasal reactivity classes obtained after nasal provocation, thus showing that 39% of patient showed very high reactivity, 37% showed high reactivity, 21% showed medium reactivity, and 3% showed low reactivity. The differences among the nasal reactivity classes so obtained were significant, that is, $p = 0.0001$ between strong and consistent reactivity and $p = 0.002$ between consistent and moderate reactivity. As to the controls, 20% of them showed low reactivity and 80% did not show any reactivity after NSNPT.

TNR values after NSNPT with histamine in the NARES group are shown in Fig. 3. The pretest average (SD) values were $0.22 \pm 0.03 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$ for the controls and $0.65 \pm 0.27 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$ for the NARES group ($p < 0.0005$). The average (SD) values after NSNPT were $4.28 \pm 2.31 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$ for the NARES group, and $0.31 \pm 0.15 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$ for the control group, and a comparison of the two groups showed significant results ($p < 0.0005$). In fact, after NSNPT, 20% of the controls showed an increase in TNR average value ($0.57 \pm 0.02 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$), that is, low reactivity, whereas 39% of the patients with NARES were in the very high reactivity class (average, $6.72 \pm 1.32 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$), 37% showed high

Table 1 Index of correlation for nasal reactivity class between Eos, MCT, TNR, and CSS in the NARES group

Nasal Reactivity Classes	Eos vs TNR	Eos vs MCT	TNR vs MCT	Eos vs CSS	TNR vs CSS	MCT vs CSS
Very high	0.66	0.37	0.84	0.29	0.24	0.32
High	0.57	0.39	0.48	0.49	0.19	0.15
Medium	0.91	0.67	0.68	0.69	0.62	0.47
Overall	0.9	0.84	0.94	0.87	0.79	0.75

Eos = nasal eosinophilia; CSS = composite symptom score; MCT = mucociliary transport; TNR = total nasal resistance.

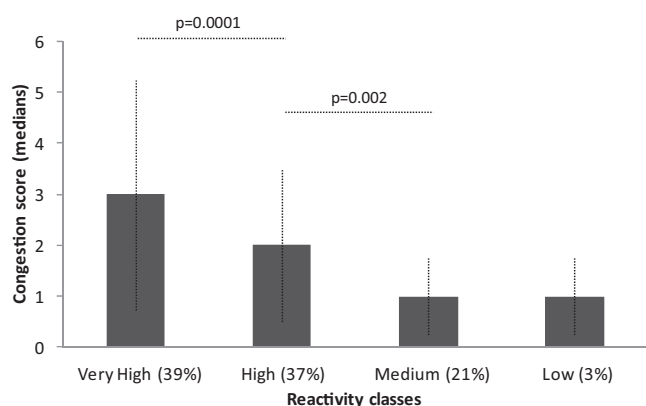


Figure 2. Reactivity classes for the nasal obstruction symptom in patients with NARES after NSNPT with histamine. Data are presented as medians with 75% confidence intervals. The percentage of patients observed in each nasal reactivity classes is reported.

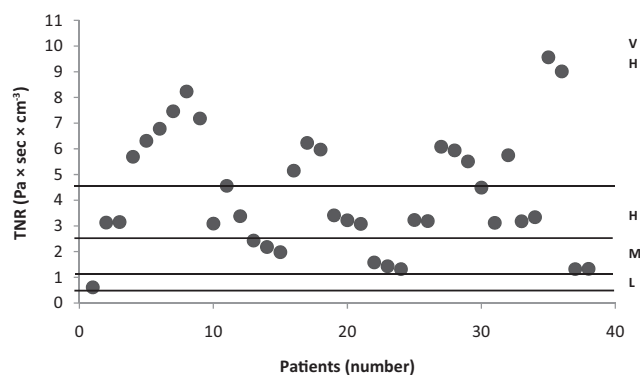


Figure 3. TNR values and nasal reactivity classes after NSNPT with histamine in patients with NARES. Horizontally, there are the following reactivity classes: very high (VH), high (H), medium (M), and low (L).

reactivity (average, $3.40 \pm 0.49 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$), 21% showed medium reactivity (average, $1.7 \pm 0.44 \text{ Pa} \times \text{sec} \times \text{cm}^{-3}$), and 3% showed low reactivity (Table 2). These percentages of the subjects were similar to those mentioned above for the nasal obstruction symptom after NSNPT. In the NARES group, the mean percentage of eosinophils in the nasal smear was $45 \pm 9\%$. The mean eosinophilia values matched with the reactivity classes were $54 \pm 4\%$ for the very high class, $45 \pm 2\%$

for the high class, $32 \pm 5\%$ for the medium class, and 25% for the low class (Table 2). The linear regression analyses of the relationship between the TNR increase after NSNPT and the eosinophils counted in the nasal smear of patients of NARES (Fig. 4) showed remarkable correlation between the two variables ($R^2 = 0.82$; $p = 0.001$). Conversely, there were no eosinophils in the nasal smear of the control group.

The MCT average (SD) values of the NARES group, expressed in minutes, was higher than that of controls (30 ± 7 minutes versus 14 ± 2 minutes; $p = 0.001$), and the related mean values matched with the reactivity classes. These values were as follows: 37 ± 4 minutes for the very high class, 27 ± 2 minutes for the high class, 24 ± 3 minutes for the medium class, and 19 minutes for the low class (Table 2). MCT was related to the percentage of eosinophils in the nasal smear of the NARES group (Fig. 5), and linear regression analysis showed a determination coefficient of the two variables, of $R^2 = 0.71$, with $p = 0.001$. MCT and TNR responses after NSNPT with histamine in the NARES group (Fig. 6) exhibited a significant linear correlation ($R^2 = 0.89$; $p = 0.001$), whereas there was no correlation for the control group ($R^2 = 0.05$; $p = \text{not significant}$). In regard to the TNR, MCT and nasal eosinophilia (Table 2) in the NARES group were observed with significant differences between before and after NSNPT for all classes of nasal reactivity ($p > 0.0005$) and for the TNR between NARES and control groups both before and after NSNPT ($p < 0.0005$). In the end, the correlations among TNR, MCT, and nasal eosinophilia were strong or moderate for every reactivity class (Table 1).

DISCUSSION

NARES is described as a chronic eosinophilic self-perpetuating inflammation with nonspecific histamine release. It is characterized by increased tryptase in nasal fluid (which also characterizes allergic rhinitis and various forms of nonallergic rhinitis) and eosinophil cationic protein levels, whereas the fluid's Immunoglobulin E level shows no significant increase. The level of eosinophils in the nasal smear, instead, is increased¹² as well as the levels of proinflammatory Th2-type cytokines, IL-6 and IL-17 in the nasal fluid, which seem to be a factor of

Table 2 Mean (SD) values of TNR before and after NSNPT with histamine, of MCT, and of nasal eosinophilia in patients with NARES and in the controls

Variables	NARES, Mean (SD)		Controls, Mean (SD)		p*	p#	p§
	Pre-NSNPT	Post-NSNPT	Pre-NSNPT	Post-NSNPT			
TNR, Pa × sec × cm ⁻³							
Overall	0.65 ± 0.27	4.28 ± 2.31	0.22 ± 0.03	0.31 ± 0.15	<0.0005	<0.0005	
Very high	0.91 ± 0.26	6.72 ± 1.32			<0.0005		<0.0005¶
High	0.78 ± 0.35	3.40 ± 0.49			<0.0005		<0.0005
Medium	0.63 ± 0.17	1.71 ± 0.44			<0.0005		<0.0005**
Low	0.29 (1 case)	0.61 (1 case)	0.24 ± 0.05	0.57 ± 0.02			
MCT, min							
Overall	30 ± 7		14 ± 2			<0.0005	
Very high	37 ± 4						<0.0005¶
High	27 ± 2						0.01
Medium	24 ± 3						<0.0005**
Low	19 (1 case)						
Eosinophils, %							
Overall	45 ± 9		0			<0.0005	
Very high	54 ± 4						<0.0005¶
High	45 ± 2						<0.0005
Medium	32 ± 5						<0.0005**
Low	25 (1 case)						

MCT = mucociliary transport; TNR = total nasal resistance; NSNPT = nonspecific nasal provocation tests; NARES = nonallergic rhinitis with eosinophilic syndrome.

*Comparisons between before and after NSNPT in the NARES.

#Comparison between NARES and control groups before NSNPT.

§Comparison among nasal reactivity classes.

¶Very high vs high.

||High vs medium.

**Very high vs medium.

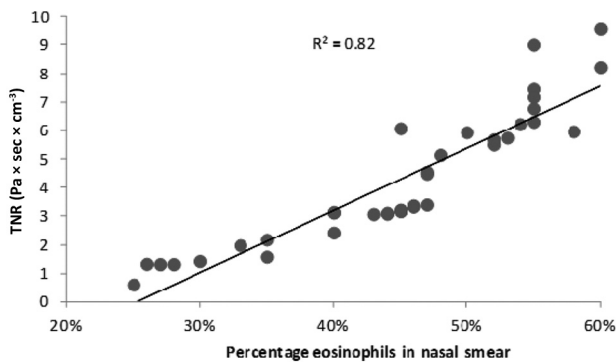


Figure 4. TNR linear regression after NSNPT with histamine versus the eosinophil percentages in nasal smear in the NARES group ($R^2 = 0.82$; $p = 0.001$).

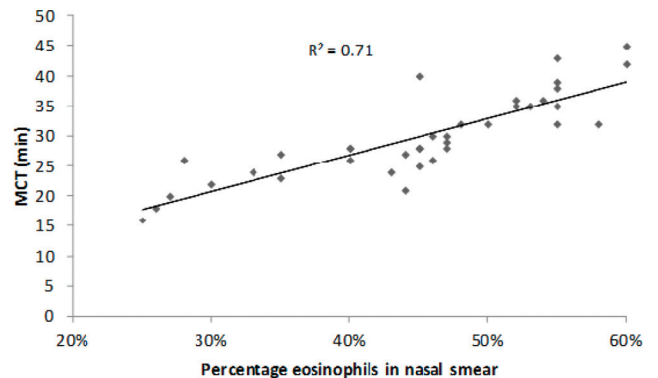


Figure 5. MCT linear regression versus the eosinophil percentages in nasal smear in the NARES group ($R^2 = 0.72$; $p = 0.001$).

neutrophil cell infiltration and NARES remodelling processes.¹³ Almost all our patients with NARES (95%) did not have clinical histories positive for allergy and allergic sensitization to conventional tests. Only two of the patients (5%) showed sensitization to perennial allergens. Hence, hypereosinophilia of the nasal mucosa proved to

be a pathognomonic sign for this disease because it can contribute to its dysfunction.¹⁴⁻¹⁶

NSNH evoked by the NSNPT with histamine^{17,18} was significantly stronger in patients with NARES than in the healthy controls. Generally, nonspecific, autonomic stimuli have a stronger effect on nonallergic rhinitis and, hence, on NARES than they have on healthy subjects or

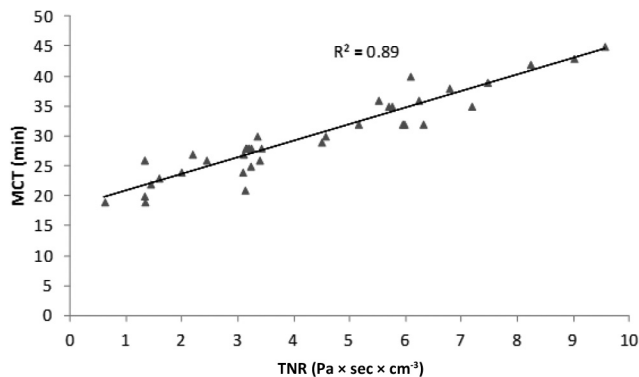


Figure 6. TNR linear regression after NSNPT with histamine versus the MCT of patients with NARES ($R^2 = 0.89$; $p = 0.001$).

on patients with allergic rhinitis,¹ and strengthen NSNH. Although the nasal response to NSNPT with histamine of patients with persistent allergic rhinitis seems to be related to the symptom score that corresponds to the number of sneezes and to the quantity of nasal secretions,¹⁹ in our study, it was not possible to differentiate the NARES group from the control group based on the symptoms. The composite symptom score¹¹ can be considered an instrument to verify the nasal reactivity in the patients for the inclusion of a variety of symptoms, which is preferably measured in a more objective and reproducible way from the variation of the TNR. Severe nasal congestion appeared to be the prevailing symptom²⁰ in the very high and high classes,⁹ whereas patients who belonged in the medium and low categories only showed partial obstruction. Most patients with NARES showed very high and high hyperreactivity (76%), whereas medium and low NSNH with no normal reactivity was observed in the others subjects (24%). All control group patients belonged to lower nasal reactivity classes; in particular, 20% of them showed low reactivity and 80% showed normal reactivity. In the NARES group, there was good correspondence between the nasal obstruction symptom score and TNR after the histamine challenge test. The patients with NARES enrolled in our study showed considerable nasal eosinophilia, in line with the definition of this special form of persistent rhinitis, and more or less significant forms of nasal eosinophilia are reported in literature,^{21–24} but they have never been related to the nasal reactivity classes obtained afterward. The nasal reactivity has been classified as very high, high, medium, and low based on reactivity coefficients.⁹

In NARES, the highest nasal eosinophilia values were observed in patients who belonged to the very high and high reactivity classes, whereas milder eosinophilia was shown by the subjects included in the lower medium and low reactivity classes. This statement was further confirmed by the linear regression analysis between the TNR after histamine and eosinophilia variables. As it has already been pointed out for late allergic reactions¹⁶ and for

persistent allergic rhinitis,²⁵ nasal eosinophilia was correlated with nasal obstruction caused by NSNPT with histamine, thus proving that the eosinophilic inflammation was the cause for nasal hyperreactivity in patients with NARES and, therefore, in the relating nasal reactivity classes. Eosinophils are important cells for the physiopathology of many forms of chronic rhinitis, and there is strong evidence that they play a crucial role in inflammation and consequential tissue lesions.²⁶ The granules of the eosinophils contain toxic basic proteins, most of which are eosinophil cationic proteins. Experimental studies on the eosinophil function revealed the cytotoxic effects on the respiratory epithelium.^{27,28} Of the patients with NARES enrolled in our study, the mucociliary clearance time showed a progressive and significant increase as the proportion of eosinophils in the nasal smear increased, which indicated ciliostasis and a possible relationship with the nasal reactivity classes.

Neural dysfunction can also contribute to NARES symptoms.²⁹ Nasal hyperreactivity shows itself with a worsening of the rhinitic symptoms after NSNPT with histamine, and it has been established that it can be caused by disequilibrium in the sympathetic or parasympathetic nerve system and by the antidromic release of some neuropeptides that cause neurogenic inflammation.^{2,30,31} This hyperreactivity was associated with an increase in the quantity of eosinophils in the smear and results in an increment in nasal resistance. The eosinophils have a cytotoxic effect on the respiratory epithelium by slowing down the clearance time; in fact, we have observed a significant correlation between MCT and TNR obtained after NSNPT with histamine in the NARES group, which was not the case for the controls, thus confirming their role in altering the respiratory function and mucociliary clearance.

CONCLUSIONS

NARES is a poorly understood clinical syndrome, with complex pathophysiogenetic implications,^{32–35} in which the absence of atopy and an increase in nasal eosinophilia was demonstrated. There is a $\geq 25\%$ of eosinophils over the cell total in the nasal smear, and it is pathognomonic for this disease. Other authors define it as a nasal hyperreactivity syndrome without atopy and with $>20\%$ nasal eosinophilia.³⁶ Eosinophils contribute to the dysfunction of the nasal mucosa. In most cases, NARES showed very high or high NSNH, and the proportion of eosinophils had strong correlations with the nasal reactivity classes. Eosinophils play an important role in causing lesions of the respiratory epithelium, which result in ciliostasis, that is, slowing down of mucociliary clearance, which is correlated with an increase in their proportion in the nasal smear. Ultimately, in NARES, NSNH is the result of the inflammation caused by eosinophils that, by damaging the respiratory epithelium, slow down MCT, cause TNR

increase after NSNPT with histamine, and, hence, higher relating nasal reactivity classes. This could have useful applications in NARES, including for classification purposes, based on the response intensity; for prognostic purposes, in that higher reactivity classes have a worse prognosis; and to control the efficacy of available treatments (antileukotriene drugs, corticosteroid, capsaicin, etc.) for NSNH^{1,37-40} by improving the nasal reactivity classes.

REFERENCES

- Ellis AK, and Keith PK. Nonallergic rhinitis with eosinophilia syndrome. *Curr Allergy Asthma Rep* 6:215-220, 2006.
- Moneret-Vautrin DA, Jankowski R, Bene MC, et al. NARES: A model of inflammation caused by activated eosinophils? *Rhinology* 30:161-168, 1992.
- Jacobs RL, Freedman PM, and Boswell RN. Nonallergic rhinitis with eosinophilia (NARES syndrome). Clinical and immunologic presentation. *J Allergy Clin Immunol* 67:253-262, 1981.
- Kramer MF, de la Chaux R, Fintelmann R, and Rasp G. NARES: A risk factor for obstructive sleep apnea? *Am J Otolaryngol* 25:173-177, 2004.
- Leone C, Teodoro C, Pelucchi A, et al. Bronchial responsiveness and airway inflammation in patients with nonallergic rhinitis with eosinophilia syndrome. *J Allergy Clin Immunol* 100:775-780, 1997.
- Moneret-Vautrin DA, Hsieh V, Wayoff M, et al. Nonallergic rhinitis with eosinophilia syndrome a precursor of the triad: Nasal polyposis, intrinsic asthma, and intolerance to aspirin. *Ann Allergy* 64:513-518, 1990.
- Andersson M, Greiff L, Svensson C, and Persson C. Mechanism of nasal hyper-reactivity. *Eur Arch Otorhinolaryngol* 252(suppl. 1):S22-S26, 1995.
- van Wijk RG, de Graaf-in 't Veld C, and Garrelds IM. Nasal hyperreactivity. *Rhinology* 37:50-55, 1999.
- Zambetti G, Moresi M, Romeo R, et al. Non-specific nasal provocation test with histamine. Analysis of the dose-response curve. *Rhinology* 37:168-174, 1999.
- Randerath W, Smith HJ, Knarr D, and Ruhle KH. Comparison of anterior rhinomanometry and impulse oscillometry in nasal provocation with histamine. *Pneumologie* 52:97-103, 1998.
- Lebel B, Bousquet J, Morel A, et al. Correlation between symptoms and the threshold for release of mediators in nasal secretions during nasal challenge with grass-pollen grains. *J Allergy Clin Immunol* 82:869-877, 1988.
- Kramer MF, Burrow G, Pfrogner E, and Rasp G. In vitro diagnosis of chronic nasal inflammation. *Clin Exp Allergy* 34:1086-1092, 2004.
- Gröger M, Klemens C, Wendt S, et al. Mediators and cytokines in persistent allergic rhinitis and nonallergic rhinitis with eosinophilia syndrome. *Int Arch Allergy Immunol* 159:171-178, 2012.
- Bousquet J, Chanez P, Lacoste JY, et al. Eosinophilic inflammation in asthma. *N Engl J Med* 323:1033-1039, 1990.
- Vange P, Dahl R, Fredens K, and Peterson CG. Epithelial injury by human eosinophils. *Am Rev Respir Dis* 138(pt. 2):S54-S57, 1988.
- Terada N, Konno A, and Togawa K. Biochemical properties of eosinophils and their preferential accumulation mechanism in nasal allergy. *J Allergy Clin Immunol* 94:629-642, 1994.
- Philip G, and Togias AG. Nonallergic rhinitis. Pathophysiology and models for study. *Eur Arch Otorhinolaryngol* 252(suppl. 1):S27-S32, 1995.
- Wuestenberg EG, Hauswald B, and Huettenbrink KB. Thresholds in nasal histamine challenge in patients with allergic rhinitis, patients with hyperreflexory rhinopathy, and healthy volunteers. *Am J Rhinol* 18:371-375, 2004.
- van Wijk RG, Mulder PGH, and Dieges PH. Nasal provocation with histamine in allergic rhinitis patients: Clinical significance and reproducibility. *Clin Exp Allergy* 19:293-298, 1989.
- Bonini S, Rasi G, Brusasco V, et al. Nonspecific provocation of target organs in allergic disease: EAACI-GA²LEN consensus report. *Allergy* 62:683-694, 2007.
- Gelardi M, Maselli Del Giudice A, Fiorella ML, et al. Non-allergic rhinitis with eosinophils and mast cells constitutes new severe nasal disorder. *Int J Immunopathol Pharmacol* 21:325-331, 2007.
- Çomoğlu S, Keles N, and Değer K. Inflammatory cell patterns in the nasal mucosa of patients with idiopathic rhinitis. *Am J Rhinol Allergy* 26:e55-e62, 2012.
- Blom HM, Godthelp T, Fokkens WJ, et al. Mast cells, eosinophils and IgE-positive cells in the nasal mucosa of patients with vasomotor rhinitis. A immunohistochemical study. *Eur Arch Otorhinolaryngol* 252(suppl. 1):S33-S39, 1995.
- Staevska M, and Baraniuk JN. Persistent nonallergic rhinosinusitis. *Curr Allergy Asthma Rep* 5:233-242, 2005.
- de Graaf-in't Veld C, Garrelds IM, Koenders S, and Gerth van Wijk R. Relationship between nasal hyperreactivity, mediator and eosinophils in patients with perennial allergic rhinitis and controls. *Clin Exp Allergy* 26:903-908, 1996.
- Togias A, Nacheiro RM, Proud D, et al. Studies on the allergic and nonallergic nasal inflammation. *J Allergy Clin Immunol* 81:782-790, 1988.
- Kramer MF, and Rasp G. Nasal polyposis: eosinophils and interleukin-5. *Allergy* 54:669-680, 1999.
- Davidson AE, Miller SD, Settupane RJ, et al. Delayed nasal mucociliary clearance in patients with nonallergic rhinitis and nasal eosinophilia. *Allergy Proc* 13:81-84, 1992.
- Numao T, and Agrawal DK. Neuropeptides modulate human eosinophil chemotaxis. *J Immunol* 149:3309-3315, 1992.
- Joos GF, Germonpre PR, and Pauwels RA. Neurogenic inflammation in human airways: Is it important? *Thorax* 50:217-219, 1995.
- Takeda N. Neurogenic inflammation in nasal hyperreactivity. *Nihon Kyobu Shikkan Gakkai Zasshi* 31 (suppl.):108-114, 1993.
- Bousquet J, Fokkens W, Burney P, et al. Important research questions in allergy and related disease: Non allergic rhinitis: A GA²LEN paper. *Allergy* 63:842-853, 2008.
- Powe DG, Kormelink TG, Sisson M, et al. Evidence for the involvement of free light chain immunoglobulins in allergic and nonallergic rhinitis. *J Allergy Clin Immunol* 125:139-145.e1-3, 2010.
- Powe DG, Bonnin AJ, and Jones NS. 'Entropy': Local allergy paradigm. *Clin Exp Allergy* 40:987-997, 2010.
- Rondon C, Canto G, and Blanca M. Local allergic rhinitis: A new entity, characterization and further studies. *Curr Opin Allergy Clin Immunol* 10:1-7, 2010.
- Moneret-Vautrin DA, Jankowski R, and Wayoff M. Clinical and pathogenic aspects of NARES (nonallergic rhinitis with eosinophilic syndrome) [in French with English abstract]. *Rev Laryngol Otol Rhinol (Bord)* 112:41-44, 1991.
- Saito H, Morikawa H, Howie K, et al. Effects of a cysteinyl leukotriene receptor antagonist on eosinophil recruitment in experimental allergic rhinitis. *Immunology* 113:246-252, 2004.
- Filiaci F, Zambetti G, Ciofalo A, et al. Local treatment of aspecific nasal hyperreactivity with capsaicin. *Allergol Immunopathol (Madr)* 22:264-268, 1994.
- Van Gerven L, Alpizar YA, Wouters MM, et al. Capsaicin treatment reduces nasal hyperreactivity and transient receptor potential cationic channel subfamily V, receptor 1 (TRPV1) overexpression in patients with idiopathic rhinitis. *J Allergy Clin Immunol* 133:1332-1339, 2014.
- Filiaci F, Zambetti G, Luce M, and Ciofalo A. Local treatment of nasal polyposis with capsaicin: Preliminary findings. *Allergol Immunopathol (Madr)* 24:13-18, 1996. □