## Supporting information

## Self-assembly of model amphiphilic peptides in non-aqueous solvents: changing driving forces, same structure?

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**Table S1** The scaling constants used for plotting the data in the insets of Figure 3A,C are tabulated (last column) together with the experimental concentrations in wt%, the estimated volume fractions  $(\phi)$ , and the ratios of the concentration between the 1 wt% sample chosen as a reference and the more dilute samples (1/Ratio wt%)

Sample	$\mathrm{wt}\%$	$\phi^*$	1/ Ratio wt%	Scaling constant**
A <sub>8</sub> K MeOH-Lab	1.01	0.0053	1	1.2
$A_8K_MeOH$	1.01	0.0053	1	1
	0.49	0.0026	2.1	2.3
	0.26	0.0014	3.9	4.8
	0.12	0.0006	8.2	10.6
	0.06	0.0003	16.7	17.9
	-1	0.0000	1	1.0
$A_8K_DMF$ -Lab	1	0.0063	1	1.3
$A_8K_DMF$	1.03	0.0000	1	1
	0.5	0.0032	2.1	2.2 E.C
	0.25	0.0016	4.2	0.0 11.0
	0.12	0.0008	8.4	11.2
	0.06	0.0004	16.7	24.2
A <sub>10</sub> K MeOH-Lab	1.03	0.0065	1	1.2
$A_{10}K$ MeOH	1.05	0.0066	1	1
10 _	0.52	0.0033	2	2.3
	0.25	0.0016	4.2	4.8
	0.12	0.0008	8.7	9.8
	0.06	0.0004	16.5	17.9
	0.00	0.0075		1.0
$A_{10}K_DMF$ -Lab	0.99	0.0075	1.1	1.3
$A_{10}K_DMF$	1.04	0.0079	1	1
	0.52	0.0039	2	2.3
	0.26	0.0019	4.1	4.7
	0.13	0.001	8.2	15.2
	0.06	0.0005	17.3	33.9

\*estimated considering a peptide density in all solvents of 1.26 g (without counterions)/ml for  $A_{10}K$  and 1.50 g/ml for  $A_8K$ .

\*\*calculated to optimize superimposition in the q range 0.635-1.227 nm<sup>-1</sup>

$\mathbf{Sample}$	Fixed	q Range (nm <sup>-1</sup> )		Optimized				$\chi^2$
	L (nm)	$\mathbf{q}_{min}$	$q_{max}$	$\operatorname{scale}$	bg	radius (nm)	axis ratio	
A <sub>8</sub> K H <sub>2</sub> O								
$1.35 \mathrm{wt}\%$	350	0.1	3.5	$8\cdot 10^{-3}$	$2 \cdot 10^{-3}$	1.64	2.3	7.2
$A_8K$ MeOH								
$0.12 \mathrm{wt}\%$	350	0.1	3.5	$9\cdot 10^{-4}$	$9\cdot 10^{-4}$	1.33	2.2	1.5
$A_8K$ DMF					_			
$0.25\mathrm{wt}\%$	350	0.06	3.5	$2 \cdot 10^{-3}$	$7 \cdot 10^{-5}$	1.33	2.4	1.8
$A_{10}KH_2O$				0				
$1.00  \operatorname{wt}\%$	60	0.1	3.5	$1 \cdot 10^{-2}$	$5 \cdot 10^{-4}$	1.75	3.2	5.5
$A_{10}K$ MeOH								
$0.12~{ m wt}\%$	350	0.1	3.5	$8\cdot 10^{-4}$	$5 \cdot 10^{-4}$	1.57	2.5	9.3
$A_{10}K$ DMF								
$0.12 \mathrm{wt}\%$	350	0.1	3.5	$7\cdot 10^{-4}$	$3\cdot 10^{-5}$	1.57	2.9	4.4

**Table S2** Results of the fitting of the  $A_8K$  and  $A_{10}K$  SAXS data shown in Figure 3. The form factor of an elliptical cylinder was used ( http://www.sasview.org/docs/user/models/elliptical\_cylinder. html). The scattering length densities were fixed:  $11.4 \cdot 10^{-4}$  nm<sup>-2</sup> for the cylinder,  $7.5 \cdot 10^{-4}$  nm<sup>-2</sup> for MeOH,  $8.8 \cdot 10^{-4}$  nm<sup>-2</sup> for DMF and  $9.4 \cdot 10^{-4}$  nm<sup>-2</sup> for water.

**Table S3** Values of the radius of gyration of the cross-section  $R_{CS}$  of a long rod, obtained from the SAXS data of  $A_8K$  and  $A_{10}K$  by applying a Guinier fit for rod-like objects to the scattering intensity multiplied by the scattering vector  $I(q) \cdot q$  in the q range indicated. From this, the number of laminated  $\beta$  sheets constituting the fibril cross section (N) is estimated within the approximation of an elliptical cross section with semi axes a and b, for which  $R_{CS} = \sqrt{(a^2 + b^2)}/2$ , and considering 2a = 3.2 nm for  $A_8K$  and 3.9 nm for  $A_{10}K$ , and 2b as  $0.54 \cdot N$  nm.

Sample	$q_{min} \ (\mathrm{nm}^{-1})$	$q_{max} (\mathrm{nm}^{-1})$	$R_{CS}$ (nm)	2b (nm)	N	$N/N_{water}$
$A_8K$ H <sub>2</sub> O 1.30 wt%	0.24	0.51	2.0	7.4	14	
$A_8K$ MeOH 0.12 wt%	0.25	0.63	1.5	5.3	10	0.72
$\rm A_8 K$ DMF 0.25 wt $\%$	0.28	0.59	1.7	6.0	11	0.81
A <sub>10</sub> K H <sub>2</sub> O 1.00 wt%	0.30	0.50	2.5	9.4	17	
$A_{10}^{10}$ K MeOH 0.12 wt%	0.33	0.50	2.0	7.1	13	0.76
$\rm A_{10}K~DMF~0.12~wt\%$	0.31	0.49	2.3	8.3	15	0.89



Figure S1 SAXS profiles of  $A_8K$  (A) and  $A_{10}K$  (B) solutions in MeOH at different concentrations obtained by dilution (approximate concentrations are 1 wt% purple, 0.5 wt% red, 0.25 wt% orange, 0.125 wt% green, 0.0625 wt% blue)



Figure S2 Pair distance distribution functions of the cross section of a long rod obtained by applying the indirect Fourier inversion to the I(q)q experimental profiles of  $A_8K$  (A) and  $A_{10}K$  (C) fibrils in the solvents water (blue), MeOH (green) and DMF (purple). The distributions are normalized by the subtended area. Fits to the experimental data are shown in (B) ( $A_8K$ ) and (D) ( $A_{10}K$ ). Sample concentrations are 1 wt% for  $A_{10}K$  in all solvents and for  $A_8K$  in MeOH and DMF; the data for  $A_8K$  in water correspond to a sample with 3 wt% concentration.



**Figure S3** WAXS data of  $A_8K$  (A,B) and  $A_{10}K$  (C,D) in water (blue), MeOH (green) and DMF (purple) at concentrations around 1 wt%. The *q* values where Bragg peaks were previously observed are marked with vertical dashed lines. In (B) and (D) the raw data before background subtraction are shown to highlight the solvent signal, which is reported as black lines (almost overlapping).



**Figure S4** Estimate of the aggregate thickness from the superimposition of several profiles taken across the cross section in several points of the micrographs (red shadows).