



# **Immunogenicity and Efficacy of Vaccination in People Living** with Human Immunodeficiency Virus

Eeva Tortellini <sup>1,\*</sup>, Yann Collins Fosso Ngangue <sup>1</sup>, Federica Dominelli <sup>1</sup>, Mariasilvia Guardiani <sup>1</sup>, Carmen Falvino <sup>1</sup>, Fabio Mengoni <sup>1</sup>, Anna Carraro <sup>1</sup>, Raffaella Marocco <sup>2</sup>, Patrizia Pasculli <sup>1</sup>, Claudio Maria Mastroianni <sup>1</sup>, Maria Rosa Ciardi <sup>1</sup>, Miriam Lichtner <sup>2,3</sup> and Maria Antonella Zingaropoli <sup>1</sup>

- <sup>1</sup> Department of Public Health and Infectious Diseases, Sapienza University of Rome, 00185 Rome, Italy; yanncollins.fosso@uniroma1.it (Y.C.F.N.); federica.dominelli@uniroma1.it (F.D.); mariasilvia.guardiani@uniroma1.it (M.G.); carmen.falvino@uniroma1.it (C.F.); fabio.mengoni@uniroma1.it (F.M.); anna.carraro@uniroma1.it (A.C.); patrizia.pasculli@uniroma1.it (P.P.); claudio.mastroianni@uniroma1.it (C.M.M.); maria.ciardi@uniroma1.it (M.R.C.); mariaantonella.zingaropoli@uniroma1.it (M.A.Z.)
  - <sup>2</sup> Infectious Diseases Unit, SM Goretti Hospital, Sapienza University of Rome, 00185 Latina, Italy; raffaella.marocco@uniroma1.it (R.M.); miriam.lichtner@uniroma1.it (M.L.)
  - <sup>3</sup> Department of Neurosciences, Mental Health, and Sense Organs, NESMOS, Sapienza University of Rome, 00185 Rome, Italy
  - \* Correspondence: eeva.tortellini@uniroma1.it

**Abstract:** People living with HIV (PLWH) remain at high risk of mortality and morbidity from vaccine-preventable diseases, even though antiretroviral therapy (ART) has restored life expectancy and general well-being. When, which, and how many doses of vaccine should be administered over the lifetime of PLWH are questions that have become clinically relevant. Immune responses to most vaccines are known to be impaired in PLWH. Effective control of viremia with ART and restored CD4+ T-cell count are correlated with an improvement in responsiveness to routine vaccines. However, the presence of immune alterations, comorbidities and co-infections may alter it. In this article, we provide a comprehensive review of the literature on immune responses to different vaccines in the setting of HIV infection, emphasizing the potential effect of HIV-related factors and presence of comorbidities in modulating such responses. A better understanding of these issues will help guide vaccination and prevention strategies for PLWH.

Keywords: HIV; PLWH; ART; vaccination; immune responses; CD4; COVID-19; HPV; influenza

# 1. Introduction

In people living with HIV (PLWH), reduced immune responses to most vaccines are known [1,2]. Antiretroviral therapy (ART) restores life expectancy and general well-being, reducing the risk of severe outcomes after infection in PLWH [3,4].

HIV infection induces a profound disruption of both the innate and adaptive immune systems leading to immunological alterations and persistent immune dysfunction [5]. Primary infection elicits systemic immune activation and inflammation followed by a progressive loss in CD4+ T-cell count and a persistent expansion of circulating CD8+ T cells [6]. Furthermore, exhaustion of T cells often recurs, together with an alteration of the innate immune cell functions [6,7]. Indeed, alterations of B-cell activity such as abnormal activation and lower antibody responses have been described [8].

ART-induced suppression of HIV replication is associated with a significant increase in absolute CD4+ T-cell and B-cell counts, including naïve and memory cells that are essential for humoral and cellular immunity to T-cell-dependent and independent immunogens [9]. However, despite effective virological suppression, chronic activation persists and antigen-specific T- and B-cell responses, including T follicular helper cell (Tfh) functions, are still



Citation: Tortellini, E.; Fosso Ngangue, Y.C.; Dominelli, F.; Guardiani, M.; Falvino, C.; Mengoni, F.; Carraro, A.; Marocco, R.; Pasculli, P.; Mastroianni, C.M.; et al. Immunogenicity and Efficacy of Vaccination in People Living with Human Immunodeficiency Virus. *Viruses* **2023**, *15*, 1844. https:// doi.org/10.3390/v15091844

Academic Editors: Pietro Hiram Guzzi, Marianna Milano and Jayanta Kumar Das

Received: 27 July 2023 Revised: 17 August 2023 Accepted: 21 August 2023 Published: 30 August 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). impaired. Furthermore, PLWH continue to have higher levels of inflammatory mediators, such as interleukin (IL)-6, tumor necrosis factor-alpha (TNF-a), soluble (s) CD163, sCD14 and C-reactive protein (CRP)-accelerated aging, and some other comorbidities may accompany this therapy [10,11]. Some molecules with immunomodulatory properties have been shown to have some beneficial effects on this residual inflammation [12].

However, in PLWH, this impairment of the immune system may affects the quantity, quality and persistence of protective immune responses induced by natural infection or vaccination, reducing responsiveness to vaccines and their effectiveness [13–15]. In addition, vaccine-induced antibodies may decline more rapidly than in the general population [16].

As reported in Table 1, guidelines recommend a proactive approach for immunizing PLWH who are susceptible to vaccine-preventable infections and at risk of exposure, including those who have received previously contraindicated live attenuated vaccines such as those against measles, mumps and rubella [17]. On the contrary, the bacillus Calmette–Guérin (BCG) continues to be contraindicated in PLWH due to its unfavorable benefit/risk profile [18,19].

Pathogen	Vaccine Platform	Absolute CD4+ T Cell Count	
		<200	>200
HAV	Inactivated	2–3 doses (varies by formulation)	
HBV	Recombinant	2–4 doses (varies by formulation)	
HPV	Recombinant	3 doses through age 26	
Influenza	Inactivated (IIV) Recombinant (RIV) Live, attenuated (LAIV)	1 dose annually 1 dose annually not recommended	
MPXV	Live, attenuated	not recommended	2 doses
SARS-CoV-2	mRNA-based Viral vector Recombinant	2 doses + booster 2 doses 2 doses	
Streptococcus pneumoniae	PCV15 PCV20 PPSV23	1 dose PCV15 followed ≥8 weeks by 1 dose PPSV23 or 1 dose PCV20	
VZV	Live, attenuated (ZVL) Recombinant (RZV)	not recommended 2 doses for 18 and older	

Table 1. Recommended PLWH Immunization.

HAV: Hepatitis A virus, HBV: Hepatitis B virus, HPV: Human Papilloma virus, MPXV: Monkeypox virus, VZV: Varicella zoster virus, SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2, PCV: pneumococcal polysaccharide vaccine.

In general, the effective viremia control by ART and the improvement in the absolute CD4+ T-cell counts are correlated with an enhancement in responsiveness to routine vaccines, although this issue continues to be of concern. Together with current CD4+ T-cell absolute count, the CD4/CD8 ratio has proved to be an accurate predictor of vaccine success [20] (Figure 1).

Finally, co-infections represent an additional factor that may influence immune responses to vaccination, due to their contribution to a persistent immune activation state and induction of immune senescence [21,22].

Overall, vaccination of PLWH remains challenging and with the present review, we summarized recent works in the literature on different vaccine responses in the setting of HIV infection (Table 1).

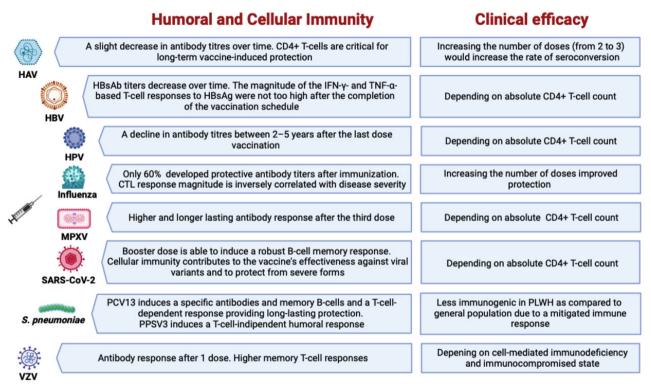


Figure 1. Summary of humoral and cellular immunity in PLWH to recommended vaccinations.

#### 2. Hepatitis A Virus Vaccination

Hepatitis A is a viral infection caused by the Hepatitis A virus (HAV), a single-stranded RNA virus from the *Picornaviridae* family [23]. HAV is commonly transmitted through the fecal–oral route (through ingestion of contaminated food or water), person-to-person contact and men who have sex with men (MSM) [24,25] and may be responsible for forms of acute hepatitis that may progress to fulminant hepatic failure in non-immune adult populations [26,27].

Hepatitis A occurs worldwide and is highly endemic in the most precarious areas of low-income countries [28,29]. It was estimated to have caused, in 2005, apart from approximately 200 million subclinical and oligo-symptomatic HAV infections, 33 million cases of symptomatic illness and 35,000 deaths [30,31].

In Europe, the HAV seroprevalence is low and observational data suggest that PLWH, especially MSM and injecting drug users (IDUs), are at increased risk of contracting HAV [24,32–34]. Additionally, a small study conducted on 15 PLWH with acute hepatitis A showed that the duration of HAV viremia was prolonged compared to the general population with acute hepatitis A, which may increase the likelihood of contracting HAV and transmission to others [35]. Overall, older age, IDUs and MSM have been identified as independent factors associated with HAV seropositivity in PLWH [36–39].

Among the preventive measures to reduce the spread of hepatitis A, vaccination against HAV remains the most effective [40,41]. Currently, two types of HAV vaccines are available: the live attenuated vaccine and the inactivated HAV vaccine [42]. Only the latter is recommended for PLWH [43] including different adult, adolescent and pediatric formulations [43].

In the literature, several studies have evaluated the effectiveness of vaccination against HAV in PLWH. A low rate of seroconversion compared to the general population was observed. Absolute CD4+ T-cell count < 200 cells/ $\mu$ L, viral load, old age, CD4/CD8 ratio, hepatitis C co-infection and gender were identified as factors of poor response after vaccination [44–47].

Several previous studies evaluating the efficacy of HAV vaccination in PLWH have shown lower seroconversion rates (vaccine efficacy) than their seronegative counterparts [48–53]. However, a recent prospective observational study in the setting of an epidemic of acute hepatitis A among MSM in Taiwan observed an overall seroconversion rate among PLWH MSM of 39.7% and 93.4% after receiving one dose and completing a two-dose series of HAV vaccination, respectively, and despite the delayed serological response, HAV vaccination resulted in a 93% reduction in the risk of acute HAV infection in HIV-positive MSM over the course of the epidemic [43]. Higher absolute CD4+ T-cell counts were consistently correlated with higher seroconversion rates [43].

In another study evaluating the efficacy of the vaccine against HAV in a group of 29 children, including 6 children living with HIV and having lost their HAV seropositivity 7 years after being vaccinated with an inactivated vaccine against HAV; after revaccination (two doses), 83% of these PLWH had a seroconversion after the first dose [54].

Other studies evaluating the efficacy of the HAV vaccine according to the increase in the number of vaccine doses have shown that increasing the number of doses from two to three increases the rate of seroconversion in PLWH [52,53,55–57].

Various studies evaluating the persistence of immune memory have demonstrated that in healthy adults following a primary two-dose regimen, anti-HAV antibodies can persist in >90% of vaccines for 40 years or more [58]. In PLWH, on the other hand, a slight decrease was observed over time. In other prospective studies investigating PLWH (all with an inactivated vaccine against HAV), better results of the persistence of immune memory were observed with a duration of seroprotection in PLWH patients equivalent to 7 years in 79% of 29 adolescents, 94 for 6–10 years in 85% of 116 adults, 117 for 3.7 years in 85% of 52 adults [59], and for 5 years in 75.5% of 49 adults [47].

Different studies assessing the persistence of immune memory have observed that in healthy adults following a primary two-dose regimen, anti-HAV antibodies can persist in >90% of vaccines for 40 years or more [58]. In prospective studies of successfully vaccinated PLWH (all with an inactivated HAV vaccine), there was a slight increase in the persistence of seroprotection, which in some of these PLWH (both adults and adolescents) oscillated between 5 and 10 years old [47,54,59,60].

The persistence of immune memory was also confirmed by a recent study comparing three-dose and two-dose HAV vaccination schedules, where a slightly higher seroprotection rate of 94% versus 88% was found after 5 years in 155 and 95 adults, respectively [61].

Despite the evidence of efficacy conferred by the HAV vaccine, PLWH remain susceptible to HAV infection in high-income countries, due to low compliance with recommended HAV vaccination guidelines, at-risk sexual behaviors and injecting drug use.

## 3. Hepatitis B Virus Vaccination

Hepatitis B is a liver viral infection caused by the Hepatitis B virus (HBV), a DNA virus belonging to the *Hepadnaviridae* family [62].

Because HIV and HBV share similar routes of transmission, co-infection with the two viruses is common [63,64]. HBV infection in PLWH is a global public health problem [65]. Globally, nearly 10% of PLWH are infected with HBV [66,67]. HBV infection in PLWH is generally characterized by an increased rate of cirrhosis (10–20%), a higher risk of hepatocellular carcinoma [63,68–70] and lastly a higher risk of liver-related death [70].

In general, HBV is not cytopathic. It causes damage through the induction of immune mechanisms. Cytotoxic CD8 cells recognize expressed HBV antigens and destroy infected hepatocytes, resulting in increased aminotransferases [71]. Thus, HBV establishes a persistent infection with a stable reservoir of genetic material in the form of circularized DNA in the cell nucleus [65]. The different phases of HBV infection are characterized by the presence of certain viral and immunological markers allowing the orientation of the therapeutic decision and the evaluation of the response to treatment. Surface antigen (HBsAg) is the first marker detected in serum [72]. Its presence indicates HBV infection [73]. The disappearance of HBsAg is followed by the appearance of anti-HBs antibodies. Anti-HBs is considered as a neutralizing antibody and is recognized as a marker of disease protection and cure [73]. In most patients, anti-HBs persists for life, confirming long-term immunity.

Anti-HBs is only serological marker in individuals who have an immune response after vaccination against hepatitis B [74].

HBV vaccination is recommended in PLWH as the most important method of prevention [75]. Despite these recommendations in 2015, only 2/3 of PLWH receive at least one dose of HBV vaccine [75]. The efficacy rate of this vaccine in terms of immune response is generally defined by seroconversion with anti-HBs antibodies > 10  $\mu$ L/mL in these subjects [76].

The first studies on the efficacy of the vaccine against HBV applying the "classical" schedule (20  $\mu$ g of HBs antigen at months 0–1–6) showed relatively low seroconversion rates in PLWH, with only 20–70% overprotected against 90–95% in the general population [77,78]. Nevertheless, in these studies, a low rate of response to vaccination could be correlated with various risk predictors of poor response, including viral load and absolute CD4+ T-cell count, CD4/CD8 ratio, co-infection with HCV, poor general health and occult hepatitis B [79–83]. In addition, the female sex [84,85], younger age [86–88], alcohol consumption [86] and smoking [88] are all factors of negative response to vaccination.

Recent studies have shown improvements in the effectiveness of the HBV vaccine in PLWH, particularly with seroconversion rates [89–91]. In Uganda, to assess the efficacy of HBV vaccines in PLWH, a cohort of 132 participants received both the base dose of the vaccine and the 1-month dose, and 127 received the 6-month dose. The 132 participants who entered the study were predominantly female and 52% had received ART for  $\geq$ 3 months and 94% had undetectable HIV RNA. The median (IQR) CD4+ T-cell count was 426 cells/µL (261–583). A high humoral response rate in PLWH was seen. Nevertheless, in this study, a variation was observed in the immune response of these PLWH to the vaccine, with 86% of participants that were high-level responders with anti-HBs titer levels  $\geq$  100 IU/L, while 6% were low-level responders (anti-HBs levels 10 to 99 IU/L) and 10 (8%) were non-responders (titer levels < 10 IU/L) [89].

This high response rate differs from some studies that have shown suboptimal seroconversion rates in response to the standard series of HBV vaccines in PLWH. These results are like a study in China that also found high response rates to the HBV vaccine in PLWH. Nevertheless, compared to the general population in China [90], the response to HB vaccination was diminished in PLWH.

In a study in Thailand, high rates of response to the HBV vaccine were also observed in PLWH with fully suppressed HIV viral load and absolute CD4+ T-cell count  $\geq 200$  cells/µL [88].

Given the importance of immune status in vaccine response, it is possible that participants' degree of immune reconstitution differed between studies, despite apparently similar current absolute CD4+ T-cell counts documented in these studies. On the other hand, in the literature, we find results of the efficacy of the vaccine against HBV, correlated with the increase in vaccination schedules against HBV. Launay et al. [92], in one study, found that PLWH vaccinated with a four-double-dose schedule had higher anti-HBV titers and stronger immune responses than those vaccinated with the standard three-dose schedule (82% versus 65%, p < 0.05). Chaiklang et al. compared the immunogenicity and safety of three standard doses and four double doses versus four standard doses in one RCT [88]. A large randomized trial evaluating three-dose 20  $\mu$ g and four-dose 40  $\mu$ g HB vaccination regimens in PLWH reported seroconversion rates of 65 and 82%, respectively [92]. Prospective studies and randomized trials have reported similar response rates of 50–62% [93,94], while other studies have reported similar or better rates, ranging from 84 to 92% [88,89,95]. However, these studies with higher seroconversion rates exclusively enrolled patients with absolute CD4+ T-cell counts of 200 cells/µL or higher or included patients regardless of whether they had received an HBV vaccine in the past.

In another study evaluating the persistence of vaccination, O'Bryan et al. [96] followed 186 HIV patients for 5 years and found that the persistence of the HBV vaccine response was longer when these patients had an undetectable or low viral load [97].

These data demonstrate an improvement in the efficacy conferred by the HBV vaccine, but achieving a long-lasting and protective level of immunity remains a challenge in patients with detectable HIV RNA or low CD4+ T-cell counts at the time of vaccination.

# 4. Human Papillomavirus Vaccination

Human papillomavirus (HPV) infection represents the most prevalent sexually transmitted infection in the world [98]. HPV is a small, non-enveloped DNA virus infecting skin or mucosal cells that belongs to the *Papillomaviridae* family. The genome encodes for six early proteins responsible for virus replication and two late proteins, L1 and L2, which are the viral structural proteins [99]. At least 13 of more than 100 known HPV genotypes can cause cancer of the cervix and are associated with other anogenital cancers and cancers of the head and neck [99]. HPV types are divided into high risk, associated with the development of anogenital cancer, and low risk, rarely associated with the development of cancers [100]. The two most common "high-risk" genotypes (HPV 16 and 18) cause approximately 70% of all cervical cancers [99].

In immunocompetent individuals, most HPV infections spontaneously resolve; however, the persistent infection with oncogenic HPV genotypes is associated with cancers of the cervix, vulva, vagina, anus, penis and the oropharynx [100]. In immunosuppressed individuals, including PLWH, HPV infection often becomes chronic [101]. In particular, PLWH have a higher incidence of HPV infection, abnormal pap smears and persistent HPV infection due to the less efficient viral clearance, leading to a high risk of HPV-related cancers [101].

Because of the more common and persistent HPV-related complications in PLWH, HPV vaccination programs are encouraged in this population. HPV vaccination represents the main preventive tool for HPV-related cancers as well as other HPV-related diseases [102]. HPV vaccines are based on the recombinant protein virus-like particle (L1 VLP) with a proprietary adjuvant. Currently, there are three licensed prophylactic L1 VLP-based vaccines that provide protection against two (bivalent, commercialized in 2008), four (quadrivalent) and nine (nonavalent) HPV genotypes [103]. Given the higher vulnerability of PLWH, particularly in men who have sex with men (MSM) who are also PLWH, to acquire multisite infections mostly characterized by various genotype combinations and the ability of the nonavalent vaccine to prevent 80% of HPV infections, vaccination programs with this nine-genotype protection should be implemented, especially among MSM [104,105].

HPV vaccines have proved their safety, efficacy and effectiveness in immunocompetent young persons, leading to a standard vaccination regime for young girls and boys (aged 9–14 years) reduced from the originally licensed three-dose regimen to two doses [106,107].

Serum neutralizing antibodies are thought to be the major protective branch of adaptative immunity afforded by L1 VLP-based vaccines, although CD4+ T cells are involved in the induction and re(activation) of antigen-specific memory B cells, leading to high antibody levels and, therefore, are critical for long-term vaccine-induced protection [108].

The immune response elicited by the quadrivalent HPV vaccine seems to persist in vaccinated individuals up to 5 years post-vaccination [109].

Concerning PLWH, licensed HPV vaccines have proven to be generally safe and well tolerated [110]. Lower rates of antibody levels elicited by similar vaccine constructs have been observed, raising concern about the efficacy of HPV vaccines in PLWH [111]. However, several studies have reported that the immune response induced in PLWH is similar to that found in general population, with high rates of seroconversion and a cellular immunogenicity comparable to that of general population [98,110,112–114]. Furthermore, antibody levels following vaccination appear to be stable over time [115].

Significant positive correlations between T-cell responses and current absolute CD4+ T-cell count together with negative correlations between such responses and HIV viremia have been observed [116,117]. Furthermore, higher seroconversion rates among PLWH with current absolute CD4+ T-cell counts >200 cells/ $\mu$ L compared with  $\leq$ 200 cells/ $\mu$ L have also been reported [116,117]. However, a possible decline in B-cell memory responses between 2 and 5 years after the last vaccination dose has been described in PLWH [116]. This is consistent with observations focused on the characterization of the immunogenicity of other vaccines, but few immunogenicity studies of HPV vaccines among PLWH include participants with low absolute CD4+ T-cell counts, supporting the need to further elucidate their immune capacity. In addition, to the best of our knowledge, few studies are focused on the immunogenicity of the nonavalent HPV vaccine, and information is lacking about the quality of such response.

#### 5. Influenza Vaccination

Influenza is an infectious respiratory disease with annual estimations of approximately 1 billion infections, 3–5 million cases of severe illness and 300,000–500,000 deaths, according to WHO [118]. Influenza viruses belong to the *Orthomyxoviridae* family and account for three different types: influenza A, B and C. All three types share common characteristics, such as the segmented genome, made of negative-sense single-stranded RNA, and the presence of an envelope (derived from host cell membrane) with glycoproteins, essential for viral entry in target cells [119]. Viral particles are enveloped, and surface glycoproteins hemagglutinin (HA) and neuraminidase (NA) represent the major antigenic determinants [120]. Influenza viruses are characterized by antigenic variation, based on two different mechanisms: antigenic drift (present in all influenza types) and antigenic shift (characteristic of influenza A only) [120].

Influenza B and C have a narrower host range (humans only and humans and swine, respectively) than influenza A, which can infect humans, swine, equine, other mammals such as ferrets, felids, mink, dogs, civets, marine mammals and avians [119]. Influenza A and B viruses are most relevant clinically, since they cause severe respiratory infections in humans [121]. Sequencing has confirmed that these viruses share a common genetic ancestry. They have genetically diverged, and an exchange of viral RNA segments between viruses has been reported occurring within each genus or type, but not across types [120]. Influenza A viruses are further characterized by the subtype of their surface glycoproteins, the HA and the NA. There are 18 different HA subtypes and 11 different NA subtypes [122]. Unlike influenza A, influenza B is not further divided into subtypes [123].

During infection, the epithelial cells are the primary targets for influenza viruses. These cells line the respiratory tract and initiate an antiviral immune response upon influenza virus detection. The first line of defense is represented by the innate immune system and constituted by physical barriers and the innate immune cellular responses [124]. A critical role is performed by the adaptive immunity in the clearance of viral pathogens during the later stages of infection. Furthermore, respiratory mucosal immunity is induced in the related mucosal tissues during influenza infection and involved in antiviral defense [125].

Generally, infections occur in children, although most of the severe cases involve very young or elderly individuals and individuals affected by chronic pulmonary or cardiac conditions, diabetes mellitus or immunocompromising conditions [126]. Considering the periodical recurrence of influenza infection and the severe complications occurring in the elderly and in patients with concomitant chronic diseases, the influenza vaccine represents an essential tool for preventing infection and limiting the burden of the disease. The choice of relevant antigens remains of paramount importance in developing influenza vaccines which are formulated every year to match the circulating strains.

Currently, there are three kinds of vaccines, inactivated, live attenuated and recombinant HA vaccines, licensed in different countries [127]. The WHO recommends seasonal influenza vaccination to children 6 months to 5 years of age, elderly individuals (>65), all persons with chronic medical conditions and pregnant women. All available influenza virus vaccines are injected intramuscularly, except for the live attenuated influenza virus vaccines, which are administered intranasally [128].

Debate is still ongoing on the efficacy and effectiveness of the licensed vaccines, although most studies find a positive effect of vaccination on vaccinated individuals [129]. The effectiveness of influenza vaccines has been found to be related predominantly to the

age and immune competence of the vaccinated individual and the antigenic relatedness of vaccine strains to circulating strains [130].

Currently licensed influenza vaccines focus on the production of antibodies against the viral HA, which binds host receptors to mediate viral entry, neutralizing the virus and preventing the infection [131]. However, the decline in vaccine-specific antibodies and the antigenic drift of influenza viruses over time leads to the necessity of annual revaccination. Targeting T-cell responses seems to be a promising technique to ameliorate influenza vaccines, although it does not prevent infection, but it can reduce the severity of the infection [131].

Indeed, the role of T-cell immunity was demonstrated during the 2001 H1N1 pandemic, where the magnitude of the pre-existing cytotoxic T-lymphocyte (CTL) response inversely correlated with disease severity in individuals without detectable neutralizing antibody [132].

PLWH experience prolonged duration of influenza infection and increased severity of illness, together with higher rates of hospitalizations compared to the general population [133]. As a result, annual vaccination against seasonal influenza is recommended by many national immunization guidelines [134].

Again, the success of influenza vaccination is related to current absolute CD4+ T-cell count [135]. Indeed, weaker response rates were observed in PLWH with lower current absolute CD4+ T-cell count, probably due to impaired function of the peripheral blood Tfh and B-cell functions [136,137].

However, the data are not always in accordance, as demonstrated by the study conducted by Tebas et al. [138], aimed at evaluating safety and immunogenicity of the H1N1 2009 vaccine in PLWH. The authors showed that only 60% of the participants developed protective antibody titers after immunization [138]. On the contrary, a clinical trial (P1088) launched by the International Maternal Pediatric and Adolescent Clinical Trials (IMPAACT) Network evaluated safety and efficacy of a monovalent pandemic H1N1 (pH1N1) vaccine in perinatally HIV-1-infected children and adolescents, showing that two doses of doublestrength pH1N1 vaccine are safe and immunogenic and may provide improved protection against influenza in this population [139].

Concerns remain about the efficacy in elderly PLWH, as observed in the general population. Alternative vaccines, dosing, adjuvants or schedule strategies may be needed to achieve effective immunization of this vulnerable population.

#### 6. Monkeypox Virus Vaccination

Monkeypox virus infection (MPXV), also commonly known as "monkey pox" or simian orthopoxvirus, is an infectious disease caused by an *Orthopoxvirus* (family *Poxviridae*). There are two genetically distinct MPXV clades that exhibit different lethality rates. Clade II comprising the first cases of infections was reported in West Africa and clade I in Central Africa. Clade IIb was responsible for the global epidemic outbreak in May 2022. Monkeypox virus infection presents a clinical picture that can vary according to the clades: clades I and IIa resemble smallpox and clade IIb is characterized by atypical presentations. During the 2022 epidemic which raged in countries where the disease was not endemic, the symptoms were very polymorphic (cutaneous and mucous membrane involvement, painful lymphadenopathy, angina, anitis or proctitis, etc.) which could lead to more complicated forms (ocular involvement, encephalitis or encephalopathies, multiorgan involvement in otherwise immunocompromised patients). Transmission to humans occurs from an animal reservoir or from human to human via direct or indirect physical contact (contaminated objects).

Despite the announcement of the end of the MPXV epidemic by the WHO in May 2023 [140], questions around MPXV remain topical in the scientific community, especially in immunocompromised subjects such as PLWH who are considered as a population at risk [141]. Nearly 957 cases of monkeypox virus (MPXV) infection in Italy and 25,887 cases of infection in Europe have been reported so far [142].

Recent studies have described the fatal nature of the MPXV infection in a subpopulation of these PLWH characterized by absolute CD4+ T-cell counts of >200 cells/ $\mu$ L, presenting a clinical picture marked by massive necrotizing skin and cutaneous, genital and non-genital mucosal lesions, which can sometimes be accompanied by pulmonary involvement with multifocal nodular opacities or respiratory failure and severe cutaneous and blood bacterial sequelae which, in 15% of cases, led to death [141,143,144].

Most cases of MPXV infections reported in Europe and North America since May 2022 were mainly transmitted among men who have sex with men (MSM) with evidence of an increased prevalence of HIV and other sexually transmitted infections (STIs). Given the morbidity and lethality in PLWH, a strong evolution of the therapeutic arsenal against MPXV with many vaccines has been made available to stem the epidemic [145].

The third-generation vaccine contains the live modified attenuated virus of vaccinia Ankara. MVA-BN is currently the only approved vaccine in areas where sufficient vaccine stocks are available. Jynneos/MVA-BN is used for pre-exposure prophylaxis to MPXV in HIV-infected individuals; it is also indicated for the prevention of MPXV in individuals 18 years of age and older who are at high risk of infection [146,147].

To date, there are very few data on an immune response of the MVA-BN vaccine against MPXV in PLWH. In a study that focused on evaluating the safety and immunogenicity of MVA-BN in immunocompromised subjects [148], a phase II trial was conducted between 2006 and 2009 in the United States and Puerto Rico; a total of 579 volunteers were recruited into the study: 439 vaccine-naïve subjects (88 immunocompetent subjects, 351 PLWH) and 140 vaccine-experienced subjects (9 immunocompetent subjects, 131 PLWH) received at least one vaccination. The results of this study demonstrated that the MVA-BN vaccine presents a better safety and tolerance profile in PLWH with absolute CD4+T-cell counts < 200 cells/ $\mu$ L than in immunocompetent subjects, regardless of their previous smallpox vaccination status. In other subsequent studies, the safety profile of the MVA-BN vaccine in immunocompromised subjects, particularly those infected with HIV, was comparable or even better in terms of local reactions than in subjects not infected with HIV [149,150].

The results of the studies showed that the safety profile of the MVA-BN vaccine in immunocompromised subjects, particularly PLWH, considered at risk for conventional smallpox vaccination, was comparable or even better in terms of local reactions than in the general population [148,150]. Antibody responses were also comparable between immunocompetent subjects and PLWH.

## 7. SARS-CoV-2 Vaccination

With more than 757 million confirmed cases, the coronavirus disease 2019 (COVID-19), caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), is the third coronavirus disease in the past 20 years [151,152].

SARS-CoV-2 is an enveloped, positive-sense, single-stranded RNA virus that belongs to the *Coronaviridae* family [153]. Its genome encodes for four major structural proteins, namely the spike surface glycoprotein (S), which is responsible for the binding to the host receptor *angiotensin-converting enzyme* 2 (ACE2), the small envelope protein (E), the matrix protein (M) and the nucleocapsid protein (N), and other non-structural proteins [154]. Viral transmission can occur by direct, indirect or close contact by infected people through secretions (saliva or respiratory droplets) [155]. SARS-CoV-2 infects bronchial epithelial cells, pneumocytes and upper respiratory tract cells in humans, developing into severe, life-threatening respiratory diseases and lung injuries [156].

Many countries have launched vaccination campaigns to prevent SARS-CoV-2 infection, and several vaccines have been approved by the World Health Organization (WHO), although many obstacles to global vaccination remain [157]. Among the vaccines based on different technologies that have been developed during the health emergency, messenger RNA (mRNA)-based vaccines have been widely used to contain the pandemic [158]. In the general population, mRNA-based vaccines have proven to elicit a robust and protective humoral and cellular response against the SARS-CoV-2 S protein, reducing mortality and morbidity related to SARS-CoV-2 infection [159–161]. In addition, the specific T-cell response induced by the vaccine, with the ability to recognize different regions of the S protein, contributes to the vaccine's effectiveness against viral variants and protects individuals from severe forms of COVID-19 [162,163].

It has been estimated that PLWH represent 1% of total hospitalized cases and, differently from HIV infection which, in the absence of ART, is invariably fatal, COVID-19 disease is highly variable, ranging from mild to severe and critical forms of illness [164,165].

During the health emergency, vaccination of PLWH became of vital significance and strongly recommended by health authorities because of the potentially worse outcomes after SARS-CoV-2 infection, although reports about the increased risk of severe COVID-19 in this population are in some cases contradictory [166]. However, PLWH may experience a higher burden of various comorbidities, many of which have emerged as risk factors for severe COVID-19. Key risk factors for severe COVID-19 include both non-HIV comorbidities known to be associated with severe disease like older age, diabetes, obesity and cardiovascular disease as well as HIV-specific risk factors such as low absolute CD4+T-cell count, viremia and *Mycobacterium tuberculosis* co-infection [167]. Furthermore, the suboptimal responses to other vaccines have raised concerns about the efficacy of vaccines against SARS-CoV-2 in this potentially more vulnerable population.

Some HIV viral blips following mRNA vaccinations have been reported; however, licensed vaccines have proven to be safe and efficacious in PLWH with stable absolute CD4+ T-cell counts and well-controlled viremia [168].

In particular, published data on the immunogenicity of mRNA vaccines show values of anti-S antibodies, neutralizing antibody activity and cellular immune responses in PLWH on ART and with current absolute CD4+ T-cell counts above 200 cells/ $\mu$ L comparable to those observed in the general population after a primary vaccination cycle [168–170]. Such a response was found to be significantly inferior in PLWH with current absolute CD4+ T-cell count < 200 cells/ $\mu$ L compared to those with >500 cell/mm<sup>3</sup> and the general population, suggesting that the immunogenicity at the time of vaccination is related to the current absolute CD4+ T-cell count < CD4+ T-cell count [171,172].

In September 2021, the administration of an additional booster of anti-SARS-CoV-2 mRNA vaccine was approved in Italy to be given after >28 days after completion of the primary vaccination cycle in PLWH depending on current absolute CD4+ T-cell count and/or detectable HIV viremia [173]. The third dose improved the responsiveness particularly in PLWH on ART with current absolute CD4+ T-cell counts < 200 cells/ $\mu$ L, improving both the rate and the magnitude of the response and supporting the additional dose strategy in this category of patients with severe immune impairment [172,174].

Some strategies aimed at increasing the tolerability of the mRNA vaccine, such as the use of pidotimod, which was able to reduce vaccination-related adverse events, could be useful to encourage people to received vaccination [175].

In PLWH with a current absolute CD4+ T-cell count > 200 cells/ $\mu$ L, T-cell response elicited by the third dose was like that induced by the primary vaccination cycle, suggesting that the first two doses were able to achieve full T-cell immunization. Furthermore, the increased humoral response is consistent with the hypothesis that the third dose is able to induce a robust B-cell memory response, previously elicited by the primary vaccination series [176].

However, questions remain about mRNA vaccine's immunogenicity in PLWH with ongoing immunosuppression and viremia who represent a particularly vulnerable group that is poorly represented in vaccine trials. Furthermore, recent studies have shown a lower polyfunctional capacity in this population, as already described in the setting of other co-infections, raising issues about the real capability of their immune response [174,177].

#### 8. Streptococcus Pneumoniae Vaccination

*Streptococcus pneumoniae* (*S. pneumoniae*), a Gram-positive bacterium, is the most significant cause of bacterial disease in humans. A variety of clinical syndromes are related to its infection, including pneumonia, meningitis, bacteremia, acute otitis media and sinusitis [178]. Despite the availability of a broad arsenal of antibiotics and a vaccine, worldwide, approximately 14.5 million cases of serious pneumococcal diseases per year have been reported, leading to approximately 826,000 deaths [179].

In PLWH, invasive pneumococcal disease (IPD) and pneumococcal pneumonia continue to pose a challenge with high recurrence rates [180], significant public health impact, morbidity and a high mortality rate of up to 25% [181].

The introduction of pneumococcal vaccines has significantly reduced morbidity, although PLWH still remain at a 30-times-higher risk of IPD as compared to the general population [182]. Specifically, an absolute CD4+ T-cell count < 200 cells/ $\mu$ L and high levels of HIV RNA have been strongly associated with the risk of IPD [182].

Four pneumococcal vaccines are currently available: PCV13, PPSV23, PCV15 and PCV20 [183]. The PCV13 contains protein-conjugated polysaccharides of 13 serotypes of pneumococci [184]. The PCV15 and PCV20 contain all the PCV13 serotypes, with two additional serotypes in PCV15 and seven additional serotypes in PCV20 [183]. The PPSV23 contains 23-valent pneumococcal polysaccharides [185]. The PCV15 is administered as a single dose with one PPSV23 follow-up dose given at least 8 weeks later; no additional doses are recommended after that. The PCV20 requires one dose only; there are no additional doses needed [183].

There are limited data on the efficacy of pneumococcal vaccination in PLWH. Retrospective studies indicate that PPSV23 alone has modest clinical benefit, if any, in reducing rates of pneumococcal infections [186,187]. The immune response induced by PPSV23 is a T-cell-independent humoral response, while PCV13 induces a T-cell-dependent response producing pneumococcal serotype-specific antibodies and memory B cells which provide long-lasting protection [188]. Sequential vaccination with PCV 13 followed by PPSV23 provides a prime boost effect on inducing and maintaining protective immunity [189].

These vaccines are less immunogenic in PLWH compared to the general population due to a mitigated immune response. The combination of PPSV23 and PCV 13 has been shown to be more immunogenic than either of the vaccines alone and is recommended internationally for prevention of IPD in PLWH [190]. Moreover, other strategies to improve the immunogenicity of pneumococcal vaccines in PLWH were performed. Indeed, in a double-blind, placebo-controlled study, the addition of adjuvant CPG 7909, a toll-like receptor agonist, significantly enhanced the proportion of high responders to the vaccine [191].

According to the Italian Vaccination Plan (2017–2019), the tetanus, diphtheria and pertussis (Tdap) vaccine is co-administered with PCV and HBV vaccines. For PLWH of  $\geq$ 11 years old who have never received any vaccine, three doses of Tdap are administered, with an interval of 0, 1 month, 6–12 months. In individuals with advanced-stage HIV, the response is suboptimal for both tetanus and diphtheria, while in subjects with CD4 > 300 cells/µL, the response against tetanus is optimal, comparable to subjects without HIV while, for diphtheria, it can remain markedly lower [192].

# 9. Varicella Zoster Virus Vaccination

The Varicella zoster virus (VZV), a double-stranded DNA ubiquitous human alphaherpesvirus [193], causes varicella, establishes lifelong latency in ganglionic neurons and reactivates later in life to cause herpes zoster, commonly associated with chronic pain [193–196].

Varicella and herpes zoster are more common and more severe in the elderly, the female sex [197] and in people who are immunocompromised, such as PLWH and people taking immunosuppressive drugs and chemotherapy [198]. The incidence of herpes zoster is more than 15 times higher in PLWH compared to age-matched immunocompetent subjects. Herpes zoster can occur in PLWH at any absolute CD4+ T-cell count, but disease frequency is highest when absolute CD4+ T-cell counts are below 200 cells/ $\mu$ L [199–201].

Despite the mandatory vaccination of children against chickenpox in the early 1995s in the United States [202] and from 2003 in Europe [203] (this led to immunization (83% to 95%) of the general population [204]), the risk of VZV reactivation remains particularly high in seropositive adults [205,206]. The incidence of herpes zoster is approximately 4–7 cases/1000 person-years and without vaccination, the lifetime herpes zoster risk is 20–30% [207,208].

Vaccination offers an option that could overcome the challenges associated with conventional antiviral prophylaxis while potentially providing longer-lasting protection against shingles [209]. The live zoster vaccine (ZVL) is a live attenuated vaccine approved for people aged  $\geq 60$  years [210]. The effectiveness of the ZVL vaccine may decrease with age and it is generally contraindicated in immunocompromised subjects due to its potential infection risks [211,212].

Recently, the recombinant zoster vaccine (RZV), an adjuvanted subunit vaccine recommended for use in adults  $\geq$  50 years of age since 2017 by the Advisory Committee on Immunization Practices (ACIP) [213], was also approved by the ACIP for the prevention of herpes zoster in adults aged 19 and older who have or will have an increased risk of shingles due to immunodeficiency or immunosuppression caused by diseases or treatment [210].

The efficacy of RZV in immunocompromised subjects is lower than in immunocompetent subjects, reflecting cell-mediated immunodeficiency and a weaker immune response due to an underlying immunocompromised state [214]. Clinical trial data comparing memory T-cell responses to both vaccines mentioned (ZVL and RZV) found higher responses in RZV recipients, and only RZV recipients had five-year persistence of higher responses [215,216]. The efficacy of RZV is high, even in people aged  $\geq$ 70 years [217]. Pooled analyses also showed that the vaccine was 91.3% effective against shingles in participants over the age of 70 [218,219]. The clinical efficacy of the RZV vaccine has also been demonstrated in various phase II and III, placebo-controlled, observer-blinded studies conducted in immunocompromised adults aged 18 years and older with two doses administered 1–2 months apart [220,221].

Regarding the safety of the RZV vaccine, results from an observational study showed no difference between immunocompetent and immunocompromised groups, indicating that immunosuppression may not be a determinant of adverse vaccine effects [221].

These preliminary data confirm the efficacy conferred by the RZV vaccine against herpes zoster. However, these data should be interpreted with caution and require indepth studies.

#### 10. Conclusions

Despite ART-induced virologic suppression, PLWH remain at increased risk of mortality and morbidity from vaccine-preventable diseases, in part because of persistent immunopathology, resulting in a compromised response to vaccination, and vaccine-induced antibodies may fade more rapidly in PLWH than in the general population [1,16].

Moreover, besides the primary response, long-term persistence of protection has been poorly documented and recommendations on the timing of booster injections are based on data collected in the general population, although patterns of antibody decay may differ. In this regard, it is necessary to estimate how seroprotection declines over time among patients who initially responded to immunization.

Many efforts have been made during the SARS-CoV-2 pandemic to evaluate vaccine efficacy in PLWH. The findings obtained further confirm the critical role of CD4+ T cells as a key factor of effective humoral responses and predictor of vaccine success. In addition, evidence about the complementary role of T cell-specific responses in mediating protection has emerged, particularly in individuals with low seroconversion rates, reducing mortality and morbidity related to SARS-CoV-2 infection. However, what constitutes protective immunity is still discussed, making it difficult to define protective efficacy of vaccines. In determining vaccine scheduling and efficacy, CD4+ T-cell count, CD4/CD8 ratio and

viremia should be considered, with the awareness that it will not capture the full immune profile of this population.

In fact, it is becoming increasingly clear that PLWH represent a diverse population in terms of immune phenotype, with the consequence that different subgroups require different vaccination strategies to improve their immunological responses.

Furthermore, the setting of co-infection poses additional concerns, particularly regarding T-cell immunity, since with the intersecting of SARS-CoV-2, HIV and TB epidemics, SARS-CoV-2-specific CD4+ T cells have shown a lower polyfunctional capacity.

In our opinion, a better understanding of these issues will help guide vaccination and prevention strategies for PLWH.

We should also consider that male adults living in Europe and in the United States are the most represented participants in the studies, which poorly reflects the global prevalence of PLWH, and that, with the pandemic, a reduction in the access to ART and in vaccine coverage may leave PLWH potentially more vulnerable.

To date, studies assessing long-term immunogenicity, planned with scientific rigor, are needed. An improvement in the field of vaccine development could bring changes in the lives of PLWH. In conclusion, the main preventive tool for many infectious diseases remains vaccination, together with counseling and screening programs. However, greater attention needs to be paid to PLWH with uncontrolled viral infection and/or low CD4+ T-cell counts and to the effects of aging and comorbidities.

Author Contributions: Conceptualization, M.L. and M.A.Z.; resources, R.M., A.C., P.P., C.F., M.G., F.D., F.M., M.R.C. and C.M.M.; writing—original draft preparation, E.T., Y.C.F.N. and M.A.Z.; writing—review and editing, supervision, M.L. and M.A.Z.; project administration, M.L. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: We would like to acknowledge the Department of Public Health and Infectious Diseases of Sapienza University of Rome.

**Conflicts of Interest:** Miriam Lichtner received department grants from Gilead outside the submitted work and support from advisory boards for attending conferences from MSD, Abbvie, Gilead, GSK, Angelini and ViiV. All other authors have no conflicts of interest to declare.

#### References

- 1. Geretti, A.M.; Doyle, T. Immunization for HIV-Positive Individuals. Curr. Opin. Infect. Dis. 2010, 23, 32–38. [CrossRef]
- Abzug, M.J. Vaccination in the Immunocompromised Child: A Probe of Immune Reconstitution. *Pediatr. Infect. Dis. J.* 2009, 28, 233–236. [CrossRef] [PubMed]
- Samji, H.; Cescon, A.; Hogg, R.S.; Modur, S.P.; Althoff, K.N.; Buchacz, K.; Burchell, A.N.; Cohen, M.; Gebo, K.A.; Gill, M.J.; et al. Closing the Gap: Increases in Life Expectancy among Treated HIV-Positive Individuals in the United States and Canada. *PLoS* ONE 2013, 8, e81355. [CrossRef] [PubMed]
- Antiretroviral Therapy Cohort Collaboration. Life Expectancy of Individuals on Combination Antiretroviral Therapy in High-Income Countries: A Collaborative Analysis of 14 Cohort Studies. *Lancet* 2008, 372, 293–299. [CrossRef] [PubMed]
- Mullender, C.; da Costa, K.A.S.; Alrubayyi, A.; Pett, S.L.; Peppa, D. SARS-CoV-2 Immunity and Vaccine Strategies in People with HIV. Oxf. Open Immunol. 2022, 3, iqac005. [CrossRef] [PubMed]
- Fenwick, C.; Joo, V.; Jacquier, P.; Noto, A.; Banga, R.; Perreau, M.; Pantaleo, G. T-Cell Exhaustion in HIV Infection. *Immunol. Rev.* 2019, 292, 149–163. [CrossRef] [PubMed]
- Sabado, R.L.; O'Brien, M.; Subedi, A.; Qin, L.; Hu, N.; Taylor, E.; Dibben, O.; Stacey, A.; Fellay, J.; Shianna, K.V.; et al. Evidence of Dysregulation of Dendritic Cells in Primary HIV Infection. *Blood* 2010, 116, 3839–3852. [CrossRef]
- 8. Bussmann, B.M.; Reiche, S.; Bieniek, B.; Krznaric, I.; Ackermann, F.; Jassoy, C. Loss of HIV-Specific Memory B-Cells as a Potential Mechanism for the Dysfunction of the Humoral Immune Response against HIV. *Virology* **2010**, *397*, 7–13. [CrossRef]

- Moir, S.; Malaspina, A.; Ho, J.; Wang, W.; Dipoto, A.C.; O'Shea, M.A.; Roby, G.; Mican, J.M.; Kottilil, S.; Chun, T.-W.; et al. Normalization of B Cell Counts and Subpopulations after Antiretroviral Therapy in Chronic HIV Disease. *J. Infect. Dis.* 2008, 197, 572–579. [CrossRef]
- Guaraldi, G.; Orlando, G.; Zona, S.; Menozzi, M.; Carli, F.; Garlassi, E.; Berti, A.; Rossi, E.; Roverato, A.; Palella, F. Premature Age-Related Comorbidities among HIV-Infected Persons Compared with the General Population. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2011, 53, 1120–1126. [CrossRef]
- 11. Meir-Shafrir, K.; Pollack, S. Accelerated Aging in HIV Patients. Rambam Maimonides Med. J. 2012, 3, e0025. [CrossRef] [PubMed]
- 12. Ucciferri, C.; Falasca, K.; Reale, M.; Tamburro, M.; Auricchio, A.; Vignale, F.; Vecchiet, J. Pidotimod and Immunological Activation in Individuals Infected with HIV. *Curr. HIV Res.* **2021**, *19*, 260–268. [CrossRef] [PubMed]
- Pallikkuth, S.; De Armas, L.R.; Pahwa, R.; Rinaldi, S.; George, V.K.; Sanchez, C.M.; Pan, L.; Dickinson, G.; Rodriguez, A.; Fischl, M.; et al. Impact of Aging and HIV Infection on Serologic Response to Seasonal Influenza Vaccination. *AIDS* 2018, 32, 1085–1094. [CrossRef]
- 14. Bonetti, T.C.S.; Succi, R.C.M.; Weckx, L.Y.; Tavares-Lopes, L.; de Moraes-Pinto, M.I. Tetanus and Diphtheria Antibodies and Response to a Booster Dose in Brazilian HIV-1-Infected Women. *Vaccine* **2004**, *22*, 3707–3712. [CrossRef]
- Avelino-Silva, V.I.; Miyaji, K.T.; Hunt, P.W.; Huang, Y.; Simoes, M.; Lima, S.B.; Freire, M.S.; Caiaffa-Filho, H.H.; Hong, M.A.; Costa, D.A.; et al. CD4/CD8 Ratio and KT Ratio Predict Yellow Fever Vaccine Immunogenicity in HIV-Infected Patients. *PLoS Negl. Trop. Dis.* 2016, 10, e0005219. [CrossRef] [PubMed]
- Kernéis, S.; Launay, O.; Turbelin, C.; Batteux, F.; Hanslik, T.; Boëlle, P.-Y. Long-Term Immune Responses to Vaccination in HIV-Infected Patients: A Systematic Review and Meta-Analysis. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2014, 58, 1130–1139. [CrossRef]
- Geretti, A.M.; BHIVA Immunization Writing Committee; Brook, G.; Cameron, C.; Chadwick, D.; Heyderman, R.S.; MacMahon, E.; Pozniak, A.; Ramsay, M.; Schuhwerk, M. British HIV Association Guidelines for Immunization of HIV-Infected Adults 2008. *HIV Med.* 2008, 9, 795–848. [CrossRef] [PubMed]
- Mansoor, N.; Scriba, T.J.; de Kock, M.; Tameris, M.; Abel, B.; Keyser, A.; Little, F.; Soares, A.; Gelderbloem, S.; Mlenjeni, S.; et al. HIV-1 Infection in Infants Severely Impairs the Immune Response Induced by Bacille Calmette-Guérin Vaccine. *J. Infect. Dis.* 2009, 199, 982–990. [CrossRef]
- Hesseling, A.C.; Cotton, M.F.; Fordham von Reyn, C.; Graham, S.M.; Gie, R.P.; Hussey, G.D. Consensus Statement on the Revised World Health Organization Recommendations for BCG Vaccination in HIV-Infected Infants. *Int. J. Tuberc. Lung Dis. Off. J. Int. Union Tuberc. Lung Dis.* 2008, 12, 1376–1379.
- Hadrup, S.R.; Strindhall, J.; Køllgaard, T.; Seremet, T.; Johansson, B.; Pawelec, G.; thor Straten, P.; Wikby, A. Longitudinal Studies of Clonally Expanded CD8 T Cells Reveal a Repertoire Shrinkage Predicting Mortality and an Increased Number of Dysfunctional Cytomegalovirus-Specific T Cells in the Very Elderly. J. Immunol. 2006, 176, 2645–2653. [CrossRef]
- Perello, R.; Vergara, A.; Monclus, E.; Jimenez, S.; Montero, M.; Saubi, N.; Moreno, A.; Eto, Y.; Inciarte, A.; Mallolas, J.; et al. Cytomegalovirus Infection in HIV-Infected Patients in the Era of Combination Antiretroviral Therapy. *BMC Infect. Dis.* 2019, 19, 1030. [CrossRef]
- Lichtner, M.; Cicconi, P.; Vita, S.; Cozzi-Lepri, A.; Galli, M.; Lo Caputo, S.; Saracino, A.; De Luca, A.; Moioli, M.; Maggiolo, F.; et al. Cytomegalovirus Coinfection Is Associated with an Increased Risk of Severe Non–AIDS-Defining Events in a Large Cohort of HIV-Infected Patients. J. Infect. Dis. 2015, 211, 178–186. [CrossRef] [PubMed]
- Feinstone, S.M.; Kapikian, A.Z.; Purceli, R.H. Hepatitis A: Detection by Immune Electron Microscopy of a Viruslike Antigen Associated with Acute Illness. *Science* 1973, 182, 1026–1028. [CrossRef] [PubMed]
- 24. McFarlane, E.S.; Embil, J.A.; Manuel, F.R.; Thiébaux, H.J. Antibodies to Hepatitis A Antigen in Relation to the Number of Lifetime Sexual Partners in Patients Attending an STD Clinic. *Br. J. Vener. Dis.* **1981**, *57*, 58–61. [CrossRef]
- Corey, L.; Holmes, K.K. Sexual Transmission of Hepatitis A in Homosexual Men: Incidence and Mechanism. N. Engl. J. Med. 1980, 302, 435–438. [CrossRef] [PubMed]
- 26. Chen, C.-M.; Chen, S.C.-C.; Yang, H.-Y.; Yang, S.-T.; Wang, C.-M. Hospitalization and Mortality Due to Hepatitis A in Taiwan: A 15-Year Nationwide Cohort Study. *J. Viral Hepat.* **2016**, *23*, 940–945. [CrossRef] [PubMed]
- Boucher, A.; Meybeck, A.; Alidjinou, K.; Huleux, T.; Viget, N.; Baclet, V.; Valette, M.; Alcaraz, I.; Sauser, E.; Bocket, L.; et al. Clinical and Virological Features of Acute Hepatitis A during an Ongoing Outbreak among Men Who Have Sex with Men in the North of France. Sex. Transm. Infect. 2019, 95, 75–77. [CrossRef]
- Lin, K.-Y.; Sun, H.-Y.; Chen, Y.-H.; Lo, Y.-C.; Hsieh, S.-M.; Sheng, W.-H.; Chuang, Y.-C.; Pan, S.-C.; Cheng, A.; Hung, C.-C.; et al. Effect of a Hepatitis A Vaccination Campaign during a Hepatitis A Outbreak in Taiwan, 2015–2017: A Modeling Study. *Clin. Infect. Dis.* 2020, 70, 1742–1749. [CrossRef]
- 29. Aggarwal, R.; Goel, A. Hepatitis A: Epidemiology in Resource-Poor Countries. *Curr. Opin. Infect. Dis.* **2015**, *28*, 488–496. [CrossRef]
- Rein, D.B.; Stevens, G.; Flaxman, A.; Wittenborn, J.S.; Timothy, N.; Wiktor, S.Z.; Wiersma, S.T. P703 the Global Burden of Hepatitis a Virus in 1990 and 2005. J. Hepatol. 2014, 1, S303. [CrossRef]
- 31. WHO Position Paper on Hepatitis A Vaccines—June 2012. *Releve Epidemiol. Hebd.* **2012**, *87*, 261–276. Available online: https://www.who.int/publications-detail-redirect/who-wer8728-29-261-276 (accessed on 6 June 2012).

- Ochnio, J.J.; Patrick, D.; Ho, M.; Talling, D.N.; Dobson, S.R. Past Infection with Hepatitis A Virus among Vancouver Street Youth, Injection Drug Users and Men Who Have Sex with Men: Implications for Vaccination Programs. CMAJ Can. Med. Assoc. J. J. Assoc. Medicale Can. 2001, 165, 293–297.
- 33. Crofts, N.; Cooper, G.; Stewart, T.; Kiely, P.; Coghlan, P.; Hearne, P.; Hocking, J. Exposure to Hepatitis A Virus among Blood Donors, Injecting Drug Users and Prison Entrants in Victoria. *J. Viral Hepat.* **1997**, *4*, 333–338. [CrossRef]
- Corona, R.; Stroffolini, T.; Giglio, A.; Cotichini, R.; Tosti, M.E.; Prignano, G.; Di Carlo, A.; Maini, A.; Mele, A. Lack of Evidence for Increased Risk of Hepatitis A Infection in Homosexual Men. *Epidemiol. Infect.* 1999, 123, 89–93. [CrossRef] [PubMed]
- Ida, S.; Tachikawa, N.; Nakajima, A.; Daikoku, M.; Yano, M.; Kikuchi, Y.; Yasuoka, A.; Kimura, S.; Oka, S. Influence of Human Immunodeficiency Virus Type 1 Infection on Acute Hepatitis A Virus Infection. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2002, 34, 379–385. [CrossRef] [PubMed]
- Sun, H.-Y.; Kung, H.-C.; Ho, Y.-C.; Chien, Y.-F.; Chen, M.-Y.; Sheng, W.-H.; Hsieh, S.-M.; Wu, C.-H.; Liu, W.-C.; Hung, C.-C.; et al. Seroprevalence of Hepatitis A Virus Infection in Persons with HIV Infection in Taiwan: Implications for Hepatitis A Vaccination. *Int. J. Infect. Dis. IJID Off. Publ. Int. Soc. Infect. Dis.* 2009, 13, e199–e205. [CrossRef]
- Lee, H.-C.; Ko, N.-Y.; Lee, N.-Y.; Chang, C.-M.; Ko, W.-C. Seroprevalence of Viral Hepatitis and Sexually Transmitted Disease among Adults with Recently Diagnosed HIV Infection in Southern Taiwan, 2000-2005: Upsurge in Hepatitis C Virus Infections among Injection Drug Users. J. Formos. Med. Assoc. 2008, 107, 404–411. [CrossRef]
- Kourkounti, S.; Paparizos, V.; Leuow, K.; Kyriakis, K.; Antoniou, C. Prevalence and Titre of Antibodies against Hepatitis A Virus in HIV-Infected Men Having Sex with Men in Greece. *Infez. Med.* 2014, 22, 206–212.
- Linkins, R.W.; Chonwattana, W.; Holtz, T.H.; Wasinrapee, P.; Chaikummao, S.; Varangrat, A.; Tongtoyai, J.; Mock, P.A.; Curlin, M.E.; Sirivongrangson, P.; et al. Hepatitis A and Hepatitis B Infection Prevalence and Associated Risk Factors in Men Who Have Sex with Men, Bangkok, 2006–2008. J. Med. Virol. 2013, 85, 1499–1505. [CrossRef]
- NIH. Immunizations for Preventable Diseases in Adults and Adolescents with HIV. Available online: https://clinicalinfo .hiv.gov/en/guidelines/hiv-clinical-guidelines-adult-and-adolescent-opportunistic-infections/immunizations (accessed on 10 June 2023).
- Doshani, M.; Weng, M.; Moore, K.L.; Romero, J.R.; Nelson, N.P. Recommendations of the Advisory Committee on Immunization Practices for Use of Hepatitis A Vaccine for Persons Experiencing Homelessness. *MMWR Morb. Mortal. Wkly. Rep.* 2019, 68, 153–156. [CrossRef]
- 42. Infection Par Le Virus de l'hépatite A: Traitement et Prévention—UpToDate. Available online: https://www.uptodate.com/conte nts/hepatitis-a-virus-infection-treatment-and-prevention (accessed on 11 June 2023).
- Lin, K.-Y.; Chen, G.-J.; Lee, Y.-L.; Huang, Y.-C.; Cheng, A.; Sun, H.-Y.; Chang, S.-Y.; Liu, C.-E.; Hung, C.-C. Hepatitis A Virus Infection and Hepatitis A Vaccination in Human Immunodeficiency Virus-Positive Patients: A Review. *World J. Gastroenterol.* 2017, 23, 3589–3606. [CrossRef]
- 44. Zimmermann, P.; Curtis, N. Factors That Influence the Immune Response to Vaccination. *Clin. Microbiol. Rev.* **2019**, *32*, e00084-18. [CrossRef] [PubMed]
- Neukam, K.; Fernández, M.D.; Quero, J.H.; Rivero-Juárez, A.; Llaves-Flores, S.; Oñate, F.J.; Gutiérrez-Valencia, A.; Espinosa, N.; Viciana, P.; López-Cortés, L.-F. Brief report: Response to hepatitis a virus vaccine in HIV-infected patients within a retrospective, multicentric cohort: Facing hepatitis a outbreaks in the clinical practice. *JAIDS J. Acquir. Immune Defic. Syndr.* 2019, 81, e1–e5. [CrossRef] [PubMed]
- 46. Fritzsche, C.; Bergmann, L.; Loebermann, M.; Glass, A.; Reisinger, E.C. Immune Response to Hepatitis A Vaccine in Patients with HIV. *Vaccine* **2019**, *37*, 2278–2283. [CrossRef]
- Jabłonowska, E.; Kuydowicz, J. Durability of Response to Vaccination against Viral Hepatitis A in HIV-Infected Patients: A 5-Year Observation. Int. J. STD AIDS 2014, 25, 745–750. [CrossRef] [PubMed]
- 48. Santagostino, E.; Gringeri, A.; Rocino, A.; Zanetti, A.; de Biasi, R.; Mannucci, P.M. Patterns of Immunogenicity of an Inactivated Hepatitis A Vaccine in Anti-HIV Positive and Negative Hemophilic Patients. *Thromb. Haemost.* **1994**, *72*, 508–510. [CrossRef]
- 49. Hess, G.; Clemens, R.; Bienzle, U.; Schönfeld, C.; Schunck, B.; Bock, H.L. Immunogenicity and Safety of an Inactivated Hepatitis A Vaccine in Anti-HIV Positive and Negative Homosexual Men. *J. Med. Virol.* **1995**, *46*, 40–42. [CrossRef]
- 50. Weissman, S.; Feucht, C.; Moore, B.A. Response to Hepatitis A Vaccine in HIV-Positive Patients. J. Viral Hepat. 2006, 13, 81–86. [CrossRef]
- 51. Neilsen, G.A.; Bodsworth, N.J.; Watts, N. Response to Hepatitis A Vaccination in Human Immunodeficiency Virus-Infected and -Uninfected Homosexual Men. J. Infect. Dis. **1997**, 176, 1064–1067. [CrossRef]
- Kemper, C.A.; Haubrich, R.; Frank, I.; Dubin, G.; Buscarino, C.; McCutchan, J.A.; Deresinski, S.C.; California Collaborative Treatment Group. Safety and Immunogenicity of Hepatitis A Vaccine in Human Immunodeficiency Virus-Infected Patients: A Double-Blind, Randomized, Placebo-Controlled Trial. J. Infect. Dis. 2003, 187, 1327–1331. [CrossRef]
- Wallace, M.R.; Brandt, C.J.; Earhart, K.C.; Kuter, B.J.; Grosso, A.D.; Lakkis, H.; Tasker, S.A. Safety and Immunogenicity of an Inactivated Hepatitis A Vaccine among HIV-Infected Subjects. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2004, 39, 1207–1213. [CrossRef]

- 54. de Fátima Thomé Barbosa Gouvêa, A.; de Moraes Pinto, M.I.; Miyamoto, M.; Machado, D.M.; Pessoa, S.D.; do Carmo, F.B.; de Vasconcelos Beltrão, S.C.; de Menezes Succi, R.C. Persistence of Hepatitis A Virus Antibodies after Primary Immunization and Response to Revaccination in Children and Adolescents with Perinatal HIV Exposure. *Rev. Paul. Pediatr.* **2015**, *33*, 142–149. [CrossRef]
- Tseng, Y.-T.; Chang, S.-Y.; Liu, W.-C.; Sun, H.-Y.; Wu, C.-H.; Wu, P.-Y.; Lu, C.-L.; Hung, C.-C.; Chang, S.-C. Comparative Effectiveness of Two Doses versus Three Doses of Hepatitis A Vaccine in Human Immunodeficiency Virus-Infected and -Uninfected Men Who Have Sex with Men. *Hepatology* 2013, *57*, 1734–1741. [CrossRef] [PubMed]
- 56. Overton, E.T.; Nurutdinova, D.; Sungkanuparph, S.; Seyfried, W.; Groger, R.K.; Powderly, W.G. Predictors of Immunity after Hepatitis A Vaccination in HIV-Infected Persons. *J. Viral Hepat.* **2007**, *14*, 189–193. [CrossRef] [PubMed]
- 57. Launay, O.; Grabar, S.; Gordien, E.; Desaint, C.; Jegou, D.; Abad, S.; Girard, P.-M.; Bélarbi, L.; Guérin, C.; Dimet, J.; et al. Immunological Efficacy of a Three-Dose Schedule of Hepatitis A Vaccine in HIV-Infected Adults: HEPAVAC Study. *J. Acquir. Immune Defic. Syndr.* 2008, 49, 272–275. [CrossRef] [PubMed]
- Theeten, H.; Van Herck, K.; Van Der Meeren, O.; Crasta, P.; Van Damme, P.; Hens, N. Long-Term Antibody Persistence after Vaccination with a 2-Dose Havrix (Inactivated Hepatitis A Vaccine): 20 Years of Observed Data, and Long-Term Model-Based Predictions. *Vaccine* 2015, *33*, 5723–5727. [CrossRef]
- Kernéis, S.; Desaint, C.; Brichler, S.; Rey, D.; Belarbi, L.; Gordien, E.; Pacanowski, J.; Lortholary, O.; Abgrall, S.; Boëlle, P.-Y.; et al. Long-Term Persistence of Humoral Immunity after Hepatitis A Vaccination in HIV-Infected Adults. *J. Acquir. Immune Defic. Syndr.* 2011, 57, e63–e66. [CrossRef]
- Crum-Cianflone, N.F.; Wilkins, K.; Lee, A.W.; Grosso, A.; Landrum, M.L.; Weintrob, A.; Ganesan, A.; Maguire, J.; Klopfer, S.; Brandt, C.; et al. Long-Term Durability of Immune Responses after Hepatitis A Vaccination among HIV-Infected Adults. *J. Infect. Dis.* 2011, 203, 1815–1823. [CrossRef]
- Cheng, A.; Chang, S.-Y.; Sun, H.-Y.; Tsai, M.-S.; Liu, W.-C.; Su, Y.-C.; Wu, P.-Y.; Hung, C.-C.; Chang, S.-C. Long-Term Durability of Responses to 2 or 3 Doses of Hepatitis A Vaccination in Human Immunodeficiency Virus-Positive Adults on Antiretroviral Therapy. J. Infect. Dis. 2017, 215, 606–613. [CrossRef]
- 62. Liang, T.J. Hepatitis B: The Virus and Disease. Hepatology 2009, 49, S13–S21. [CrossRef]
- 63. Kellerman, S.E.; Hanson, D.L.; McNaghten, A.D.; Fleming, P.L. Prevalence of Chronic Hepatitis B and Incidence of Acute Hepatitis B Infection in Human Immunodeficiency Virus-Infected Subjects. J. Infect. Dis. 2003, 188, 571–577. [CrossRef] [PubMed]
- 64. Bruguera, M.; Cremades, M.; Salinas, R.; Costa, J.; Grau, M.; Sans, J. Impaired Response to Recombinant Hepatitis B Vaccine in HIV-Infected Persons. J. Clin. Gastroenterol. **1992**, 14, 27–30. [CrossRef] [PubMed]
- Soriano, V.; Puoti, M.; Bonacini, M.; Brook, G.; Cargnel, A.; Rockstroh, J.; Thio, C.; Benhamou, Y. Care of Patients with Chronic Hepatitis B and HIV Co-Infection: Recommendations from an HIV-HBV International Panel. *AIDS* 2005, *19*, 221–240. [CrossRef] [PubMed]
- 66. Thio, C.L. Hepatitis B in the Human Immunodeficiency Virus-Infected Patient: Epidemiology, Natural History, and Treatment. *Semin. Liver Dis.* **2003**, *23*, 125–136. [CrossRef] [PubMed]
- 67. Homann, C.; Krogsgaard, K.; Pedersen, C.; Andersson, P.; Nielsen, J.O. High Incidence of Hepatitis B Infection and Evolution of Chronic Hepatitis B Infection in Patients with Advanced HIV Infection. J. Acquir. Immune Defic. Syndr. 1991, 4, 416–420. [PubMed]
- 68. Sterling, R.K.; Wahed, A.S.; King, W.C.; Kleiner, D.E.; Khalili, M.; Sulkowski, M.; Chung, R.T.; Jain, M.K.; Lisker-Melman, M.; Wong, D.K.; et al. Spectrum of Liver Disease in Hepatitis B Virus (HBV) Patients Co-Infected with Human Immunodeficiency Virus (HIV): Results of the HBV-HIV Cohort Study. Am. J. Gastroenterol. 2019, 114, 746–757. [CrossRef]
- 69. Kottilil, S.; Jackson, J.O.; Polis, M.A. Hepatitis B & Hepatitis C in HIV-Infection. Indian J. Med. Res. 2005, 121, 424-450.
- Thio, C.L.; Seaberg, E.C.; Skolasky, R.; Phair, J.; Visscher, B.; Muñoz, A.; Thomas, D.L. Multicenter AIDS Cohort Study HIV-1, Hepatitis B Virus, and Risk of Liver-Related Mortality in the Multicenter Cohort Study (MACS). *Lancet* 2002, 360, 1921–1926. [CrossRef]
- Zarski, J.P.; Thelu, M.A.; Rachail, M.; Seigneurin, J.M. Molecular biology of the hepatitis B virus. II: Importance of the detection of DNA of the hepatitis B virus in serum and liver. *Gastroenterol. Clin. Biol.* 1991, 15, 497–508.
- 72. Krugman, S.; Overby, L.R.; Mushahwar, I.K.; Ling, C.M.; Frösner, G.G.; Deinhardt, F. Viral Hepatitis, Type B. Studies on Natural History and Prevention Re-Examined. *N. Engl. J. Med.* **1979**, *300*, 101–106. [CrossRef]
- 73. Bowden, S. Serological and Molecular Diagnosis. Semin. Liver Dis. 2006, 26, 97–103. [CrossRef]
- Tsang, T.K.; Blei, A.T.; O'Reilly, D.J.; Decker, R. Clinical Significance of Concurrent Hepatitis B Surface Antigen and Antibody Positivity. *Dig. Dis. Sci.* 1986, 31, 620–624. [CrossRef] [PubMed]
- 75. Mast, E.E.; Weinbaum, C.M.; Fiore, A.E.; Alter, M.J.; Bell, B.P.; Finelli, L.; Rodewald, L.E.; Douglas, J.M.; Janssen, R.S.; Ward, J.W.; et al. A Comprehensive Immunization Strategy to Eliminate Transmission of Hepatitis B Virus Infection in the United States: Recommendations of the Advisory Committee on Immunization Practices (ACIP) Part II: Immunization of Adults. *MMWR Recomm. Rep. Morb. Mortal. Wkly. Rep. Recomm. Rep.* 2006, 55, 1–33.
- Martins, S.; do Livramento, A.; Andrigueti, M.; Kretzer, I.F.; Machado, M.J.; Spada, C.; Treitinger, A. Vaccination Coverage and Immunity against Hepatitis B among HIV-Infected Patients in South Brazil. *Braz. J. Infect. Dis. Off. Publ. Braz. Soc. Infect. Dis.* 2015, 19, 181–186. [CrossRef] [PubMed]

- 77. Rey, D.; Krantz, V.; Partisani, M.; Schmitt, M.P.; Meyer, P.; Libbrecht, E.; Wendling, M.J.; Vetter, D.; Nicolle, M.; Kempf-Durepaire, G.; et al. Increasing the Number of Hepatitis B Vaccine Injections Augments Anti-HBs Response Rate in HIV-Infected Patients. Effects on HIV-1 Viral Load. *Vaccine* **2000**, *18*, 1161–1165. [CrossRef] [PubMed]
- 78. Wilson, C.M.; Ellenberg, J.H.; Sawyer, M.K.; Belzer, M.; Crowley-Nowick, P.A.; Puga, A.; Futterman, D.C.; Peralta, L.; Adolescent Medicine HIV/AIDS Research Network. Serologic Response to Hepatitis B Vaccine in HIV Infected and High-Risk HIV Uninfected Adolescents in the REACH Cohort. Reaching for Excellence in Adolescent Care and Health. J. Adolesc. Health Off. Publ. Soc. Adolesc. Med. 2001, 29, 123–129. [CrossRef]
- 79. Laurence, J.C. Hepatitis A and B Immunizations of Individuals Infected with Human Immunodeficiency Virus. *Am. J. Med.* 2005, *118* (Suppl. 10A), 75S–83S. [CrossRef] [PubMed]
- Wong, E.K.; Bodsworth, N.J.; Slade, M.A.; Mulhall, B.P.; Donovan, B. Response to Hepatitis B Vaccination in a Primary Care Setting: Influence of HIV Infection, CD4+ Lymphocyte Count and Vaccination Schedule. *Int. J. STD AIDS* 1996, 7, 490–494. [CrossRef]
- Collier, A.C.; Corey, L.; Murphy, V.L.; Handsfield, H.H. Antibody to Human Immunodeficiency Virus (HIV) and Suboptimal Response to Hepatitis B Vaccination. *Ann. Intern. Med.* 1988, 109, 101–105. [CrossRef]
- Gandhi, R.T.; Wurcel, A.; Lee, H.; McGovern, B.; Shopis, J.; Geary, M.; Sivamurthy, R.; Sax, P.E.; Ukomadu, C. Response to Hepatitis B Vaccine in HIV-1-Positive Subjects Who Test Positive for Isolated Antibody to Hepatitis B Core Antigen: Implications for Hepatitis B Vaccine Strategies. J. Infect. Dis. 2005, 191, 1435–1441. [CrossRef] [PubMed]
- Tedaldi, E.M.; Baker, R.K.; Moorman, A.C.; Wood, K.C.; Fuhrer, J.; McCabe, R.E.; Holmberg, S.D.; HIV Outpatient Study (HOPS) Investigators. Hepatitis A and B Vaccination Practices for Ambulatory Patients Infected with HIV. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2004, 38, 1478–1484. [CrossRef] [PubMed]
- Bauer, S.; Kirschning, C.J.; Häcker, H.; Redecke, V.; Hausmann, S.; Akira, S.; Wagner, H.; Lipford, G.B. Human TLR9 Confers Responsiveness to Bacterial DNA via Species-Specific CpG Motif Recognition. *Proc. Natl. Acad. Sci. USA* 2001, *98*, 9237–9242. [CrossRef] [PubMed]
- Kim, J.H.; Psevdos, G.; Groce, V.; Sharp, V. Persistence of Protective Hepatitis B Surface Antibody Titers after Successful Double-Dose Hepatitis B Virus Rescue Vaccination in HIV-Infected Patients. *Gut Liver* 2012, *6*, 86–91. [CrossRef] [PubMed]
- Shafran, S.D.; Mashinter, L.D.; Lindemulder, A.; Taylor, G.D.; Chiu, I. Poor Efficacy of Intradermal Administration of Recombinant Hepatitis B Virus Immunization in HIV-Infected Individuals Who Fail to Respond to Intramuscular Administration of Hepatitis B Virus Vaccine. *HIV Med.* 2007, *8*, 295–299. [CrossRef]
- de Vries-Sluijs, T.E.M.S.; Hansen, B.E.; van Doornum, G.J.J.; Springeling, T.; Evertsz, N.M.; de Man, R.A.; van der Ende, M.E. A Prospective Open Study of the Efficacy of High-Dose Recombinant Hepatitis B Rechallenge Vaccination in HIV-Infected Patients. J. Infect. Dis. 2008, 197, 292–294. [CrossRef]
- Chaiklang, K.; Wipasa, J.; Chaiwarith, R.; Praparattanapan, J.; Supparatpinyo, K. Comparison of Immunogenicity and Safety of Four Doses and Four Double Doses vs. Standard Doses of Hepatitis B Vaccination in HIV-Infected Adults: A Randomized, Controlled Trial. *PLoS ONE* 2013, *8*, e80409. [CrossRef] [PubMed]
- Seremba, E.; Ocama, P.; Ssekitoleko, R.; Mayanja-Kizza, H.; Adams, S.; Orem, J.; Katabira, E.; Reynolds, S.; Nabatanzi, R.; Casper, C.; et al. Immune Response to the Hepatitis B Vaccine Among HIV-Infected Adults in Uganda. *Vaccine* 2021, 39, 1265–1271. [CrossRef]
- 90. Nie, L.; Hua, W.; Liu, X.; Pang, X.; Guo, C.; Zhang, W.; Tian, Y.; Qiu, Q. Associated Factors and Immune Response to the Hepatitis B Vaccine with a Standard Schedule: A Prospective Study of People with HIV in China. *Vaccines* **2023**, *11*, 921. [CrossRef]
- 91. Xu, L.; Zhang, L.; Kang, S.; Li, X.; Lu, L.; Liu, X.; Song, X.; Li, Y.; Li, X.; Lyu, W.; et al. Immune Responses to HBV Vaccine in People Living with HIV (PLWHs) Who Achieved Successful Treatment: A Prospective Cohort Study. *Vaccines* 2023, *11*, 400. [CrossRef]
- 92. Launay, O.; van der Vliet, D.; Rosenberg, A.R.; Michel, M.-L.; Piroth, L.; Rey, D.; Colin de Verdière, N.; Slama, L.; Martin, K.; Lortholary, O.; et al. Safety and Immunogenicity of 4 Intramuscular Double Doses and 4 Intradermal Low Doses vs Standard Hepatitis B Vaccine Regimen in Adults with HIV-1: A Randomized Controlled Trial. *JAMA* 2011, 305, 1432–1440. [CrossRef]
- Fuster, F.; Vargas, J.I.; Jensen, D.; Sarmiento, V.; Acuña, P.; Peirano, F.; Fuster, F.; Arab, J.P.; Martínez, F.; Fuster, F.; et al. CD4/CD8 Ratio as a Predictor of the Response to HBV Vaccination in HIV-Positive Patients: A Prospective Cohort Study. *Vaccine* 2016, 34, 1889–1895. [CrossRef] [PubMed]
- 94. de Vries-Sluijs, T.E.M.S.; Hansen, B.E.; van Doornum, G.J.J.; Kauffmann, R.H.; Leyten, E.M.S.; Mudrikova, T.; Brinkman, K.; den Hollander, J.G.; Kroon, F.P.; Janssen, H.L.A.; et al. A Randomized Controlled Study of Accelerated Versus Standard Hepatitis B Vaccination in HIV-Positive Patients. J. Infect. Dis. 2011, 203, 984–991. [CrossRef] [PubMed]
- 95. Feng, Y.; Yao, T.; Chang, Y.; Gao, L.; Shao, Z.; Dong, S.; Wu, Y.; Shi, X.; Shi, J.; Feng, D.; et al. Immunogenicity and Persistence of High-Dose Recombinant Hepatitis B Vaccine in Adults Infected with Human Immunodeficiency Virus in China: A Randomized, Double-Blind, Parallel Controlled Trial. *Vaccine* 2021, *39*, 3582–3589. [CrossRef]
- O'Bryan, T.A.; Rini, E.A.; Okulicz, J.F.; Messner, O.; Ganesan, A.; Lalani, T.; Bavaro, M.F.; O'Connell, R.J.; Agan, B.K.; Landrum, M.L. HIV Viraemia during Hepatitis B Vaccination Shortens the Duration of Protective Antibody Levels. *HIV Med.* 2015, 16, 161–167. [CrossRef] [PubMed]
- 97. Fonseca, M.O.; Pang, L.W.; de Paula Cavalheiro, N.; Barone, A.A.; Lopes, M.H. Randomized Trial of Recombinant Hepatitis B Vaccine in HIV-Infected Adult Patients Comparing a Standard Dose to a Double Dose. *Vaccine* 2005, 23, 2902–2908. [CrossRef]

- Zurek Munk-Madsen, M.; Toft, L.; Kube, T.; Richter, R.; Ostergaard, L.; Søgaard, O.S.; Tolstrup, M.; Kaufmann, A.M. Cellular Immunogenicity of Human Papillomavirus Vaccines Cervarix and Gardasil in Adults with HIV Infection. *Hum. Vaccines Immunother.* 2017, 14, 909–916. [CrossRef]
- Doorbar, J.; Egawa, N.; Griffin, H.; Kranjec, C.; Murakami, I. Human Papillomavirus Molecular Biology and Disease Association. *Rev. Med. Virol.* 2015, 25 (Suppl. S1), 2–23. [CrossRef]
- 100. Melbye, M.; Frisch, M. The Role of Human Papillomaviruses in Anogenital Cancers. *Semin. Cancer Biol.* **1998**, *8*, 307–313. [CrossRef]
- 101. Phanuphak, N.; Teeraananchai, S.; Hansudewechakul, R.; Gatechompol, S.; Chokephaibulkit, K.; Dang, H.L.D.; Tran, D.N.H.; Achalapong, J.; Teeratakulpisarn, N.; Chalermchockcharoenkit, A.; et al. Incidence and Persistence of High-Risk Anogenital Human Papillomavirus Infection Among Female Youth with and without Perinatally Acquired Human Immunodefiency Virus Infection: A 3-Year Observational Cohort Study. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2020, 71, e270–e280. [CrossRef]
- 102. Garland, S.M.; Hernandez-Avila, M.; Wheeler, C.M.; Perez, G.; Harper, D.M.; Leodolter, S.; Tang, G.W.K.; Ferris, D.G.; Steben, M.; Bryan, J.; et al. Quadrivalent Vaccine against Human Papillomavirus to Prevent Anogenital Diseases. N. Engl. J. Med. 2007, 356, 1928–1943. [CrossRef]
- 103. Mo, Y.; Ma, J.; Zhang, H.; Shen, J.; Chen, J.; Hong, J.; Xu, Y.; Qian, C. Prophylactic and Therapeutic HPV Vaccines: Current Scenario and Perspectives. *Front. Cell. Infect. Microbiol.* **2022**, *12*, 909223. [CrossRef] [PubMed]
- 104. Ucciferri, C.; Tamburro, M.; Falasca, K.; Sammarco, M.L.; Ripabelli, G.; Vecchiet, J. Prevalence of Anal, Oral, Penile and Urethral Human Papillomavirus in HIV Infected and HIV Uninfected Men Who Have Sex with Men. *J. Med. Virol.* 2018, 90, 358–366. [CrossRef] [PubMed]
- 105. Tartaglia, E.; Falasca, K.; Vecchiet, J.; Sabusco, G.P.; Picciano, G.; Di Marco, R.; Ucciferri, C. Prevalence of HPV Infection among HIV-positive and HIV-negative Women in Central/Eastern Italy: Strategies of Prevention. *Oncol. Lett.* 2017, 14, 7629–7635. [CrossRef]
- 106. Lacey, C.J.N. HPV Vaccination in HIV Infection. Papillomavirus Res. 2019, 8, 100174. [CrossRef] [PubMed]
- 107. Olsson, S.-E.; Villa, L.L.; Costa, R.L.R.; Petta, C.A.; Andrade, R.P.; Malm, C.; Iversen, O.-E.; Høye, J.; Steinwall, M.; Riis-Johannessen, G.; et al. Induction of Immune Memory Following Administration of a Prophylactic Quadrivalent Human Papillomavirus (HPV) Types 6/11/16/18 L1 Virus-like Particle (VLP) Vaccine. *Vaccine* 2007, 25, 4931–4939. [CrossRef]
- 108. Einstein, M.H.; Baron, M.; Levin, M.J.; Chatterjee, A.; Fox, B.; Scholar, S.; Rosen, J.; Chakhtoura, N.; Meric, D.; Dessy, F.J.; et al. Comparative Immunogenicity and Safety of Human Papillomavirus (HPV)-16/18 Vaccine and HPV-6/11/16/18 Vaccine: Followup from Months 12–24 in a Phase III Randomized Study of Healthy Women Aged 18–45 Years. *Hum. Vaccin.* 2011, 7, 1343–1358. [CrossRef]
- 109. Villa, L.L.; Ault, K.A.; Giuliano, A.R.; Costa, R.L.R.; Petta, C.A.; Andrade, R.P.; Brown, D.R.; Ferenczy, A.; Harper, D.M.; Koutsky, L.A.; et al. Immunologic Responses Following Administration of a Vaccine Targeting Human Papillomavirus Types 6, 11, 16, and 18. Vaccine 2006, 24, 5571–5583. [CrossRef] [PubMed]
- 110. Giacomet, V.; Penagini, F.; Trabattoni, D.; Viganò, A.; Rainone, V.; Bernazzani, G.; Bonardi, C.M.; Clerici, M.; Bedogni, G.; Zuccotti, G.V. Safety and Immunogenicity of a Quadrivalent Human Papillomavirus Vaccine in HIV-Infected and HIV-Negative Adolescents and Young Adults. *Vaccine* 2014, 32, 5657–5661. [CrossRef] [PubMed]
- 111. Overton, E.T.; Sungkanuparph, S.; Powderly, W.G.; Seyfried, W.; Groger, R.K.; Aberg, J.A. Undetectable Plasma HIV RNA Load Predicts Success after Hepatitis B Vaccination in HIV-Infected Persons. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2005, 41, 1045–1048. [CrossRef] [PubMed]
- 112. Pinto, L.A.; Wilkin, T.J.; Kemp, T.J.; Abrahamsen, M.; Isaacs-Soriano, K.; Pan, Y.; Webster-Cyriaque, J.; Palefsky, J.M.; Giuliano, A.R. Oral and Systemic HPV Antibody Kinetics Post-Vaccination Among HIV-Positive and HIV-Negative Men. *Vaccine* 2019, 37, 2502–2510. [CrossRef]
- 113. Wilkin, T.; Lee, J.Y.; Lensing, S.Y.; Stier, E.A.; Goldstone, S.E.; Berry, J.M.; Jay, N.; Aboulafia, D.; Cohn, D.L.; Einstein, M.H.; et al. Safety and Immunogenicity of the Quadrivalent Human Papillomavirus Vaccine in HIV-1–Infected Men. J. Infect. Dis. 2010, 202, 1246–1253. [CrossRef] [PubMed]
- 114. Denny, L.; Hendricks, B.; Gordon, C.; Thomas, F.; Hezareh, M.; Dobbelaere, K.; Durand, C.; Hervé, C.; Descamps, D. Safety and Immunogenicity of the HPV-16/18 AS04-Adjuvanted Vaccine in HIV-Positive Women in South Africa: A Partially-Blind Randomised Placebo-Controlled Study. *Vaccine* 2013, *31*, 5745–5753. [CrossRef] [PubMed]
- 115. Chow, E.P.F.; Fairley, C.K.; Zou, H.; Wigan, R.; Garland, S.M.; Cornall, A.M.; Atchison, S.; Tabrizi, S.N.; Chen, M.Y. Human Papillomavirus Antibody Levels Following Vaccination or Natural Infection Among Young Men Who Have Sex with Men. *Clin. Infect. Dis.* 2022, 75, 323–329. [CrossRef] [PubMed]
- Weinberg, A.; Huang, S.; Moscicki, A.-B.; Saah, A.; Levin, M.J. Persistence of Memory B-Cell and T-Cell Responses to the Quadrivalent HPV Vaccine in HIV-Infected Children. *AIDS* 2018, 32, 851–860. [CrossRef]
- 117. Kojic, E.M.; Kang, M.; Cespedes, M.S.; Umbleja, T.; Godfrey, C.; Allen, R.T.; Firnhaber, C.; Grinsztejn, B.; Palefsky, J.M.; Webster-Cyriaque, J.Y.; et al. Immunogenicity and Safety of the Quadrivalent Human Papillomavirus Vaccine in HIV-1-Infected Women. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2014, 59, 127–135. [CrossRef] [PubMed]
- 118. Krammer, F.; Smith, G.J.D.; Fouchier, R.A.M.; Peiris, M.; Kedzierska, K.; Doherty, P.C.; Palese, P.; Shaw, M.L.; Treanor, J.; Webster, R.G.; et al. Influenza. *Nat. Rev. Dis. Primer* **2018**, *4*, 3. [CrossRef] [PubMed]
- 119. Sandrock, C.; Kelly, T. Clinical Review: Update of Avian Influenza A Infections in Humans. Crit. Care 2007, 11, 209. [CrossRef]

- 120. Bouvier, N.M.; Palese, P. The Biology of Influenza Viruses. Vaccine 2008, 26 (Suppl. 4), D49–D53. [CrossRef]
- 121. Hampson, A.W.; Mackenzie, J.S. The Influenza Viruses. Med. J. Aust. 2006, 185, S39–S43. [CrossRef] [PubMed]
- 122. Kosik, I.; Yewdell, J.W. Influenza Hemagglutinin and Neuraminidase: Yin<sup>-</sup>Yang Proteins Coevolving to Thwart Immunity. *Viruses* **2019**, *11*, 346. [CrossRef]
- 123. Biere, B.; Bauer, B.; Schweiger, B. Differentiation of Influenza B Virus Lineages Yamagata and Victoria by Real-Time PCR. J. Clin. Microbiol. 2010, 48, 1425–1427. [CrossRef] [PubMed]
- 124. Wei, H.; Wang, S.; Chen, Q.; Chen, Y.; Chi, X.; Zhang, L.; Huang, S.; Gao, G.F.; Chen, J.-L. Suppression of Interferon Lambda Signaling by SOCS-1 Results in Their Excessive Production during Influenza Virus Infection. *PLoS Pathog.* 2014, 10, e1003845. [CrossRef]
- 125. Mifsud, E.J.; Kuba, M.; Barr, I.G. Innate Immune Responses to Influenza Virus Infections in the Upper Respiratory Tract. *Viruses* **2021**, *13*, 2090. [CrossRef]
- 126. Jain, S.; Kamimoto, L.; Bramley, A.M.; Schmitz, A.M.; Benoit, S.R.; Louie, J.; Sugerman, D.E.; Druckenmiller, J.K.; Ritger, K.A.; Chugh, R.; et al. Hospitalized Patients with 2009 H1N1 Influenza in the United States, April–June 2009. N. Engl. J. Med. 2009, 361, 1935–1944. [CrossRef] [PubMed]
- 127. Yamayoshi, S.; Kawaoka, Y. Current and Future Influenza Vaccines. Nat. Med. 2019, 25, 212–220. [CrossRef] [PubMed]
- 128. Influenza (Seasonal). Available online: https://www.who.int/health-topics/influenza-seasonal?gclid=CjwKCAjwwb6lBhBJ EiwAbuVUSm7fCcN6ZB7s7RGpY4-ANtolRWn13UBuJy8xRZgEyrQyj-xsF6eZqRoCQoEQAvD\_BwE#tab=tab\_1 (accessed on 13 July 2023).
- DiazGranados, C.A.; Denis, M.; Plotkin, S. Seasonal Influenza Vaccine Efficacy and Its Determinants in Children and Non-Elderly Adults: A Systematic Review with Meta-Analyses of Controlled Trials. *Vaccine* 2012, 31, 49–57. [CrossRef]
- Fiore, A.E.; Bridges, C.B.; Cox, N.J. Seasonal Influenza Vaccines. In Vaccines for Pandemic Influenza; Compans, R.W., Orenstein, W.A., Eds.; Current Topics in Microbiology and Immunology; Springer: Berlin/Heidelberg, Germany, 2009; pp. 43–82, ISBN 978-3-540-92165-3.
- 131. Houser, K.; Subbarao, K. Influenza Vaccines: Challenges and Solutions. Cell Host Microbe 2015, 17, 295–300. [CrossRef]
- 132. Sridhar, S.; Begom, S.; Bermingham, A.; Hoschler, K.; Adamson, W.; Carman, W.; Bean, T.; Barclay, W.; Deeks, J.J.; Lalvani, A. Cellular Immune Correlates of Protection against Symptomatic Pandemic Influenza. *Nat. Med.* **2013**, *19*, 1305–1312. [CrossRef]
- 133. Tempia, S.; Walaza, S.; Moyes, J.; Cohen, A.L.; von Mollendorf, C.; McMorrow, M.L.; Mhlanga, S.; Treurnicht, F.K.; Venter, M.; Pretorius, M.; et al. The Effects of the Attributable Fraction and the Duration of Symptoms on Burden Estimates of Influenzaassociated Respiratory Illnesses in a High HIV Prevalence Setting, South Africa, 2013–2015. *Influenza Other Respir. Viruses* 2018, 12, 360–373. [CrossRef]
- 134. Ministero della Salute. Piano Nazionale Prevenzione Vaccinale. Available online: https://www.salute.gov.it/portale/vaccinazio ni/dettaglioContenutiVaccinazioni.jsp?lingua=italiano&id=4828&area=vaccinazioni&menu=vuoto (accessed on 13 July 2023).
- Kroon, F.P.; van Dissel, J.T.; de Jong, J.C.; Zwinderman, K.; van Furth, R. Antibody Response after Influenza Vaccination in HIV-Infected Individuals: A Consecutive 3-Year Study. *Vaccine* 2000, 18, 3040–3049. [CrossRef]
- 136. Pallikkuth, S.; Parmigiani, A.; Silva, S.Y.; George, V.K.; Fischl, M.; Pahwa, R.; Pahwa, S. Impaired Peripheral Blood T-Follicular Helper Cell Function in HIV-Infected Nonresponders to the 2009 H1N1/09 Vaccine. *Blood* 2012, 120, 985–993. [CrossRef]
- 137. Pallikkuth, S.; Kanthikeel, S.P.; Silva, S.Y.; Fischl, M.; Pahwa, R.; Pahwa, S. Innate Immune Defects Correlate with Failure of Antibody Responses to H1N1/09 Vaccine in HIV-Infected Patients. J. Allergy Clin. Immunol. 2011, 128, 1279–1285. [CrossRef]
- Tebas, P.; Frank, I.; Lewis, M.; Quinn, J.; Zifchak, L.; Thomas, A.; Kenney, T.; Kappes, R.; Wagner, W.; Maffei, K.; et al. Poor Immunogenicity of the H1N1 2009 Vaccine in Well Controlled HIV-Infected Individuals. *AIDS* 2010, 24, 2187–2192. [CrossRef] [PubMed]
- Flynn, P.M.; Nachman, S.; Muresan, P.; Fenton, T.; Spector, S.A.; Cunningham, C.K.; Pass, R.; Yogev, R.; Burchett, S.; Heckman, B.; et al. Safety and Immunogenicity of 2009 Pandemic H1N1 Influenza Vaccination in Perinatally HIV-1–Infected Children, Adolescents, and Young Adults. J. Infect. Dis. 2012, 206, 421–430. [CrossRef] [PubMed]
- 140. Fifth Meeting of the International Health Regulations (2005) (IHR) Emergency Committee on the Multi-Country Outbreak of Mpox (Monkeypox). Available online: https://www.who.int/news/item/11-05-2023-fifth-meeting-of-the-international-health-r egulations-(2005)-(ihr)-emergency-committee-on-the-multi-country-outbreak-of-monkeypox-(mpox) (accessed on 16 May 2023).
- 141. Mitjà, O.; Alemany, A.; Marks, M.; Mora, J.I.L.; Rodríguez-Aldama, J.C.; Silva, M.S.T.; Herrera, E.A.C.; Crabtree-Ramirez, B.; Blanco, J.L.; Girometti, N.; et al. Mpox in People with Advanced HIV Infection: A Global Case Series. *Lancet* 2023, 401, 939–949. [CrossRef]
- Joint ECDC-WHO Regional Office for Europe Mpox Surveillance Bulletin. Available online: https://monkeypoxreport.ecdc.europa.eu/ (accessed on 16 May 2023).
- 143. Ortiz-Saavedra, B.; Montes-Madariaga, E.S.; Cabanillas-Ramirez, C.; Alva, N.; Ricardo-Martínez, A.; León-Figueroa, D.A.; Barboza, J.J.; Mohanty, A.; Padhi, B.K.; Sah, R. Epidemiologic Situation of HIV and Monkeypox Coinfection: A Systematic Review. *Vaccines* 2023, 11, 246. [CrossRef]
- Núñez, I.; Valdés-Ferrer, S.I. Fulminant Mpox as an AIDS-Defining Condition: Useful or Stigmatising? Lancet 2023, 401, 881–884. [CrossRef] [PubMed]
- 145. Martín-Delgado, M.C.; Martín Sánchez, F.J.; Martínez-Sellés, M.; Molero García, J.M.; Moreno Guillén, S.; Rodríguez-Artalejo, F.J.; Ruiz-Galiana, J.; Cantón, R.; De Lucas Ramos, P.; García-Botella, A.; et al. Monkeypox in Humans: A New Outbreak. *Rev. Esp. Quimioter. Publ. Of. Soc. Espanola Quimioter.* 2022, 35, 509–518. [CrossRef]

- 146. Public Health—European Commission. Available online: https://ec.europa.eu/health/documents/community-register/html/ h855.htm (accessed on 12 July 2023).
- 147. Rizk, J.G.; Lippi, G.; Henry, B.M.; Forthal, D.N.; Rizk, Y. Prevention and Treatment of Monkeypox. *Drugs* **2022**, *82*, 957–963. [CrossRef]
- 148. Overton, E.T.; Stapleton, J.; Frank, I.; Hassler, S.; Goepfert, P.A.; Barker, D.; Wagner, E.; von Krempelhuber, A.; Virgin, G.; Weigl, J.; et al. Safety and Immunogenicity of Modified Vaccinia Ankara-Bavarian Nordic Smallpox Vaccine in Vaccinia-Naive and Experienced Human Immunodeficiency Virus-Infected Individuals: An Open-Label, Controlled Clinical Phase II Trial. *Open Forum Infect. Dis.* 2015, 2, ofv040. [CrossRef]
- 149. Frey, S.E.; Winokur, P.L.; Salata, R.A.; El-Kamary, S.S.; Turley, C.B.; Walter, E.B.; Hay, C.M.; Newman, F.K.; Hill, H.R.; Zhang, Y.; et al. Safety and Immunogenicity of IMVAMUNE®Smallpox Vaccine Using Different Strategies for a Post Event Scenario. Vaccine 2013, 31, 3025–3033. [CrossRef]
- 150. Greenberg, R.N.; Overton, E.T.; Haas, D.W.; Frank, I.; Goldman, M.; von Krempelhuber, A.; Virgin, G.; Bädeker, N.; Vollmar, J.; Chaplin, P. Safety, Immunogenicity, and Surrogate Markers of Clinical Efficacy for Modified Vaccinia Ankara as a Smallpox Vaccine in HIV-Infected Subjects. *J. Infect. Dis.* 2013, 207, 749–758. [CrossRef]
- 151. Cui, J.; Li, F.; Shi, Z.-L. Origin and Evolution of Pathogenic Coronaviruses. *Nat. Rev. Microbiol.* **2019**, *17*, 181–192. [CrossRef] [PubMed]
- 152. WHO Coronavirus (COVID-19) Dashboard. Available online: https://covid19.who.int (accessed on 10 May 2023).
- Lu, R.; Zhao, X.; Li, J.; Niu, P.; Yang, B.; Wu, H.; Wang, W.; Song, H.; Huang, B.; Zhu, N.; et al. Genomic Characterisation and Epidemiology of 2019 Novel Coronavirus: Implications for Virus Origins and Receptor Binding. *Lancet* 2020, 395, 565–574. [CrossRef] [PubMed]
- 154. Wu, A.; Peng, Y.; Huang, B.; Ding, X.; Wang, X.; Niu, P.; Meng, J.; Zhu, Z.; Zhang, Z.; Wang, J.; et al. Genome Composition and Divergence of the Novel Coronavirus (2019-NCoV) Originating in China. *Cell Host Microbe* **2020**, *27*, 325–328. [CrossRef]
- 155. Zhou, L.; Ayeh, S.K.; Chidambaram, V.; Karakousis, P.C. Modes of Transmission of SARS-CoV-2 and Evidence for Preventive Behavioral Interventions. *BMC Infect. Dis.* **2021**, *21*, 496. [CrossRef] [PubMed]
- 156. V'kovski, P.; Kratzel, A.; Steiner, S.; Stalder, H.; Thiel, V. Coronavirus Biology and Replication: Implications for SARS-CoV-2. *Nat. Rev. Microbiol.* **2021**, *19*, 155–170. [CrossRef] [PubMed]
- 157. COVID-19 Vaccines. Available online: https://www.who.int/emergencies/diseases/novel-coronavirus-2019/covid-19-vaccines (accessed on 10 May 2023).
- Zhang, H.; Zhang, L.; Lin, A.; Xu, C.; Li, Z.; Liu, K.; Liu, B.; Ma, X.; Zhao, F.; Jiang, H.; et al. Algorithm for Optimized MRNA Design Improves Stability and Immunogenicity. *Nature* 2023, 1–3. [CrossRef]
- 159. Polack, F.P.; Thomas, S.J.; Kitchin, N.; Absalon, J.; Gurtman, A.; Lockhart, S.; Perez, J.L.; Pérez Marc, G.; Moreira, E.D.; Zerbini, C.; et al. Safety and Efficacy of the BNT162b2 MRNA Covid-19 Vaccine. *N. Engl. J. Med.* **2020**, *383*, 2603–2615. [CrossRef]
- 160. Sahin, U.; Muik, A.; Derhovanessian, E.; Vogler, I.; Kranz, L.M.; Vormehr, M.; Baum, A.; Pascal, K.; Quandt, J.; Maurus, D.; et al. COVID-19 Vaccine BNT162b1 Elicits Human Antibody and TH1 T Cell Responses. *Nature* 2020, *586*, 594–599. [CrossRef]
- 161. Kalimuddin, S.; Tham, C.Y.L.; Qui, M.; de Alwis, R.; Sim, J.X.Y.; Lim, J.M.E.; Tan, H.-C.; Syenina, A.; Zhang, S.L.; Le Bert, N.; et al. Early T Cell and Binding Antibody Responses Are Associated with COVID-19 RNA Vaccine Efficacy Onset. *Med* 2021, 2, 682–688.e4. [CrossRef]
- 162. Tarke, A.; Sidney, J.; Methot, N.; Yu, E.D.; Zhang, Y.; Dan, J.M.; Goodwin, B.; Rubiro, P.; Sutherland, A.; Wang, E.; et al. Impact of SARS-CoV-2 Variants on the Total CD4+ and CD8+ T Cell Reactivity in Infected or Vaccinated Individuals. *Cell Rep. Med.* 2021, 2, 100355. [CrossRef]
- Woldemeskel, B.A.; Garliss, C.C.; Blankson, J.N. SARS-CoV-2 MRNA Vaccines Induce Broad CD4+ T Cell Responses That Recognize SARS-CoV-2 Variants and HCoV-NL63. J. Clin. Investig. 2021, 131, e149335. [CrossRef]
- 164. Ambrosioni, J.; Blanco, J.L.; Reyes-Urueña, J.M.; Davies, M.-A.; Sued, O.; Marcos, M.A.; Martínez, E.; Bertagnolio, S.; Alcamí, J.; Miro, J.M.; et al. Overview of SARS-CoV-2 Infection in Adults Living with HIV. *Lancet HIV* 2021, 8, e294–e305. [CrossRef] [PubMed]
- Clinical Spectrum. Available online: https://www.covid19treatmentguidelines.nih.gov/overview/clinical-spectrum/ (accessed on 10 May 2023).
- 166. Eisinger, R.W.; Lerner, A.M.; Fauci, A.S. Human Immunodeficiency Virus/AIDS in the Era of Coronavirus Disease 2019: A Juxtaposition of 2 Pandemics. J. Infect. Dis. 2021, 224, 1455–1461. [CrossRef]
- Spinelli, M.A.; Jones, B.L.H.; Gandhi, M. COVID-19 Outcomes and Risk Factors Among People Living with HIV. *Curr. HIV/AIDS Rep.* 2022, 19, 425–432. [CrossRef] [PubMed]
- 168. Levy, I.; Wieder-Finesod, A.; Litchevsky, V.; Biber, A.; Indenbaum, V.; Olmer, L.; Huppert, A.; Mor, O.; Goldstein, M.; Levin, E.G.; et al. Immunogenicity and Safety of the BNT162b2 MRNA COVID-19 Vaccine in People Living with HIV-1. *Clin. Microbiol. Infect. Off. Publ. Eur. Soc. Clin. Microbiol. Infect. Dis.* 2021, 27, 1851–1855. [CrossRef]
- Woldemeskel, B.A.; Karaba, A.H.; Garliss, C.C.; Beck, E.J.; Wang, K.H.; Laeyendecker, O.; Cox, A.L.; Blankson, J.N. The BNT162b2 MRNA Vaccine Elicits Robust Humoral and Cellular Immune Responses in People Living with Human Immunodeficiency Virus (HIV). *Clin. Infect. Dis.* 2021, 74, ciab648. [CrossRef]

- Ruddy, J.A.; Boyarsky, B.J.; Bailey, J.R.; Karaba, A.H.; Garonzik-Wang, J.M.; Segev, D.L.; Durand, C.M.; Werbel, W.A. Safety and Antibody Response to Two-Dose SARS-CoV-2 Messenger RNA Vaccination in Persons with HIV. *AIDS* 2021, 35, 2399–2401. [CrossRef]
- 171. Antinori, A.; Cicalini, S.; Meschi, S.; Bordoni, V.; Lorenzini, P.; Vergori, A.; Lanini, S.; De Pascale, L.; Matusali, G.; Mariotti, D.; et al. Humoral and Cellular Immune Response Elicited by MRNA Vaccination against SARS-CoV-2 in People Living with HIV (PLWH) Receiving Antiretroviral Therapy (ART) According with Current CD4 T-Lymphocyte Count. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2022, 75, ciac238. [CrossRef]
- 172. Vergori, A.; Cozzi Lepri, A.; Cicalini, S.; Matusali, G.; Bordoni, V.; Lanini, S.; Meschi, S.; Iannazzo, R.; Mazzotta, V.; Colavita, F.; et al. Immunogenicity to COVID-19 MRNA Vaccine Third Dose in People Living with HIV. *Nat. Commun.* 2022, 13, 4922. [CrossRef]
- 173. Governo Italiano—Report Vaccini Anti COVID-19. Available online: https://www.governo.it/it/cscovid19/report-vaccini/ (accessed on 8 March 2022).
- 174. Tortellini, E.; Zingaropoli, M.A.; Mancarella, G.; Marocco, R.; Carraro, A.; Jamhour, M.; Barbato, C.; Guardiani, M.; Dominelli, F.; Pasculli, P.; et al. Quality of T-Cell Response to SARS-CoV-2 MRNA Vaccine in ART-Treated PLWH. *Int. J. Mol. Sci.* 2022, 23, 14988. [CrossRef] [PubMed]
- Ucciferri, C.; Vecchiet, J.; Auricchio, A.; Falasca, K. Improving BNT162b2 MRNA Vaccine Tolerability without Efficacy Loss by Pidotimod Supplementation. *Mediterr. J. Hematol. Infect. Dis.* 2022, 14, e2022023. [CrossRef] [PubMed]
- 176. Turner, J.S.; Kim, W.; Kalaidina, E.; Goss, C.W.; Rauseo, A.M.; Schmitz, A.J.; Hansen, L.; Haile, A.; Klebert, M.K.; Pusic, I.; et al. SARS-CoV-2 Infection Induces Long-Lived Bone Marrow Plasma Cells in Humans. *Nature* 2021, 595, 421–425. [CrossRef] [PubMed]
- 177. Riou, C.; du Bruyn, E.; Stek, C.; Daroowala, R.; Goliath, R.T.; Abrahams, F.; Said-Hartley, Q.; Allwood, B.W.; Hsiao, N.-Y.; Wilkinson, K.A.; et al. Relationship of SARS-CoV-2-Specific CD4 Response to COVID-19 Severity and Impact of HIV-1 and Tuberculosis Coinfection. *J. Clin. Investig.* **2021**, *131*, 149125. [CrossRef]
- 178. CDC. Clinical Features of Pneumococcal Disease. Available online: https://www.cdc.gov/pneumococcal/clinicians/clinical-features.html (accessed on 10 July 2023).
- 179. O'Brien, K.L.; Wolfson, L.J.; Watt, J.P.; Henkle, E.; Deloria-Knoll, M.; McCall, N.; Lee, E.; Mulholland, K.; Levine, O.S.; Cherian, T.; et al. Burden of Disease Caused by Streptococcus Pneumoniae in Children Younger than 5 Years: Global Estimates. *Lancet* **2009**, *374*, 893–902. [CrossRef]
- Grau, I.; Ardanuy, C.; Liñares, J.; Podzamczer, D.; Schulze, M.H.; Pallares, R. Trends in Mortality and Antibiotic Resistance among HIV-Infected Patients with Invasive Pneumococcal Disease. *HIV Med.* 2009, 10, 488–495. [CrossRef]
- 181. van Aalst, M.; Lötsch, F.; Spijker, R.; van der Meer, J.T.M.; Langendam, M.W.; Goorhuis, A.; Grobusch, M.P.; de Bree, G.J. Incidence of Invasive Pneumococcal Disease in Immunocompromised Patients: A Systematic Review and Meta-Analysis. *Travel Med. Infect. Dis.* 2018, 24, 89–100. [CrossRef]
- French, N.; Gordon, S.B.; Mwalukomo, T.; White, S.A.; Mwafulirwa, G.; Longwe, H.; Mwaiponya, M.; Zijlstra, E.E.; Molyneux, M.E.; Gilks, C.F. A Trial of a 7-Valent Pneumococcal Conjugate Vaccine in HIV-Infected Adults. *N. Engl. J. Med.* 2010, 362, 812–822. [CrossRef]
- 183. Kobayashi, M.; Farrar, J.L.; Gierke, R.; Britton, A.; Childs, L.; Leidner, A.J.; Campos-Outcalt, D.; Morgan, R.L.; Long, S.S.; Talbot, H.K.; et al. Use of 15-Valent Pneumococcal Conjugate Vaccine and 20-Valent Pneumococcal Conjugate Vaccine among U.S. Adults: Updated Recommendations of the Advisory Committee on Immunization Practices—United States, 2022. MMWR Morb. Mortal. Wkly. Rep. 2022, 71, 109–117. [CrossRef]
- 184. Plosker, G.L. 13-Valent Pneumococcal Conjugate Vaccine: A Review of Its Use in Adults. Drugs 2015, 75, 1535–1546. [CrossRef] [PubMed]
- CDC. Pneumococcal Polysaccharide Vaccine Information Statement. Available online: https://www.cdc.gov/vaccines/hcp/vis /vis-statements/ppv.html (accessed on 22 July 2023).
- Rodriguez-Barradas, M.C.; Alexandraki, I.; Nazir, T.; Foltzer, M.; Musher, D.M.; Brown, S.; Thornby, J. Response of Human Immunodeficiency Virus-Infected Patients Receiving Highly Active Antiretroviral Therapy to Vaccination with 23-Valent Pneumococcal Polysaccharide Vaccine. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2003, *37*, 438–447. [CrossRef] [PubMed]
- 187. Dworkin, M.S.; Ward, J.W.; Hanson, D.L.; Jones, J.L.; Kaplan, J.E.; Adult and Adolescent Spectrum of HIV Disease Project. Pneumococcal Disease among Human Immunodeficiency Virus-Infected Persons: Incidence, Risk Factors, and Impact of Vaccination. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2001, 32, 794–800. [CrossRef] [PubMed]
- 188. Clutterbuck, E.A.; Lazarus, R.; Yu, L.-M.; Bowman, J.; Bateman, E.A.L.; Diggle, L.; Angus, B.; Peto, T.E.; Beverley, P.C.; Mant, D.; et al. Pneumococcal Conjugate and Plain Polysaccharide Vaccines Have Divergent Effects on Antigen-Specific B Cells. J. Infect. Dis. 2012, 205, 1408–1416. [CrossRef]
- Farmaki, P.F.; Chini, M.C.; Mangafas, N.M.; Tzanoudaki, M.T.; Piperi, C.P.; Lazanas, M.Z.; Spoulou, V.S. Immunogenicity and Immunological Memory Induced by the 13-Valent Pneumococcal Conjugate Followed by the 23-Valent Polysaccharide Vaccine in HIV-Infected Adults. J. Infect. Dis. 2018, 218, 26–34. [CrossRef]
- Lopez, A.; Mariette, X.; Bachelez, H.; Belot, A.; Bonnotte, B.; Hachulla, E.; Lahfa, M.; Lortholary, O.; Loulergue, P.; Paul, S.; et al. Vaccination Recommendations for the Adult Immunosuppressed Patient: A Systematic Review and Comprehensive Field Synopsis. J. Autoimmun. 2017, 80, 10–27. [CrossRef]

- 191. Søgaard, O.S.; Lohse, N.; Harboe, Z.B.; Offersen, R.; Bukh, A.R.; Davis, H.L.; Schønheyder, H.C.; Østergaard, L. Improving the Immunogenicity of Pneumococcal Conjugate Vaccine in HIV-Infected Adults with a Toll-like Receptor 9 Agonist Adjuvant: A Randomized, Controlled Trial. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2010, *51*, 42–50. [CrossRef]
- 192. Kroon, F.P.; van Dissel, J.T.; Labadie, J.; van Loon, A.M.; van Furth, R. Antibody Response to Diphtheria, Tetanus, and Poliomyelitis Vaccines in Relation to the Number of CD4+ T Lymphocytes in Adults Infected with Human Immunodeficiency Virus. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 1995, 21, 1197–1203. [CrossRef]
- 193. Gershon, A.A.; Breuer, J.; Cohen, J.I.; Cohrs, R.J.; Gershon, M.D.; Gilden, D.; Grose, C.; Hambleton, S.; Kennedy, P.G.E.; Oxman, M.N.; et al. Varicella Zoster Virus Infection. *Nat. Rev. Dis. Primer* **2015**, *1*, 15016. [CrossRef]
- 194. Clinical Practice: Herpes Zoster—PubMed. Available online: https://pubmed.ncbi.nlm.nih.gov/23863052/ (accessed on 4 June 2023).
- 195. CDC. Clinical Overview of Herpes Zoster (Shingles). Available online: https://www.cdc.gov/shingles/hcp/clinical-overview.h tml (accessed on 11 June 2023).
- 196. Chawki, S.; Vilcu, A.-M.; Etienne, C.; Finet, F.; Blanchon, T.; Souty, C.; Hanslik, T. Incidence of Complications of Herpes Zoster in Individuals on Immunosuppressive Therapy: A Register-Based Population Study. J. Infect. 2022, 84, 531–536. [CrossRef]
- 197. Weitzman, D.; Shavit, O.; Stein, M.; Cohen, R.; Chodick, G.; Shalev, V. A Population Based Study of the Epidemiology of Herpes Zoster and Its Complications. J. Infect. 2013, 67, 463–469. [CrossRef] [PubMed]
- 198. Jansen, K.; Haastert, B.; Michalik, C.; Guignard, A.; Esser, S.; Dupke, S.; Plettenberg, A.; Skaletz-Rorowski, A.; Brockmeyer, N.H. Incidence and Risk Factors of Herpes Zoster among Hiv-Positive Patients in the German Competence Network for HIV/AIDS (KompNet): A Cohort Study Analysis. BMC Infect. Dis. 2013, 13, 372. [CrossRef]
- Gershon, A.A.; Mervish, N.; LaRussa, P.; Steinberg, S.; Lo, S.H.; Hodes, D.; Fikrig, S.; Bonagura, V.; Bakshi, S. Varicella-Zoster Virus Infection in Children with Underlying Human Immunodeficiency Virus Infection. *J. Infect. Dis.* 1997, 176, 1496–1500. [CrossRef] [PubMed]
- 200. Donahue, J.G.; Choo, P.W.; Manson, J.E.; Platt, R. The Incidence of Herpes Zoster. Arch. Intern. Med. 1995, 155, 1605–1609. [CrossRef]
- Buchbinder, S.P.; Katz, M.H.; Hessol, N.A.; Liu, J.Y.; O'Malley, P.M.; Underwood, R.; Holmberg, S.D. Herpes Zoster and Human Immunodeficiency Virus Infection. J. Infect. Dis. 1992, 166, 1153–1156. [CrossRef]
- Successes and Challenges in Varicella Vaccine—PMC. Available online: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC39911 54/ (accessed on 5 June 2023).
- Spoulou, V.; Alain, S.; Gabutti, G.; Giaquinto, C.; Liese, J.; Martinon-Torres, F.; Vesikari, T. Implementing Universal Varicella Vaccination in Europe: The Path Forward. *Pediatr. Infect. Dis. J.* 2019, 38, 181–188. [CrossRef]
- 204. Lee, Y.H.; Choe, Y.J.; Lee, J.; Kim, E.; Lee, J.Y.; Hong, K.; Yoon, Y.; Kim, Y.-K. Global Varicella Vaccination Programs. *Clin. Exp. Pediatr.* 2022, *65*, 555–562. [CrossRef]
- Zou, J.; Krentz, H.B.; Lang, R.; Beckthold, B.; Fonseca, K.; Gill, M.J. Seropositivity, Risks, and Morbidity from Varicella-Zoster Virus Infections in an Adult PWH Cohort From 2000–2020. *Open Forum Infect. Dis.* 2022, 9, ofac395. [CrossRef] [PubMed]
- Muñoz-Quiles, C.; López-Lacort, M.; Díez-Domingo, J.; Orrico-Sánchez, A. Herpes Zoster Risk and Burden of Disease in Immunocompromised Populations: A Population-Based Study Using Health System Integrated Databases, 2009–2014. BMC Infect. Dis. 2020, 20, 905. [CrossRef] [PubMed]
- BMJ Open. Burden of Herpes Zoster in 16 Selected Immunocompromised Populations in England: A Cohort Study in the Clinical Practice Research Datalink 2000–2012. Available online: https://bmjopen.bmj.com/content/8/6/e020528.abstract (accessed on 5 June 2023).
- Brisson, M.; Edmunds, W.J.; Law, B.; Gay, N.J.; Walld, R.; Brownell, M.; Roos, L.L.; Serres, G.D. Epidemiology of Varicella Zoster Virus Infection in Canada and the United Kingdom. *Epidemiol. Infect.* 2001, 127, 305–314. [CrossRef]
- Sullivan, K.M.; Farraye, F.A.; Winthrop, K.L.; Willer, D.O.; Vink, P.; Tavares-Da-Silva, F. Safety and Efficacy of Recombinant and Live Herpes Zoster Vaccines for Prevention in At-Risk Adults with Chronic Diseases and Immunocompromising Conditions. *Vaccine* 2023, 41, 36–48. [CrossRef] [PubMed]
- 210. Anderson, T.C.; Masters, N.B.; Guo, A.; Shepersky, L.; Leidner, A.J.; Lee, G.M.; Kotton, C.N.; Dooling, K.L. Use of Recombinant Zoster Vaccine in Immunocompromised Adults Aged ≥19 Years: Recommendations of the Advisory Committee on Immunization Practices—United States, 2022. MMWR Morb. Mortal. Wkly. Rep. 2022, 71, 80–84. [CrossRef]
- 211. Schmader, K.E.; Levin, M.J.; Gnann, J.W.; McNeil, S.A.; Vesikari, T.; Betts, R.F.; Keay, S.; Stek, J.E.; Bundick, N.D.; Su, S.-C.; et al. Efficacy, Safety, and Tolerability of Herpes Zoster Vaccine in Persons Aged 50-59 Years. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2012, 54, 922–928. [CrossRef]
- 212. Oxman, M.N.; Levin, M.J.; Johnson, G.R.; Schmader, K.E.; Straus, S.E.; Gelb, L.D.; Arbeit, R.D.; Simberkoff, M.S.; Gershon, A.A.; Davis, L.E.; et al. A Vaccine to Prevent Herpes Zoster and Postherpetic Neuralgia in Older Adults. *N. Engl. J. Med.* 2005, 352, 2271–2284. [CrossRef]
- Recommendations of the Advisory Committee on Immunization Practices for Use of Herpes Zoster Vaccines—PubMed. Available online: https://pubmed.ncbi.nlm.nih.gov/29370152/ (accessed on 5 June 2023).
- 214. Effect of Recombinant Zoster Vaccine on Incidence of Herpes Zoster after Autologous Stem Cell Transplantation: A Randomized Clinical Trial—PubMed. Available online: https://pubmed.ncbi.nlm.nih.gov/31287523/ (accessed on 5 June 2023).
- Levin, M.J.; Kroehl, M.E.; Johnson, M.J.; Hammes, A.; Reinhold, D.; Lang, N.; Weinberg, A. Th1 Memory Differentiates Recombinant from Live Herpes Zoster Vaccines. J. Clin. Investig. 2018, 128, 4429–4440. [CrossRef]

- Johnson, M.J.; Liu, C.; Ghosh, D.; Lang, N.; Levin, M.J.; Weinberg, A. Cell-Mediated Immune Responses After Administration of the Live or the Recombinant Zoster Vaccine: 5-Year Persistence. J. Infect. Dis. 2022, 225, 1477–1481. [CrossRef]
- 217. Le, P.; Rothberg, M. Herpes Zoster Infection. BMJ 2019, 364, k5095. [CrossRef]
- 218. Lal, H.; Cunningham, A.L.; Godeaux, O.; Chlibek, R.; Diez-Domingo, J.; Hwang, S.-J.; Levin, M.J.; McElhaney, J.E.; Poder, A.; Puig-Barberà, J.; et al. Efficacy of an Adjuvanted Herpes Zoster Subunit Vaccine in Older Adults. *N. Engl. J. Med.* 2015, 372, 2087–2096. [CrossRef] [PubMed]
- Cunningham, A.L.; Lal, H.; Kovac, M.; Chlibek, R.; Hwang, S.-J.; Díez-Domingo, J.; Godeaux, O.; Levin, M.J.; McElhaney, J.E.; Puig-Barberà, J.; et al. Efficacy of the Herpes Zoster Subunit Vaccine in Adults 70 Years of Age or Older. *N. Engl. J. Med.* 2016, 375, 1019–1032. [CrossRef] [PubMed]
- 220. Benson, C.A.; Andersen, J.W.; Macatangay, B.J.C.; Mailliard, R.B.; Rinaldo, C.R.; Read, S.; Bozzolo, D.R.; Purdue, L.; Jennings, C.; Keefer, M.C.; et al. Safety and Immunogenicity of Zoster Vaccine Live in Human Immunodeficiency Virus-Infected Adults with CD4+ Cell Counts > 200 Cells/ML Virologically Suppressed on Antiretroviral Therapy. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2018, 67, 1712–1719. [CrossRef]
- 221. Mok, C.C.; Chan, K.H.; Ho, L.Y.; Fung, Y.F.; Fung, W.F.; Woo, P.C.Y. Safety and Immune Response of a Live-Attenuated Herpes Zoster Vaccine in Patients with Systemic Lupus Erythematosus: A Randomised Placebo-Controlled Trial. *Ann. Rheum. Dis.* 2019, 78, 1663–1668. [CrossRef] [PubMed]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.