B R I E F   R E P O R T

Severe Acute Respiratory Syndrome Coronavirus 2 Monoclonal Antibody Combination Therapy in Patients With Coronavirus Disease 2019 and Primary Antibody Deficiency

Federica Pulvirenti,[1,a] Cinzia Milito,[2,a] Francesco Cinetto,[3]
Ane Fernandez Salinas,[4] Sara Terreri,[5] Eva Piano Mortari,[5] Stefania Auria,[2]
Valentina Soccodato,[2] Lichtner Miriam,[6,7] Emanuele Nicastri,[8] Laura Vincenzi,[9]
Rita Carsetti,[4] Gianpiero D'Offizi,[9] and Isabella Quinti[2]

[1]Primary Immune Deficiencies Unit, Azienda Ospedaliera Universitaria Policlinico Umberto I, Rome, Italy, [2]Department of Molecular Medicine, Sapienza University of Rome, Rome, Italy, [3]Department of Medicine-DIMED, University of Padova, and Internal Medicine I, Ca' Foncello Hospital, AULSS2 Marca Trevigiana, Treviso, Italy, [4]Department of Molecular Medicine, Sapienza University of Rome and Diagnostic Immunology Research Unit, Multimodal Medicine Research Area, Bambino Gesù Children's Hospital, IRCCS, Rome, Italy, [5]Diagnostic Immunology Research Unit, Multimodal Medicine Research Area, Bambino Gesù Children's Hospital, IRCCS, Rome, Italy, [6]Department of Public Health and Infectious Diseases, Sapienza University of Rome, Italy, [7]Infectious Diseases Unit, SM Goretti Hospital, Sapienza University of Rome, Latina, Italy, [8]National Institute for Infectious Diseases "Lazzaro Spallanzani" IRCCS, Rome, Italy, [9]POIT-INMI Spallanzani Infectious Diseases/Hepatology Unit, Rome, Italy

Background. Previous reports highlighted the efficacy of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-specific monoclonal antibodies (mAbs) against coronavirus disease 2019.

Methods. We conducted a prospective study on the clinical outcome and antiviral effects of mAbs added to standard of care therapy in SARS-CoV-2-infected patients with primary antibody defects.

Results. Median time of SARS-CoV-2 quantitative polymerase chain reaction (qPCR) positivity was shorter in 8 patients treated with mAbs (22 days) than in 10 patients treated with standard of care therapy only (37 days, $P = .026$). Median time of SARS-CoV-2 qPCR positivity from mAb administration was 10 days.

Conclusions. The SARS-CoV-2 mAbs treatment was effective and well tolerated in patients with primary antibody defects.

Keywords. common variable immunodeficiency; COVID-19; monoclonal antibodies; primary antibody deficiencies; SARS-CoV-2.

Due to the severely impaired immune response to immunization, primary antibody deficiency (PAD) patients represent a potential at-risk group in the coronavirus disease 2019 (COVID-19) pandemic. Early reports on cohorts of PAD patients described a low number of patients infected by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and with a variability of clinical manifestations ranging from asymptomatic to death, with the fatality rate accounting for 10% [1–4]. Patients with common variable immunodeficiencies (CVIDs) are the predominant group, showing younger age at death [3] and different risk factors predisposing to severe course in comparison to the general population [4]. Common variable immunodeficiency is the most frequent symptomatic PAD in adults and children, with a wide spectrum of clinical complications including recurrent infection and autoimmune or inflammatory manifestations. Surveys reported a severe clinical course in some CVID patients who have more severe defects in antibody responses, dysfunctional B cells, and immune dysregulation. In these patients, the defective B- and T-cell cellular immune responses might account for an increased risk for prolonged infections, leading to the possible emergence of dangerous vaccine-evasion mutants [5, 6].

At present, COVID-19-directed treatments are limited, including the antiviral agent remdesivir as the first approved therapeutic option for the treatment of COVID-19. More recently, monoclonal antibodies (mAbs) have been developed with the aim to neutralize the SARS-CoV-2 spike protein, thus preventing viral binding to host cells [7]. Given the poor specific antibody responses, PAD patients may be ideal candidates for SARS-CoV-2-based mAbs treatment.

In Italy, this new therapeutic approach has been available since March 2021 when the Italian Agency for Drugs (AIFA) approved the first SARS-CoV-2 mAbs for treatment in patients >12 years old at high-risk for severe COVID-19. Treatment was approved for mild to moderate COVID-19 within 10 days of symptom onset, with the exception for those with immunodeficiency, for which mAb administration was allowed over 10 days (https://www.aifa.gov.it/uso-degli-anticorpi-monoclonali). Data regarding mAbs-based therapy in the PAD population are lacking. The purpose of this study was to describe the clinical response and safety profile in SARS-CoV-2-positive PAD treated with mAbs and to compare data to SARS-CoV-2 PAD patients treated with COVID-19 standard of therapies that did not include mAbs.

METHODS

Patients

Adult (≥18 years old) PAD patients with SARS-CoV-2 infection and routinely observed by Care Centers of Rome and

Treviso were enrolled in this prospective study. Diagnosis of PAD was done according to the ESID criteria (www.ESID.com). Symptomatic patients were considered to be positive for SARS-CoV-2 by quantitative polymerase chain reaction (qPCR) on nasopharyngeal swabs obtained within 1 day after onset of symptoms. Asymptomatic patients were identified by screening patients attending the hospital or in case of family contact. The duration of viral positivity was calculated as the interval between the first positive and the first negative nasopharyngeal swab for SARS-CoV-2. Participants were grouped on the basis of the treatment with mAbs in 2 groups: standard of care treatment (Group 1) and standard of care treatment plus mAbs (Group 2). During the study time, patients were allowed to continue their therapies, including immunoglobulin (Ig) replacement for the underlying antibody deficiency, and were monitored for their clinical status. During SARS-CoV-2 infection, patients were tested using quantitative reverse-transcription PCR (qRT-PCR) every 7–10 days until a negative result was confirmed.

The study was approved by the Ethical Committee of the Sapienza University of Rome (Prot. 0521/2020, July 13, 2020). The study was performed in accordance with the Good Clinical Practice guidelines, the International Conference on Harmonization guidelines, and the most recent version of the Declaration of Helsinki.

Enzyme-Linked Immunosorbent Assay for Specific Immunoglobulin Detection

A semiquantitative in vitro determination of human IgG antibodies against the SARS-CoV-2 (S1) was performed on serum samples by using the Anti-SARS-CoV-2 Spike enzyme-linked immunosorbent assay (EUROIMMUN), according to the manufacturer's instructions. Values were then normalized for comparison with a calibrator. Results are reported as the ratio between optical density (OD) sample and OD calibrator. The ratio interpretation was as follows: <0.8 = negative, ≥0.8 to <1.1 = borderline, ≥1.1 = positive.

Statistical Analysis

Patient data were analyzed with standard descriptive statistics. Baseline characteristics of patients treated with COVID-19 standard of care and patients treated with standard of care plus mAbs were compared with χ^2 test for categorical variables and Mann-Whitney for continuous variables. Differences were deemed significant when $P < .05$. Statistical Package for Social Sciences version 15 (SPSS Inc., Chicago, IL) has been used for the analysis.

RESULTS

From March 2020 to May 2021, a total of 18 PAD patients, 7 females and 11 males, aged 51.5 years (range 26–71) tested positive for SARS-CoV-2 by qPCR from nasopharyngeal swabs. Four patients were asymptomatic. Fourteen patients were symptomatic: 6 of 14 patients required hospitalization due to progression of COVID-19 symptoms. The most common COVID-19 symptoms recorded were fever (72%), cough (61%), and dyspnea (39%). Chest computed tomography scan showed pneumonia in 5 patients (28%).

Demographic and clinical data of SARS-CoV-2-infected patients are provided in Table 1. Ten patients received the standard of care for COVID-19 (Group 1): 1 patient (no. 9) received dexamethasone; 1 patient (no. 10) received dexamethasone and remdesivir; 2 patients (nos. 1 and 5) were treated with tocilizumab, dexamethasone, and lopinavir/ritonavir; and 6 patients did not require treatment (nos. 2, 3, 4, 6, 7, 8). Eight PAD patients received the standard of care for COVID-19 plus mAb-based therapy (Group 2): 2 patients (nos. 11 and 12) were treated with bamlanivimab only; 6 patients (nos. 13, 15, 18) received bamlanivimab plus etesevimab and patient no. 16 bamlanivimab plus etesevimab and dexamethasone; patient no. 14 received double therapy with casirivimab/imdevimab plus dexamethasone and remdesivir; and patient no. 17 received double therapy with casirivimab/imdevimab plus dexamethasone. Demographics and clinical pictures of COVID-19 were comparable between the 2 groups (Table 2).

Treatment with mAbs was initiated within a range of 2–15 days after the first positive nasopharyngeal swab (Table 1). Infusion of mAbs was well tolerated in all patients. In 7 of 8 participants symptoms disappeared within 2–5 days, and all but 1 (no. 14) remained asymptomatic thereafter. Nasopharyngeal swab SARS-CoV-2 qPCR was repeated every 7–10 days until negative. Treatment with mAbs significantly diminished the duration of qRT PCR positivity (22 days, range 7–40), that was longer in patients treated with standard of care therapy only (37.5 days, range 21–81, [$P = .026$]) (Table 2). In mAbs recipients a negative result of qRT PCR was recorded a median time of 10 days (range 5–30) after mAbs administration. As expected, SARS-CoV-2 IgG antibodies, which were checked after a comparable median time from the first positive nasopharyngeal swab, were higher in mAbs-treated patients (median: 6.9 OD ratio; range: 6.4–11.7 OD ratio) than in patients receiving standard of care therapy only (median: 1.9 OD ratio; range 0.9–6.4 OD ratio [$P = .001$]).

DISCUSSION

Patients with primary antibody deficiencies are considered as a vulnerable population in the COVID-19 pandemic because they might be unprotected from vaccination, and they might have prolonged COVID-19 course and SARS-Cov-2 recurrences [3, 5–7]. We have recently shown that two thirds of PAD patients are unable to produce specific antibodies after 2 doses of SARS-CoV-2 vaccine, and, instead of generating classic specific memory B cells, they developed atypical memory B cells, short-lived plasma blasts, and variable T-cell response [7]. Atypical memory B cells are mostly generated during extrafollicular reactions without the involvement of antigen selection in the

Table 1. Demographic and Clinical Characteristics of the Cohort of Primary Antibody Deficiency SARS-CoV-2-Infected Patients

ID	IEI	Sex	Age	Underlying Chronic Lung Disease	Time of SARS-CoV-2 Infection	SARS-CoV-2-Associated Symptoms	Pneumonia	Days of SARS-CoV-2 Symptoms	SaO$_2$%	Days of Hospital Admission	mAbs Therapy	Time Point of mAbs Administration (Days From First SARS-CoV-2 qPCR Positivity)	Additional COVID-19-Specific Therapy	Days of SARS-CoV-2 qPCR Positivity	Days of SARS-CoV-2 qPCR Positivity After mAbs Therapy	Outcome
1	SIgAD	F	33	Bronchiectasis	March 2020	Fever, cough dyspnea	Yes	30	85	28	No	-	Lopinavir/Ritonavir, Tocilizumab, Dexamethasone	45	-	Recovery
2	CVID	M	55	COPD bronchiectasis	September 2020	Asymptomatic	No	0	97		No	-	No	51	-	Recovery
3	CVID	M	57	No	September 2020	Fever, cough	No	17	98		No	-	No	79	-	Recovery
4	CVID	F	61	Bronchiectasis	November 2020	Fever	No	2	97		No	-	No	30	-	Recovery
5	CVID	F	47	COPD, bronchiectasis	November 2020	Fever, cough, dyspnea	Yes	95	81	90	No	-	Lopinavir/Ritonavir, Tocilizumab, Dexamethasone	75	-	Dyspnea
6	CVID	M	55	Bronchiectasis	December 2020	Asymptomatic	No	0	98		No	-	No	81	-	Recovery
7	CVID	M	38	Bronchiectasis	January 2021	Fever, cough	No	30	94		No	-	No	23	-	Recovery
8	CVID	M	46	COPD	January 2021	Cough	No	10	95		No	-	No	30	-	Recovery
9	CVID	M	33	COPD, bronchiectasis	February 2021	Fever, cough, dyspnea	No	14	95	12	No	-	Dexamethasone	21	-	Recovery
10	CVID	M	46	No	February 2021	Fever, cough, dyspnea	Yes	13	90	12	No	-	Remdesivir, Dexamethasone	22	-	Recovery
11	CVID	M	52	COPD, bronchiectasis	March 2021	Asymptomatic	No	0	95		Bamlanivimab	12	No	23	11	Recovery
12	CVID	F	51	No	March 2021	Fever, cough	No	13	93		Bamlanivimab	2	Dexamethasone	7	5	Recovery
13	CVID	M	60	Bronchiectasis	March 2021	Fever, cough	No	2	98		Bamlanivimab/Etesevimab	10	No	17	7	Recovery
14	CVID	F	49	GLILD, bronchiectasis, COPD	March 2021	Fever, cough, dyspnea	Yes	40	87	35	*Casirivimab/Imdevimab*	14	Remdesivir, Dexamethasone	40	26	Dyspnea
15	CVID	F	68	COPD, bronchiectasis	March 2021	Asymptomatic	No	0	96		Bamlanivimab/Etesevimab	2	No	32	30	Recovery
16	CVID	F	70	GLILD, bronchiectasis	April 2021	Fever, cough, dyspnea	Yes	28	88	26	Bamlanivimab/Etesevimab	15	Remdesivir, Tocilizumab, Dexamethasone	21	6	Recovery
17	CVID	M	71	No	April, 2021	Fever	No	9	98		*Casirivimab/Imdevimab*	2	Dexamethasone	30	28	Recovery
18	CVID	M	26	No	May 2021	Fever	No	1	99		Bamlanivimab/Etesevimab	2	No	11	9	Recovery

Abbreviations: CVID, common variable immune deficiency; COPD, chronic obstructive pulmonary disease; COVID-19, coronavirus disease 2019; GLILD, granulomatous and lymphocytic lung interstitial disease; IEI, inborn errors of immunity; mAb, monoclonal antibody; qPCR, quantitative polymerase chain reaction; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; SIgAD, selective IgA deficiency.

Table 2. Comparison of Data Between SARS-CoV-2-Infected PAD Patients Grouped by COVID-19 Treatment

Patients' Data	Standard of Care (n = 10)	Standard of Care + mAbs (n = 8)	P Value
Female, n (%)	3 (30)	4 (50)	.630
Age (years), median (IQR)	64.5 (40–55)	56 (50.5–68.5)	.156
Fever, n (%)	7 (70)	6 (75)	1.000
Cough, n (%)	7 (70)	4 (50)	.630
Dyspnea, n (%)	4 (40)	3 (37)	1.000
Pneumonia, n (%)	3 (33)	2 (25)	1.000
Hospital admission, n (%)	4 (40)	2 (25)	.638
Days of SARS-CoV-2 qPCR positivity, median (range)	37.5 (21–81)	22 (7–40)	.026
Days of symptoms, median (range)	13.6 (0–95)	5.5 (0–40)	.393
Spike specific (IgG1) IgG (OD ratio), median (range)	1.9 (0.9–6.4)	6.9 (6.4–11.7)	.001

Abbreviations: Ig, immunoglobulin; IQR, interquartile range; mAb, monoclonal antibody; OD, optical density; PAD, primary antibody deficiency; qPCR, quantitative polymerase chain reaction; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

germinal center reaction [8] and are considered short-lived activated memory B cells. In patients unable to mount an adequate antibody response, additional strategies for protection are needed. So far, prevention of SARS-CoV-2 infection could not be achieved by immunoglobulin replacement treatment, due to the lack of specific antibodies in the current lots of gamma globulins [9]. One controversial option is the possibility of substituting the defective antibody production with convalescent plasma. The administration of convalescent plasma <72 hours after the onset of symptoms has been shown to reduce disease progression in immunocompetent patients with mild disease and at high risk for disease progression only [10]. Moreover, the low neutralizing potency of convalescent plasma therapy is difficult to be standardized. In addition, plasma and transfusion usage in patients lacking immunoglobulins and, in particular, in patients without serum IgA should be limited because it might cause adverse reactions [11].

Monoclonal antibody-based therapies might be a promising option for patients with antibody defects [12]. During the past year, an unprecedentedly large number of mAbs have been developed to fight COVID-19 [13]. Overall, this study confirmed the previous report [1–4] showing a wide range of COVID-19 spectrum of clinical conditions in PAD patients. We showed a positive clinical and antiviral response due to treatment with mAbs added to conventional therapy. Consistent with a previous report [14], the treatment was without severe side effects in all patients. Although the majority of our cohort continued to be COVID-19 symptom-free, 2 patients with severe underlying PAD-related comorbidities required hospitalization: a patient with a pre-existing severe pulmonary involvement and a patient with enteropathy.

CONCLUSIONS

The positive outcome in antibody-deficient patients was restricted to an early time point of monoclonal administration during SARS-CoV-2 infection. Therefore, we suggest regular follow-ups of PAD patients by SARS-CoV-2 qPCR and to consider an early administration of mAbs to avoid COVID-19 evolution and to shorten the time of SARS-CoV-2 positivity. The shift of mAbs administration from intravenous to different routes, such as intramuscular or subcutaneous [15], is under evaluation and will possibly contribute to an easier access to these treatments for PAD.

References

1. Meyts I, Bucciol G, Quinti I, et al; IUIS Committee of Inborn Errors of Immunity. Coronavirus disease 2019 in patients with inborn errors of immunity: an international study. J Allergy Clin Immunol **2021**; 147:520–31.
2. Ho HE, Mathew S, Peluso MJ, Cunningham-Rundles C. Clinical outcomes and features of COVID-19 in patients with primary immunodeficiencies in New York City. J Allergy Clin Immunol Pract **2021**; 9:490–3.e2.
3. Milito C, Lougaris V, Giardino G, et al. Clinical outcome, incidence, and SARS-CoV-2 infection-fatality rates in Italian patients with inborn errors of immunity. J Allergy Clin Immunol Pract **2021**: S2213-2198(21)00457-8.
4. Milito C, Soccodato V, Auria S, Pulvirenti F, Quinti I. COVID-19 in complex common variable immunodeficiency patients affected by lung diseases. Curr Opin Allergy Clin Immunol **2021**. doi: 10.1097/ACI.0000000000000789.
5. Choi B, Choudhary MC, Regan J, et al. Persistence and evolution of SARS-CoV-2 in an immunocompromised host. N Engl J Med **2020**; 383:2291–3.

6. Minotti C, Tirelli F, Barbieri E, Giaquinto C, Donà D. How is immunosuppressive status affecting children and adults in SARS-CoV-2 infection? a systematic review. J Infect 2020; 81:e61–6.

7. Salinas AF, Mortari EP, Terreri S, et al. SARS-CoV-2 vaccine q defects: everybody does their best. J Clin Immunol 2021; 20:1–14.

8. Braddom AE, Batugedara G, Bol S, Bunnik EM. Potential functions of atypical memory B cells in *Plasmodium*-exposed individuals. Int J Parasitol 2020; 50:1033–42.

9. Schwaiger J, Karbiener M, Aberham C, Farcet MR, Kreil TR. No SARS-CoV-2 neutralization by intravenous immunoglobulins produced from plasma collected before the 2020 pandemic. J Infect Dis 2020; 222:1960–4.

10. Libster R, Pérez Marc G, Wappner D, et al; Fundación INFANT–COVID-19 Group. Early high-titer plasma therapy to prevent severe Covid-19 in older adults. N Engl J Med 2021; 384:610–8.

11. Quinti I, Mortari EP, Fernandez Salinas A, Milito C, Carsetti R. IgA antibodies and IgA deficiency in SARS-CoV-2 infection. Front Cell Infect Microbiol 2021; 11:655896.

12. Van Damme KFA, Tavernier S, Van Roy N, et al. Case report: convalescent plasma, a targeted therapy for patients with CVID and severe COVID-19. Front Immunol 2020; 11:596761.

13. Kelley B. Developing therapeutic monoclonal antibodies at pandemic pace. Nat Biotechnol 2020; 38:540–5.

14. Corti D, Purcell LA, Snell G, Veesler D. Tackling COVID-19 with neutralizing monoclonal antibodies. Cell 2021; 184:3086–108.

15. Cohen MS, Wohl DA, Fischer WA, Smith DM, Eron JJ. Outpatient treatment of SARS-CoV-2 infection to prevent COVID-19 progression. Clin Infect Dis 2021; 73:1717–21.