## Appendix

## Towards natural care products: structural and deposition studies of bio-based polymer and surfactant mixtures

Alessandra Del Giudice, <sup>a§</sup> Marta Gubitosi, <sup>b§\*</sup> Adrien Sthoer,<sup>b</sup> Sebastian Köhler,<sup>c</sup> Sophie Ayscough, <sup>d</sup> Maximilian W. A. Skoda, <sup>e</sup> Tommy Nylander, <sup>c,d,f,g</sup> Tobias Halthur<sup>b,h,i</sup>

<sup>a</sup>Sapienza University of Rome, P.le A. Moro 5, 00185 Rome, Italy  ${}^{\bar{b}}$ CR Competence AB, POBox 124, SE-221 00 Lund, Sweden <sup>c</sup>LINXS Institute of Advanced Neutron and X-ray Science, Lund, Sweden  $d$ Division of Physical Chemistry, Lund University, POBox 124, SE-221 00 Lund, Sweden e ISIS Neutron and Muon Source, Harwell Science and Innovation Campus, Didcot OX110QX, U.K. <sup>f</sup>NanoLund, Lund University, Lund, Sweden <sup>g</sup>School of Chemical Engineering and Translational Nanobioscience Research Center, Sungkyunkwan University, Suwon 16419, Republic of Korea hBiomedical Science, Faculty of Health and Society, Malmö University, SE-205 06 Malmö, Sweden  $i$ Biofilms – Research Center for Biointerfaces (BRCB), Malmö University, SE-205 06 Malmö, Sweden.

§These authors equally contributed to the work

\*Corresponding author: marta@crcom.se

## Supporting methods

SAXS models for NaC10 micelles. The core-shell sphere model for surfactant micelles involves highly correlated parameters (the Scattering Length Density - SLD - of core and shell domains with their relative size, and the absolute SLD values with the micelle concentration) [1]. It is therefore necessary to check the consistency of the best-fit parameters with the molecular model and the known sample concentrations. The known concentration of surfactant should let us estimate the expected volume fraction of aggregated objects in the sample.

One possible approach is to assume the macroscopic partial molar volume of NaC10 reported in the literature for the micellar form  $(175.0 \text{ cm}^3/\text{mol}$  [2]) to approximately convert the mass concentration into volume fraction, and thus subtract the free surfactant concentration assumed equal to the CMC. The CMC has been assessed to be 1 wt% by the SAXS dilution series in the Tris buffer 20 mM pH 9 and in the presence of 0.09 M NaCl), and such value compares well to those reported in the literature for the surfactant in pure water, e.g. 1.6 wt% [3] or 2 wt% [2]). The density of a 10 wt% solution of NaC10 in pure water – approximately equal to 0.01 mole fraction – is reported to be less than  $1\%$ different from assuming that 1 g is equal to 1 cm<sup>3</sup> or 1 ml [4]. In this assumption, micellar volume fractions expected for 10, 5, 2 wt% samples would be respectively 0.081, 0.036 and 0.009. In Figure A.1a the SAXS data have been scaled according to these values, showing clearly that neither the intensity of the lower  $q$  maximum (mostly affected by the inter-micellar interference) or the intensity and position of the higher  $q$  maximum (mostly related to the micellar form factor) are perfectly superimposable. In particular, if we consider the data at 5 wt% as a reference, the intensity in the lower q for the 10 wt% sample is much lower than expected, and it is a clear indication of the effect of destructive interference due to the inter-micellar spatial correlations. In addition, the secondary maximum at higher q slightly shifts to lower q values with increasing concentration, suggesting a slight change of the average micelle form factor with concentration.

Zemb and Charpin [1] reported combined SANS and SAXS data fitting for the lower analogue sodium octanoate at concentration 200 g/L, suggesting that for this surfactant the scattering profiles at all accessible concentrations are always due to a combination of micellar form factor and inter-particle structure factor, and they suggest that the parameter values reported in the literature based on SAXS data fitting only at the time were erroneous since neglected the interference effects. Imposing for the core and shell SLD values of  $6.57 \cdot 10^{-4}$  nm<sup>-2</sup> and  $10.3 \cdot 10^{-4}$  nm<sup>-2</sup> as reported for sodium octanoate [1], the best fit for NaC10 at 5 wt% gives a radius of the core of 1.35 nm and a shell thickness of 1.00 nm, also considering a volume fraction of 0.05 and the presence of inter-micellar interactions modelled by a Hayter-Penfold MSA structure factor with charge -7. Using the same SLD values for describing the SAXS data at 10 wt%, leads to a reasonable description of the secondary maximum position and height with similar core radius and shell thickness values (1.33 nm and 0.96 nm), but a much higher volume fraction of 0.167 (more than 3 times the value obtained for the 5 wt% sample). Such assumption also allows for reproducing the main oscillation in the SAXS profile collected at 2 wt% (results reported in Table A.2).

The experimental data for  $q > 2.5$  nm<sup>-1</sup> deviate from the simple form factor of a core-shell sphere and this could be for several reasons. First, the presence of free sodium decanoate at a relatively high concentration can give a contribution to the SAXS intensity in this region, as suggested by the experimental data collected for samples at concentration below 1.5 wt% (Figure 3a in the main text). In addition, even in the absence of free surfactant monomers in a significant amount the form factor of a simple geometrical solid might lose accuracy to reproduce the micelles at the single surfactant molecule resolution (for  $q > 2.5$  nm<sup>-1</sup>, the characteristic distances are  $\leq 2.5$  nm) [5]. Introducing polydispersity or ellipticity of the micellar core sizes can usually improve the model, but in this context we decided as a first approach not to introduce additional parameters due to the low information content of the data in the low  $q$  region where the forward scattering is greatly suppressed due to both possible inter-particle interference and the low overall contrast of the micelles compared to the aqueous background, despite the well-defined contrast of the hydrophilic and counter-ion rich shell. An attempt to improve the fits presented in Figure 3a for NaC10 2% and 5% is presented in Figure A.1b: a background signal in the high  $q$  region was considered to account for the scattering due to free monomers (black dashed line), and, in addition, a deviation from the spherical shape of the micelle form factor (purple solid line) was assumed. The assumption of ellipsoidal shape rather than spherical shape improves indeed the agreement between model and data, whereas the introduction of polydispersity of core radius or shell thickness mainly reduces the depth of the minimum around 0.8 nm<sup>-1</sup> but cannot change the shape of the high q portion of the secondary maximum  $(q > 2.5 \text{ nm}^{-1})$ .

From the assumed value of the core SLD  $(6.57 \cdot 10^{-4} \text{ nm}^{-2})$  i.e. an electron density of 234 electrons/nm<sup>3</sup>), considering the atomic composition of the CH3(CH2)8- hydrophobic tails (73 electrons), a tail volume of 0.313 nm<sup>3</sup> can be estimated (compared with the estimate with the Tanford formula giving  $V_{tail}$  $(nm<sup>3</sup>) = 0.0274 + 0.0269$   $n<sub>C</sub> = 0.270$  nm<sup>3</sup>), and the aggregation number can be calculated from the ratio between the core volume of the fit models and the individual tail volume.

**Model-independent evaluation of SAXS data.** To help the assignment of a  $q_{peak}$  value, the first derivative of the smoothed SAXS data was calculated with the Savitsky-Golay method implemented in Matlab. We assigned the maximum position to the  $q$  value at which the first derivative value crosses zero, and estimated error bars of  $\pm$  0.05 nm.

The estimate of the radius of gyration  $R_g$  and intensity extrapolated at zero angle  $I(0)$  by means of the Guinier approximation was performed with the *datrg* tool of the ATSAS package [6] imposing the same q interval for all data  $(0.16-0.29 \text{ nm}^{-1}$  or  $(0.66-1.1 \text{ nm}^{-1})$  or by using the *autorg* tool to automatically select the optimal interval for Guinier fit linearity for each experimental profile.

The indirect Fourier transform method implemented in the BayesApp code [7] was employed to obtain the pair distance distribution functions  $P(r)$  allowing for negativity, together with additional estimates of  $R_g$  and  $I(0)$ .

For Quatin samples the molecular weight was estimated from the SAXS intensity I(0) according to:  $MW[Da] = \frac{I(q)[cm^{-1}] \cdot N_A[mol^{-1}]}{\Delta \rho_M^2 [cm^2 \cdot g^{-2}] \cdot c[g \cdot cm^{-3}]}$  (eq. A1)

 $\Delta \rho_M^2$  is the squared scattering contrast per mass of polymer, which is obtained as:

$$
\Delta \rho_M^2 = r_e (\rho_{M, dry} - \rho_{solv} \cdot \tilde{v})
$$
 (eq. A2)

where  $\rho_{solv}$  (3.34 $\cdot 10^{23}$  electrons/cm<sup>3</sup>) is the electron density of the water solvent,  $r_e = 2.8179 \cdot 10^{-13}$ cm is the scattering length of the electron,  $\rho_{M, dry}$  in e/g is the number of electrons per mass of dry polymer, obtained from the chemical structure and known degree of cationic substitution, and  $\tilde{v}$  is the partial specific volume of the polymer, estimated using an empirical method [8] and compared with an experimental value reported for inulin in water at 20 °C [9] (Table A.4a).



Figure A.1. a) SAXS patterns of NaC10 samples at concentrations 10, 5 and 2 wt% at 25 °C scaled according to the calculated relative volume fraction of micelles taking the 5% sample as a reference. b) Modelling of the SAXS data from NaC10 micelles up to 5 nm<sup>-1</sup>: background contribution from free NaC10 molecules modelled as a broad peak (dotted black lines), core-shell sphere including background contribution for the 2 wt% sample (black solid line), and core-shell ellipsoid models including background contribution for both 2 wt% and 5 wt% samples (purple solid lines). c) Model fits reported in Figure 2a of the main text (black lines) with highlight on the form factor and structure factor contributions: form factor contributions are reported as dashed colored lines, and in the panel below the structure factor contributions for the three NaC10 concentrations (colored solid lines) are obtained as the overall model intensity divided by the form factor.



Figure A2. a) SAXS patterns of Q680 samples at concentrations from 2 to 0.04 wt% at 25 °C scaled to the data for 1 wt% to optimize overlap in the q region 0.9-2.6 nm<sup>-1</sup>. b) Estimate of the structure factor contribution to the scattering profiles for the most concentrated samples obtained as a pointby-point division of the I(q) of samples with polymer content  $> 0.2$  wt% by the experimental I(q) for the sample at 0.2 wt%, assumed as coincident with the average form factor. The same is shown for the dilution series of  $Q350$  in panels c) and d).



Figure A3. Pair distance distribution functions (normalized by the  $I(0)$ ) obtained by IFT of the SAXS data of mixtures in dilutions series, according to the colour code reported in Figure 4 in the main text. Simulated data of mixtures in the polymer dilution series as a simple sum of surfactant and polymer components, compared with the experimental trend of scattering profiles.



Figure A4. a) SAXS patterns at 25 °C of the NaC10-Q680 2wt% mixtures with NaC10 concentration above and within the coacervation region shown in Figure 5a-b of the main text (colored dots), compared with model profiles obtained considering a monodisperse core-shell sphere model with sticky hard sphere structure factor (black solid line), a monodisperse core-shell ellipsoid model with sticky hard sphere structure factor (grey solid line), and polydisperse core-shell sphere form factors with sticky hard sphere structure factor (dotted lines) considering the SLD of the shell to be fixed (black) or free to be optimized (grey). b) For the model profiles reported in Figure 5a-b of the main text, the colored dashed lines show the form factor contribution, while the solid lines in the panel below represent the structure factor contribution. c) The same is reported for the data shown in Figure 5c of the main text.



(a) Sample details for the SAXS experiments



|   | capillary thickness estimated from alignment scans |
|---|--|
| Capillary path length (mm)  | 1.5 (disposable)                                   |
| Sample volume (µl)  | 150  |
| Exposure time (s)   | 3600, 14400  |
| Number of exposures   | 1  |
| Sample temperature (°C)   | room temperature (20-25 °C)                        |
| (c) SAXS data collection parameters - Synchrotron                                     |  |
| Source, instrument  | BM29 (ESRF)  |
| Detector  | <b>PILATUS 2M</b>                                  |
| Beam geometry $(\overline{mm^2})$   | $0.2 \times 0.2$                                   |
| Wavelength $(\AA)$  | 0.99186  |
| sample-to-detector distance (m)   | 2.813  |
| q-measurement range $(nm^{-1})$   | $0.06 - 5.2$                                       |
| water scattering $I(0) = 0.01632$ cm <sup>-1</sup> at 20°C<br>Absolute scaling method |  |
| Capillary path length (mm)  | $\mathbf{1}$                                       |
| Sample volume (µl)  | 50   |
| Exposure time (s)   | $\mathbf{1}$                                       |
| Number of exposures   | 10   |
| Extra flow time $(s)$   | 15   |
| Sample temperature (°C)   | $20^{\circ}$ C                                     |
| (e) Software employed for SAS data reduction, analysis, and interpretation            |  |
| Solvent subtraction, averaging and basic analysis (Guinier fit)                       | ATSAS 3.2  |
| Indirect Fourier transform  | BayesApp   |
| Theoretical intensity calculations  | SasView, Sasfit                                    |

Table A.2. Details of the models and parameters used for obtaining the calculated model SAXS profiles related to pure NaC10 and quaternary ammonium inulin samples. Error estimates can be considered of  $\pm 1$  on the last shown digit. Parameters that have not been modified in the optimization are shown in grey colour.













Table A.3. Summary of the parameters used to calculate the model SAXS profiles related to mixtures of NaC10 and Quatin680. Parameters that have not been modified in the optimization are shown in grey colour.



Model spherical shell (SASfit 0.94.11, model 3.1.4) with sticky hard sphere S(q) (model 11.4.1)



Model ellipsoidal shell (SASfit 0.94.11, model 8.6.3) with sticky hard sphere S(q) (model 11.4.1) + Generalized Gaussian coil (model 8.4.1.5) and broad peak background (model 7.12.1)



b)

Model spherical shell (SASfit 0.94.11, model 3.1.4) with Lognormal polydispersity of the core radius and with sticky hard sphere S(q) (model 11.4.1)



Table A.4. Estimate of the average molecular weight of quaternary ammonium inulin samples from the SAXS intensity extrapolated at zero angle. a) assumed mass and volumes based on the chemical structure. b) Calculations of molecular weight ( $M_W$ ) and polymerization degree (n) based on the experimental values of I(0) according to equation A.1. The values obtained by Guinier fit for the samples with mass concentration greater than 0.2 wt% were not considered in the average due to possible influence of a repulsive structure factor.







Table A.5. Thickness, solvent content, composition, and adsorbed mass of the layers of the models that best fit the experimental neutron reflectometry data.



 $\mathbf{h}$ 

## **References**

- [1] T. Zemb, P. Charpin, Micellar Structure From Comparison Of X-Ray And Neutron Small-Angle Scattering., J. Phys. Paris. 46 (1985) 249–256. https://doi.org/10.1051/jphys:01985004602024900.
- [2] E. Vikingstad, A. Skauge, H. Høil and, Partial molal volumes and compressibilities of the homologous series of sodium alkylcarboxylates, R6COONaR13COONa, in aqueous solution, J. Colloid Interface Sci. 66 (1978) 240–246. https://doi.org/10.1016/0021-9797(78)90301-6.
- [3] R. De Lisi, G. Perron, J.E. Desnoyers, Volumetric and thermochemical properties of ionic surfactants: sodium decanoate and octylamine hydrobromide in water, Can. J. Chem. 58 (1980) 959–969. https://doi.org/10.1139/v80-152.
- [4] S.M. Calderón, N.L. Prisle, Composition dependent density of ternary aqueous solutions of ionic surfactants and salts: Capturing the effect of surfactant micellization in atmospheric droplet model solutions, J. Atmos. Chem. 78 (2021) 99–123. https://doi.org/10.1007/s10874- 020-09411-8.
- [5] M.T. Ivanović, M.R. Hermann, M. Wójcik, J. Pérez, J.S. Hub, Small-Angle X-ray Scattering Curves of Detergent Micelles: Effects of Asymmetry, Shape Fluctuations, Disorder, and Atomic Details, J. Phys. Chem. Lett. 11 (2020) 945–951. https://doi.org/10.1021/acs.jpclett.9b03154.
- [6] K. Manalastas-Cantos, P. V. Konarev, N.R. Hajizadeh, A.G. Kikhney, M. V. Petoukhov, D.S. Molodenskiy, A. Panjkovich, H.D.T. Mertens, A. Gruzinov, C. Borges, C.M. Jeffries, D.I. Sverguna, D. Frankea, ATSAS 3.0: Expanded functionality and new tools for smallangle scattering data analysis, J. Appl. Crystallogr. 54 (2021) 343–355. https://doi.org/10.1107/S1600576720013412.
- [7] S. Hansen, BayesApp : a web site for indirect transformation of small-angle scattering data, J. Appl. Crystallogr. 45 (2012) 566–567. https://doi.org/10.1107/S0021889812014318.
- [8] H. Durchschlag, P. Zipper, Calculation of the partial volume of organic compounds and polymers, Springer-Verlag GmbH & Company KG, 1994. https://doi.org/10.1007/bfb0115599.
- [9] H. Durchschlag, Thermodynamic Data for Biochemistry and Biotechnology, Springer-Verlag, Berlin - Heidelberg - New York - Tokyo, 1986. https://doi.org/10.1016/0302- 4598(88)80044-8.