# **Supplementary material**

Performance of bees and beehive products as indicators of elemental tracers of atmospheric pollution in sites of the Rome province (Italy)

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## **1S. Quality Assurance and Control**

The analytical performance parameters of linearity, detection and quantification limit (LOD and LOQ, respectively), precision, and accuracy were evaluated. With the lack of suitable certified reference material, spiked samples were used to determine the elemental recoveries by used methods (Astolfi et al., 2020a). Method blanks, in-house quality control samples, and spiked and non-spiked real samples (three replicates each) were prepared along with every digested sample batch (see for details Astolfi et al., 2020a). Recoveries for all elements fell within 20% of the expected value, with many of the elements recovering within 10%. The within-run precision for all the elements in honey and most of the elements in other matrices was less than 10%. The intermediate precision was less than 15% for most of the elements in all matrices. After digestion, all the samples were left to cool The volume was completed to 20 mL H<sub>2</sub>O for inductively coupled plasma mass spectrometry (ICP-MS) analysis (Astolfi et al., 2020a).

#### 1S.1. Digestion efficiency

A selected tolerance level of the residual carbon content (RCC) in a solution lower than 200 mg  $L^{-1}$  was considered appropriate for the subsequent analyses by ICP-MS (Astolfi et al., 2020a).

#### 1S.2. The ICP-MS analysis

The LODs were calculated with three times the relative standard deviation percentage (RSD%) of ten method blanks multiplied by the background equivalent concentration (BEC)/100 (Astolfi et al., 2020b) and the dilution factor used for sample preparation. At regular intervals (every 20 samples) during all analyses, an intermediate calibration standard was analysed as a sample to monitor the instrument drift. A maximum percentage drift of  $\pm$  10% was considered acceptable for all the elements. Furthermore, calibration blanks (3% HNO<sub>3</sub>) were frequently analysed alongside samples to check for any loss or cross-contamination. An internal standardisation monitored the matrix effects on the sample uptake, and nebulisation with Y, Sc, Rh, In and Th, and measurements

were automatically corrected by the respective ICP software. Yttrium was not used as an internal standard for the ICP-MS analyses of the propolis samples because it is contained in the propolis at a concentration of ~0.25 mg kg<sup>-1</sup> (~0.0025 mg L<sup>-1</sup> in digests).



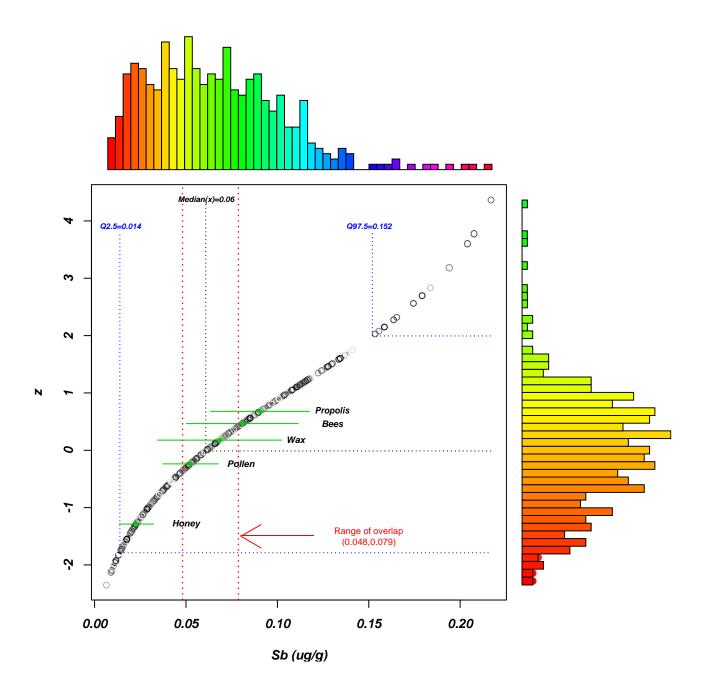


Figure S1. Control chart for Sb built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

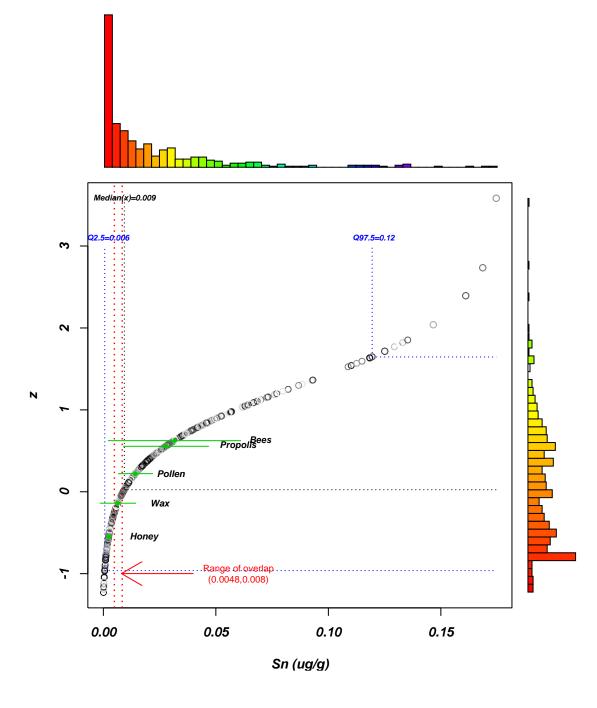


Figure S2. Control chart for Sn built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

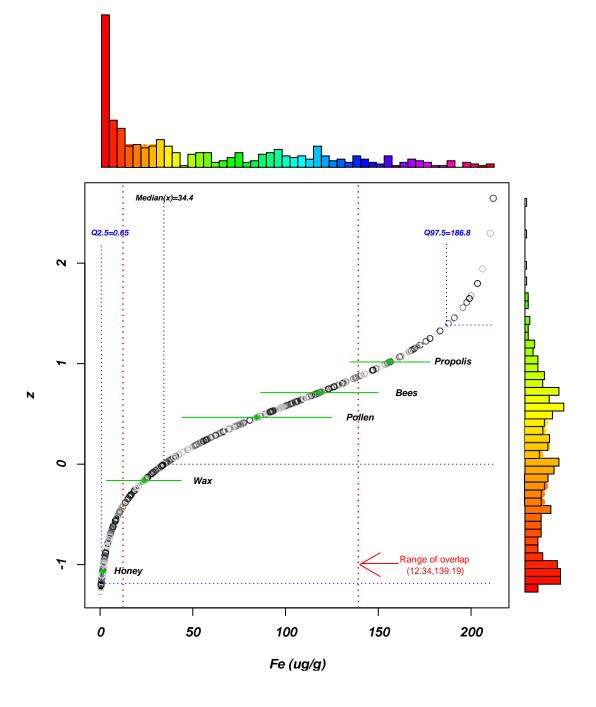


Figure S3. Control chart for Fe built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

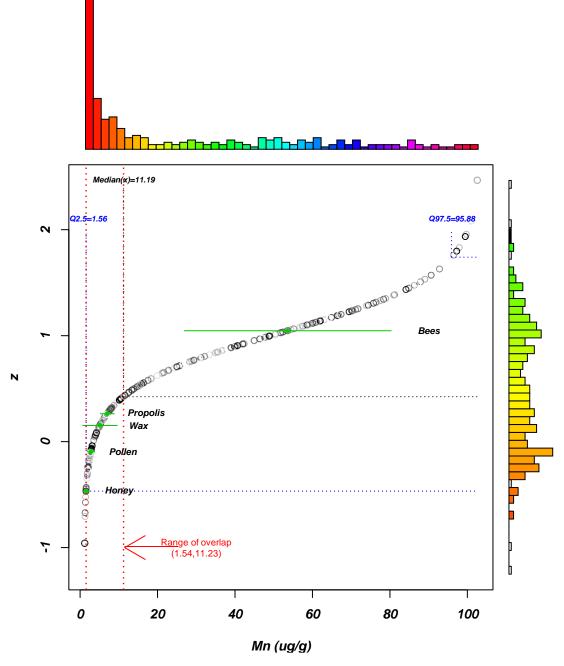


Figure S4. Control chart for Mn built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	0.04	0.14	2.0	2.9
Wax	0.03	0.29	2.6	5.9
Honey	<0.03	0.05	2.6	1.0
Pollen	0.03	0.12	2.6	2.5
Propolis	0.08	0.20	1.0	4.1
Range of overlap	0.05	-0.08		

Table S1. Q2.5 and Q97.5 percentiles of Sb data distribution ( $\mu$ g/g) and Sb OBI index.

Table S2. Q2.5 and Q97.5 percentiles of Sn data distribution ( $\mu g/g$ ) and Sn OBI index.

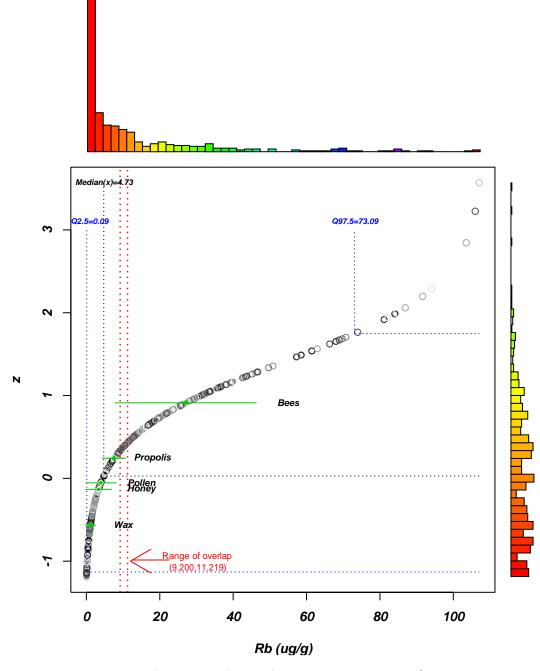
Matrice	Q2.5	Q97.5	OBI-L	OBI-U
Bees	0.0049	0.368	1.000	46.0
Wax	0.0016	0.062	3.06	7.75
Honey	0.0015	0.008	3.26	1.000
Pollen	0.0039	0.187	1.26	23.37
Propolis	0.0036	0.094	1.37	11.75
Range of overlap	0.008-0.0049	·		

Table S3. Q2.5 and Q97.5 percentiles of Fe data distribution ( $\mu\text{g/g})$  and Fe OBI index.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	69.7	247	2.0	20.0
Wax	3.0	70.3	46.3	5.7
Honey	<0.9	12.3	154.4	1.0
Pollen	33.1	154	4.2	12.5
Propolis	139	347	1.0	28.8
Range of overlap	12.3	-139		

Table S4. Q2.5 and Q97.5 percentiles of Mn data distribution ( $\mu g/g)$  and Mn OBI index.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	11.2	139	1.0	92.7
Wax	0.4	28.8	28.0	19.2
Honey	0.1	1.5	112.0	1.0
Pollen	1.9	5.3	5.8	3.5
Propolis	2.9	13.7	3.8	9.1
Range of overlap	1.5-	11.2		



## 3S. Figures and Tables for biomass burning tracers

Figure S5. Control chart for Rb built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

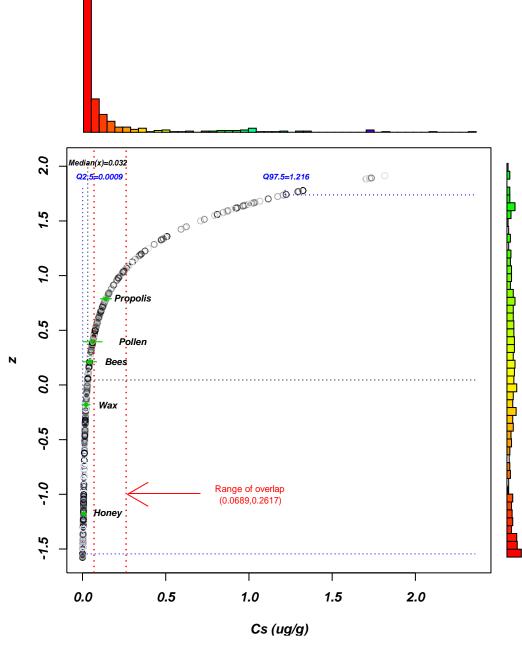


Figure S6. Control chart for Cs built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

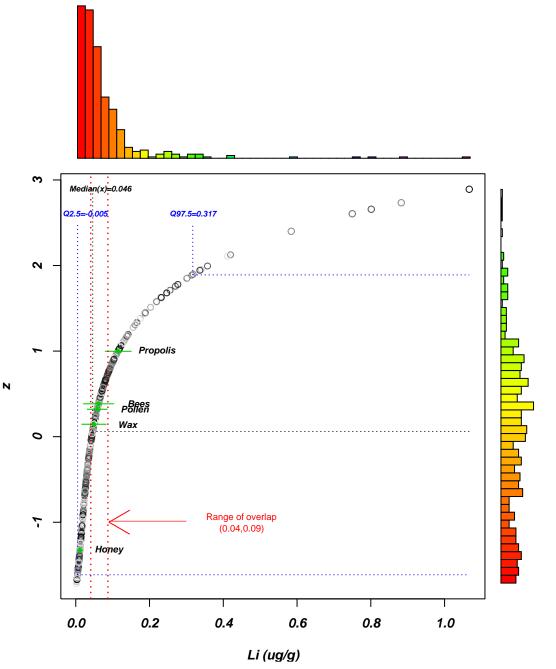


Figure S7.

Control chart for Li built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	9.20	91.9	1.0	8.2
Wax	0.08	43.4	115.0	3.9
Honey	0.32	14.8	29.0	1.3
Pollen	0.95	18.7	9.7	1.7
Propolis	3.75	11.2	2.5	1.0
Range of overlap	9.20	-11.2		

Table S5. Q2.5 and Q97.5 percentiles of Rb data distribution ( $\mu$ g/g) and Rb OBI index.

Table S6. Q2.5 and Q97.5 percentiles of Cs data distribution ( $\mu$ g/g) and Cs OBI index.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	0.027	1.70	2.6	6.5
Wax	0.002	1.01	34.5	3.9
Honey	0.008	0.262	8.6	1.0
Pollen	0.051	1.30	1.4	5.0
Propolis	0.069	0.308	1.0	1.2
Range of overlap	0.069	-0.262		

Table S7. Q2.5 and Q97.5 percentiles of Li data distribution ( $\mu$ g/g) and Li OBI index.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	0.01	0.34	9.0	8.4
Wax	<0.01	0.39	9.0	9.6
Honey	<0.01	0.04	9.0	1.0
Pollen	0.02	0.10	4.5	2.5
Propolis	0.09	0.17	1.0	4.2
Range of overlap	0.04	-0.09		

## 4S. Figures and Tables for soil tracers

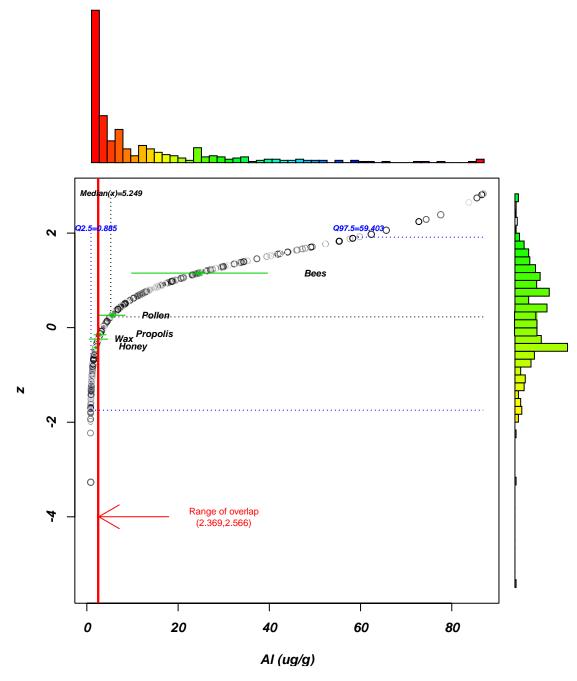


Figure S8. Control chart for Al built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

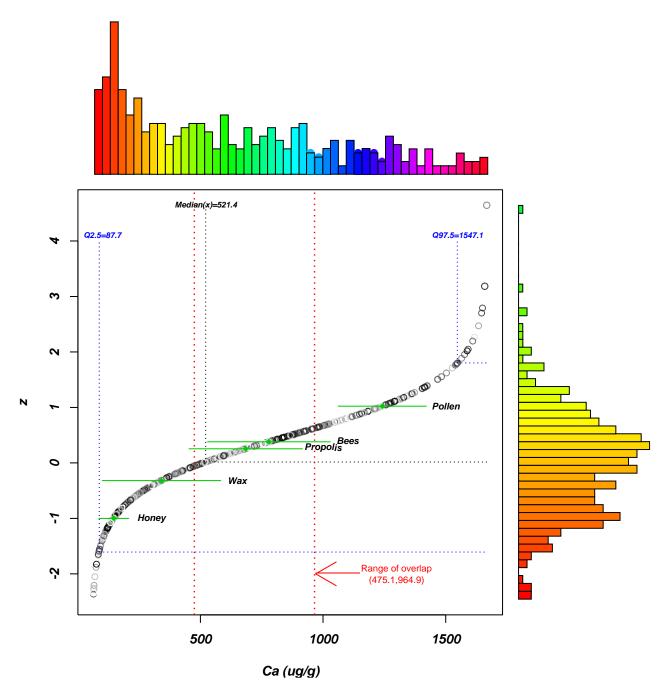


Figure S9. Control chart for Ca built for the five selected biomonitor/indicators with their obtained overlap metal concentrations ( $\mu$ g/g). Observed values are on x-axes, and values calculated by Johnson's method are on y-axes. Inside the plot are reported: the medians ± m.a.d. (median absolute deviation, i.e. green line), the lower and upper bounds of baseline range (Q2.5 and Q97.5), and the range of overlap (i.e., the common elements concentration range for the five biomonitor/indicators, see red arrow). The histograms of values are shown outside of the plot.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	3	85	1.0	42.5
Wax	<1	17	3.0	8.5
Honey	<1	2	3.0	1.0
Pollen	<1	8	3.0	3.3
Propolis	2	7	1.5	3.1
Range of overlap	2	2-3		

Table S8. Q2.5 and Q97.5 percentiles of AI data distribution ( $\mu$ g/g) and AI OBI index.

Table S9. Q2.5 and Q97.5 percentiles of Ca data distribution ( $\mu g/g$ ) and Ca OBI index.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	433	1560	2.2	3.3
Wax	129	1670	7.5	3.5
Honey	52	475	18.6	1.0
Pollen	965	1870	1.0	3.9
Propolis	410	1459	2.4	3.1
Range of overlap	475	-965		

Table S10. Q2.5 and Q97.5 percentiles of Ti data distribution ( $\mu$ g/g) and Ti OBI index.

Matrix	Q2.5	Q97.5	OBI-L	OBI-U
Bees	1.196	4.253	4.7	44.6
Wax	0.0852	2.179	66.5	22.8
Honey	0.005	0.095	1116	1.0
Pollen	0.8027	6.363	7.1	66.7
Propolis	5.58	29.73	1.0	312.9
Range of overlap	0.095	5-5.58		

Table SA. Elements' concentrations in bees, honey, pollen, wax and propolis of selected papers, expressed as µg/g (mean ± S.D., range)

			Non-exhaust traffic tracers Biomass burning tracers								Soil t	racers				
	Reference	Site	Cu	Sb	Sn	Fe	Mn	к	Rb	Cs	Li	ті	Si	AI	Ca	Ti
	This work*	Lazio, Italy	23.7 ± 4.7 (13.0 – 36.8)	0.08 ± 0.03 (<0.03 - 0.21)	0.09 ± 0.11 (0.004 – 0.62)	135 ± 46 (54 – 258)	64 ± 34 (4 – 168)	8340 ± 1400 (5360 – 13600)	37 ± 29 (6 - 160)	0.24 ± 0.43 (0.01 – 1.74)	0.08 ± 0.11 (<0.01 – 1.07)	0.033 ± 0.11 (<0.001 – 0.87)	156 ± 45 (66 – 369)	30 ± 20 (<1 – 98)	834 ± 298 (418 – 2040)	2.4 ± 1.1 (1.2 -6.8)
Bees	(Grainger et al., 2020)	New Zealand (undisclosed hive)	11 -18			48 – 97	11 - 123	5151 – 10713	8.56 - 16.3	0.030 – 0.121		0.0098 – 0.0637		6.7 – 21.4	537 - 734	
	(Zarić et al., 2022)*	Serbia (protected, city centre and thermal power plant) and Austria (city centre)	15.3 – 31.3	0.003 – 0.3		102 -265	32 -131	7281 – 10397	3.0 -10.3	0.004 – 0.042	0.02 – 0.19	0.0001 - 0.0065		16 - 125	855 -1532	
	This work**	Lazio, Italy	0.6 ± 0.3 (<0.3 – 1.5)	<0.03	0.008 ± 0.002 (<0.003 - 0.098)	2.2 ± 2.8 (<0.9 – 16.7)	0.5 ± 0.4 (<0.1 – 2.5)	1030 ± 704 (162 – 3270)	4.9 ± 5.3 (0.2 – 26.1)	0.06 ± 0.11 (0.001 – 0.53)	<0.01	0.012 ± 0.020 (<0.001 - 0.111)	43 ± 8 (14 – 61)	<1	171 ± 114 (<50 – 679)	0.03 ± 0.02 (<0.01 – 0.15)
	(Grainger et al., 2020) - multifloral	New Zealand (undisclosed hive)	0.131 - 0.577			0.80 - 4.06	0.18 – 23.5	134 – 3477	0.15 – 9.46	< 0.003 - 0.000851		< 0.018– 0.00282		0.6 - 14.5	366 -377	
Honey	(Conti et al., 2018) – multifloral**	Lazio, Italy (five different provinces, different areas of	0.06 – 5.4	<0.0008	0.001 - 0.246	<1-4.4	0.09 – 2.8	237 – 6520			0.0011 - 0.024	0.0001 – 0.150		<0.3 - 9.2	<43 - 283	0.001 - 0.230
	(Khuder et al., 2010) – monofloral (2 different ones) and multifloral **	urbanization) Syria (pollution free places and not)	0.60 – 2.20			1.4 -17.2	0.17 – 1.48	46 - 188	0.186 – 0.932						45 - 125	
	(Astolfi et al., 2020a)** - multifloral	Italy (market samples)	0.06 – 1.01			0.58 – 4.28		89 - 4661		0.0006 – 0.787				0.09 – 6.33	39 -181	
	(Sajtos et al., 2022) – monofloral (3	Hungary (five different sites)	<0.04 – 2.65			0.29 -5.94	0.13 – 0.58	254 – 4455			<0.0138			0.08 - 1.49	15– 141	

different ones)														
(Varga et al., 2020)*	Hungary (three different sites)	< 0.0442			<0.37– 7.09	0.040 – 0.307	222 – 494			<0.0138		<0.23 1.02		
(Sajtos et al., 2019) * – monofloral (10 different ones) and multifloral	Hungary (seven different sites)					0.6 -11.9	322 -2466					0.5 -11	5 24 – 218	
(Camiña et al., 2008)	La Pampa, Argentina (capital and non-capital)				1.1 - 10.3							n.d 5.22		
(Voica et al., 2020)* – monofloral (4 different ones)	Romania (three different sites)	0.04– 1.19*				0.02 – 6.14*	60 – 859*					1.19 3.26		0.07 0.36
(Hungerford et al., 2020)	Australia (urban, rural, peri-rural)	0.05 – 4.8	0.005 – 0.01	0.025 - 0.48	0.2 - 99.0	0.4 - 38.0	202 - 4600					0.05		
(Gohar & Shakeel, 2021)	Pakistan (3 different sites) and Turkey (2 different sites)	0 – 3.90	n.d. – 9.56		3.8 - 50.5	0.31 – 3.98	224 – 12197	88 – 16703	1.8 - 84.0	64 - 515			15.6 – 83.6	
(Stihi et al., 2016) – monofloral (9 different ones)	Romania (different sites)	0.10 - 2.04			1.8 - 10.8		187 - 486						34 - 98	
(Atanassova et al., 2016) - multifloral	Bulgaria (two different sites)	0.05 – 0.49			0.7 -19.2	0.3 - 4.7	136 - 1900						24 - 94	
(Squadrone et al., 2020)** – monofloral (4 different ones) and multifloral	Piedmont, Italy	0.30 – 0.76		0.007 – 0.07	1.1 - 2.0	0.61 – 2.2		0.8 – 23				0.5 - 2	6	
(Grainger et al., 2021)	New Zealand (bush, farm, orchard, rural and urban sites)	0.13 – 1.76			0.33 – 3.05	0.3 - 18.9	230 - 2030	0.64 - 9.78			0.006 – 0.153	0.3 - 1	9.3 24 - 173	
(Đogo Mračević et al., 2020) – monofloral (7 different ones)	Serbia (five different sites)	n.d. – 1.6			0.79 - 5.51	0.21 – 7.96	46 – 467					n.d 4.56		

	(Tahboub et al.,	Jordan (market	0.14 -		0.034-	7.0 – 49.3	0.13 -	27 - 2404		0.0002 -	0.0003 -	0.0002 -		2.7 – 21.9	27 - 205	
	2022)	samples)	2.34		0.186		13.4	442 5400	0.45 4.00	0.0379	0.0729	0.0134	0. 170		5.445	
	(Bayram et al., 2020) - monofloral (13 different ones)	Turkey (seven different sites)	0.08 – 2.42	n.d.	n.d. – 0.3		0.1 – 27.8	442 – 5400	0.15 - 4.33	n.d. – 0.08	n.d. – 0.60		3 – 172		5 -115	
	(Meister et al., 2021) – unifloral (3 different ones)	Wairarapa, New Zealand (five different sites)	0.13 – 0.30			0.7 - 1.2	1.1 - 4.2	463 - 1108						5 - 11	60 – 68	
	(Hategan et al., 2021)	French and Romania – monofloral (11 different ones)	n.d. – 8.67	0.0005 0.040	0.01 - 242.05	0.3 - 148	0.2 – 36.5	64 – 3238	0.1 - 14.5		n.d. – 0.68			0.02 – 90.19		
	(Kędzierska- Matysek et al., 2021) – monofloral (4 different ones) and multifloral	Lublin, Poland	0.47 – 0.95			0.00139 – 0.00185	0.72 – 5.58	0.7 - 1.2								
	This work**	Lazio, Italy	1.0 ± 0.4 (0.5 – 2.3)	0.06 ± 0.02 ( <0.03 – 0.13)	0.042 ± 0.032 (0.006 – 0.159)	86 ± 40 (20 -197)	3.2 ± 1.5 (1.6 – 8.1)	2030 ± 484 (1200- 31400)	6.1 ± 5.3 (0.6 – 19.5)	0.28 ± 0.40 (0.01 – 1.32)	0.06 ± 0.03 ( <0.01 - 0.10)	0.035 ± 0.090 (<0.001– 0.429)	129 ± 32 ( 48 – 221)	4 ± 2 (<1 - 9)	1310 ± 218 (664 – 2000)	2.7 ± 1.6 (0.64 – 8.92)
	(Grainger et al., 2020)	New Zealand (undisclosed hive)	9.0-25.0			65-385	32.4- 61.5	5275 – 7077	6.0-18.0	<0.003				26.5-68.4	1730 - 4059	
	(Matuszewska et al., 2021)	Poland (inner village)	4.33- 5.17	0.008- 0.009		47.0-51.3	16.7- 33.3	4166-4300					33.8– 46.7	22.6 – 29.7	1226 – 1250	
Pollen	(Astolfi et al., 2020a)**	Italy (market samples)	5.82			24.0		3703		0.0142				3.68	547	
	(Lilek et al., 2021)*	Slovenia (four macro-regions)				81 - 145	14.2 – 93.1	2710-11400	12.4-43.1						1080 – 2360	
	(Pavlova et al., 2022)	Albania and Bulgaria (serpentine sites)	9.45 – 9.59			103 -246	29.5 - 29.6	5234-5260							1506 – 1811	
	(Swiatly- Blaszkiewicz et al., 2021)	Poland (west- central)	4.4 - 5.7			49.0 -68.0	16.0- 62.0	4400 - 4600							680 - 1500	
	(Mayda et al.,	Turkey (five	9.3 –	0.01 -	16 - 105	84 - 258	17 - 110	5429 – 8994	6.1-30.1	0.03 -0.11	0.01 -	n.d. –	47 -	3 -341	236 - 447	

	2020)	different sites)	20.0	0.08							2.38	0.75	416			
	(Pohl et al., 2020)	Various countries	0.1 – 27.7	0.01 – 0.50	1.72 – 6.46	2.6 -1180	0.1 - 367	0.4-38.0	29.4 -89.7		1.91 – 2.35	n.d. – 0.02	0.1- 10.5	0.1 - 836	0.13-5.19	
	(Atanassova et al., 2016)	Bulgaria (two different sites)	3 - 17			53 - 79	15 -17	3533 -3746							1500 - 2211	
Wax	This work*	Lazio, Italy	2.2 ± 4.6 (<0.3 - 37.0)	0.08 ± 0.03 (<0.03 – 0.37)	0.343 ± 0.023 (0.016 – 6.92)	29 ± 23 (2 -129)	3.8 ± 10.7 (<0.1 – 61.0)	1400 ± 1960 (19 -9480)	6.6 ± 12.8 (0.05 – 179)	0.14 ± 0.34 (0.001 – 2.36)	0.08 ± 0.13 (<0.01 - 1.07)	0.028 ± 0.069 (<0.001 – 0.474)	37 ± 21 (4 – 128)	5 ± 9 (<1 – 58)	559 ± 414 (88 – 1930)	0.69 ± 0.59 (0.03 – 2.47)
	(Astolfi et al. <i>,</i> 2020a)*	Italy (market samples)				0.15				0.000091						
	(Gajger et al., 2019)	Croatia (rural, city and industrial sites)	12.8 – 40.9			56 -286	16.6 – 32.9		11.1 - 15.0							
	This work**	Lazio, Italy	3.1 ± 1.7 (0.9 – 7.9)	0.12 ± 0.04 (0.04 – 0.25)	0.970 ± 0.026 (0.10 – 3.69)	243 ± 25 (137 – 528)	7.3 ± 3.0 (2.7 – 15.7)	1250 ± 822 (539 – 3910)	7.7 ± 3.5 (2.6 – 14.9)	0.156 ± 0.067 (0.063 – 0.321)	0.12 ± 0.04 (0.07 – 0.22)	0.009 ± 0.004 (0.004 – 0.019)	354 ± 31 (321 – 1080)	4 ± 3 (2 – 12)	785 ± 327 (398 – 1630)	11.6 ± 7.7 (5.1 – 35.4)
	(Matuszewska et al., 2021)	Poland (inner village)	1.60 – 1.69	0.031 – 0.042		86 - 143	6.87 – 7.53	647 – 767					101 – 157	86-127	293 – 453	
Propolis	(Arslan et al., 2021)	Turkey (two different sites)	2.01 – 2.45			428 - 508	5.30 – 7.47	1156 – 2607			0.20 – 0.22			407 - 408	269 - 429	
	(Hodel et al., 2020)*	Brazil (nine different sites)	0.6 -11.6					0.23 – 7.94								
	(Mutlu et al., 2022)	Turkey (seven different regions)			n.d 3.69	117 - 627	3.4 -14.1	952 - 1636					268 - 785	122 -475	665 - 2974	n.d. – 62.6

\* dry weight, \*\* wet weight

## References

- Arslan, M., Sevgiler, Y., Güven, C., Murathan, Z. T., Erbil, N., Ylldlrlm, D., Büyükleyla, M., Karadaş, Ş., Çelik, R., & Rencüzoğullarl, E. (2021). Chemical and biological characteristics of propolis from Apis mellifera caucasica from the Ardahan and Erzurum provinces of Turkey: A comparative study. *Arhiv* Za Higijenu Rada i Toksikologiju. <u>https://doi.org/10.2478/aiht-2021-72-3492</u>
- Astolfi, M. L., Conti, M. E., Marconi, E., Massimi, L., & Canepari, S. (2020a). Effectiveness of different sample treatments for the elemental characterisation of bees and beehive products. *Molecules*, 25(18), 4263. https://doi.org/10.3390/molecules25184263
- Astolfi, M.L., Protano, C., Marconi, E., Massimi, L., Brunori, M., Piamonti, D., Migliara, G., Vitali, M., & Canepari, S. (2020b). A new treatment of human hair for elemental determination by inductively coupled mass spectrometry. *Analytical Methods*, 12, 1906–1918. https://doi.org/10.1039/C9AY01871A
- Atanassova, J., Pavlova, D., Lazarova, M., & Yurukova, L. (2016). Characteristics of Honey from Serpentine Area in the Eastern Rhodopes Mt., Bulgaria. *Biological Trace Element Research*. https://doi.org/10.1007/s12011-015-0616-9
- Bayram, N. E., Canli, D., Gercek, Y. C., Bayram, S., Çelik, S., Güzel, F., Morgil, H., & Oz, G. C. (2020). Macronutrient and micronutrient levels and phenolic compound characteristics of monofloral honey samples. *Journal of Food and Nutrition Research*.
- Camiña, J. M., Cantarelli, M. A., Lozano, V. A., Boeris, M. S., Irimia, M. E., Gil, R. A., & Marchevsky, E. J. (2008). Chemometric tools for the characterisation of honey produced in la pampa, argentina, from their elemental content, using inductively coupled plasma optical emission spectrometry (ICP-OES). *Journal of Apicultural Research*. https://doi.org/10.1080/00218839.2008.11101434
- Conti, M. E., Canepari, S., Finoia, M. G., Mele, G., & Astolfi, M. L. (2018). Characterisation of Italian multifloral honeys on the basis of their mineral content and some typical quality parameters. *Journal of Food Composition and Analysis*. https://doi.org/10.1016/j.jfca.2018.09.002
- Đogo Mračević, S., Krstić, M., Lolić, A., & Ražić, S. (2020). Comparative study of the chemical composition and biological potential of honey from different regions of Serbia. *Microchemical Journal*. https://doi.org/10.1016/j.microc.2019.104420
- Gajger, I. T., Kosanović, M., Oreščanin, V., Kos, S., & Bilandžić, N. (2019). Mineral Content in Honeybee Wax Combs as a Measurement of the Impact of Environmental Factors. *Bulletin of Environmental Contamination and Toxicology*. https://doi.org/10.1007/s00128-019-02713-y
- Gohar, A., & Shakeel, M. (2021). Assessment of environmental impact on essential and toxic elements composition in natural honeys by using inductively coupled plasma mass spectrometry. *Environmental Science and Pollution Research*. https://doi.org/10.1007/s11356-020-11688-x
- Grainger, M. N. C., Hewitt, N., & French, A. D. (2020). Optimised approach for small mass sample preparation and elemental analysis of bees and bee products by inductively coupled plasma mass spectrometry. *Talanta*. https://doi.org/10.1016/j.talanta.2020.120858
- Grainger, M. N. C., Klaus, H., Hewitt, N., & French, A. D. (2021). Investigation of inorganic elemental content of honey from regions of North Island, New Zealand. *Food Chemistry*. https://doi.org/10.1016/j.foodchem.2021.130110
- Hategan, A. R., Puscas, R., Cristea, G., Dehelean, A., Guyon, F., Molnar, A. J., Mirel, V., & Magdas, D. A. (2021). Opportunities and constraints in applying artificial neural networks (Anns) in food authentication. honey—a case study. *Applied Sciences (Switzerland)*. https://doi.org/10.3390/app11156723
- Hodel, K. V. S., MacHado, B. A. S., Santos, N. R., Costa, R. G., Menezes-Filho, J. A., & Umsza-Guez, M. A. (2020). Metal Content of Nutritional and Toxic Value in Different Types of Brazilian Propolis. *Scientific World Journal*. https://doi.org/10.1155/2020/4395496

- Hungerford, N. L., Tinggi, U., Tan, B. L. L., Farrell, M., & Fletcher, M. T. (2020). Mineral and trace element analysis of Australian/Queensland Apis mellifera honey. *International Journal of Environmental Research and Public Health*. https://doi.org/10.3390/ijerph17176304
- Kędzierska-Matysek, M., Teter, A., Stryjecka, M., Skałecki, P., Domaradzki, P., Rudaś, M., & Florek, M. (2021). Relationships linking the colour and elemental concentrations of blossom honeys with their antioxidant activity: A chemometric approach. *Agriculture (Switzerland)*. https://doi.org/10.3390/agriculture11080702
- Khuder, A., Ahmad, M., Hasan, R., & Saour, G. (2010). Improvement of X-ray fluorescence sensitivity by dry ashing method for elemental analysis of bee honey. *Microchemical Journal*. https://doi.org/10.1016/j.microc.2009.11.001
- Lilek, N., Kandolf Borovšak, A., Bertoncelj, J., Vogel Mikuš, K., & Nečemer, M. (2021). Use of EDXRF elemental fingerprinting for discrimination of botanical and geographical origin of Slovenian bee pollen. *X-Ray Spectrometry*. https://doi.org/10.1002/xrs.3250
- Matuszewska, E., Klupczynska, A., Maciołek, K., Kokot, Z. J., & Matysiak, J. (2021). Multielemental analysis of bee pollen, propolis, and royal jelly collected in west-central poland. *Molecules*. https://doi.org/10.3390/molecules26092415
- Mayda, N., Özkök, A., Ecem Bayram, N., Gerçek, Y. C., & Sorkun, K. (2020). Bee bread and bee pollen of different plant sources: determination of phenolic content, antioxidant activity, fatty acid and element profiles. *Journal of Food Measurement and Characterization*. https://doi.org/10.1007/s11694-020-00427-y
- Meister, A., Gutierrez-Gines, M. J., Maxfield, A., Gaw, S., Dickinson, N., Horswell, J., & Robinson, B. (2021). Chemical elements and the quality of mānuka (Leptospermum scoparium) honey. *Foods*. https://doi.org/10.3390/foods10071670
- Mutlu, C., Özer-Atakoğlu, Ö., Erbaş, M., & Yalçın, M. G. (2022). Advances in the Elemental Composition Analysis of Propolis Samples from Different Regions of Turkey by X-Ray Fluorescence Spectrometry. *Biological Trace Element Research*, 0123456789. https://doi.org/10.1007/s12011-022-03152-3
- Pavlova, D., Atanassova, J., Karadjova, I., & Bani, A. (2022). Pollen and Chemical Content of Beebreads from Serpentine Areas in Albania and Bulgaria. *Biological Trace Element Research*. https://doi.org/10.1007/s12011-021-02638-w
- Pohl, P., Dzimitrowicz, A., Greda, K., Jamroz, P., Lesniewicz, A., Szymczycha-Madeja, A., & Welna, M. (2020). Element analysis of bee-collected pollen and bee bread by atomic and mass spectrometry Methodological development in addition to environmental and nutritional aspects. In *TrAC Trends in Analytical Chemistry*. https://doi.org/10.1016/j.trac.2020.115922
- Sajtos, Z., Herman, P., Harangi, S., & Baranyai, E. (2019). Elemental analysis of Hungarian honey samples and bee products by MP-AES method. *Microchemical Journal*. https://doi.org/10.1016/j.microc.2019.103968
- Sajtos, Z., Varga, T., Gajdos, Z., Burik, P., Csontos, M., Lisztes-Szabó, Z., Jull, A. J. T., Molnár, M., & Baranyai, E. (2022). Rape, sunflower and forest honeys for long-term environmental monitoring: Presence of indicator elements and non-photosynthetic carbon in old Hungarian samples. *Science of the Total Environment*, 808. https://doi.org/10.1016/j.scitotenv.2021.152044
- Squadrone, S., Brizio, P., Stella, C., Pederiva, S., Brusa, F., Mogliotti, P., Garrone, A., & Abete, M. C. (2020). Trace and rare earth elements in monofloral and multifloral honeys from Northwestern Italy; A first attempt of characterisation by a multi-elemental profile. *Journal of Trace Elements in Medicine* and Biology. https://doi.org/10.1016/j.jtemb.2020.126556
- Stihi, C., Chelarescu, E. D., Duliu, O. G., & Toma, L. G. (2016). Characterisation of Romanian honey using physico-chemical parameters and the elemental content determined by analytical techniques. *Romanian Reports in Physics*.

- Swiatly-Blaszkiewicz, A., Pietkiewicz, D., Matysiak, J., Czech-Szczapa, B., Cichocka, K., & Kupcewicz, B. (2021). Rapid and accurate approach for honeybee pollen analysis using ed-xrf and ftir spectroscopy. *Molecules*. https://doi.org/10.3390/molecules26196024
- Tahboub, Y. R., Al-Ghzawi, A. A. M. A., Al-Zayafdneh, S. S., & AlGhotani, M. S. (2022). Levels of trace elements and rare earth elements in honey from Jordan. *Environmental Science and Pollution Research*. https://doi.org/10.1007/s11356-021-16460-3
- Varga, T., Sajtos, Z., Gajdos, Z., Jull, A. J. T., Molnár, M., & Baranyai, E. (2020). Honey as an indicator of long-term environmental changes: MP-AES analysis coupled with 14C-based age determination of Hungarian honey samples. *Science of the Total Environment*. https://doi.org/10.1016/j.scitotenv.2020.139686
- Voica, C., Iordache, A. M., & Ionete, R. E. (2020). Multielemental characterisation of honey using inductively coupled plasma mass spectrometry fused with chemometrics. *Journal of Mass Spectrometry*. https://doi.org/10.1002/jms.4512
- Zarić, N. M., Brodschneider, R., & Goessler, W. (2022). Honey bees as biomonitors Variability in the elemental composition of individual bees. *Environmental Research*. https://doi.org/10.1016/j.envres.2021.112237